Ameliorating effect of quercetin on epilepsy by inhibition of inflammation in glial cells

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Abstract. Epilepsy is a prevalent neurological disorder and it is a significant health risk, affecting >50 million people worldwide. The development of novel and appropriate strategies is required for ameliorating the progression and/or limiting the detrimental consequences of epilepsy. In the current study, kainic acid (KA), a neurotoxin, was used to induce seizures in mice. The flavonoid quercetin has recently been reported to have neuroprotective effects. Therefore, the effects of quercetin on KA-induced epilepsy and the potential underlying molecular mechanisms were examined. It was noted that quercetin attenuated the KA-induced seizure score and proinflammatory cytokine production, including tumor necrosis factor-α (TNF-α) and interleukin-1β (IL-1β), and activation of nuclear factor κB (NF-κB) in mice. Quercetin attenuated KA-induced proinflammatory cytokine (TNF-α and IL-1β) release from microglia cells, as well as activation of NF-κB and ionized calcium binding adapter molecule 1 in microglia cells. Therefore, quercetin inhibited KA-induced epilepsy by microglia cell inactivation and the production of NF-κB, TNF-α and IL-1β.

Introduction

Epilepsy is part of a group of neurological disorders characterized by a long-term risk of recurrent unprovoked seizures (1).

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According to statistics from a meta-analysis, >65 million people suffer from epilepsy worldwide (2). The majority of patients with epilepsy response well to anti-epileptic drugs, which control or reduce epileptic seizure occurrence. However, a number of patients gradually develop recurrent seizures and a resistance to anti-epileptic drugs (3). Further investigation is required to examine the underlying molecular mechanism of epilepsy and provide novel therapeutic approaches to fulfill clinical needs. Seizures are initiated by neuronal abnormality (4). Kainic acid (KA) is an agonist of kainite glutamate receptors and can induce their overstimulation, eventually leading to neuronal excitotoxicity and neuronal cell death (5). Therefore, KA is used in model systems to establish temporal lobe epilepsy, in order to study the mechanism and efficacy of drugs in epileptic seizures (6).

Inflammation processes have been clinically and experimentally reported to serve a pivotal role in the generation of seizures (7,8). It was observed that inflammatory mediators are upregulated during the development of seizures in the mouse model (9). In patients with intractable epilepsy, the elevation of cytokines, including interleukin (IL)-1β was detected in the brain tissues (10). IL-1β blockers are therefore considered as a potential treatment approach for patients with epilepsy (11). Results from studies using KA model indicated that KA administration activated microglia and the release of several important cytokines, including tumor necrosis factor-α (TNF-α), IL-1β, IL-12 and IL-18 (12-15). IL-1β serves a key role in the recurrence of seizures by mediating enhanced calcium influx to induce proconvulsant effects (16).

Quercetin is derived from *Quercetum* (oaks) and is categorized as a flavonol (17). Previous studies suggested that quercetin shows anti-carcinogenic, anti-inflammatory, antiviral, antioxidant and psychostimulant activities (17,18). For example, quercetin administration can decrease histological signs of acute inflammation in animals in a dose-dependent manner by inhibiting the release of chemokine and the lipid peroxidation end-product malondialdehyde, and increasing antioxidant enzyme activity (19). Quercetin also exhibits a neuro-protective function in several central nervous system disorders, including seizures and Huntington's disease (20,21). Moghbelinejad *et al* (22) has

suggested that quercetin regulated GABAA receptor α5, as well as β1 and β3, in a KA-induced seizure model of mice. However, the potential molecules regulated by quercetin in a KA-induced seizure remain to be investigated.

Materials and methods

Mouse model. A total of 30 male BALB/c mice (weight 20-22 g; 8 weeks old) were purchased and housed in laboratory conditions (relative humidity of 45-55%, 12-h-light/dark cycle, freely available food and water) at room temperature of 20-23˚C. Experiments were carried out in accordance with the International Guidelines for Animal Studies regarding the care and use of animals for experimental purposes (23). The study was approved by the Ethics Committee of School of Life Science at the Jiangsu Normal University. KA and quercetin were bought from Sigma-Aldrich (Merck KGaA).

KA and quercetin were dissolved in saline (0.9% w/v) and Tween-80 (0.8% v/v), respectively. The mice were divided into three groups consisting of 10 mice per group. The control group was intraperitoneally administered with saline (10 μl, 0.9% w/v, i.p.) + Tween-80 (10 μl, 0.8% v/v, i.p.) daily for one week and on the last day they were injected with saline (10 μl, 0.9% w/v, i.p.) + Tween-80 (10 μl, 0.8% v/v, i.p.) followed by saline (10 μl, 0.9% w/v, i.p.) injection 30 min later. The mice in the KA group were injected daily with saline (10 μl, 0.9% w/v, i.p.) + Tween-80 (10 μl, 0.8% v/v, i.p.) for one week and on the last day, the mice were injected with saline, and KA (10 μl, 10 mg/kg, i.p.) was subsequently intraperitoneally administered. In the KA+quercetin group, the mice were intraperitoneally injected with quercetin (10 μl, 100 mg/kg, i.p.) daily for one week and on the last day, KA (10 μl, 10 mg/kg, i.p.) was administered 30 min following injection with quercetin (10 μl, 100 mg/kg, i.p.). Following injection of KA, mice were observed for behavioral changes over a period of 2 h. In accordance with a previous study, the behavioral tests were scored from 0-6 according to the following criteria: 0, No response; 1, immobility; 2, rigid posture; 3, scratching/circling/head bobbing; 4, forelimb clonus/rearing/falling; 5, repetitive pattern of 4; and 6, severe tonic-clonic seizures (24). Following observation, mice were deeply anesthetized with sodium pentobarbital (65 mg/kg, intraperitoneally) and sacrificed using cervical dislocation. The hippocampus of each mouse was collected, cleaned with chilled saline at 4˚C and frozen for subsequent experimentation.

Primary glial cell culture. Experiments were carried out in accordance with the International Guidelines for Animal Studies regarding the care and use of animals for experimental purposes (23). The study was approved by the Ethics Committee of School of Life Science at the Jiangsu Normal University. Glial cells were derived from 20 postnatal day 1-3 BALB/c mice purchased from the Branch of National Breeder Center of Rodents. Briefly, 5 neonatal mice were rinsed in 70% ethanol, followed by a quick decapitation. Afterwards, cerebral cortices were isolated, meninges were removed and tissue was minced and incubated with trypsin (0.025%) for 15 min at 37˚C, followed with a trituration in the presence of DNAse I (50 μg/ml; Sigma-Aldrich; Merck KGaA) and 20% fetal bovine serum (FBS) in Ca^{2+}-/Mg^{2+}-free PBS. Cells were suspended in DMEM (Gibco; Thermo Fisher Scientific, Inc.) supplemented

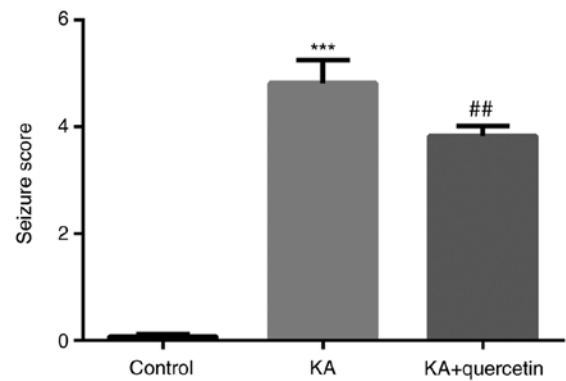
Figure 1. Effects of quercetin (100 mg/kg) on seizure score of mice with KA (10 mg/kg)-induced seizures. ***P<0.001 KA group vs. control group, ##P<0.01 KA+quercetin group vs. KA group. n=10 mice in each group. KA, kainic acid.

with 10% FBS (Gibco; Thermo Fisher Scientific, Inc.), 2 mM L-glutamine, 100 U/ml penicillin and 100 g/ml streptomycin, thereafter, cells ($2x10^6$ cells/well) were seeded onto poly-L-ornithine coated 6-cm diameter Petri dishes and incubated in 95% humidity and 5% CO_2 at 37˚C. After incubation for 14 days, microglia were collected by shaking the mixed glial cell cultures for 1 h. Thereafter, microglial cells ($5x10^4$ cells/well) were seeded into 96-well plates and incubated in 95% humidity and 5% CO_2 at 37˚C for 1 h, followed by the removal of non-adhering cells by washing the plates. The final purity of microglia cells was ~80%.

Isolated microglia cells were then placed in DMEM followed by treatment with saline + Tween-80 (control group) or KA alone (100 μM; KA group) or 100 μM KA preceded by 10 nM quercetin (KA+quercetin group). Treatment with quercetin was performed 30 min prior to KA treatment. Cells were treated with KA for 24 h then subjected to the following experiments.

Western blot analysis. TNF-α (cat. no. 11948; 1:1,000), IL-1β (cat. no. 31202; 1:1,000) and NF-κB p65 (cat. no. 8242; 1:2,000) and phospho-p65 (p-p65; cat. no. 3031; 1:1,000) antibodies were purchased from Cell Signaling Technology, Inc. Ionized calcium-binding adapter molecule 1 (IBA1; cat. no. ab178846; 1:2,000) and GAPDH (cat. no. ab181602; 1:10,000) antibodies were purchased from Abcam. HRP-conjugated secondary antibodies anti-rabbit (cat. no. ab7083; 1:10,000) and anti-goat (cat. no. ab7125; 1:10,000) were purchased from Abcam. Hippocampi were dissected and snap frozen in liquid N_2 for subsequent protein extraction. Proteins from tissues and cells were isolated with RIPA lysis buffer (Sigma-Aldrich; Merck KGaA). The concentration of protein lysates was determined by a bicinchoninic acid kit (Sigma-Aldrich; Merck KGaA). Equal amount of proteins (20 μg) were loaded and separated on an 8% SDS-PAGE gel. The proteins were transferred to a PVDF membrane, blocked by 5% non-fat milk at room temperature for 2 h and washed by TBST (TBS contained 0.1% Tween-20) for 3 times (10 min/time). The membranes were then incubated with primary antibodies at 4˚C overnight, washed by TBST for 3 times (10 min/time) and incubated with secondary antibodies at room temperature for 1 h, washed by TBST for 3 times (10 min/time), successively. The blots were developed with ECL Western Blot kit (Pierce;
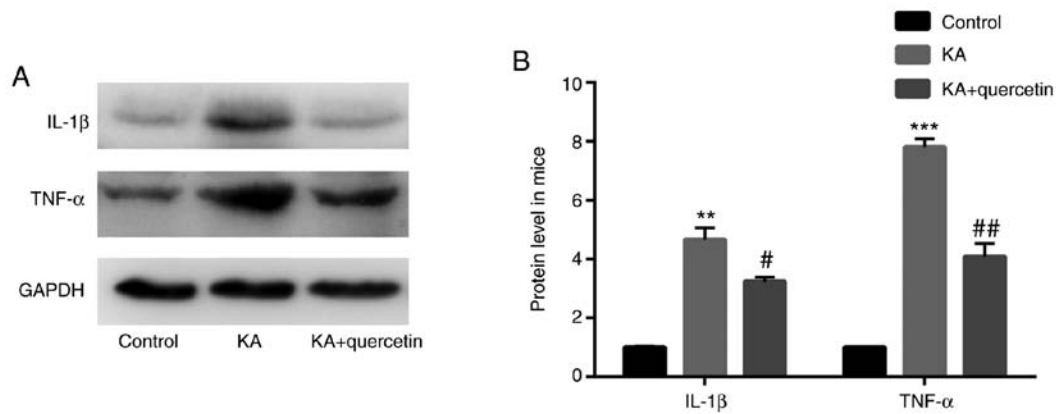
Figure 2. Effects of quercetin (100 mg/kg) on proinflammatory cytokine production in mice with KA (10 mg/kg)-induced seizures. (A) Western blotting and (B) quantification. $^{**}P<0.01$ and $^{***}P<0.001$ KA group vs. control group, $^{#}P<0.05$ and $^{##}P<0.01$ KA+quercetin group vs. KA group. n=10 mice in each group. KA, kainic acid; IL, interleukin; TNF, tumor necrosis factor.
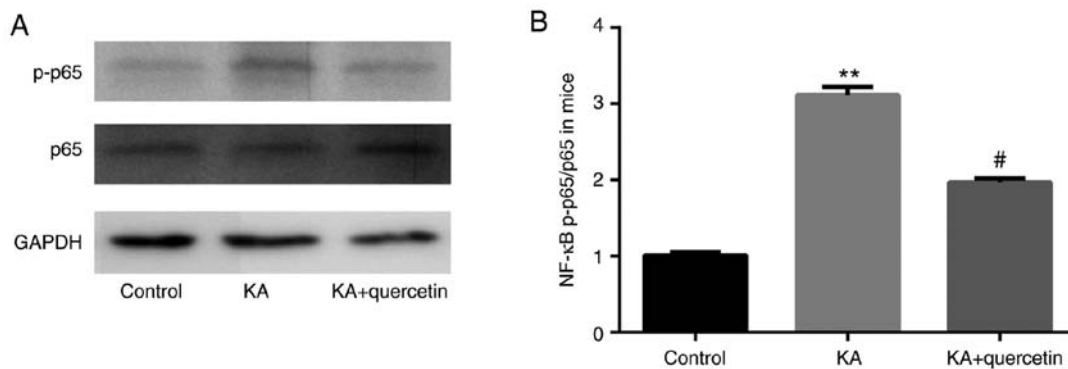

Figure 3. Effects of quercetin (100 mg/kg) on NF-κB production in mice with KA (10 mg/kg)-induced seizures). (A) Western blotting and (B) quantification. $^{**}P<0.01$ KA group vs. control group, $^{#}P<0.05$ KA+quercetin group vs. KA group. n=10 mice in each group. KA, kainic acid; NF, nuclear factor; p, phosphorylated.

Thermo Fisher Scientific, Inc.), according to the manufacturer's protocol. ImageJ software version 1.8.0 (National Institutes of Health) was used for densitometry analysis.

ELISA. The microglia cells were collected by scraping before the removal of the culture medium. Cells were washed with cold phosphate buffer saline (PBS), re-suspended in Tris-Cl (10 mM; pH 7.4) and lysed in triplicate. Protein content in the culture medium was then analyzed by ELISA using the Quantikine TNF-α (cat. no. MTA00B) and IL-1β (cat. no. MLB00C) ELISA kit (R&D Systems, Inc.), according to the manufacturer's protocol.

Statistical analysis. The experiment was repeated three times and all data were calculated by GraphPad Prism 6.0 (GraphPad Software, Inc.) and presented as mean ± standard deviation. Two groups were compared with unpaired Student's t-test. Three groups were firstly compared with one-way ANOVA followed by Newman Keuls analysis. P<0.05 was considered to indicate a statistically significant difference.

Results

Quercetin attenuates KA-induced seizure score in mice. Compared with the control group, KA administration

(10 mg/kg) successfully caused seizures with a significantly increased seizure score (P<0.001, 4.82±0.44 vs. 0.08±0.04), which in turn was significantly reduced by quercetin administration (100 mg/kg) in the KA+quercetin group (P<0.01, 3.83±0.18) (Fig. 1). The results suggested the potential role of quercetin in relieving KA-induced epilepsy in mice.

Quercetin attenuates KA-induced proinflammatory cytokine production in mice. The results of the western blot analysis showed that mice in the KA group (10 mg/kg) exhibited increased TNF-α (P<0.001) and IL-1β (P<0.01) protein expression levels, when compared with those of the control group. In addition, TNF-α (P<0.01) and IL-1β (P<0.05) protein levels were lower in mice treated with quercetin (100 mg/kg) in the KA+quercetin group, when compared with those in the KA group (Fig. 2).

Quercetin attenuates KA-induced activation of NF-κB in mice. The results of the western blot analysis showed that there was increased NF-κB phosphorylated (p)-p65 protein expression level in the KA group (P<0.01, 10 mg/kg), when compared with the control group, which was subsequently significantly decreased by quercetin treatment (P<0.05, 100 mg/kg). Meanwhile, there was no significant difference in NF-κB p65 protein expression level between the 3 groups (Fig. 3).
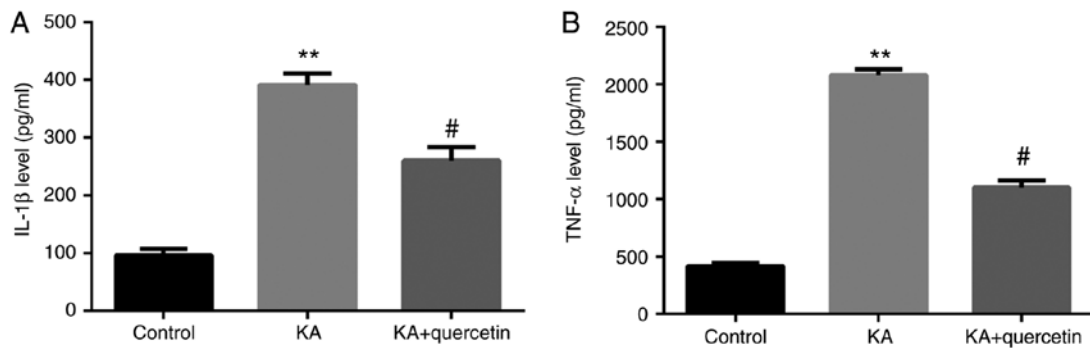
Figure 4. Effects of quercetin (10 nM) on KA (100 μM)-induced proinflammatory cytokine expression in microglia cells (A) IL-1β and (B) TNF-α. **P<0.01 KA group vs. control group, #P<0.05 KA+quercetin group vs. KA group. n=6, results are collected from different time of repeats. KA, kainic acid; IL, interleukin; TNF-α, tumor necrosis factor-α.
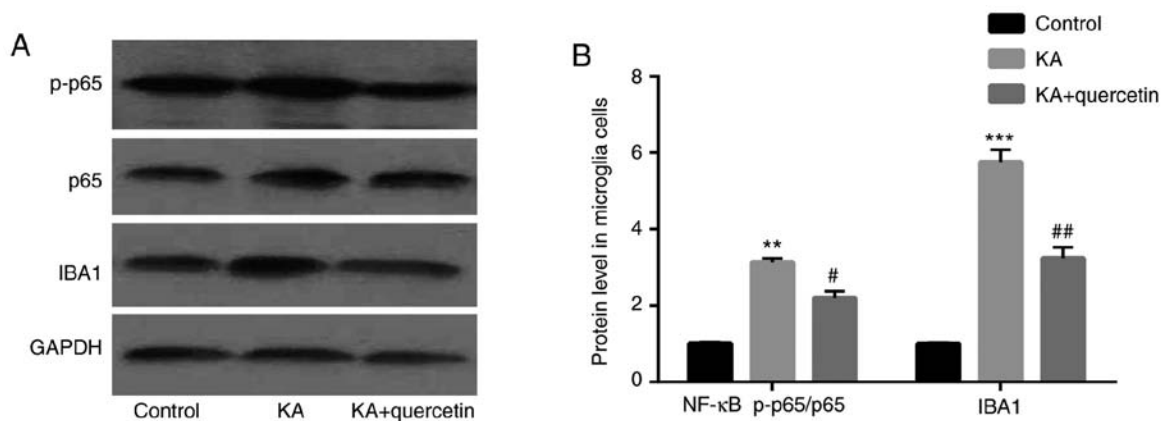

Figure 5. Effects of quercetin (10 nM) on KA (100 μM)-induced NF-κB activation and IBA1 in microglia cells. (A) Western blotting and (B) quantification. **P<0.01 and ***P<0.001 KA group vs. control group, #P<0.05 and ##P<0.01 KA+quercetin group vs. KA group. n=6, results are collected from different time of repeats. KA, kainic acid; NF, nuclear factor; IBA1, ionized calcium-binding adapter molecule 1; p, phosphorylated.

Quercetin attenuates KA-induced proinflammatory cytokine release from microglia cells. Results from the ELISA demonstrated that the culture medium of KA (P<0.01, 100 μM)-treated microglia cells expressed significantly increased protein levels of TNF-α and IL-1β compared with those of the non-treated group. Furthermore, in the culture medium of microglia cells, which were pre-treated with quercetin (P<0.05, 10 nM) prior to KA, TNF-α and IL-1β expression levels were significantly decreased compared with those of the KA group (Fig. 4).

Quercetin attenuates KA-induced activation of NF-κB and microglia cells. The results of the western blot analysis showed that compared with the control group, NF-κB p-p65 (P<0.01) expression levels were increased in the KA (100 μM) group, which were decreased by quercetin treatment (P<0.05, 10 nM). There was no significant difference of NF-κB p65 protein expression level among the 3 groups (Fig. 5). In addition, compared with the control group, IBA1 (P<0.001) expression levels were higher in the KA (100 μM) group, which were decreased by quercetin treatment (P<0.01, 10 nM).

Discussion

The functions of quercetin in animal models of seizures have been studied and its anticonvulsant properties have

been indicated in rats and mice (25-27). In addition, clinical and experimental evidence has been reported, which indicates the association between epilepsy and inflammation (28,29). In addition, quercetin has been reported to serve anti-inflammatory roles by inhibiting the expression of pro-inflammatory cytokines, such as TNF-α and IL-1β, in neurodegenerative diseases (30), including Parkinson's disease (31) and Alzheimer's disease (32). The aim of the present study was to investigate the potential therapeutic effects of quercetin and the underlying molecules that were regulated by quercetin in KA-induced epilepsy. The present study indicated that KA increased behavioral seizure activities and pro-inflammatory cytokines in the hippocampus of mice compared with the control group. However, quercetin decreased both of the aforementioned effects.

Neuroinflammation is characterized by the activation of microglia cells (33). Therefore, once activated, microglial cells take part in the inflammation process and facilitate the release of cytokines and chemokines, such as NF-κB, TNF-α and IL-1β (34). Quercetin inhibits the production of nitric oxide in BV2 microglial cells via NF-κB pathway inactivation (35). In addition, KA activates the NF-κB pathway prior to seizure occurrence (36).

Throughout the present study, in comparison with the control group, it was noted that KA increased TNF-α, IL-1β

and NF-κB expression levels in the hippocampus of mice, and these were subsequently decreased with quercetin treatment.

KA has been reported to activate microglia cells in epilepsy (37). The aforementioned experiments were also repeated in microglia cells *in vitro*, in order to examine whether quercetin attenuates KA-induced epilepsy by inhibiting the activation of microglia cells. It was indicated that compared with the control group, there were higher TNF-α and IL-1β expression levels in the culture medium of microglia cells in the KA group, which were subsequently diminished by quercetin. Meanwhile, compared with the control group, there were higher NF-κB and IBA1 expression levels in microglia cells in the KA group, which were abolished by quercetin, indicating that it attenuates KA-induced epilepsy by inhibiting the activation of microglia cells.

In conclusion, the findings of the present study provide evidence on the role of quercetin in inhibiting KA-induced epilepsy by microglia cell inactivation and the production of NF-κB, TNF-α and IL-1β. These findings highlight the potential role of quercetin in the treatment of epilepsy.

However, there are two limitations of the present study: One is the lack of analysis of different quercetin doses, the other is the lack of *in vivo* and pathology experiments, these will be the subject of future studies.

Acknowledgements

Not applicable.

Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

DW, JL, YZ designed the experiments. DW, ZZhe, SF, XW, XH, SW collected samples and performed experiments. DW, ZZhe, YW, ZZha, QS, ML, BH collected and assembled data. DW, ZZhe, JL, YZ analyzed and interpreted the data. DW, ZZhe, SF, XW, XH, SW, YW, ZZha, QS, ML, BH were involved in drafting the manuscript and revising it critically for important intellectual content. All the authors read and approved the final manuscript.

Ethics approval and consent to participate

Experiments were carried out in accordance with the International Guidelines for Animal Studies regarding the care and use of animals for experimental purposes. The study was approved by the Ethics Committee of School of Life Science at the Jiangsu Normal University.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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