Gastrointestinal cancer - only a deregulation of stem cell differentiation? (Review)

DANIEL NEUREITER¹, CHRISTOPH HEROLD² and MATTHIAS OCKER²

¹Institute of Pathology, Salzburger Landeskliniken, Paracelsus Private Medical University, Müllnerstrasse 48, A-5020 Salzburg, Austria; ²Department of Medicine I, University Hospital Erlangen, Ulmenweg 18, D-91054 Erlangen, Germany

Received October 4, 2005; Accepted November 18, 2005

Abstract. Recent research on embryonic and adult stem cells questions the currently accepted models of multi-step carcinogenesis in solid cancer. Accordingly, differentiated epithelial cells are considered to be the main target for mutational steps, leading to a growth and survival advantage of malignantly transformed cells. In contrast, the stem cell model of carcinogenesis emphasizes the role of stem cells as the initiating structure for tumor development. Yet, it is unclear if tumors contain dysregulated (embryonic) stem cells or if tumors consist of differentiated adult cells that obtained a de-differentiated stem cell-like phenotype. Here, we review the current knowledge on the roles of stem cells in gastrointestinal cancer formation and the implication on future diagnostic and therapeutic strategies.

Contents

- 1. Introduction
- 2. The current concept: multistep carcinogenesis
- 3. Constraints of multistep models
- 4. Stem cells in the gastrointestinal system
- 5 Contribution of stem (cell-like) cells in gastrointestinal carcinogenesis
- 6. Modulation of differentiation status of gastrointestinal tumors
- 7. Summary and future directions

1. Introduction

Gastrointestinal malignancies are among the most frequent cancer diseases in men and women and represent leading

E-mail: matthias.ocker@med1.imed.uni-erlangen.de

causes of cancer-related deaths worldwide (1,2). Especially colorectal (3,4), liver (5) and pancreatic cancers (6) still remain urgent medical problems. Although some progress has been made in treatment of early disease stages, advanced and metastasized carcinomas can only be treated palliatively. Several risk factors like infectious agents (e.g. Hepatitis C virus), chronic inflammation (e.g. colitis ulcerosa or chronic pancreatitis) or genetic predispositions (e.g. familial forms of colorectal cancer such as familial adenomatous polyposis) have been identified in the past. Additionally, different genetic changes like activation of oncogenes as well as genetic or epigenetic inactivation of tumor suppressor genes were commonly observed in several cancer types, leading to the postulation of the so-called multistep carcinogenesis models (7,8). While these models are attractive in supporting morphologic and molecular diagnosis at early stages, they fail to explain several commonly observed features of tumor development and progression, dissemination or relapse.

2. The current concept: multistep carcinogenesis

The present textbook knowledge of tumor development favors the consecutive accumulation of oncogenic events in preneoplastic cells that finally leads to a malignant phenotype. In these models, several activating mutations in cellular protooncogenes as well as the genetic or epigenetic inactivation of tumor suppressor genes lead to a stepwise alteration of the morphologic appearance of normal epithelium to dysplasia, adenoma or intraepithelial premalignant lesions, and finally to invasive carcinoma (Fig. 1). Several distinct oncogenic pathways are commonly altered during the course of this development, e.g. changes in ß-catenin/Wnt-signaling, inactivation of endogenous cell cycle regulators such as Rb or p16^{ink4a}, re-activation of telomerase or mutations in K-ras or p53. These genetic or epigenetic events lead to the acquisition of growth promoting features defined as the 'hallmarks of cancer' by Hanahan and Weinberg in their outstanding review (9). In particular, these include: 1) the independence of the tumor cell to growth-regulating signals (e.g. constitutive activation of K-ras by point mutations in codon 12; mutations in p53 or Rb) (10-18), 2) deficiency in apoptosis (e.g. overexpression of bcl-2) (19-24), 3) tissue remodeling and metastasis (e.g. ß-catenin/Wnt) (25-29), 4) unrestricted proliferation (activation of telomerase) (30-32) and 5) induction of neo-angiogenesis (e.g. by autocrine or

Correspondence to: Dr Matthias Ocker, Department of Medicine I, University Hospital Erlangen, Ulmenweg 18, D-91054 Erlangen, Germany

Key words: cancer stem cells, multistep carcinogenesis, adenoma carcinoma sequence, embryonic differentiation pathways

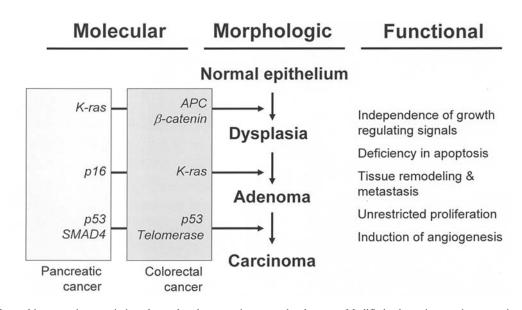


Figure 1. Model for multistep carcinogenesis in colorectal and pancreatic cancer development. Modified schematic overview over the current models of colorectal (7) and pancreatic (8) cancer development. Several oncogenic events are acquired during malignant transformation which provide a survival and growth advantage for the transformed cell, leading to the functional state of a cancer cell (9). The genetic events depicted here are variable in different tumor entities and some of the morphologic or genetic features can also be omitted during tumor development.

paracrine secretion of VEGF) (33-35). It is important to note that during this process of malignant transformation not all of these steps have to be fulfilled and that there is a huge variety of possibilities for the timing and the order of the above described genetic or epigenetic changes. Additionally, these multistep models should not be considered as a linear process but should be viewed as a non-linear dynamic network of intertwining and interdependent steps of malignant transformation (36,37).

3. Constraints of multistep models

The acquisition of several mutations that synergistically provide the epithelial cell with a growth advantage is of low statistical probability as the differentiated epithelium in the gastrointestinal tract is predominantly in the G₀ state of the cell cycle. Most of the genetic material is present as tightly packed and highly organized chromatin in the center of the nucleus, thus being protected from mutational influences, e.g. by irradiation or chemical carcinogens. Additionally, most genetic programs regulating cell growth and migration are inactivated during differentiation and maturation (38-40) and can be rescued by numerous redundant pathways. Gastrointestinal epithelia, especially of large and small intestine, are regenerated by asymmetric division of peripheral stem cells at the crypt basis, giving rise to a small number of highly proliferating progenitor cells that migrate and differentiate to terminal epithelial cells in a course of 2-3 days. Overall, the life-time of differentiated epithelial cells is limited to about 7-10 days in humans, which further decreases the statistical probability for acquiring targeted mutations (41-43).

Recent research efforts therefore focused 1) on the role of long-living and persistent stem and progenitor cells during carcinogenesis, an already accepted concept for hematopoietic neoplasias (44-46), but 2) also on the possibility of trans- or de-differentiation of mature cells during carcinogenesis (44,47-49).

4. Stem cells in the gastrointestinal system

The organs of the gastrointestinal system possess a high capability of continuous tissue regeneration (e.g. mucosa of small and large intestine) or in response to acute or chronic injuries or inflammatory conditions (e.g. chronic helicobacter pylori gastritis; inflammatory bowel diseases, such as colitis ulcerosa; chronic viral hepatitis). These processes are largely maintained by a stable pool of peripheral stem cells that are tightly regulated in their proliferative capacity and give rise to a pool of highly proliferating progenitor cells (42,43,50). These progenitor cells already lost some of the key features of true stem cells, predominantly the capabilities to asymmetric self-renewal and limitless replication with extended life span. Intestinal stem cells are phenotypically characterized by nuclear expression of ß-catenin (51) and of the RNA-binding protein Musashi-1 (52-54). These factors are involved in Wnt/APCand Notch-signaling, respectively, which have also been shown to be important pathways for colorectal carcinogenesis (29,55-58). Some other factors identified in stem cell differentiation and carcinogenesis are Oct-4, Notch, bone morphogenic protein (BMP), janus family kinase (JAK) or sonic hedgehog (Shh) (9,44,58-60) in different anatomic gastrointestinal sites indicating the close relationship between the tightly regulated physiologic process of stem cell maintenance and differentiation and the dysregulated malignant transformation. During embryonic development, but also during the process of cell maturation and migration from the colonic crypt basis, stem cells and their offspring are able to induce angiogenesis and matrix remodeling (60).

A population of progenitor cells has been identified in different forms of liver regeneration and has also been associated with the development of hepatocellular carcinomas. These so-called oval cells form a distinct population in the periductular region of the liver lobules and express markers specific for hepatocytes (e.g. AFP, albumin), as well as biliary tract markers such as cytokeratin 19 or γ -GT, and

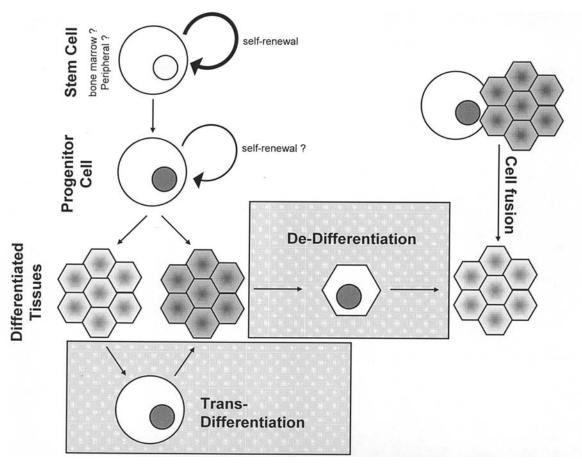


Figure 2. Tissue and stem cell plasticity. True stem cells have the capability of self-renewal and asymmetric division into a resting stem cell and an already committed progenitor cell, which can either give rise to one or to multiple cell types, and has only a limited capacity of self-renewal. Trans-differentiation describes the re-programming of progenitor cells to form different cell types or tissues, while de-differentiation denotes the acquisition or re-activation of immature (stem or progenitor) cell capabilities to give rise to new phenotypes. Adapted from (49,60).

classical bone marrow markers like Thy-1, CD34, c-kit or flk-2 (61). In addition to this phenotypic characterization, experimental evidence from rodents and humans shows that bone marrow cells are capable of repopulating the liver (62-64) and that these progenitor cells are involved in different liver diseases such as hepatocellular carcinoma or cirrhosis (65-69). The notion of bone marrow-derived cells contributing to peripheral tissue maintenance in the gastrointestinal tract has been strengthened by observations of female patients receiving a mixed-sex bone marrow transplantation from male donors. Differentiated Y-chromosome-positive cells have been detected in various organs of the gastrointestinal tract (64,70,71). Yet, the exact mechanism of how bone marrowderived cells support peripheral tissue regeneration is still under debate and several models (e.g. cell fusion, transdifferentiation, de-differentiation, phenocopying) are controversially discussed in the literature (47,60,72) (Fig. 2).

5. Contribution of stem (cell-like) cells in gastrointestinal carcinogenesis

Comparing the properties of cancer cells and (embryonic) stem cells, there seem to be more similarities than differences and of course, more similarities than to resting and differentiated adult cells (Fig. 3). Therefore, two main questions arise: 1) Do stem cells really participate in tumorigenesis and 2) are these cells true stem cells or de-differentiated adult cells with a temporary stem cell-like phenotype?

Morphological comparisons between gastrointestinal embryogenesis and carcinogenesis displayed a similar patterning, i.e. organized arrangement of cells and tissues. In the colon, embryologic development is characterized by the buildup of primitive tubules. Adenomas and adenocarcinomas of the colon show the same pattern with tubular branching imitating colon embryology in an apparently uncontrolled fashion (73,74).

These findings are further supported by the observation that relevant markers of embryogenesis are expressed in early or late stages of gastrointestinal tumorigenesis. While cytokeratin 7 (CK7) is abundantly expressed in the fetal stomach, it is barely detectable in adult gastric tissue. Yet, during gastric carcinogenesis, a neo-expression of CK7 is observed. These cells are considered as de-differentiated cells resembling a stem cell-like phenotype (75). Similar results regarding the nuclear expression of B-catenin were obtained from colorectal adenomas (73). In pancreatic cancer development, re-expression of the pancreatic duodenal homeobox gene 1 (PDX-1), a regulator of exocrine/endocrine development during embryogenesis, is commonly observed (76,77). A further regulator of embryonic gut development is the family of Hedgehog (Hh) proteins that is required for correct specification and patterning (78). Especially, expression

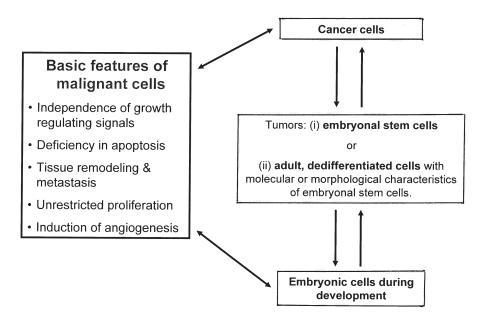


Figure 3. Schematic comparison of tumor and stem cell properties. Both cell types posses growth-promoting features that distinguish them from terminally differentiated cells. It is thus conceivable that tumors either harbor a stem cell population with reactivated embryonic properties or that epithelial cells dedifferentiate to a more stem cell-like phenotype by the acquisition of several genetic or epigenetic events.

of Sonic Hedgehog (Shh) has been attributed to gastrointestinal stem cells and its overexpression has now been described in colorectal, pancreatic and other digestive tract tumors (79-83). The expression of early markers of hepatic development has been observed in dysplastic foci of the liver and in hepatocellular adenomas. This has been linked to the presence of hepatic progenitor cells or the fusion of progenitor cells with mature hepatocytes, but not to the reactivation of embryonic programs in differentiated liver parenchyma (84-86).

Recently, a novel class of non-coding RNAs, the so-called micro-RNAs (miRNA), have been identified as important regulators of gene expression, cellular differentiation and survival (87,88) and might be involved in tumorigenesis by dysregulating oncogenes such as ras (89-95). However, the exact meaning and impact of miRNAs on stem cell biology, differentiation, and cancer development remains to be clarified.

In summary, the processes of determination in embryonic differentiation as well as in gastrointestinal carcinogenesis are relevantly influenced by various signaling pathways (e.g. Wnt/ß-catenin, sonic hedgehog proteins). Additionally, morphology studies showed major morphological similarities and analogies of these two scenes. Overall, tumor cells display morphological phenotypes and molecular markers of early embryonic development with the complete, possibly dangerous, potency of this de-differentiation status. Besides the classical hallmarks of cancer such as proliferation, apoptosis, tissue remodeling and metastasis as well as induction of neoangiogenesis, it seems that de-differentiation and re-activation of embryonic-signaling pathways plays an additional role in gastrointestinal carcinogenesis. With reference to our questions at the beginning of this paragraph, we suggest that de-differentiation processes are essentially involved in this process, as it is not possible to distinguish embryonic from adult stem cells with our currently established techniques. Several studies revealed a relevant and reciprocal inter-acting association between the hallmarks of malignancy, treatment outcome and patient survival with markers of differentiation, e.g. the Wnt/β-catenin pathway (96-98), which could be used for novel targeted approaches.

6. Modulation of differentiation status of gastrointestinal tumors

Presuming that human gastrointestinal tumors resemble de-differentiated states of embryonic or adult stem cells, it will be an interesting therapeutic approach to induce differentiation of these tumor cells into normal resting adult cells or to reduce the malignant potency of de-differentiation.

Several classes of differentiation modulating agents have been examined and tested in pre-clinical or clinical settings. Among these, natural or synthetic derivatives of retinoic acids (e.g. all-trans retinoic acid, ATRA), epigenetic modulators such as the DNA methyltransferase inhibitor zebularine, inhibitors of histone deacetylases like suberoylanilide hydroxamic acid (SAHA) or Trichostatin A (TSA), as well as 'specific' inhibitors of WNT/ß-catenin or hedgehogsignaling-like cyclopamine, are the most prominent. Besides inhibition of proliferation and induction of apoptosis, retinoids interact with nuclear receptors forcing differentiation of cells in several non-gastrointestinal malignancies such as acute promyelocytic leukemia, teratocarcinomas and different solid tumors (e.g. squamous cell carcinoma and breast carcinoma) (99). Several natural and synthetic derivatives are currently tested in clinical trials. Additionally, retinoids have the potential of chemoprevention (100). Epigenetic modulators (e.g. zebularine or SAHA) regulate gene transcription via inhibition of DNA methylation or deacetylation of lysine residues in core histones. Hypermethylation and hypoacetylation are observed in many solid tumors, especially in gastrointestinal tumors. These phenomena have been linked

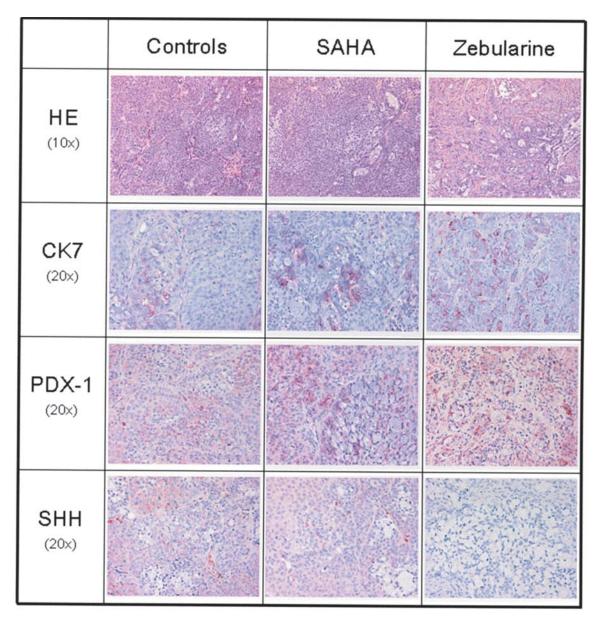


Figure 4. Morphologic and molecular stabilization of pancreatic cancer xenografts after treatment with epigenetic modulators. Tumor-bearing nude mice were treated intraperitoneally with the DNMT inhibitor zebularine or the HDAC inhibitor SAHA and parameters of differentiation [cytokeratin (CK) 7] and embryonic-signaling pathways (pdx-1 and shh) were assessed by immunohistochemistry in paraffin-embedded specimens as described (77; unpublished data) compared to untreated controls. In short, our experiments show a morphological shift from a solid (controls) to a more ductal phenotype (zebularine, H&E-staining), associated with an upregulation of CK7 and PDX-1 and a downregulation of SHH.

to the inactivation of tumor suppressor genes (e.g. transcriptional repression of p16^{ink4a} by promoter hypermethylation), while the inactivation of genes by these processes is also commonly observed during embryogenesis and cellular differentiation. In vitro and in vivo experiments confirmed that these two classes of drugs (inhibitors of DNA methylation and histone deacetylation) have anti-proliferative and proapoptotic capabilities as well as pro-differentiation potency (101-104). Our experience with zebularine and SAHA confirmed earlier findings, that these compounds have antiproliferative and pro-apoptotic effects (105). Additionally, pancreatic carcinoma xenografts in nude mice show a morphological and molecular stability after this treatment, especially regarding the expression of different cytokeratins or PDX-1 (Fig. 4) (77; unpublished data). Different agents (e.g. cyclopamine or sulindac) have been identified which selectively inhibit components of the Wnt/β-catenin or hedgehog signaling pathways; thus, inhibiting proliferation and inducing apoptosis and differentiation (106). These results indicate that interference with these embryonic pathways, which represent early changes during the process of carcinogenesis, might be promising approaches for the development of future therapies.

7. Summary and future directions

The challenge is to gain better understanding and deeper knowledge on mechanisms and impact of differentiation and its dysregulation in the process of malignant transformation of embryonic or adult stem cells. As current therapy options aim at tumor cells at the end of a differentiation process, novel therapies should concentrate on the other side of this scale, i.e. early changes in stem cell-like tumor progenitor cells that could revert the instability of the differentiation status and lead to a phenotypic stabilization.

References

- 1. Greenlee RT, Hill-Harmon MB, Murray T and Thun M: Cancer statistics. CA Cancer J Clin 51: 15-36, 2001.
- 2. Jemal A, Clegg LX, Ward E, Ries LA, Wu X, Jamison PM, et al: Annual report to the nation on the status of cancer, 1975-2001, with a special feature regarding survival. Cancer 101: 3-27, 2004
- 3. Schmiegel W, Pox C, Adler G, Fleig W, Folsch UR, Fruhmorgen P, et al: S3-Guidelines Conference 'Colorectal Carcinoma' 2004. Z Gastroenterol 42: 1129-1177, 2004.
- 4. Weitz J, Koch M, Debus J, Hohler T, Galle PR and Buchler MW: Colorectal cancer. Lancet 365: 153-165, 2005.
- 5. Tang ZY: Hepatocellular carcinoma surgery-review of the past and prospects for the 21st century. J Surg Oncol 91: 95-96, 2005.
- 6. Carpelan-Holmstrom M, Nordling S, Pukkala E, Sankila R, Luttges J, Kloppel G, et al: Does anyone survive pancreatic ductal adenocarcinoma? A nationwide study re-evaluating the data of the Finnish Cancer Registry. Gut 54: 385-387, 2005
- 7. Fearon ER and Vogelstein B: A genetic model for colorectal tumorigenesis. Cell 61: 759-767, 1990.
- 8. Hruban RH, Goggins M, Parsons J and Kern SE: Progression model for pancreatic cancer. Clin Cancer Res 6: 2969-2972, 2000.
- 9. Hanahan D and Weinberg RA: The hallmarks of cancer. Cell 100: 57-70, 2000.
- 10. Hruban RH, van Mansfeld AD, Offerhaus GJ, van Weering DH, Allison DC, Goodman SN, et al: K-ras oncogene activation in adenocarcinoma of the human pancreas. A study of 82 carcinomas using a combination of mutant-enriched polymerase chain reaction analysis and allele-specific oligonucleotide hybridization. Am J Pathol 143: 545-554, 1993.
- 11. Bos JL: Ras oncogenes in human cancer: a review. Cancer Res 49: 4682-4689, 1989.
- 12. Bos JL: The ras gene family and human carcinogenesis. Mutat Res 195: 255-271, 1988.
- 13. Ellis CA and Clark G: The importance of being K-ras. Cell Signal 12: 425-434, 2000.
- 14. Kinzler KW and Vogelstein B: Lessons from hereditary colorectal cancer. Cell 87: 159-170, 1996.
- 15. Levine AJ: p53, the cellular gatekeeper for growth and division. Cell 88: 323-331, 1997.
- 16. Chau BN and Wang JY: Coordinated regulation of life and death by RB. Nat Rev Cancer 3: 130-138, 2003.
- 17. Sherr CJ: Principles of tumor suppression. Cell 116: 235-246, 2004.
- 18. Sherr CJ and McCormick F: The RB and p53 pathways in cancer. Cancer Cell 2: 103-112, 2002. 19. Ghobrial IM, Witzig TE and Adjei AA: Targeting apoptosis
- pathways in cancer therapy. CA Cancer J Clin 55: 178-194, 2005.
- 20. Piro LD: Apoptosis, Bcl-2 antisense, and cancer therapy. Oncology 18 (Suppl 10): 5-10, 2004.
- 21. Gross A, McDonnell JM and Korsmeyer SJ: BCL-2 family members and the mitochondria in apoptosis. Genes Dev 13: 1899-1911, 1999.
- 22. Cory S, Huang DC and Adams JM: The Bcl-2 family: roles in cell survival and oncogenesis. Oncogene 22: 8590-8607, 2003. 23. Kim R, Emi M, Tanabe K and Toge T: Therapeutic potential of
- antisense Bcl-2 as a chemosensitizer for cancer therapy. Cancer 101: 2491-2502, 2004.
- 24. Kountouras J, Zavos C and Chatzopoulos D: Apoptotic and anti-angiogenic strategies in liver and gastrointestinal malignancies. J Surg Oncol 90: 249-259, 2005.
- 25. Christofori G and Semb H: The role of the cell-adhesion molecule E-cadherin as a tumour-suppressor gene. Trends Biochem Sci 24: 73-76, 1999.
- 26. Perl AK, Wilgenbus P, Dahl U, Semb H and Christofori G: A causal role for E-cadherin in the transition from adenoma to carcinoma. Nature 392: 190-193, 1998.
- 27. Reya T and Clevers H: Wnt signalling in stem cells and cancer. Nature 434: 843-850, 2005.
- 28. Moon RT, Kohn AD, De Ferrari GV and Kaykas A: WNT and beta-catenin signalling: diseases and therapies. Nat Rev Genet 5: 691-701, 2004.

- 29. Behrens J and Lustig B: The Wnt connection to tumorigenesis. Int J Dev Biol 48: 477-487, 2004.
- 30. Rudolph KL, Millard M, Bosenberg MW and De Pinho RA: Telomere dysfunction and evolution of intestinal carcinoma in mice and humans. Nat Genet 28: 155-159, 2001. 31. Sharpless NE and De Pinho RA: Telomeres, stem cells,
- senescence, and cancer. J Clin Invest 113: 160-168, 2004.
- 32. Artandi SE and Attardi LD: Pathways connecting telomeres and p53 in senescence, apoptosis, and cancer. Biochem Biophys Res Commun 331: 881-890, 2005.
- 33. Folkman J: Tumor angiogenesis: therapeutic implications. N Engl J Med 285: 1182-1186, 1971.
- 34. Von Marschall Z, Cramer T, Hocker M, Burde R, Plath T, Schirner M, et al: De novo expression of vascular endothelial growth factor in human pancreatic cancer: evidence for an autocrine mitogenic loop. Gastroenterology 119: 1358-1372, 2000.
- 35. Hicklin DJ and Ellis LM: Role of the vascular endothelial growth factor pathway in tumor growth and angiogenesis. J Clin Oncol 23: 1011-1027, 2005.
- Barker N and Clevers H: Tumor environment: a potent driving force in colorectal cancer? Trends Mol Med 7: 535-537, 2001.
- 37. Brabletz T, Jung A, Reu S, Porzner M, Kunz-Schughart LA, Hlubek F, *et al*: Variable beta-catenin expression in colorectal cancers indicates tumor progression driven by the tumor environment. Proc Natl Acad Sci USA 98: 10356-10361, 2001.
- 38. Morgan HD, Santos F, Green K, Dean W and Reik W: Epigenetic reprogramming in mammals. Hum Mol Genet 14 Spec No 1: R47-R58, 2005.
- 39. Cremer T and Cremer C: Chromosome territories, nuclear architecture and gene regulation in mammalian cells. Nat Rev Genet 2: 292-301, 2001.
- 40. Fisher AG and Merkenschlager M: Gene silencing, cell fate and nuclear organisation. Curr Opin Genet Dev 12: 193-197, 2002.
- 41. Heath JP: Epithelial cell migration in the intestine. Cell Biol Int 20: 139-146, 1996.
- 42. Radtke F and Clevers H: Self-renewal and cancer of the gut: two sides of a coin. Science 307: 1904-1909, 2005.
- 43. Okamoto R and Watanabe M: Molecular and clinical basis for the regeneration of human gastrointestinal epithelia. J Gastroenterol 39: 1-6, 2004. 44. Reya T, Morrison SJ, Clarke MF and Weissman IL: Stem cells,
- cancer, and cancer stem cells. Nature 414: 105-111, 2001.
- 45. Beachy PA, Karhadkar SS and Berman DM: Tissue repair and stem cell renewal in carcinogenesis. Nature 432: 324-331, 2004.
- 46. Huntly BJ and Gilliland DG: Leukaemia stem cells and the evolution of cancer-stem-cell research. Nat Rev Cancer 5: 311-321, 2005.
- 47. Tosh D and Slack JM: How cells change their phenotype. Nat Rev Mol Cell Biol 3: 187-194, 2002.
- 48. Weissman IL: Stem cells: units of development, units of regeneration, and units in evolution. Cell 10: 157-168, 2000.
- 49. Wagers AJ and Weissman IL: Plasticity of adult stem cells. Cell 116: 639-648, 2004.
- 50. Brittan M and Wright NA: Stem cell in gastrointestinal structure and neoplastic development. Gut 53: 899-910, 2004.
- 51. Van de Wetering M, Sancho E, Verweij C, De Lau W, Oving I, Hurlstone A, et al: The beta-catenin/TCF-4 complex imposes a crypt progenitor phenotype on colorectal cancer cells. Cell 111: 241-250, 2002.
- 52. Kayahara T, Sawada M, Takaishi S, Fukui H, Fukuzawa H, Seno H, et al: Candidate markers for stem and early progenitor cells, Musashi-1 and Hes1, are expressed in crypt base columnar cells of mouse small intestine. FEBS Lett 535: 131-135, 2003.
- 53. Nishimura S, Wakabayashi N, Toyoda K, Kashima K and Mitsufuji S: Expression of Musashi-1 in human normal colon crypt cells: a possible stem cell marker of human colon epithelium. Dig Dis Sci 48: 1523-1529, 2003.
 54. Potten CS, Booth C, Tudor GL, Booth D, Brady G, Hurley P, *et al*:
- Identification of a putative intestinal stem cell and early lineage marker; musashi-1. Differentiation 71: 28-41, 2003
- 55. Batlle E, Henderson JT, Beghtel H, van den Born MM, Sancho E, Huls G, et al: Beta-catenin and TCF mediate cell positioning in the intestinal epithelium by controlling the expression of EphB/ephrinB. Cell 111: 251-263, 2002.
- 56. Pinto D and Clevers H: Wnt control of stem cells and differentiation in the intestinal epithelium. Exp Cell Res 306: 357-363, 2005.
- 57. Pinto D, Gregorieff A, Begthel H and Clevers H: Canonical Wnt signals are essential for homeostasis of the intestinal epithelium. Genes Dev 17: 1709-1713, 2003.

- Nicolas M, Wolfer A, Raj K, Kummer JA, Mill P, van Noort M, et al: Notch1 functions as a tumor suppressor in mouse skin. Nat Genet 33: 416-421, 2003.
- Taipale J and Beachy PA: The Hedgehog and Wnt signalling pathways in cancer. Nature 411: 349-354, 2001.
- Sell S: Stem cell origin of cancer and differentiation therapy. Crit Rev Oncol Hematol 51: 1-28, 2004.
- 61. Fausto N: Liver regeneration and repair: hepatocytes, progenitor cells, and stem cells. Hepatology 39: 1477-1487, 2004.
- 62. Theise ND, Badve S, Saxena R, Henegariu O, Sell S, Crawford JM, *et al*: Derivation of hepatocytes from bone marrow cells in mice after radiation-induced myeloablation. Hepatology 31: 235-240, 2000.
- 63. Theise ND, Nimmakayalu M, Gardner R, Illei PB, Morgan G, Teperman L, *et al*: Liver from bone marrow in humans. Hepatology 32: 11-16, 2000.
- 64. Korbling M, Katz RL, Khanna A, Ruifrok AC, Rondon G, Albitar M, *et al*: Hepatocytes and epithelial cells of donor origin in recipients of peripheral-blood stem cells. N Engl J Med 346: 738-746, 2002.
- 65. Xiao JC, Ruck P, Adam A, Wang TX and Kaiserling E: Small epithelial cells in human liver cirrhosis exhibit features of hepatic stem-like cells: immunohistochemical, electron microscopic and immunoelectron microscopic findings. Histopathology 42: 141-149, 2003.
- 66. Tan J, Hytiroglou P, Wieczorek R, Park YN, Thung SN, Arias B, et al: Immunohistochemical evidence for hepatic progenitor cells in liver diseases. Liver 22: 365-373, 2002.
- 67. Forbes SJ, Russo FP, Rey V, Burra P, Rugge M, Wright NA, et al: A significant proportion of myofibroblasts are of bone marrow origin in human liver fibrosis. Gastroenterology 126: 955-963, 2004.
- 68. Forbes SJ, Poulsom R and Wright NA: Hepatic and renal differentiation from blood-borne stem cells. Gene Ther 9: 625-630, 2002.
- 69. Theise ND, Yao JL, Harada K, Hytiroglou P, Portmann B, Thung SN, *et al*: Hepatic 'stem cell' malignancies in adults: four cases. Histopathology 43: 263-271, 2003.
- Matsumoto T, Okamoto R, Yajima T, Mori T, Okamoto S, Ikeda Y, et al: Increase of bone marrow-derived secretory lineage epithelial cells during regeneration in the human intestine. Gastroenterology 128: 1851-1867, 2005.
 Okamoto R, Yajima T, Yamazaki M, Kanai T, Mukai M,
- 71. Okamoto R, Yajima T, Yamazaki M, Kanai T, Mukai M, Okamoto S, *et al*: Damaged epithelia regenerated by bone marrow-derived cells in the human gastrointestinal tract. Nat Med 8: 1011-1017, 2002.
- Alison MR, Poulsom R, Otto WR, Vig P, Brittan M, Direkze NC, et al: Recipes for adult stem cell plasticity: fusion cuisine or readymade? J Clin Pathol 57: 113-1120, 2004.
- 73. Kirchner T and Brabletz T: Patterning and nuclear beta-catenin expression in the colonic adenoma-carcinoma sequence. Analogies with embryonic gastrulation. Am J Pathol 157: 1113-1121, 2000.
- Kirchner T and Brabletz T: Tumor patterning: analogies of neoplastic morphogenesis with embryogenesis. Verh Dtsch Ges Pathol 84: 22-27, 2000.
- 75. Kirchner T, Muller S, Hattori T, Mukaisyo K, Papadopoulos T, Brabletz T, *et al*: Metaplasia, intraepithelial neoplasia and early cancer of the stomach are related to dedifferentiated epithelial cells defined by cytokeratin-7 expression in gastritis. Virchows Arch 439: 512-522, 2001.
- 76. Koizumi M, Doi R, Toyoda E, Masui T, Tulachan SS, Kawaguchi Y, *et al*: Increased PDX-1 expression is associated with outcome in patients with pancreatic cancer. Surgery 134: 260-266, 2003.
- 77. Neureiter D, Zopf S, Dimmler A, Stintzing S, Hahn EG, Kirchner T, *et al*: Different capabilities of morphological pattern formation and its association with the expression of differentiation markers in a xenograft model of human pancreatic cancer cell lines. Pancreatology 5: 387-397, 2005.
- Watkins DN and Peacock CD: Hedgehog signalling in foregut malignancy. Biochem Pharmacol 68: 1055-1060, 2004.
- 79. Berman DM, Karhadkar SS, Maitra A, Montes De Oca R, Gerstenblith MR, Briggs K, *et al*: Widespread requirement for Hedgehog ligand stimulation in growth of digestive tract tumours. Nature 425: 846-851, 2003.
- Kayed H, Kleeff J, Keleg S, Guo J, Ketterer K, Berberat PO, *et al*: Indian hedgehog signaling pathway: expression and regulation in pancreatic cancer. Int J Cancer 110: 668-676, 2004.

- 81.Qualtrough D, Buda A, Gaffield W, Williams AC and Paraskeva C: Hedgehog signalling in colorectal tumour cells: induction of apoptosis with cyclopamine treatment. Int J Cancer 110: 831-837, 2004.
- Thayer SP, Di Magliano MP, Heiser PW, Nielsen CM, Roberts D, Lauwers GY, *et al*: Hedgehog is an early and late mediator of pancreatic cancer tumorigenesis. Nature 425: 851-856, 2003.
- 83. Van Eyll JM, Pierreux CE, Lemaigre FP and Rousseau GG: Shh-dependent differentiation of intestinal tissue from embryonic pancreas by activin A. J Cell Sci 117: 2077-2086, 2004.
 84. Libbrecht L, De Vos R, Cassiman D, Desmet V, Aerts R and
- 84. Libbrecht L, De Vos R, Cassiman D, Desmet V, Aerts R and Roskams T: Hepatic progenitor cells in hepatocellular adenomas. Am J Surg Pathol 25: 1388-1396, 2001.
- 85.Libbrecht L, Desmet V, van Damme B and Roskams T: The immunohistochemical phenotype of dysplastic foci in human liver: correlation with putative progenitor cells. J Hepatol 33: 76-84, 2000.
- 86. Roskams TA, Libbrecht L and Desmet VJ: Progenitor cells in diseased human liver. Semin Liver Dis 23: 385-396, 2003.
- Miska EA: How microRNAs control cell division, differentiation and death. Curr Opin Genet Dev 15: 563-568, 2005.
- Mendell JT: MicroRNAs: Critical regulators of development, cellular physiology and malignancy. Cell Cycle 4: 1179-1184, 2005.
- 89.Iorio MV, Ferracin M, Liu CG, Veronese A, Spizzo R, Sabbioni S, *et al*: MicroRNA gene expression deregulation in human breast cancer. Cancer Res 65: 7065-7070, 2005.
- 90. Caldas C and Brenton JD: Sizing up miRNAs as cancer genes. Nat Med 11: 712-714, 2005.
- 91.Croce CM and Calin GA: miRNAs, cancer, and stem cell division. Cell 122: 6-7, 2005.
- 92. Lu J, Getz G, Miska EA, Alvarez-Saavedra E, Lamb J, Peck D, et al: MicroRNA expression profiles classify human cancers. Nature 435: 834-838, 2005.
- 93.Eder M and Scherr M: MicroRNA and lung cancer. N Engl J Med 352: 2446-2448, 2005.
- 94. Gregory RI and Shiekhattar R: MicroRNA biogenesis and cancer. Cancer Res 65: 3509-3512, 2005.
- 95.Johnson SM, Grosshans H, Shingara J, Byrom M, Jarvis R, Cheng A, *et al*: RAS is regulated by the let-7 microRNA family. Cell 120: 635-647, 2005.
- 96. Shibata T, Chuma M, Kokubu A, Sakamoto M and Hirohashi S: EBP50, a beta-catenin-associating protein, enhances Wnt signaling and is over-expressed in hepatocellular carcinoma. Hepatology 38: 178-186, 2003.
 97. Ueda Y, Hijikata M, Takagi S, Takada R, Takada S, Chiba T,
- 97.Ueda Y, Hijikata M, Takagi S, Takada R, Takada S, Chiba T, et al: Wnt/beta-catenin signaling suppresses apoptosis in low serum medium and induces morphologic change in rodent fibroblasts. Int J Cancer 99: 681-688, 2002.
- 98. Chen S, Guttridge DC, You Z, Zhang Z, Fribley A, *et al*: Wnt-1 signaling inhibits apoptosis by activating beta-catenin/T cell factor-mediated transcription. J Cell Biol 152: 87-96, 2001.
- 99. Niles RM: Recent advances in the use of vitamin A (retinoids) in the prevention and treatment of cancer. Nutrition 16: 1084-1089, 2000.
- 100. Okuno M, Kojima S, Matsushima-Nishiwaki R, Tsurumi H, Muto Y, Friedman SL, *et al*: Retinoids in cancer chemoprevention. Curr Cancer Drug Targets 4: 285-298, 2004.
- 101. Fang JY: Histone deacetylase inhibitors, anticancerous mechanism and therapy for gastrointestinal cancers. J Gastroenterol Hepatol 20: 988-994, 2005.
- 102. Munster PN, Troso-Sandoval T, Rosen N, Rifkind R, Marks PA and Richon VM: The histone deacetylase inhibitor suberoylanilide hydroxamic acid induces differentiation of human breast cancer cells. Cancer Res 61: 8492-8497, 2001.
- 103. Cheng JC and Yoo CB, Weisenberger DJ, Chuang J, Wozniak C, Liang G, et al: Preferential response of cancer cells to zebularine. Cancer Cell 6: 151-158, 2004.
- 104. Yoo CB, Cheng JC and Jones PA: Zebularine: a new drug for epigenetic therapy. Biochem Soc Trans 32: 910-912, 2004.
- 105. Ocker M, Alajati A, Ganslmayer M, Zopf S, Luders M, Neureiter D, *et al*: The histone-deacetylase inhibitor SAHA potentiates proapoptotic effects of 5-fluorouracil and irinotecan in hepatoma cells. J Cancer Res Clin Oncol 131: 385-394, 2005.
- 106.Li H, Pamukcu R and Thompson WJ: beta-Catenin signaling: therapeutic strategies in oncology. Cancer Biol Ther 1: 621-625, 2002.