# Melanoma: Molecular pathogenesis and emerging target therapies (Review)

ALESSIA E. RUSSO<sup>1\*</sup>, ELENA TORRISI<sup>1\*</sup>, YLENIA BEVELACQUA<sup>2</sup>, ROSARIO PERROTTA<sup>2</sup>, MASSIMO LIBRA<sup>1</sup>, JAMES A. McCUBREY<sup>3</sup>, DEMETRIOS A. SPANDIDOS<sup>4</sup>, FRANCA STIVALA<sup>1</sup> and GRAZIA MALAPONTE<sup>1</sup>

<sup>1</sup>Department of Biomedical Sciences, <sup>2</sup>Plastic Surgery Section, Department of Medicine and Surgery Specialities, University of Catania, Catania, Italy; <sup>3</sup>Department of Microbiology and Immunology, Brody School of Medicine at East Carolina University, Greenville, NC, USA; <sup>4</sup>Department of Virology, Medical School, University of Crete, Heraklion, Greece

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Abstract. Malignant melanoma is an aggressive tumor of the skin with a poor prognosis for patients with advanced disease. It is resistant to current therapeutic approaches. In melanoma, both the Ras/Raf/MEK/ERK (MAPK) and the PI3K/AKT (AKT) signalling pathways are constitutively activated through multiple mechanisms. Mutations of BRAF have been proposed to contribute to melanoma development. Increased activity of the MAPK pathway prevents apoptosis and induces cell cycle progression. PTEN deletion results in Akt activation. Akt activation can result in the phosphorylation and inactivation of Raf. This decrease in downstream MEK and ERK activation may lead to loss of differentiation or senescence. This review summarizes the most relevant studies focused on the signalling pathways involved in melanomagenesis. New therapeutic strategies are also reported.

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*Correspondence to*: Dr Massimo Libra, Department of Biomedical Sciences, University of Catania, Via Androne 83, I-95124 Catania, Italy

E-mail: mlibra@unict.it

\*Contributed equally

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#### 1. Introduction

Melanoma is the most aggressive form of skin cancer (1). Its incidence has increased dramatically worldwide over the last 50 years (2); currently, the risk of developing melanoma is 1/58 for males in the United States and 1/25 for Australian males (3). It affects predominantly caucasian young and middle-aged adults (4). The main risk factors for cutaneous melanoma are intense and intermittent ultraviolet radiation exposures, phenotypic characteristics (fair skin and blond or red hair), melanocytic nevi's number, type and location (lower limbs in females, posterior trunk in males), personal or family history of melanoma (5).

If melanoma is diagnosed early it can be cured by surgical excision, and about 80% of cases are dealt with in this way (6). However, metastatic malignant melanoma is refractory to current therapies and has a very poor prognosis, with a median survival rate of 6 months (1). Recent discoveries in the complex networks involved in melanoma proliferation, progression and survival have created many opportunities for targeted drugs and new therapeutic approaches for this disease. These new targets include signal transduction pathways, oncogenes, growth factors and their receptors (7). In this review we summarize the most important studies on signalling pathways implicated in the pathogenesis of melanoma. New therapeutic strategies are also reported.

#### 2. Ras/Raf/MEK/ERK pathway

The Ras/Raf/MEK/ERK pathway, also known as the MAPK (mitogen-activated protein kinase) pathway, is a signal transduction cascade relaying extracellular signals from plasma membrane to nucleus via an ordered series of consecutive phosphorylation events (8). In response to a variety of cellular stimuli, including growth factor-mediated activation of receptor tyrosine kinases (RTKs), Ras assumes an activated, GTP-bound state, leading to recruitment of Raf from the cytosol to the cell membrane where it becomes activated, likely via an Src-family tyrosine kinase (9-11). Activated Raf

causes the phosphorylation and activation of MAP kinase extracellular signal regulated kinases 1 and 2 (MEK1/MEK2), which in turn phosphorylate and activate extracellular signal-regulated kinases 1 and 2 (ERK1/ERK2) at specific Thr and Tyr residues (12-14). Activated ERK translocate to the nucleus and phosphorylate several nuclear transcription factors (Elk-1, Myc, CREB, Fos and others) which bind promoters of many genes, including growth factor and cytokine genes that are important for stimulating the cellular proliferation, differentiation, and survival of multiple cell types (15-35).

Dysregulation of Ras/Raf/MEK/ERK pathway plays a key role in pathogenesis of several human cancers (36); mutations at upstream membrane receptors, Ras and B-Raf as well as genes in other pathways (e.g., PI3K, PTEN, Akt), which serve to regulate Raf activity, promote constitutive ERK signalling, stimulating proliferation and survival and providing essential tumor growth and maintenance functions (37). Effects of PTEN deletion on PI3K/Akt and Raf/MEK/ERK activation in melanoma cancer are shown in Fig. 1. Therapies targeting mutant activity of components of the MAP kinase cascade could stop progression of malignant tumors by slowing tumor growth and inducing tumor cell death (36).

Abnormal activation of the MAP kinase cascade in melanoma. The MAPK pathway plays an important role in melanoma cell proliferation and survival, with ERK being constitutively activated in up to 90% of melanomas (38). In this disease, ERK hyperphosphorylation is most commonly due to mutations of NRAS (15-30%) and especially BRAF (50-70%) genes (39,40). The aberration of NRAS often is a substitution of leucine for glutamine at residue 61, this change impairs GTP hydrolysis and maintains the protein in a state of constitutive activation (41). Mutations in other Ras isoforms are rare in melanoma, suggesting an activity context dependent on specific Ras isoforms (42).

The most frequent BRAF mutation, which accounts for more than 90% of melanomas with alteration of B-Raf, is a glutamic acid for valine substitution at codon 600 in exon 15 (Val600Glu; B-RafV600E) (39); this mutation introduces a conformational change in protein structure due to glutamic acid that acts as a phosphomimetic between the Thr598 and Ser<sup>601</sup> phosphorylation sites, leading to constitutive activation of the protein with a substantial increase in the basal kinase activity (43); the resulting hyperactivity of the MAP kinase pathway promotes tumor development (39,44,45). V600EBRAF also promotes vascular development by stimulating autocrine vascular endothelial growth factor (VEGF) secretion (46). Mutations in ARAF and CRAF have not been found in this tumor type. Likely, this pattern of mutations is due to the different mechanism of activation of the three Raf genes: BRAF requires one genetic mutation for oncogenic activation, while ARAF and CRAF require two mutations (47,48), and this is a very rare.

Interestingly, genetic alterations in NRAS and BRAF rarely coexist in melanoma (39,49,50), suggesting that mutant BRAF or NRAS alone is able to activate the MEK/ERK pathway. In Fig. 2 Raf/MEK/ERK and PI3K/Akt pathways and gene alterations that activate these pathways in melanoma are described.

### 3. Additional genetic insults involving other signalling networks are needed for melanoma tumorigenesis and progression

The mechanisms by which NRAS or BRAF mutations promote melanoma cell cycle progression and/or survival remain unclear. Several studies have shown the presence of the same mutations also in a high percentage of benign nevi, suggesting activation of the MAPK pathway is a necessary event for melanoma development, but it is not sufficient for malignant transformation (51,52). Therefore, oncogenic BRAF and NRAS must cooperate with additional genetic insults to induce invasive cancer development in melanocytes. Several candidate cooperative genetic changes have been identified in melanoma including MITF amplification and mutation and/or deletion of PTEN, p53 and p16<sup>INK4a</sup> (53).

Role of Microphthalmia-associated transcription factor (MITF) in melanomagenesis. The connection between MITF and melanoma development is complex. MITF acts as a master regulator of melanocyte development, function and survival (54,55); it plays a double role of inducer/repressor of cellular proliferation (56). High levels of MITF expression lead to G1 cell-cycle arrest and differentiation, through induction of the cell cycle inhibitors p16<sup>INK4a</sup> and p21<sup>Cip</sup> (57,58), whereas very low, or null, expression levels predispose to apoptosis (6). Only inter-mediate levels promote cell proliferation. Therefore, it is thought that melanoma cells have developed strategies to maintain MITF levels in the range compatible with tumorigenesis. It has been shown that constitutive ERK activity, stimulated by V600EBRAF in melanoma cells, is associated with MITF ubiquitin-dependent degradation (59).

Nevertheless, continued expression of MITF is necessary for proliferation and survival of melanoma cells, because it regulates CDK2 and BCL-2 genes, respectively (60,61); furthermore, BRAF mutation is associated with MITF amplification in 10-15% of melanomas (62). However, other mechanisms likely counteract MITF degradation stimulated by ERK-dependent proteasomal degradation, since MITF amplification occurs only in few cases of melanomas in which BRAF and NRAS are mutated. MITF is a downstream target of \( \beta\)-catenin, a key effector of the Wnt signalling pathway, able to stimulate growth of melanoma cells (63); thus, an alternative mechanism of MITF recovery could involve stabilizing mutations in \( \mathbb{B}\)-catenin leading to induction of MITF (64-66). Another mechanism could involve the same mutant BRAF; it has been recently shown that oncogenic BRAF controls MITF on two levels. It downregulates the protein by stimulating its degradation, but then counteracts this by increasing MITF expression through the transcription factor BRN2 (67).

Role of the PI3K pathway in melanoma tumorigenesis. In response to activated growth factor receptors, the phosphoinositide-3-OH kinase (PI3K) phosphorylates phosphatidylinositol-4,5-biphosphate (PIP2) to phosphatidylinositol-3,4,5-triphosphate (PIP3), leading to activation of the major downstream effector of the PI3K pathway, Akt (68); once activated, Akt phosphorylates the downstream cellular proteins

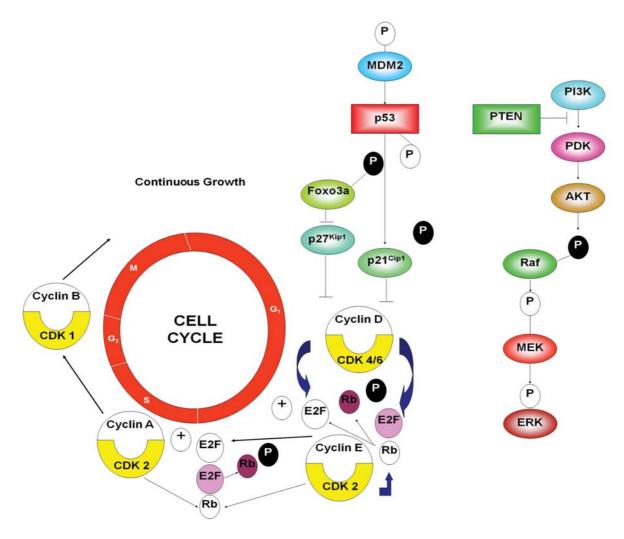


Figure 1. Effects of PTEN deletion on PI3K/Akt and Raf/MEK/ERK activation in melanoma. AKT activation results in inhibition of Raf and downstream MEK/ERK. Raf inactivation leads to decreased  $p21^{Cip1}$  levels and increased cell cycle progression. Akt activation results in  $p21^{Cip1}$  phosphorylation and FOXO3A phosphorylation which prevents transcription of  $p27^{Kip1}$ . Inactivation of Raf/MEK/ERK leads to decreased differentiation and proliferation of immature cells. Decreasing Raf/MEK/ERK cascade by PTEN deletion and Akt hyperactivation results in the continuous proliferation of immature melanoma cells.

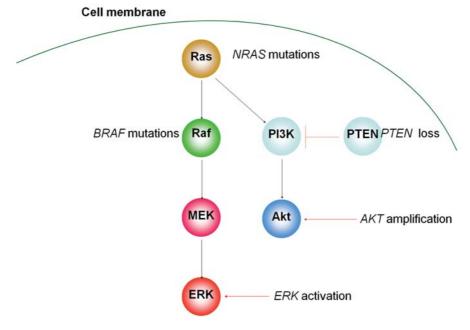


Figure 2. Raf/MEK/ERK and PI3K/Akt pathways and the mutations activating these pathways in melanoma.

that promote cell proliferation and survival (68-70). The lipid phosphatase PTEN negatively regulates this cascade through dephosphorylation of PIP3 (70).

Recent studies have revealed deregulation of the PI3K signalling in a high proportion of melanomas. Indeed, in about 45% of melanomas, PTEN is deleted and the downstream Akt gene is amplified (71,72). Both of these mutations result in overexpression of Akt3 (72), an isoform of Akt. Increased phospho-Akt expression in melanoma is associated with tumor progression and shorter survival (73,74). Oncogenic RAS can also bind and activate PI3K, resulting in increased AKT activity (75). These data suggest that loss of PTEN and oncogenic activation of RAS are largely equivalent with regard to their ability to increase oncogenic signalling through the PI3K pathway (76). This hypothesis is supported by the finding that PTEN somatic mutations are seen in melanomas harbouring mutations in BRAF but not NRAS (77). This is consistent with the ability of NRAS to activate both the PI3K and MAPK cascades, so in the presence of oncogenic NRAS additional mutations in BRAF and PTEN are unnecessary (6,74). In the recent evaluation of genomic alterations in primary melanomas, tumors with BRAF mutations had fewer copies of PTEN than those with NRAS mutations, suggesting that dual activation of the PI3K and MAPK pathways are important events in melanoma development (74,78). In Fig. 3 the frequency of NRAS, BRAF, PI3K mutations in our melanoma case series is reported. The preliminary data showed that the MAPK and AKT pathways were activated in all samples having gene mutations (Fig. 4). However, only the AKT pathway was highly expressed in tumor tissues with both PIK3CA and BRAF mutations suggesting that some PIK3CA mutations may block the MAPK pathway by activating AKT which phosphorylates and inactivates Raf.

Role of the p16(INK4a)-Rb (retinoblastoma protein) pathway in melanoma tumorigenesis. The p16(INK4a)-Rb pathway is a critical gatekeeper for cell cycle progression; in the Cdk4/6-mediated phosphorylated state, Rb drives cells towards G1/S-phase transition, while in the hypophosphorylated state, Rb binds and represses the E2F transcription factor and prevents the progression through the S-phase (79). p16<sup>INK4a</sup> stops cell cycle inhibiting the cyclin D/CDK4 complex, thereby preventing it from phosphorylating Rb (80).

The exit of cells from cell cycle is a physiological process; indeed, normal somatic cells have a finite lifespan, and after a finite number of divisions they exit from the cell cycle and enter a state known as senescence (6,81); senescence also occurs in response to oncogenic stress, so acting as a cellular protection mechanism against cancer formation (82,83). It has been shown that abnormally high activation of the MAP kinase pathway can inhibit cellular growth in a wide variety of normal and cancer cells by promoting cellular senescence (84,85); notably, V600EBRAF was recently found to induce p16<sup>INK4a</sup> expression and senescence in primary human melanocytes in vitro (84,86). Therefore, senescence can be overcome only if the p16(INK4a)-Rb pathway is not fully engaged, and this may occur when p16<sup>INK4a</sup> is inactivated (87,88). It has been reported that germline mutations in  $p16^{\text{INK4a}}$  are linked to familial melanoma susceptibility (89-91); somatic mutations in gene encoding p16<sup>INK4a</sup> are also found in most sporadic

melanomas (92,93). p16<sup>INK4a</sup> is inactivated by deletions, point mutations, promoter methylation (94,95) or through transcriptional silencing by overexpression of the transcriptional suppressor, inhibitor of differentiation 1 (ID1) (96). Given that p16<sup>INK4a</sup> needs to directly interact with the cyclin-Cdk complex in order to inhibit its protein kinase activity, changes in CDK4 that render it resistant to p16<sup>INK4a</sup> mimic p16<sup>INK4a</sup> loss (66). Both somatic and germline mutations in CDK4 have been detected in melanoma cell lines (97) and in familial melanomas (98).

Recently, another way to circumvent oncogene-induced senescence during melanoma progression has been discovered, Akt3 in early melanocytic lesions has been shown to phosphorylate V600EBRAF to reduce its activity and the MAP kinase pathway activity to levels promoting, rather than inhibiting, proliferation to overcome the senescence block (85,99).

#### 4. Resistance to apoptosis and chemotherapy

It has been shown that melanoma cells have low levels of spontaneous apoptosis *in vivo* compared with other tumor cell types, and are relatively resistant to drug-induced apoptosis *in vitro* (6,100). As most chemotherapeutic drugs function by inducing apoptosis in malignant cells, resistance to apoptosis is thought to be the main cause of drug resistance in melanoma (100).

Dysregulation of the intrinsic (mitochondrial-dependent) apoptotic pathway form the basis for melanoma's resistance to apoptosis and chemotherapy (101-107). The p53/Bcl-2 signalling network is one of the most important regulators of cell apoptosis; the Bcl-2 superfamily includes proapoptotic (BAX, BAK, BAD, BID, Bim, NOXA, PUMA) and antiapoptotic (Bcl-2, Bcl-xL, Mcl-1, BCL-w, and A1) members. In response to irreversible DNA damage, p53 becomes activated and induces the expression of proapoptotic members of the Bcl-2 family; these effectors promote mitochondrial membrane permeabilization and release of cytochrome c, which binds to Apaf-1 leading to the activation of effector caspases that result in apoptosis (108).

The loss of p53 function allows the cells that have suffered DNA damage to survive and divide, propagating pro-cancerous mutations; unlike many other chemoresistant cancers, melanomas harbour a very low frequency of p53 mutations (109-112). Therefore, other components of the p53 pathway, either upstream or downstream of p53 are likely defective in melanoma. It has been shown that aberrant methylation lead to loss of Apaf-1 expression rendering the cells unable to execute the normal apoptotic response following p53 activation (66,104). It has also been reported that decreased levels of Apaf-1 correlate with advanced disease and chemoresistance (66,113).

High levels of Bcl-2 expression have been found in melanoma and melanocytes (100,114). Aberrations in various signalling pathways contribute to elevated Bcl-2 levels in melanoma (66). In 1999, Borner *et al* found that mutant NRAS upregulates the expression of Bcl-2 *in vitro* and in SCID mice (101). MITF may also contribute to survival by the transactivation of Bcl-2 (61,66).

Alterations in other members of the Bcl-2 family were demonstrated to be involved in melanoma progression and

## Gene mutations in melanoma

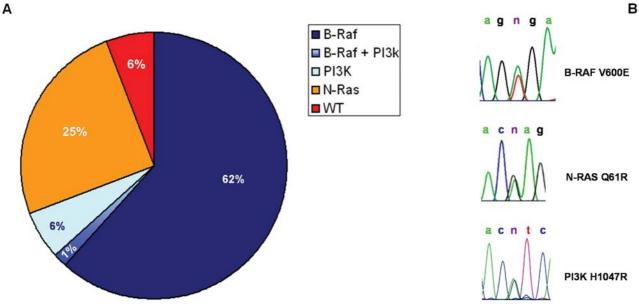
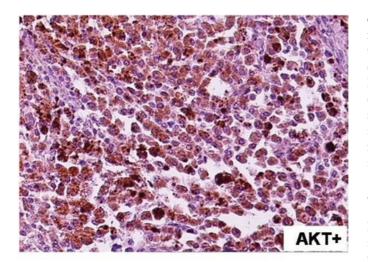


Figure 3. (A), NRAS, BRAF, PI3K mutations in a melanoma case series. (B), Hot-spot mutations of these genes.



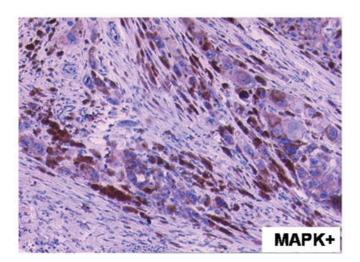


Figure 4. AKT and MAPK activation in a representative melanoma sample with both BRAF and PI3K mutations.

chemoresistance. Several studies have demonstrated that resistance to a variety of traditional and targeted chemotherapeutic agents is largely mediated by Mcl-1 overexpression (66,115-121); unlike other antiapoptotic Bcl-2 family members, Mcl-1 suppresses apoptosis induced by BAK but not BAX (122); Mcl-1 also has the unique property of rapid steady-state turnover due to proteasomal degradation (66,123); thus, in the presence of chemotherapeutics that inhibit proteasome function, such as bortezomib, Mcl-1 can accumulate and result in decreased sensitivity to these agents (66,118).

#### 5. Emerging target therapies

Recent progress in understanding the signalling pathways involved in melanomagenesis has led researchers to develop targeted therapies for this disease. These include selective inhibitors of the RAF and MEK kinases, inhibitors of the PI3K pathway and the Hsp90 chaperone protein.

Inhibitors of the RAF kinases. Sorafenib (BAY43-9006) is an oral multi-kinase inhibitor that decreases activity of RAF, VEGF receptor 1, 2 and 3, PDGFR, Flt-3, p38, c-kit, and FGFR-1 (124), so inhibiting both tumor cell growth and angiogenesis (46,125,126). It has been shown that sorafenib inhibits the growth of melanoma xenografts in mice (46), while it has little or no antitumor activity in advanced melanoma patients as a single agent (127). The reasons why sorafenib failed in clinical trials are not clear; perhaps it is unable to reach a concentration sufficient to inhibit B-Raf or it is possible that proliferation of melanoma cells is driven by alternative signalling pathways after signalling through RAF/ MEK/ERK has been blocked (127). To improve the efficacy of sorafenib in the therapy of melanoma, it is being combined with standard chemotherapeutic drugs; preliminary results combining sorafenib with carboplatin and paclitaxel were

encouraging (128). However, phase III trials have shown that this combination failed to improve progression-free survival of patients with advanced melanoma (128). Recently, it has been seen that sorafenib activates glycogen synthase kinase-3ß (GSK-3ß) in melanoma cell lines (129,130); constitutive activation of this kinase correlates with a marked increase in basal levels of Bcl-2, Bcl-x(L) and decreased antitumor efficacy of sorafenib. Therefore, sorafenib given in conjunction with targeted therapies against glycogen synthase kinase-3ß or the antiapoptotic Bcl-2 family members may prove useful (66,130).

The limited activity of sorafenib in tumors with oncogene BRAF prompted the evaluation of the efficacy of more specific BRAF inhibitors, such as RAF-265 (CHIR-265) and PLX-4032 (Plexikkon), in a phase I study for stage III/IV melanoma (http://www.clinicaltrial.gov/ct2/show/NCT 00304525?term=NCT00304525&rank=1) and advanced solid tumors, respectively (Tsai J, *et al*, Proc Am Assoc Cancer Res 47: abs. 571, 2006).

Inhibitors of the MEK kinases. Recently, it has been shown that melanoma cell lines with mutant BRAF are more sensitive to MEK inhibition than lines harboring oncogene RAS (131). In BRAF mutant tumors, MEK inhibition results in down-regulation of cyclin D1, upregulation of p27, hypophosphorylation of RB and growth arrest in G1. MEK inhibition also induces differentiation and senescence of BRAF mutant cells and apoptosis in some but not all V600E BRAF mutant models (53,131,132). Two MEK inhibitors are currently being tested in clinical trials: PD0325901 (Pfizer Oncology) and ARRY-142886 (AZD6244).

Inhibitors of the PI3K pathway. CCI-779 (Temsirolimus) and RAD001 (Everolimus), are the most advanced agents in the attack on the PI3K pathway (133). They target mTOR, a serine/threonine kinase downstream of Akt that modulates protein synthesis, cell-cycle progression, and angiogenesis (134). Since mTOR is a cytosolic protein expressed by all tissues, these inhibitors do not have high specificity in targeting melanoma tumor cells (66). Furthermore, it has been determined that the mTOR pathway has a complicated feedback loop that involves suppression of Akt, hence mTOR inhibitors would potentially activate Akt in some cells (36).

The MAPK and the PI3K signalling pathways both play a key role in melanoma cell proliferation and survival (77) suggesting that parallel inhibition of targets in both pathways may result in synergistic inhibition of growth in melanomas (135).

Inhibitors of the Hsp90 chaperone protein. The molecular chaperone heat-shock protein 90 (Hsp90) is required for the folding, conformational maturation, and stability of a subset of signalling molecules, including CRAF, mutant BRAF, HER2 and AKT. Exposure of melanoma cells to the Hsp90 inhibitor benzoquinone anisomycin 17AAG results in the proteasomal degradation of mutant BRAF, inhibition of mitogen-activated protein kinase activation and cell proliferation, induction of apoptosis, and antitumor activity (136,137). Furthermore, clinical activity has been shown with 17-AAG in patients with HER2 amplified breast cancer

and multiple myeloma (53,138,139). Though promising, 17-AAG has limited oral bioavailability and is poorly soluble. This has necessitated the use of intermittent intravenous dosing (once or twice weekly) likely limiting its efficacy in cancer patients. Novel small molecule Hsp90 inhibitors with improved oral bioavailability have recently entered phase I clinical testing and clinical evaluation of these compounds in tumors with a high frequency of mutated BRAF is warranted (53,140-142).

#### 6. Conclusions

The recent identification of several key molecular pathways implicated in the pathogenesis of melanoma and induction of chemotherapeutic drug resistance has led to the development of new targeted therapies for this devastating disease. Targeting various effectors of these pathways with pharmacologic inhibitors may inhibit melanoma cell growth and angiogenesis; the specific action of these new molecular targeted agents minimizes unexpected toxicity that is typical of systemic chemotherapy. Ongoing clinical trials provide hope to improve progression-free survival of patients with advanced melanoma.

#### References

- 1. Miller AJ and Mihm MC Jr: Melanoma. N Engl J Med 355: 51-65, 2006.
- Carlson JA, Slominski A, Linette GP, Mysliborski J, Hill J, Mihm MC Jr and Ross JS: Malignant melanoma 2003: predisposition, diagnosis, prognosis, and staging. Am J Clin Pathol 120: 101-127, 2003.
- 3. Jemal A, Siegel R, Ward E, Hao Y, Xu J, Murray T and Thun MJ: Cancer statistics, 2008. CA Cancer J Clin 58: 71-96, 2008.
- 4. Tsao H, Atkins MB and Sober AJ: Management of cutaneous melanoma. N Engl J Med 351: 998-1012, 2004.
- Moan J, Porojnicu AC and Dahlback A: Ultraviolet radiation and malignant melanoma. Adv Exp Med Biol 624: 104-116, 2008.
- 6. Gray-Schopfer V, Wellbrock C and Marais R: Melanoma biology and new targeted therapy. Nature 445: 851-857, 2007.
- 7. Chabner BA: Biological basis for cancer treatment. Ann Intern Med 118: 633-637, 1993.
- 8. Garnett MJ and Marais R: Guilty as charged: B-RAF is a human oncogene. Cancer Cell 6: 313-319, 2004.
- Minden A, Lin A, McMahon M, Lange-Carter C, Dérijard B, Davis RJ, Johnson GL and Karin M: Differential activation of ERK and JNK mitogen-activated protein kinases by Raf-1 and MEKK. Science 266: 1719-1723, 1994.
- Lange-Carter CA and Johnson GL: Ras-dependent growth factor regulation of MEK kinase in PC12 cells. Science 265: 1458-1461, 1994.
- 11. Marais R, Light Y, Paterson HF and Marshall CJ: Ras recruits Raf-1 to the plasma membrane for activation by tyrosine phosphorylation. EMBO J 14: 3136-3145, 1995.
- Marais R, Light Y, Paterson HF, Mason CS and Marshall CJ: Differential regulation of Raf-1, A-Raf, and B-Raf by oncogenic ras and tyrosine kinases. J Biol Chem 272: 4378-4383, 1007
- Mason CS, Springer CJ, Cooper RG, Superti-Furga G, Marshall CJ and Marais R: Serine and tyrosine phosphorylations cooperate in Raf-1, but not B-Raf activation. EMBO J 18: 2137-2148, 1999.
- 14. Xu S, Robbins D, Frost J, Dang A, Lange-Carter C and Cobb MH: MEKK1 phosphorylates MEK1 and MEK2 but does not cause activation of mitogen-activated protein kinase. Proc Natl Acad Sci USA 92: 6808-6812, 1995.
- Deng T and Karin M: c-Fos transcriptional activity stimulated by H-Ras-activated protein kinase distinct from JNK and ERK. Nature 371: 171-175, 1994.
- Davis RJ: Transcriptional regulation by MAP kinases. Mol Reprod Dev 42: 459-467, 1995.

- 17. Robinson MJ, Stippec SA, Goldsmith E, White MA and Cobb MH: A constitutively active and nuclear form of the MAP kinase ERK2 is sufficient for neurite outgrowth and cell transformation. Curr Biol 8: 1141-1150, 1998.
- Aplin AE, Stewart SA, Assoian RK and Juliano RL: Integrinmediated adhesion regulates ERK nuclear translocation and phosphorylation of Elk-1. J Cell Biol 153: 273-282, 2001.
- McCubrey JA, May WS, Duronio V and Mufson A: Serine/ threonine phosphorylation in cytokine signal transduction. Leukemia 14: 9-21, 2000.
- 20. Tresini M, Lorenzini A, Frisoni L, Allen RG and Cristofalo VJ: Lack of Elk-1 phosphorylation and dysregulation of the extracellular regulated kinase signaling pathway in senescent human fibroblast. Exp Cell Res 269: 287-300, 2001.
- Eblen ST, Catling AD, Assanah MC and Weber MJ: Biochemical and biological functions of the N-terminal, noncatalytic domain of extracellular signal-regulated kinase 2. Mol Cell Biol 21: 249-259, 2001.
- 22. Adachi T, Kar S, Wang M and Carr BI: Transient and sustained ERK phosphorylation and nuclear translocation in growth control. J Cell Physiol 192: 151-159, 2002.
- 23. Wang CY, Bassuk AG, Boise LH, Thompson CB, Bravo R and Leiden JM: Activation of the granulocyte-macrophage colony-stimulating factor promoter in T cells requires cooperative binding of Elf-1 and AP-1 transcription factors. Mol Cell Biol 14: 1153-1159, 1994.
- 24. Thomas RS, Tymms MJ, McKinlay LH, Shannon MF, Seth A and Kola I: ETS1, NFkappaB and AP1 synergistically transactivate the human GM-CSF promoter. Oncogene 14: 2845-2855, 1997.
- Ponti C, Gibellini D, Boin F, Melloni E, Manzoli FA, Cocco L, Zauli G and Vitale: Role of CREB transcription factor in c-fos activation in natural killer cells. Eur J Immunol 32: 3358-3365, 2002.
- 26. Fry TJ and Mackall CL: Interleukin-7: from bench to clinic. Blood 99: 3892-3904, 2002.
- Deng X, Kornblau SM, Ruvolo PP and May WS Jr: Regulation of Bcl2 phosphorylation and potential significance for leukemic cell chemoresistance. J Natl Cancer Inst Monogr 28: 30-37, 2001
- 28. Carter BZ, Milella M, Tsao T, McQueen T, Schober WD, Hu W, Dean NM, Steelman L, McCubrey JA and Andreeff M: Regulation and targeting of antiapoptotic XIAP in acute myeloid leukemia. Leukemia 17: 2081-2089, 2003.
- 29. Jia W, Yu C, Rahmani M, Krystal G, Sausville EA, Dent P and Grant S: Synergistic antileukemic interactions between 17-AAG and UCN-01 involve interruption of RAF/MEK- and AKTrelated pathways. Blood 102: 1824-1832, 2003.
- 30. Troppmair J and Rapp UR: Raf and the road to cell survival: a tale of bad spells, ring bearers and detours. Biochem Pharmacol 66: 1341-1345, 2003.
- 31. Harada H, Quearry B, Ruiz-Vela A and Korsmeyer SJ: Survival factor-induced extracellular signal-regulated kinase phosphorylates BIM, inhibiting its association with BAX and proapoptotic activity. Proc Natl Acad Sci USA 101: 15313-15317, 2004.
- 32. Marani M, Hancock D, Lopes R, Tenev T, Downward J and Lemoine NR: Role of Bim in the survival pathway induced by Raf in epithelial cells. Oncogene 23: 2431-2441, 2004.
- 33. Ley R, Balmanno K, Hadfield K, Weston C and Cook SJ: Activation of the ERK1/2 signaling pathway promotes phosphorylation and proteasome-dependent degradation of the BH3-only protein, Bim. J Biol Chem 278: 18811-18816, 2003.
- 34. Weston CR, Balmanno K, Chalmers C, Hadfield K, Molton SA, Ley R, Wagner EF and Cook SJ: Activation of ERK1/2 by deltaRaf-1:ER\* represses Bim expression independently of the JNK or PI3K pathways. Oncogene 22: 1281-1293, 2003.
- JNK or PI3K pathways. Oncogene 22: 1281-1293, 2003.
  35. Domina AM, Vrana JA, Gregory MA, Hann SR and Craig RW: MCL1 is phosphorylated in the PEST region and stabilized upon ERK activation in viable cells, and at additional sites with cytotoxic okadaic acid or taxol. Oncogene 23: 5301-5315, 2004.
- McCubrey JA, Milella M, Tafuri A, Martelli AM, Lunghi P, Bonati A, Cervello M, Lee JT and Steelman LS: Targeting the Raf/MEK/ERK pathway with small-molecule inhibitors. Curr Opin Investig Drugs 9: 614-630, 2008.
- 37. McCubrey JA, Steelman LS, Chappell WH, Abrams SL, Wong EW, Chang F, Lehmann B, Terrian DM, Milella M, Tafuri A, Stivala F, Libra M, Basecke J, Evangelisti C, Martelli AM and Franklin RA: Roles of the Raf/MEK/ERK pathway in cell growth, malignant transformation and drug resistance. Biochim Biophys Acta 1773: 1263-1284, 2007.

- 38. Cohen C, Zavala-Pompa A, Sequeira JH, Shoji M, Sexton DG, Cotsonis G, Cerimele F, Govindarajan B, Macaron N and Arbiser JL: Mitogen-actived protein kinase activation is an early event in melanoma progression. Clin Cancer Res 8: 3728-3733, 2002.
- 39. Davies H, Bignell GR, Cox C, *et al*: Mutations of the BRAF gene in human cancer. Nature 417: 949-954, 2002.
- Libra M, Malaponte G, Navolanic PM, Gangemi P, Bevelacqua V, Proietti L, Bruni B, Stivala F, Mazzarino MC, Travali S and McCubrey JA: Analysis of BRAF mutation in primary and metastatic melanoma. Cell Cycle 4: 1382-1384, 2005.
- 41. Dahl C and Guldberg P: The genome and epigenome of malignant melanoma. APMIS 115: 1161-1176, 2007.
- 42. Whitwam T, Vanbrocklin MW, Russo ME, Haak PT, Bilgili D, Resau JH, Koo HM and Holmen SL: Differential oncogenic potential of activated RAS isoforms in melanocytes. Oncogene 26: 4563-4570, 2007.
- 43. Wan PT, Garnett MJ, Roe SM, Lee S, Niculescu-Duvaz D, Good VM, Jones CM, Marshall CJ, Springer CJ, Barford D and Marais R: Mechanism of activation of the RAF-ERK signaling pathway by oncogenic mutations of B-RAF. Cell 116: 855-867, 2004.
- 44. Tuveson DA, Weber BL and Herlyn M: BRAF as a potential therapeutic target in melanoma and other malignancies. Cancer Cell 4: 95-98, 2003.
- Hoeflich KP, Eby MT, Forrest WF, Gray DC, Tien JY, Stern HM, Murray LJ, Davis DP, Modrusan Z and Seshagiri S: Regulation of ERK3/MAPK6 expression by BRAF. Int J Oncol 29: 839-849, 2006.
- Sharma A, Trivedi NR, Zimmerman MA, Tuveson DA, Smith CD and Robertson GP: Mutant V599EB-Raf regulates growth and vascular development of malignant melanoma tumors. Cancer Res 65: 2412-2421, 2005.
- 47. Pritchard CA, Samuels ML, Bosch E and McMahon M: Conditionally oncogenic forms of the A-Raf and B-Raf protein kinases display different biological and biochemical properties in NIH 3T3 cells. Mol Cell Biol 15: 6430-6442, 1995.
- 48. Emuss V, Garnett M, Mason C and Marais R: Mutations of C-RAF are rare in human cancer because C-RAF has a low basal kinase activity compared with B-RAF. Cancer Res 65: 9719-9726, 2005.
- Christensen C and Guldberg P: Growth factors rescue cutaneous melanoma cells from apoptosis induced by knock-down of mutated (V600E) BRAF. Oncogene 24: 6292-6302, 2005.
- Goel VK, Lazar AJ, Warneke CL, Redston MS and Haluska FG: Examination of mutations in BRAF, NRAS, and PTEN in primary cutaneous melanoma. J Invest Dermatol 126: 154-160, 2006.
- 51. Pollock PM, Harper UL, Hansen KS, *et al*: High frequency of BRAF mutations in nevi. Nat Genet 33: 19-20, 2003.
- Yazdi AS, Palmedo G, Flaig MJ, et al: Mutations of the BRAF gene in benign and malignant melanocytic lesions. J Invest Dermatol 121: 1160-1162, 2003.
- Halilovic E and Solit DB: Therapeutic strategies for inhibiting oncogenic BRAF signaling. Curr Opin Pharmacol 8: 419-426, 2008.
- 54. Widlund HR and Fisher DE: Microphthalamia-associated transcription factor: a critical regulator of pigment cell development and survival. Oncogene 22: 3035-3041, 2003.
- Levy C, Khaled M and Fisher DE: MITF: master regulator of melanocyte development and melanoma oncogene. Trends Mol Med 12: 406-414, 2006.
- Denat L and Larue L: Malignant melanoma and the role of the paradoxal protein Microphthalmia transcription factor. Bull Cancer 94: 81-92, 2007.
- Loercher AE, Tank EM, Delston RB and Harbour JW: MITF links differentiation with cell cycle arrest in melanocytes by transcriptional activation of INK4A. J Cell Biol 168: 35-40, 2005.
- Carreira S, Goodall J, Aksan I, et al: Mitf cooperates with Rb1 and activates p21Cip1 expression to regulate cell cycle progression. Nature 433: 764-769, 2005.
- Wellbrock C and Marais R: Elevated expression of MITF counteracts B-RAF stimulated melanocyte and melanoma cell proliferation. J Cell Biol 170: 703-708, 2005.
- Du J, Widlund HR, Horstmann MA, et al: Critical role of CDK2 for melanoma growth linked to its melanocyte-specific transcriptional regulation by MITF. Cancer Cell 6: 565-576, 2004.
- 61. McGill GG, Horstmann M, Widlund HR, *et al*: Bcl2 regulation by the melanocyte master regulator Mitf modulates lineage survival and melanoma cell viability. Cell 109: 707-718, 2002.

- 62. Garraway LA, Widlund HR, Rubin MA, *et al*: Integrative genomic analyses identify MITF as a lineage survival oncogene amplified in malignant melanoma. Nature 436: 117-122, 2005.
- 63. Widlund HR, Horstmann MA, Price ER, *et al*: Betacatenin-induced melanoma growth requires the downstream target Microphthalmia-associated transcription factor. J Cell Biol 158: 1079-1087, 2002.
- 64. Rubinfeld B, Robbins P, El-Gamil M, Albert I, Porfiri E and Polakis P: Stabilization of beta-catenin by genetic defects in melanoma cell lines. Science 275: 1790-1792, 1997.
- Rimm DL, Caca K, Hu G, Harrison FB and Fearon ER: Frequent nuclear/cytoplasmic localization of beta-catenin without exon 3 mutations in malignant melanoma. Am J Pathol 154: 325-329, 1999.
- 66. Hocker TL, Singh MK and Tsao H: Melanoma genetics and therapeutic approaches in the 21st century: moving from the benchside to the bedside. J Invest Dermatol 128: 2575-2595, 2008
- Wellbrock C, Rana S, Paterson H, Pickersgill H, Brummelkamp T and Marais R: Oncogenic BRAF regulates melanoma proliferation through the lineage specific factor MITF. PLoS ONE 3: e2734, 2008.
- 68. Robertson GP: Functional and therapeutic significance of akt deregulation in malignant melanoma. Cancer Metastasis Rev 24: 273-285, 2005.
- 69. Cully M, You H, Levine AJ and Mak TW: Beyond PTEN mutations: the PI3K pathway as an integrator of multiple inputs during tumorigenesis. Nat Rev Cancer 6: 184-192, 2006.
- 70. Hennessy BT, Smith DL, Ram PT, Lu Y and Mills GB: Exploiting the PI3K/AKT pathway for cancer drug discovery. Nat Rev Drug Discov 4: 988-1004, 2005.
- 71. Zhou XP, Gimm O, Hampel H, Niemann T, Walker MJ and Eng C: Epigenetic PTEN silencing in malignant melanomas without PTEN mutation. Am J Pathol 157: 1123-1128, 2000.
- Stahl JM, Sharma A, Cheung M, Zimmerman M, Cheng JQ, Bosenberg MW, Kester M, Sandirasegarane L and Robertson GP: Deregulated Akt3 activity promotes development of malignant melanoma. Cancer Res 64: 7002-7010, 2004.
- 73. Meier F, Schittek B, Busch S, Garbe C, Smalley K, Satyamoorthy K, Li G and Herlyn M: The RAS/RAF/MEK/ERK and PI3K/AKT signaling pathways present molecular targets for the effective treatment of advanced melanoma. Front Biosci 10: 2986-3001, 2005.
- 74. Fecher LA, Cummings SD, Keefe MJ and Alani RM: Toward a molecular classification of melanoma. J Clin Oncol 25: 1606-1620, 2007.
- 75. Sekulic A, Haluska P Jr, Miller AJ, *et al*: Malignant melanoma in the 21st century: the emerging molecular landscape. Mayo Clin Proc 83: 825-846, 2008.
- Tsao H, Zhang X, Fowlkes K and Haluska FG: Relative reciprocity of NRAS and PTEN/MMAC1 alterations in cutaneous melanoma cell lines. Cancer Res 60: 1800-1804, 2000.
- 77. Tsao H, Goel V, Wu H, Yang G and Haluska FG: Genetic interaction between NRAS and BRAF mutations and PTEN/ MMAC1 inactivation in melanoma. J Invest Dermatol 122: 337-341, 2004.
- Curtin JA, Fridlyand J, Kageshita T, Patel HN, Busam KJ, Kutzner H, Cho KH, Aiba S, Bröcker EB, Le Boit PE, Pinkel D and Bastian BC: Distinct sets of genetic alterations in melanoma. N Engl J Med 353: 2135-2147, 2005.
- Harbour JW and Dean DC: The Rb/E2F pathway: expanding roles and emerging paradigms. Genes Dev 14: 2393-2409, 2000.
- Serrano M, Hannon GJ and Beach D: A new regulatory motif in cell-cycle control causing specific inhibition of cyclin D/CDK4. Nature 366: 704-707, 1993.
- 81. Bennett DC: Human melanocyte senescence and melanoma susceptibility genes. Oncogene 22: 3063-3069, 2003.
- Mooi WJ and Peeper DS: Oncogene-induced cell senescence-halting on the road to cancer. N Engl J Med 355: 1037-1046, 2006.
- 83. Bartkova J, Rezaei N, Liontos M, *et al*: Oncogene-induced senescence is part of the tumorigenesis barrier imposed by DNA damage checkpoints. Nature 444: 633-637, 2006.
- 84. Michaloglou C, Vredeveld LC, Soengas MS, Denoyelle C, Kuilman T, van der Horst CM, Majoor DM, Shay JW, Mooi WJ and Peeper DS: BRAFE600-associated senescence-like cell cycle arrest of human naevi. Nature 436: 720-724, 2005.
- 85. Madhunapantula SV and Robertson GP: Is B-Raf a good therapeutic target for melanoma and other malignancies? Cancer Res 68: 5-8, 2008.

- 86. Gray-Schopfer VC, Cheong SC, Chong H, Chow J, Moss T, Abdel-Malek ZA, Marais R, Wynford-Thomas D and Bennett DC: Cellular senescence in naevi and immortalisation in melanoma: a role for p16? Br J Cancer 95: 496-505, 2006.
- 87. Haferkamp S, Becker TM, Scurr LL, Kefford RF and Rizos H: p16INK4a-induced senescence is disabled by melanoma-associated mutations. Aging Cell 7: 733-745, 2008.
  88. Bachmann IM, Straume O and Akslen LA: Altered expression
- 88. Bachmann IM, Straume O and Akslen LA: Altered expression of cell cycle regulators Cyclin D1, p14, p16, CDK4 and Rb in nodular melanomas. Int J Oncol 25: 1559-1565, 2004.
- 89. Hussussian CJ, Struewing JP, Goldstein AM, Higgins PA, Ally DS, Sheahan MD, Clark WH Jr, Tucker MA and Dracopoli NC: Germline p16 mutations in familial melanoma. Nat Genet 8: 15-21, 1994.
- 90. Eliason MJ, Larson AA, Florell SR, Zone JJ, Cannon-Albright LA, Samlowski WE and Leachman SA: Population-based prevalence of CDKN2A mutations in Utah melanoma families. J Invest Dermatol 126: 660-666, 2006.
- 91. Goldstein AM, Chan M, Harland M, et al: Features associated with germline CDKN2A mutations: a GenoMEL study of melanoma-prone families from three continents. J Med Genet 44: 99-106, 2007.
- 92. Bartkova J, Lukas J, Guldberg P, Alsner J, Kirkin AF, Zeuthen J and Bartek J: The p16-cyclin D/Cdk4-pRb pathway as a functional unit frequently altered in melanoma pathogenesis. Cancer Res 56: 5475-5483, 1996.
- 93. Walker GJ, Flores JF, Glendening JM, Lin AH, Markl ID and Fountain JW: Virtually 100% of melanoma cell lines harbor alterations at the DNA level within CDKN2A, CDKN2B, or one of their downstream targets. Genes Chromosomes Cancer 22: 157-163, 1998.
- 94. Sharpless E and Chin L: The INK4a/ARF locus and melanoma. Oncogene 22: 3092-3098, 2003.
- 95. Zhang H and Rosdahl I: Deletion in p16INK4a and loss of p16 expression in human skin primary and metastatic melanoma cells. Int J Oncol 24: 331-335, 2004.
- 96. Polsky D, Young AZ, Busam KJ and Alani RM: The transcriptional repressor of p16/Ink4a, Id1, is up-regulated in early melanomas. Cancer Res 61: 6008-6011, 2001.
- 97. Tsao H, Benoit E, Sober AJ, Thiele C and Haluska FG: Novel mutations in the p16/CDKN2A binding region of the cyclin-dependent kinase-4 gene. Cancer Res 58: 109-113, 1998.
- 98. Zuo L, Weger J, Yang Q, Goldstein AM, Tucker MA, Walker GJ, Hayward N and Dracopoli NC: Germline mutations in the p16INK4a binding domain of CDK4 in familial melanoma. Nat Genet 12: 97-99, 1996.
- 99. Cheung M, Sharma A, Madhunapantula SV and Robertson GP: Akt3 and mutant V600E B-Raf cooperate to promote early melanoma development. Cancer Res 68: 3429-3439, 2008.
- 100. Soengas MS and Lowe SW: Apoptosis and melanoma chemoresistance. Oncogene 22: 3138-3151, 2003.
- 101. Borner C, Schlagbauer-Wadl H, Fellay I, Selzer E, Polterauer P and Jansen B: Mutated N-ras upregulates Bcl-2 in human melanoma in vitro and in SCID mice. Melanoma Res 9: 347-350, 1999.
- 102. Jansen B, Wacheck V, Heere-Ress E, Schlagbauer-Wadl H, Hoeller C, Lucas T, Hoermann M, Hollenstein U, Wolff K and Pehamberger H: Chemosensitisation of malignant melanoma by BCL2 antisense therapy. Lancet 356: 1728-1733, 2000.
- 103. Helmbach H, Rossmann E, Kern MA and Schadendorf D: Drug-resistance in human melanoma. Int J Cancer 93: 617-622, 2001.
- 104. Soengas MS, Capodieci P, Polsky D, Mora J, Esteller M, Opitz-Araya X, McCombie R, Herman JG, Gerald WL, Lazebnik YA, Cordón-Cardó C and Lowe SW: Inactivation of the apoptosis effector Apaf-1 in malignant melanoma. Nature 409: 207-211, 2001.
- 409: 207-211, 2001.

  105. Heere-Ress E, Thallinger C, Lucas T, Schlagbauer-Wadl H, Wacheck V, Monia BP, Wolff K, Pehamberger H and Jansen B: Bcl-X(L) is a chemoresistance factor in human melanoma cells that can be inhibited by antisense therapy. Int J Cancer 99: 29-34, 2002.
- 106. Thallinger C, Wolschek MF, Wacheck V, Maierhofer H, Gunsberg P, Polterauer P, Pehamberger H, Monia BP, Selzer E, Wolff K and Jansen B: Mcl-1 antisense therapy chemosensitizes human melanoma in a SCID mouse xenotransplantation model. J Invest Dermatol 120: 1081-1086, 2003.
- 107. Galluzzi L, Larochette N, Zamzami N and Kroemer G: Mitochondria as therapeutic targets for cancer chemotherapy. Oncogene 25: 4812-4830, 2006.

- 108. Hengartner MO: The biochemistry of apoptosis. Nature 407: 770-776, 2000
- 109. Albino AP, Vidal MJ, McNutt NS, Shea CR, Prieto VG, Nanus DM, Palmer JM and Hayward NK: Mutation and expression of the p53 gene in human malignant melanoma. Melanoma Res 4: 35-45, 1994.
- 110. Barnhill RL, Castresana JS, Rubio MP, Martin MT, Idoate M, Vazquez JJ and Thor AD: p53 expression in cutaneous malignant melanoma: an immunohistochemical study of 87 cases of primary, recurrent, and metastatic melanoma. Mod Pathol 7: 533-535, 1994.
- 111. Lubbe J, Reichel M, Burg G and Kleihues P: Absence of p53 gene mutations in cutaneous melanoma. J Invest Dermatol 102: 819-821, 1994.
- 112. Petitjean A, Mathe E, Kato S, Ishioka C, Tavtigian SV, Hainaut P and Olivier M: Impact of mutant p53 functional properties on TP53 mutation patterns and tumor phenotype: lessons from recent developments in the IARC TP53 database. Hum Mutat 28: 622-629, 2007.
- 113. Mustika R, Budiyanto A, Nishigori C, Ichihashi M and Ueda M: Decreased expression of Apaf-1 with progression of melanoma. Pigment Cell Res 18: 59-62, 2005.
- 114. Selzer E, Schlagbauer-Wadl H, Okamoto I, Pehamberger H, Potter R and Jansen B: Expression of Bcl-2 family members in human melanocytes, in melanoma metastases and in melanoma cell lines. Melanoma Res 8: 197-203, 1998.
- 115. Fernandez Y, Verhaegen M, Miller TP, Rush JL, Steiner P, Opipari AW Jr, Lowe SW and Soengas MS: Differential regulation of noxa in normal melanocytes and melanoma cells by proteasome inhibition: therapeutic implications. Cancer Res 65: 6294-6304, 2005.
- 116. Qin JZ, Xin H, Sitailo LA, Denning MF and Nickoloff BJ: Enhanced killing of melanoma cells by simultaneously targeting Mcl-1 and NOXA. Cancer Res 66: 9636-9645, 2006.
- 117. Verhaegen M, Bauer JA, Martin de la Vega C, et al: A novel BH3 mimetic reveals a mitogenactivated protein kinasedependent mechanism of melanoma cell death controlled by p53 and reactive oxygen species. Cancer Res 66: 11348-11359, 2006.
- 118. Nguyen M, Marcellus RC, Roulston A, *et al*: Small molecule obatoclax (GX15-070) antagonizes MCL-1 and overcomes MCL-1-mediated resistance to apoptosis. Proc Natl Acad Sci USA 104: 19512-19517, 2007.
- 119. Wang YF, Jiang CC, Kiejda KA, Gillespie S, Zhang XD and Hersey P: Apoptosis induction in human melanoma cells by inhibition of MEK is caspase-independent and mediated by the Bcl-2 family members PUMA, Bim, and Mcl-1. Clin Cancer Res 13: 4934-4942, 2007.
- 120. Wolter KG, Verhaegen M, Fernandez Y, Nikolovska-Coleska Z, Riblett M, De la Vega CM, Wang S and Soengas MS: Therapeutic window for melanoma treatment provided by selective effects of the proteasome on Bcl-2 proteins. Cell Death Differ 14: 1605-1616, 2007.
- 121. Chetoui N, Sylla K, Gagnon-Houde JV, Alcaide-Loridan C, Charron D, Al-Daccak R and Aoudjit F: Down-regulation of mcl-1 by small interfering RNA sensitizes resistant melanoma cells to fas mediated apoptosis. Mol Cancer Res 6: 42-52, 2008.
- 122. Zhai D, Jin C, Huang Z, Satterthwait AC and Reed JC: Differential regulation of Bax and Bak by anti-apoptotic Bcl-2family proteins, Bcl-B and Mcl-1. J Biol Chem 283: 9580-9586, 2008.
- 123. Nijhawan D, Fang M, Traer E, Zhong Q, Gao W, Du F and Wang X: Elimination of Mcl-1 is required for the initiation of apoptosis following ultraviolet irradiation. Genes Dev 17: 1475-1486, 2003.
- 124. Wilhelm SM, Carter C, Tang L, et al: BAY 43-9006 exhibits broad spectrum oral antitumor activity and targets the RAF/ MEK/ERK pathway and receptor tyrosine kinases involved in tumor progression and angiogenesis. Cancer Res 64: 7099-7109, 2004.

- 125. Liu L, Cao Y, Chen C, Zhang X, McNabola A, Wilkie D, Wilhelm S, Lynch M and Carter C: Sorafenib blocks the RAF/MEK/ERK pathway, inhibits tumor angiogenesis, and induces tumor cell apoptosis in hepatocellular carcinoma model PLC/PRF/5. Cancer Res 66: 11851-11858, 2006.
- 126.Kim S, Yazici YD, Calzada G, Wang ZY, Younes MN, Jasser SA, El-Naggar AK and Myers JN: Sorafenib inhibits the angiogenesis and growth of orthotopic anaplastic thyroid carcinoma xenografts in nude mice. Mol Cancer Ther 6: 1785-1792, 2007.
- 127. Eisen T, Ahmad T, Flaherty KT, *et al*: Sorafenib in advanced melanoma: a Phase II randomised discontinuation trial analysis. Br J Cancer 95: 581-586, 2006.
- 128. Flaherty KT: Chemotherapy and targeted therapy combinations in advanced melanoma. Clin Cancer Res 12: 2366-2370, 2006.
- 129. Lejeune FJ, Rimoldi D and Speiser D: New approaches in metastatic melanoma: biological and molecular targeted therapies. Expert Rev Anticancer Ther 7: 701-713, 2007.
- 130.Panka DJ, Cho DC, Atkins MB and Mier JW: GSK-3beta inhibition enhances sorafenib induced apoptosis in melanoma cell lines. J Biol Chem 283: 726-732, 2008.
  131.Solit DB, Garraway LA, Pratilas CA, et al: BRAF mutation
- 131. Solit DB, Garraway LA, Pratilas CA, et al: BRAF mutation predicts sensitivity to MEK inhibition. Nature 439: 358-362, 2006.
- 132. Solit DB, Santos E, Pratilas CA, Lobo J, Moroz M, Cai S, Blasberg R, Sebolt-Leopold J, Larson S and Rosen N: 30-deoxy-30-[18F]fluorothymidine positron emission tomography is a sensitive method for imaging the response of BRAF dependent tumors to MEK inhibition. Cancer Res 67: 11463-11469, 2007.
- 133. Dancey JE: Therapeutic targets: MTOR and related pathways. Cancer Biol Ther 5: 1065-1073, 2006.
- 134. Granville CA, Memmott RM, Gills JJ and Dennis PA: Handicapping the race to develop inhibitors of the phosphoinositide 3-kinase/Akt/mammalian target of rapamycin pathway. Clin Cancer Res 12: 679-689, 2006.
- 135. Molhoek KR, Brautigan DL and Slingluff CL Jr: Synergistic inhibition of human melanoma proliferation by combination treatment with B-Raf inhibitor BAY43-9006 and mTOR inhibitor rapamycin. J Transl Med 3: 39, 2005.
- 136. Grbovic OM, Basso AD, Sawai A, Ye Q, Friedlander P, Solit D and Rosen N: V600E B-Raf requires the Hsp90 chaperone for stability and is degraded in response to Hsp90 inhibitors. Proc Natl Acad Sci USA 103: 57-62, 2006.
- 137. Rocha Dias S, Friedlos F, Light Y, Springer C, Workman P and Marais R: Activated B-RAF is an Hsp90 client protein that is targeted by the anticancer drug 17-allylamino-17-demethoxygeldanamycin. Cancer Res 65: 10686-10691, 2005.
- 138. Solit DB, Ivy SP, Kopil C, *et al*: Phase I trial of 17-allylamino-17-demethoxygeldanamycin in patients with advanced cancer. Clin Cancer Res 13: 1775-1782, 2007.
- 139. Modi S, Stopeck AT, Gordon MS, *et al*: Combination of trastuzumab and tanespimycin (17-AAG, KOS-953) is safe and active in trastuzumab-refractory HER-2 overexpressing breast cancer: a phase I dose-escalation study. J Clin Oncol 25: 5410-5417, 2007.
- 140. Chiosis G, Timaul MN, Lucas B, Munster PN, Zheng FF, Sepp-Lorenzino L and Rosen N: A small molecule designed to bind to the adenine nucleotide pocket of Hsp90 causes Her2 degradation and the growth arrest and differentiation of breast cancer cells. Chem Biol 8: 289-299, 2001.
- 141. Chandarlapaty S, Sawai A, Ye Q, Scott A, Silinski M, Huang K, Fadden P, Partdrige J, Hall S, Steed P, Norton L, Rosen N and Solit DB: SNX2112, a synthetic heat shock protein 90 inhibitor, has potent antitumor activity against HER kinase-dependent cancers. Clin Cancer Res 14: 240-248, 2008.
- 142. Eccles SA, Massey A, Raynaud FI, *et al*: NVPAUY922: a novel heat shock protein 90 inhibitor active against xenograft tumor growth, angiogenesis, and metastasis. Cancer Res 68: 2850-2860, 2008.