Vascular endothelial growth factor -634G/C polymorphism is associated with increased breast cancer risk and aggressiveness

DOONYAPAT SA-NGUANRAKSA¹, TUENJAI CHUANGSUWANICH², TAWATCHAI PONGPRUTTIPAN², TANAWAN KUMMALUE³, SUPAKORN ROJANANIN¹, ADUNE RATANAWICHHITRASIN¹, PORAMAPORN PRASARTTONG-OSOTH¹, SUEBWONG CHUTHATISITH¹, PONGTHEP PISARNTURAKIT¹, WARAPORN AEUMRITHAICHAROENCHOK¹, PRADIT RUSHATAMUKAYANUNT¹, VISNU LOHSIRIWAT¹, MONGKOL BOONSRIPITAYANON¹, PRIDA MALASIT⁴ and PORNCHAI O-CHAROENRAT¹

¹Division of Head, Neck and Breast Surgery, Department of Surgery; Departments of ²Pathology and ³Clinical Pathology; ⁴Dengue Hemorrhagic Fever Research Unit, Office for Research and Development, Faculty of Medicine, Siriraj Hospital, Mahidol University, Bangkok 10700, Thailand

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Abstract. Polymorphisms in the promoter and 5' untranslated region of vascular endothelial growth factor (VEGF) have been associated with VEGF levels. To investigate the role of VEGF polymorphisms in breast cancer, the VEGF -2578C/A, -1498C/T, -1154G/A and -634G/C polymorphisms were genotyped in 483 breast cancer patients and 524 healthy controls. VEGF mRNA levels in breast cancer tissue were determined using semi-quantitative RT-PCR. The genotypes, -634G/C and -634C/C, were associated with an increased risk for breast cancer when compared with the -634G/G genotype. The VEGF -634G/C genotype was associated with tumor size >20 mm, perineural invasion and stage II-IV. Individuals with -634C/C had lower disease-free survival. Patients with the VEGF -634C/C genotype exhibited the highest VEGF mRNA levels. High VEGF mRNA expression correlated with tumor size >20 mm, presence of lymphovascular invasion and axillary nodal metastasis. These observations suggested that VEGF -634G/C polymorphisms have a significant role in breast cancer susceptibility and aggressiveness.

Introduction

Breast cancer is the most prevalent cancer and the leading cause of cancer mortality in females worldwide, as well as in Thailand. Angiogenesis is the formation of new blood vessels and has been involved in the initiation and aggressiveness of breast cancer (1-3). The most important key modulator in this complex process is vascular endothelial growth factor (*VEGF*). *VEGF* plays a role in breast cancer (4) and the *VEGF* pathway is targeted in the treatment of breast cancer (5).

Human VEGF is localized on chromosome 6p21.3 and organized as eight exons, separated by seven introns (6,7). Several polymorphisms in the promoter and 5' untranslated region of (UTR) of VEGF have been identified (8,9). Awata et al previously identified seven polymorphisms in the promoter region as well as 5' and 3'UTR of VEGF in a Japanese population. Serum VEGF levels have also been found to be significantly higher in healthy subjects with the -634C/C genotype (10). However, in vitro experiments using lipopolysaccharide-stimulated peripheral blood mononuclear cells demonstrated that -634 G/G correlates with elevated VEGF production (9). In non-small cell lung cancer, a low VEGF expression in cancer tissue was significantly associated with the presence of the -2578C/C, -634G/G and -1154A/A and GA alleles in the VEGF promoter (11). The association between VEGF polymorphisms and breast cancer has been previously investigated (12). Based on these observations, we hypothesized that polymorphisms in the VEGF promoter and 5'UTR contribute to varied levels of VEGF expression, subsequently leading to susceptibility to aggressive breast cancer. To investigate this hypothesis, the association between VEGF polymorphisms and breast cancer susceptibility and aggressiveness was investigated, as well as mRNA expression in breast cancer tissue.

Materials and methods

Study population. The study population was recruited from the Division of Head, Neck and Breast Surgery (Siriraj Hospital, Bangkok, Thailand) between 2000 and 2003. Patients with histopathologically confirmed breast carcinoma were included in this prospective study and newly diagnosed breast cancer patients were included in the case group. Patients with histories

Correspondence to: Professor Pornchai O-Charoenrat, Division of Head, Neck and Breast Surgery, Department of Surgery, Faculty of Medicine, Siriraj Hospital, Mahidol University, 2 Prannok Road, Bangkok 10700, Thailand E-mail: sipoc@mahidol.ac.th

Key words: angiogenesis, breast cancer, polymorphisms, vascular endothelial growth factor

Polymorphisms	Primers	Product size, bp	PCR conditions (T, MgCl ₂) ^a	Alleles	Restriction enzyme	DNA fragment sizes, bp
-634G/C	F: CATTGATCCGGGTTTTATCCC	282	65-60 ^b , 1			
	G: CACTCACTTTGTCCCTGTAG			G	-	-
	C: CACTCACTTTGTCCCTGTAC			С		-
	Inc F: AGATGGTCCCTCACCTTCCT	352				
	Inc R: GTCTACCCTCCTGAGCTTGC					
-1154G/A	F: GTCCGCACGTAACCTCACTT	220	62, 1.5			
	G: GACAGGCGAGCTTCAGCACC			G	-	-
	A: GACAGGCGAGCTTCAGCACT			А	-	
	Inc F: AGATGGTCCCTCACCTTCCT	352				
	Inc R: GTCTACCCTCCTGAGCTTGC					
-1498C/T	F: TGTGCGTGTGGGGGTTGAGCG	175	60, 1.5	Т	<i>Bst</i> UI	175
	R: TACGTGCGGACAGGGCCTGA			С		155,20
-2578C/A	F: ATTGCTGCATTCCCATTCTC	251	60, 2.5	С	<i>Bst</i> YI	251
	R: CCCTTTTCCTCCAACTCTC	268		А		180,88

Table I. PCR primer pairs and conditions for VEGF genotyping.

^aT, annealing temperature (°C) and MgCl₂ concentrations (mM). ^bWith decrement of temperature of 0.5°C for each cycle until annealing temperature reached 60°C and reaction contained 2% DMSO. *VEGF*, vascular endothelial growth factor. F, forward; R, reverse; Inc, internal control.

	Table II. PCR primer	pairs and conditions	for semi-quantitative	RT-PCR of VEGF	and <i>β-actin</i> mRNA
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Gene	Primers	Product size, bp	$\begin{array}{c} PCR \ conditions \\ (T, MgCl_2)^a \end{array}$	PCR cycles
VEGF	F: CTCACCAAGGCCAGCACATAGG	159	55, 2.5	32
	R: ATCTGGTTCCCGAAACCCTGAG	291		
		363		
		414		
β-actin	F: TCGACAACGGCTCCGGCAT	239	50, 2.5	26
	R: AAGGTGTGGTGCCAGATTTTC			

^aT, annealing temperature (°C) and MgCl₂ concentrations (mM), VEGF, vascular endothelial growth factor. F, forward; R, reverse.

of other types of cancer were excluded. Healthy individuals and patients who attended the hospital due to benign conditions with no previous diagnosis of cancer were included in the control group and frequency matched to the breast carcinoma cases with regard to age (± 5 years). Informed consent was obtained from all subjects. This study was approved by the ethics committee of the Siriraj Hospital, Mahidol University, Bangkok, Thailand.

Genotyping of VEGF polymorphisms. VEGF -634G/C and -1154G/A were genotyped by the allele refractory mutation system-PCR. PCR was performed as summarized in Table I. VEGF -1498C/T and -2578C/A were genotyped by the PCR-restriction fragment length polymorphisms. Representative PCR products were sequenced to validate the assay.

Assessment of VEGF mRNA expression levels. The correlation between VEGF polymorphisms and expression levels was determined in breast tissue. VEGF mRNA expression was assayed by semi-quantitative RT-PCR as described previously (13). A primer pair that amplified β -actin was employed to check RNA integrity and used as an internal control. Primer sequences, PCR conditions and the number of PCR cycles are presented in Table II. To account for variability between gels, an RT-PCR product from the MDA-MB-231 cell line was electrophoresed as a control on each gel. PCR product intensity was analyzed using GeneTools® software (Syngene, Cambridge, UK). mRNA levels were calculated as the ratio of tissue sample to corresponding β -actin and then corrected as a ratio to the MDA-MB-231 sample on the same scan. Each RNA sample was assayed in duplicate and in two separate settings.

VEGF genotype polymorphisms	Controls, n (%)	Cases, n (%)	OR (95% CI), P-value
-634			
GG	234 (65.91)	223 (46.17)	1 (ref.)
GC	81 (22.82)	199 (41.20)	2.544 (1.852-3.496), 0.001
CC	40 (11.27)	61 (12.63)	1.600 (1.030-2.485), 0.036
-1154			
GG	279 (69.23)	318 (65.84)	1 (ref.)
GA	118 (29.28)	149 (30.85)	1.096 (0.819-1.467), 0.538
AA	6 (1.49)	16 (3.31)	2.320 (0.819-6.014), 0.084
-1498			
TT	215 (52.31)	243 (50.31)	1 (ref.)
CT	172 (41.85)	214 (44.31)	1.097 (0.835-1.441), 0.507
CC	24 (5.84)	26 (5.38)	1.002 (0.557-1.802), 0.995
-2578			
CC	214 (51.69)	240 (49.69)	1 (ref.)
AC	173 (41.79)	213 (44.10)	1.091 (0.830-1.435), 0.530
AA	27 (6.52)	30 (6.21)	1.016 (0.584-1.767), 0.957

Table III. Correlation between VEGF genotype and breast cancer susceptibility.

ORs and 95% CIs calculated by logistic regression, age-adjusted. VEGF, vascular endothelial growth factor; OR, odds ratio; CI, confidence interval.

Statistical analysis. Distribution of VEGF allele frequencies and genotypes among the case and control groups was analyzed using the χ^2 test. Odds ratios (ORs) and 95% confidence intervals (CIs), obtained from unconditional logistic regression, were used to measure the strength of the association between VEGF polymorphisms and susceptibility and aggressiveness of breast cancer. Individual haplotypes were determined using the PHASE program available at http://www.stat.washington.edu/stephens/phase.html (14). The end-point of overall survival (OS) analysis was breast cancer-associated mortality. The disease-free survival (DFS) analysis end-point was cancer recurrent/metastasis or breast cancer-associated mortality. DFS and OS time was calculated as the time from diagnosis to the end point of the study, censoring at the date of last contact or non-cancer mortality. The survival curves were determined using a Kaplan-Meier product-limit method. Statistical significance between the survival curves was assessed using the log-rank test. Multivariate analysis was performed to evaluate the effect of prognostic factors on OS, using the Cox proportional hazards model. P<0.05 was considered to indicate a statistically significant difference. mRNA levels were calculated as the ratio of tissue sample to corresponding β -actin and corrected as a ratio to the MDA-MB-231 cell line on the same scan. Each RNA sample was assayed in duplicate and in two separate settings.

Results

Correlation between VEGF genotypes and breast cancer susceptibility. Genotyping was performed on 483 breast cancer patients and 524 controls. -634C allele distribution was significantly higher in breast cancer patients compared with control subjects (33.23 vs. 23.71%; P<0.001).

VEGF -634G/C and -634C/C genotype distributions were significantly higher in breast cancer patients (GC, 41.20 and CC, 12.63 vs. GC, 25.06 and CC, 11.18%; P<0.001). Allele and genotype frequency distributions of other loci were not found to be different. The mean age of the control group was 48.56±14.45 years (SEM). The mean age of the breast cancer patients was 50.8±11.326 years (SEM). The mean age of the breast cancer and control groups were statistically different [OR, -2.238; 95% CI, (-5.20)-(-3.957); P=0.011], thus, ORs and 95% CIs calculated by logistic regression were adjusted for age. Individuals with the -634G/C genotype had an increased risk of breast cancer when compared with the -634G/G genotype (OR, 2.544; 95% CI, 1.852-3.496; P<0.001). Homozygous CC had an increased risk when compared with -634G/G (OR, 1.600; 95% CI, 1.030-2.485; P=0.036; Table III). VEGF polymorphisms in other loci did not demonstrate any increased risk for breast cancer.

Correlation between VEGF genotypes and clinicopathological parameters. Table IV shows known clinicopathological parameters and demographic factors of the breast cancer patients. Numerous patients received surgery (mastectomy in 385 patients and wide excision in 91 patients). The patients with invasive carcinoma who underwent wide excision received radiotherapy. VEGF -634G/C genotype was associated with tumor size >20 mm (OR, 1.638; 95% CI, 1.103-2.434; P=0.015), perineural invasion (OR, 2.261; 95% CI, 1.217-4.202; P=0.010) and stage II-IV at diagnosis (OR, 1.915; 95% CI, 1.255-2.944; P=0.003). Separate analysis of invasive ductal carcinoma revealed a marked association with tumor size >20 mm (OR, 1.722; 95% CI, 1.097-2.703; P=0.018), perineural invasion (OR, 2.36; 95% CI, 1.227-4.539; P=0.010) and stage II-IV at diagnosis (OR, 2.078; 95% CI, 1.237-3.490; P=0.006). The VEGF -1498C/C genotype correlated with decreased

Characteristics	Breast cancer patients, n (%)
Age at diagnosis, years	
≤40	68 (14.08)
40-49	178 (36.85)
50-59	137 (28.36)
>60	100 (20.70)
Tumor type	
Ductal carcinoma in situ	36 (7.45)
Invasive ductal carcinoma	396 (81.99)
Invasive lobular carcinoma	12 (2.48)
Others	39 (8.07)
Tumor size, mm	
In situ	30 (6.21)
≤20	159 (32.92)
>20-50	250 (51.76)
>50	44 (9.11)
Axillary nodal metastasis	
No	271 (56.11)
Yes	205 (42.44)
Unknown	7 (1.45)
Distant metastasis	
No	466 (96.48)
Yes	17 (3.52)
Stage at diagnosis	
0	28 (5.80)
Ι	115 (23.81)
II	214 (44.31)
III	109 (22.57)
IV	17 (3.52)
Histological grading	
Well-differentiated	36 (7.45)
Moderately differentiated	230 (47.62)
Poorly differentiated	146 (30.23)
Unknown	71 (14.70)
ER	
Negative	191 (39.54)
Positive	261 (54.04)
Unknown	31 (6.42)
PR	
Negative	236 (48.86)
Positive	210 (43.48)
Unknown	37 (7.66)
Surgery	
Yes	476 (98.55)
No	3 (0.62)
Unknown	4 (0.83)
Chemotherapy	
Yes	308 (63.77)
No	166 (34.37)
Unknown	9 (1.86)

Table IV. Clinicopathological parameters and demographic factors of breast cancer.

Table IV Continued.

Characteristics	Breast cancer patients, n (%)
Radiotherapy	
Yes	198 (40.99)
No	272 (56.31)
Unknown	13 (2.69)

ER, estrogen receptors; PR, progesterone receptor.

risk of lymphovascular invasion (LVI; OR, 0.308; 95% CI, 0.102-0.927; P=0.036).

Haplotype analysis. The -2578C/-1498T/-1154G/-634G haplotype was the most common haplotype in the two groups (frequency, 0.3778 and 0.4739, respectively). Permutation testing revealed a significant difference between haplotype frequencies in the breast cancer and control groups (P=0.01). CTGG and CTGC haplotype copy number distributions were significantly different between the groups. Bearing 1 or 2 copies of the CTGG haplotype had a protective effect against breast cancer (OR, 0.55; 95% CI, 0.42-0.73; P<0.001). By contrast, bearing 1 or 2 copies of the CTGC haplotype was found to have an increased risk for breast cancer (OR, 1.81; 95% CI, 1.39-2.35; P<0.001). Patients with 1 or 2 copies of the CTGC haplotype significantly correlated with a tumor size >20 mm, (OR, 1.60; 95% CI, 1.09-2.35; P=0.0126), presence of perinural invasion (PNI; OR, 1.84; 95% CI, 0.97-3.52; P=0.046) and stage II-IV (OR, 1.73; 95% CI, 1.14-2.61; P=0.0062). Patients with 1 or 2 copies of ACAG haplotype exhibited a reduced risk for LVI and poorly differentiated cell types (OR, 0.74; 95% CI, 0.48-1.15; P=0.1656 and OR, 0.74; 95% CI, 0.46-1.17; P=0.1768, respectively).

Survival analysis. The median follow-up time was 65 months (range, 1.25-136.7 months). Among 446 patients, there were 37 mortalities during the study period, 30 of which were breast cancer-related. Locoregional recurrence was observed in 29 patients. Distant metastasis occurred in 86 patients. The Kaplan-Meier survival curves of various VEGF genotypes are shown in Fig. 1. Patients with the -634CC genotype (5 mortalities of 57 patients) succumbed to breast cancer in the first 21 months following breast cancer diagnosis. Univariate analysis between clinicopathological parameters, VEGF genotypes and corresponding 5-year survival are provided in Table V. Age >50 years correlated with lower 5-year OS. Larger tumor size, LVI, PNI, estrogen receptor- and progesterone receptor-negative, regional lymph-node positive, distant metastasis and advanced staging were the major prognostic factors for OS and DFS. No statistically significant correlation was found between VEGF genotypes and 5-year survival in the univariate analysis. Factors with a P-value ≤0.2 were included in the Cox regression analysis and poor differentiation, presence of PNI, PR-negative, axillary nodal metastasis and having the -634C/C and -1154G/A genotypes were found to significantly correlate with increased hazard ratio (HR) for DFS. HRs were 3.050 (95% CI, 1.354-6.871; P=0.007) and 2.452 (95% CI, 1.384-4.343; P=0.002) for the -634C/C and -1154G/A genotypes, respectively.

Table V. DFS an	d OS by	 clinicopathologica 	l parameters and VEGF	polymorphisms.
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ParametersCases, nEvent5 year survival, %P-valueCases, nEvent5 year survival, %P-valueAge at diagnosis, years ≤ 50 2505079.700.8632531295.020.047>501974078.462051990.891047Imor size, mmnnn1293.13<0.001180497.660.001>202747870.032852790.1811411141Histological gradingWell-/moderately differentiated2454481.480.0142531594.930.158Poorty differentiated1243870.2311371388.4511595.220.047Present2773984.80<0.0012841595.220.047Present1474569.311541589.1515Present3225183.79<0.0013431595.800.003Regative1794747.4481832088.190.007Present19474.4881.32088.1990.072282289.38Present1942482.240.001204797.390.007Negative2225972.062282289.3888.19PaPositive1962587.31<0.001262797.41<0.001 <th></th> <th></th> <th></th> <th>DFS</th> <th></th> <th colspan="3">OS</th> <th></th>				DFS		OS			
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Parameters	Cases, n	Event	5-year survival, %	P-value	Cases, n	Event	5-year survival, %	P-value
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	Age at diagnosis, years								
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	≤50	250	50	79.70	0.863	253	12	95.02	0.047
Tumor size, mm In situ and $\pounds 20$ 177 12 93.13 <0.001 180 4 97.66 0.001 >20 274 78 70.03 285 27 90.18 0.001 Histological grading	>50	197	40	78.46		205	19	90.89	
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Tumor size, mm								
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	In situ and ≤ 20	177	12	93.13	< 0.001	180	4	97.66	0.001
Histological gradingVell-'moderately differentiated2454481.480.0142531594.930.158Poorly differentiated1343870.231371388.450.158LVI	>20	274	78	70.03		285	27	90.18	
Well-imoderately differentiated 245 44 81.48 0.014 253 15 94.93 0.158 Poorly differentiated 134 38 70.23 137 13 88.45 LVI Absent 277 39 84.80 <0.001	Histological grading								
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Well-/moderately differentiated	245	44	81.48	0.014	253	15	94.93	0.158
LVT Absent 277 39 84.80 <0.001 284 15 95.22 0.047 Present 147 45 69.31 154 15 89.15 $^{\circ}$ PNI Absent 332 51 83.79 <0.001 343 15 95.80 0.003 Present 49 20 55.11 52 7 85.92 $^{\circ}$ PR PR PR PR PR PR Positive 244 42 82.24 0.032 254 10 96.36 0.003 Negative 179 44 74.48 183 20 88.19 $^{\circ}$ PR PR PR PR Positive 222 59 72.06 228 22 89.38 $^{\circ}$ Regional nodal metastasis Regional nodal metastasis No 258 25 89.24 <0.001 262 7 97.41 <0.001 Yes 187 64 65.05 197 23 87.48 $^{\circ}$ Distant metastasis No N/A N/A N/A N/A 449 28 93.72 0.007 Yes N/A N/A N/A N/A 449 28 93.72 0.007 Yes N/A N/A N/A N/A 449 28 93.72 0.007 Yes N/A N/A N/A N/A 449 28 93.72 0.001 Staging 0-1 134 8 94.06 <0.001 133 1 99.25 0.001 ILV 312 82 72.73 324 30 90.55 634G/C GG 209 35 82.87 0.088 215 12 94.18 0.490 GC 187 40 77.54 193 14 92.98 CC 55 15 71.14 57 5 90.29 -1154G/A GG 297 52 81.23 0.166 304 18 93.99 0.627 GG 297 52 81.23 0.166 304 18 93.99 0.52 CC 25 4 87.50 26 1 94.74 -2578C/A CC 223 39 81.82 0.207 229 13 93.67 0.162	Poorly differentiated	134	38	70.23		137	13	88.45	
Absent2773984.80<0.0012841595.220.047Present1474569.311541589.15PNI	INI								
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Absent	277	39	84.80	< 0.001	284	15	95.22	0.047
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Present	147	45	69.31		154	15	89.15	
Absent33251 83.79 <0.001 343 15 95.80 0.003 Present4920 55.11 52 7 85.92 ERPositive 244 42 82.24 0.032 254 10 96.36 0.003 Negative17944 74.48 18320 88.19 PRRegional nodal metastasisNo25825 89.24 <0.001 262 No25825 89.24 <0.001 262 No25825 89.24 <0.001 262 NoN/AN/AN/AN/AN/AStagingO-1134 8 94.06 <0.001 1331GG20935 82.87 0.0882151294.180.490GC18740GG20935 82.87 0.0882151294.180.490GC18740 </td <td>PNI</td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td>	PNI								
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Absent	332	51	83.79	< 0.001	343	15	95.80	0.003
ER No 244 42 82.24 0.032 254 10 96.36 0.033 Negative 179 44 74.48 183 20 88.19 0.033 PR	Present	49	20	55.11		52	7	85.92	
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	ER								
Negative1794474.481832088.19PRPR2587.31<0.001	Positive	244	42	82.24	0.032	254	10	96.36	0.003
P Positive 196 25 87.31 <0.001 204 7 97.39 0.007 Negative 222 59 72.06 228 22 89.38 Regional nodal metastasis No 258 25 89.24 <0.001 262 7 97.41 <0.001 Yes 187 64 65.05 197 23 87.48 Distant metastasis No N/A N/A N/A N/A 16 3 74.29 Staging 0-I 134 8 94.06 <0.001 133 1 99.25 0.001 II-IV 312 82 72.73 324 30 90.55 634G/C GG 209 35 82.87 0.088 215 12 94.18 0.490 GC 187 40 77.54 193 14 92.98 CC 55 15 71.14 57 5 90.29 -1154G/A GG 297 52 81.23 0.166 304 18 93.99 0.627 GA 138 35 74.45 145 12 91.83 AA 16 3 81.25 16 1 92.31 -1498C/T TT 225 38 82.33 0.224 231 13 93.72 0.542 CC 25 4 87.50 26 1 94.74 -1498C/T TT 225 38 82.33 0.224 231 13 93.72 0.542 CC 25 4 87.50 26 1 94.74 -2578C/A CC 223 39 81.82 0.207 229 13 93.67 0.162	Negative	179	44	74.48		183	20	88.19	
Positive 196 25 87.31 <0.001 204 7 97.39 0.007 Negative 222 59 72.06 228 22 89.38 Regional nodal metastasis 225 89.24 <0.001	PR								
Negative 222 59 72.06 228 22 89.38 Regional nodal metastasis No 258 25 89.24 <0.001	Positive	196	25	87.31	< 0.001	204	7	97.39	0.007
Regional nodal metastasis No 258 25 89.24 <0.001 262 7 97.41 <0.001 Yes 187 64 65.05 197 23 87.48 Distant metastasis 93.72 0.007 Yes N/A N/A N/A N/A N/A 0.01 3 74.29 Staging 90.1 133 1 99.25 0.001 II-IV 312 82 72.73 324 30 90.55 634G/C 93.72 0.001 GC 209 35 82.87 0.088 215 12 94.18 0.490 GC 187 40 77.54 193 14 92.98 CC 55 15 71.14 57 5 90.29 -1154G/A 31.8 35 74.45	Negative	222	59	72.06		228	22	89.38	
No2582589.24<0.001262797.41<0.001Yes1876465.051972387.48Distant metastasisNoN/AN/AN/AN/A4492893.720.007YesN/AN/AN/AN/AN/A16374.29Oti134894.06<0.001	Regional nodal metastasis								
Yes 187 64 65.05 197 23 87.48 Distant metastasis No N/A N/A N/A N/A 449 28 93.72 0.007 Yes N/A N/A N/A N/A N/A 16 3 74.29 Staging 0-I 134 8 94.06 <0.001	No	258	25	89.24	< 0.001	262	7	97.41	< 0.001
Distant metastasis No N/A N/A N/A N/A N/A A 449 28 93.72 0.007 Yes N/A N/A N/A N/A N/A N/A 16 3 74.29 Staging	Yes	187	64	65.05		197	23	87.48	
No N/A N/A N/A N/A N/A N/A N/A N/A N/A 16 3 74.29 Staging	Distant metastasis								
YesN/AN/AN/AN/A16374.29Staging0-I134894.06 <0.001 133199.250.001II-IV3128272.733243090.55634G/C62093582.870.0882151294.180.490GC1874077.541931492.980.627CC551571.1457590.29-1154G/A62975281.230.1663041893.990.627GA1383574.451451291.830.62764192.3114-1498C/T72253882.330.2242311393.720.5420.542CT2014874.662081792.520.5422570.26194.74-2578C/A72233981.820.2072291393.670.162	No	N/A	N/A	N/A	N/A	449	28	93.72	0.007
Staging 0.1 134 8 94.06 <0.001 133 1 99.25 0.001 II-IV 312 82 72.73 324 30 90.55 0.001 634G/C	Yes	N/A	N/A	N/A	N/A	16	3	74.29	
0-I 134 8 94.06 <0.001	Staging								
II-IV 312 82 72.73 324 30 90.55 634G/C GG 209 35 82.87 0.088 215 12 94.18 0.490 GC 187 40 77.54 193 14 92.98 CC 55 15 71.14 57 5 90.29 -1154G/A GG 297 52 81.23 0.166 304 18 93.99 0.627 GA 138 35 74.45 145 12 91.83 0.490 -1498C/T TT 225 38 82.33 0.224 231 13 93.72 0.542 CT 201 48 74.66 208 17 92.52 0.542 CT 201 48 74.66 208 17 92.52 0.542 CC 25 4 87.50 26 1 94.74 -25578C/A CC 223 39 81.82 0.207 29 13 93.67 0.162	0-I	134	8	94.06	< 0.001	133	1	99.25	0.001
634G/C 209 35 82.87 0.088 215 12 94.18 0.490 GC 187 40 77.54 193 14 92.98 CC 55 15 71.14 57 5 90.29 -1154G/A - - - - - - GA 297 52 81.23 0.166 304 18 93.99 0.627 GA 138 35 74.45 145 12 91.83 AA 16 3 81.25 16 1 92.31 -1498C/T TT 225 38 82.33 0.224 231 13 93.72 0.542 CT 201 48 74.66 208 17 92.52 0.542 CT 201 48 74.66 208 17 92.52 0.542 CC 25 4 87.50 26 1 94.74 -2578C/A CC 223 39 81.82 0.207 229	II-IV	312	82	72.73		324	30	90.55	
GG 209 35 82.87 0.088 215 12 94.18 0.490 GC 187 40 77.54 193 14 92.98 CC 55 15 71.14 57 5 90.29 -1154G/A GG 297 52 81.23 0.166 304 18 93.99 0.627 GA 138 35 74.45 145 12 91.83 AA 16 3 81.25 16 1 92.31 -1498C/T T 225 38 82.33 0.224 231 13 93.72 0.542 CT 201 48 74.66 208 17 92.52 CC 25 4 87.50 26 1 94.74 -2578C/A CC 223 39 81.82 0.207 229 13 93.67 0.162	634G/C								
GC1874077.541931492.98CC551571.1457590.29-1154G/AGG2975281.230.1663041893.990.627GA1383574.451451291.83AA16381.2516192.31-1498C/T772253882.330.2242311393.720.542CT2014874.662081792.520.54225487.5026194.74-2578C/ACC2233981.820.2072291393.670.162	GG	209	35	82.87	0.088	215	12	94.18	0.490
CC 55 15 71.14 57 5 90.29 -1154G/A GG 297 52 81.23 0.166 304 18 93.99 0.627 GA 138 35 74.45 145 12 91.83 AA 16 3 81.25 16 1 92.31 -1498C/T TT 225 38 82.33 0.224 231 13 93.72 0.542 CT 201 48 74.66 208 17 92.52 0.542 CC 25 4 87.50 26 1 94.74 -2578C/A CC 223 39 81.82 0.207 229 13 93.67 0.162	GC	187	40	77.54		193	14	92.98	
-1154G/A GG 297 52 81.23 0.166 304 18 93.99 0.627 GA 138 35 74.45 145 12 91.83 AA 16 3 81.25 16 1 92.31 -1498C/T TT 225 38 82.33 0.224 231 13 93.72 0.542 CT 201 48 74.66 208 17 92.52 CC 25 4 87.50 26 1 94.74 -2578C/A CC 223 39 81.82 0.207 229 13 93.67 0.162	CC	55	15	71.14		57	5	90.29	
GG2975281.230.1663041893.990.627GA1383574.451451291.83AA16381.2516192.31-1498C/T712253882.330.2242311393.720.542CT2014874.662081792.520.5420.20726194.74-2578C/ACC2233981.820.2072291393.670.162	-1154G/A								
GA 138 35 74.45 145 12 91.83 AA 16 3 81.25 16 1 92.31 -1498C/T TT 225 38 82.33 0.224 231 13 93.72 0.542 CT 201 48 74.66 208 17 92.52 CC 25 4 87.50 26 1 94.74 -2578C/A CC 223 39 81.82 0.207 229 13 93.67 0.162	GG	297	52	81.23	0.166	304	18	93.99	0.627
AA16381.2516192.31-1498C/TTT2253882.330.2242311393.720.542CT2014874.662081792.52CC25487.5026194.74-2578C/ACC2233981.820.2072291393.670.162	GA	138	35	74.45		145	12	91.83	
-1498C/T TT 225 38 82.33 0.224 231 13 93.72 0.542 CT 201 48 74.66 208 17 92.52 CC 25 4 87.50 26 1 94.74 -2578C/A CC 223 39 81.82 0.207 229 13 93.67 0.162	AA	16	3	81.25		16	1	92.31	
TT 225 38 82.33 0.224 231 13 93.72 0.542 CT 201 48 74.66 208 17 92.52 CC 25 4 87.50 26 1 94.74 -2578C/A CC 223 39 81.82 0.207 229 13 93.67 0.162	-1498C/T								
CT 201 48 74.66 208 17 92.52 CC 25 4 87.50 26 1 94.74 -2578C/A -223 39 81.82 0.207 229 13 93.67 0.162	TT	225	38	82.33	0.224	231	13	93.72	0.542
CC 25 4 87.50 26 1 94.74 -2578C/A 223 39 81.82 0.207 229 13 93.67 0.162	СТ	201	48	74.66		208	17	92.52	
-2578C/A CC 223 39 81.82 0.207 229 13 93.67 0.162	CC	25	4	87.50		26	1	94.74	
CC 223 39 81.82 0.207 229 13 93.67 0.162	-2578C/A								
	CC	223	39	81.82	0.207	229	13	93.67	0.162
AC 199 47 74.82 206 18 91.73	AC	199	47	74.82		206	18	91.73	
AA 29 4 88.82 30 0 100.00	AA	29	4	88.82		30	0	100.00	

Proportion of survival obtained from Kaplan-Meier analysis. *VEGF*, vascular endothelial growth factor; DFS, disease-free survival; OS, overall survival; ER, estrogen receptors; PR, progesterone receptors; LVI, lymphovascular invasion; PNI, perinural invasion.

Table VI. Characteristics of 124 breast cancer patients included in *VEGF* mRNA evaluation.

Characteristics	Patients, n (%)
Age at diagnosis, years	
≤50	66 (53.23)
>50	58 (47.77)
Tumor type	
Ductal carcinoma in situ	2 (1.61)
Invasive ductal carcinoma	108 (87.10)
Invasive lobular carcinoma	4 (3.23)
Others	9 (7.26)
Tumor size, mm	
In situ	2 (1.61)
≤20	28 (22.58)
>20-50	68 (54.84)
>50	26 (20.97)
Axillary nodal metastasis	
No	59 (47.58)
Yes	65 (52.42)
Distant metastasis	
No	117 (94.35)
Yes	7 (5.65)
Stage at diagnosis	
0	2 (1.61)
Ι	20 (16.13)
II	61 (49.19)
III	34 (27.42)
IV	7 (5.65)
Histological grading	
Well differentiated	3 (2.61)
Moderately differentiated	69 (60.00)
Poorly differentiated	43 (37.39)
LVI	
Absent	68 (57.14)
Present	51 (42.86)
PNI	
Absent	93 (83.78)
Present	18 (16.22)
ER	
Negative	60 (48.78)
Positive	63 (51.22)
PR	、/
Negative	70 (56.91)
Positive	53 (43.09)



VEGF mRNA expression in breast cancer tissue. VEGF mRNA expression was evaluated in 124 breast cancer tissues. Characteristics of breast cancer patients are provided



Figure 1. Patient survival following diagnosis by *VEGF* genotypes. (A) Patients with the -634C/C genotype correlated with lower DFS compared with those with the -634G/G genotype. (B) Lower OS rate correlated with -634G/C and CC compared with -634G/G genotype. *VEGF*, vascular endothelial growth factor; DFS, disease-free survival; OS, overall survival.



Figure 2. *VEGF* mRNA expression in patients with -634G/C polymorphisms, presented as a ratio to β -actin mRNA levels. One-way analysis of variance revealed that the three genotypes exhibited significantly different *VEGF* mRNA expression levels (P<0.001). Patients with the -634C/C genotype exhibited a significantly higher *VEGF* mRNA expression when compared with -634G/G and -634G/C genotype (Scheffe post test; P<0.001 and 0.001, respectively). The number of patients was 57, 55 and 12 for the -634GG, GC and CC genotypes, respectively. *P<0.001, vs. -634G/G; **P=0.001, vs. -634G/C. *VEGF*, vascular endothelial growth factor.

in Table VI. Expression ranged between 0 and 3.27 with a median of 1.10. Patients with the VEGF -634C/C genotype had significantly higher VEGF mRNA in breast cancer tissue compared with those with the -634G/G or -634G/C genotype (Fig. 2). Patients with heterozygous -1154G/A, -1498C/T and -2578A/C exhibited lower VEGF mRNA when compared with homozygous -1154G/G, -1498T/T and -2578C/C. Due to the presence of outliers and a small number of patients with the homozygous -1154A/A, -1498C/C and -2578A/A genotypes,

Table VII.	Correlation betwee	n VEGF mRNA ex	pression and cl	inicopatholog	gical parameters.

	VEGF mRN	A expression		
Characteristics	Low	High	OR (95% CI)	P-value
Age, years				
≤50	36 (58.06)	30 (48.39)		
>50	26 (41.94)	32 (51.61)	1.477 (0.727-3.001)	0.281
Tumor size, mm				
≤20	20 (32.26)	10 (16.13)		
>20	42 (67.74)	52 (83.87)	2.476 (1.047-5.858)	0.039
Axillary nodal				
metastasis				
No	36 (58.06)	23 (37.70)		
Yes	26 (41.94)	38 (62.30)	2.288 (1.110-4.713)	0.025
Distant				
metastasis				
No	60 (96.77)	56 (91.80)		
Yes	2 (3.23)	5 (8.20)	2.679 (0.499-14.369)	0.250
Staging				
0-II	46 (74.19)	37 (59.68)		
III-IV	16 (25.81)	25 (40.32)	1.943 (0.906-4.163)	0.088
Histological				
grading				
Well-/moderately				
differentiated	38 (65.52)	34 (59.65)		
Poorly				
differentiated	20 (34.48)	23 (40.35)	1.285 (0.603-2.740)	0.516
LVI				
Absent	40 (67.80)	28 (46.67)		
Present	19 (32.20)	32 (53.33)	2.406 (1.142-5.070)	0.021
PNI				
Absent	48 (87.27)	45 (80.36)		
Present	7 (12.73)	11 (19.64)	1.676 (0.598-4.701)	0.326
ER				
Positive	36 (58.06)	27 (44.26)		
Negative	26 (41.94)	34 (55.74)	0.574 (0.281-1.171)	0.127
PR				
Positive	31 (50.00)	22 (36.07)		
Negative	31 (50.00)	39 (63.93)	0.564 (0.274-1.161)	0.120
Hormone receptor	· · · ·		· / /	
Negative	39 (62.90)	30 (49.18)		
Positive	23 (37.10)	31 (50.82)	0.571 (0.278-1.172)	0.126

VEGF, vascular endothelial growth factor; OR, odds ratio; ER, estrogen receptor; PR, progesterone receptor; LVI, lymphovascular invasion; PNI, perinural invasion; CI, confidence interval.

VEGF mRNA expression in these groups appeared to be high. Following exclusion of the outlier, *VEGF* mRNA expression was decreased in patients with homozygous -1154A/A, -1498C/C and -2578A/A (data not shown).

Correlation between VEGF mRNA levels and clinicopathological parameters. Breast cancer patients were classified into low and high expression groups using a median value of 1.10. Patient distribution in each group and clinicopathological parameters are provided in Table VII. Elevated *VEGF* expression correlated with a tumor size >20 mm (OR, 2.476; 95% CI, 1.047-5.858; P=0.039), axillary nodal metastasis (OR, 2.288; 95% CI, 1.110-4.713; P=0.025) and presence of LVI (OR, 2.406; 95% CI, 1.142-5.070; P=0.021).

Discussion

In the current study, it was observed that alteration of nucleotides from G to C at -634, resulted in an increased risk of breast cancer. However, previous studies have not reported this correlation in breast cancer (15-20). In the present study, the -634G/C genotype was significantly associated with more aggressive features. Due to a limited number of patients with the -634C/C genotype, the difference was not observed to be statistically significant. This was consistent with a previous study by Balasubramanian et al and Jin et al which reported that an alteration of G to C at this position was associated with a larger tumor size and high grade breast cancer (16,18). By contrast, Langsenlehner et al observed a significant correlation between the -634G/C and -634C/C genotypes and smaller tumor size (19). Survival analysis revealed a significant correlation between the -634C/C genotype and lower DFS. However, OS of the patients with different -634G/C genotypes was similar, which may be due to the relatively short term follow up of this study. By contrast, the survival analysis of 1,455 Chinese breast cancer patients revealed that patients with the -634G/G genotype had a lower OS compared with those with the -634C/C genotype, however, this polymorphism was not found to correlate with DFS (21). The variance in the demographic results of the cancer population may contribute to discrepancies observed. In the present study, 26.09% of the patients were diagnosed as stage III and IV and 63.77% of the patients received chemotherapy, while in a study by Lu et al, only 11.34% of the patients were diagnosed as stage III and IV and a large number of patients received chemotherapy (93.95%) (21).

In the present study, no correlation was found between -1154G/A polymorphisms and breast cancer risk, consistent with previous studies in Caucasian populations (18,20,22). No correlation between breast cancer aggressiveness and -1154G/A polymorphisms was observed, consistent with a study by Jin et al (18). Breast cancer susceptibility or aggressiveness was not associated with -1498C/T polymorphisms, consistent with previous large case-control studies in Asian and Caucasian populations (15,16). There was no association between -2578A/C polymorphisms and breast cancer susceptibility/aggressiveness as previously observed in Caucasian populations by Jin et al and Langsenlehner et al (18,19). By contrast, two additional studies in Caucasian populations revealed conflicting results. Schneider et al reported an association between the -2578A/A genotype and breast cancer risk while Jacobs et al reported that -2578C was associated with an increased risk of invasive breast cancer (17,20). However, the latter study stated the importance of LD of -2578A/C and -1154G/A. Thus, the association of the two polymorphisms and breast cancer risk may not be individually demonstrated.

Haplotype analysis revealed that the -2578C/-1498T/ -1154G/-634G haplotypes had a protective effect against breast cancer. Patients with the -2578C/-1498T/-1154G/-634C haplotype had an increased risk of breast cancer and were associated with a tumor size >20 mm, stage II-IV and PNI. In a Swedish population, haplotypes -2578C/-634C were significantly associated with a large tumor size and higher grade. Having 2 copy numbers of haplotypes -2578A/-634G was associated with lower tumor grade (18). Jacobs *et al* reported that the -2578A/-1154A/-634G haplotypes correlated with a reduced risk of breast cancer in an American population (17). These observations and the observations of the present study indicate that haplotypes bearing the-634C allele contribute to increased breast cancer risk and aggressiveness. Non-replication of genetic association results is common in genetic epidemiology. In addition, the polymorphisms on the promoter region and 5'UTR were in LD. Interpretation of haplotype analysis revealed that the alleles included in the haplotypes may be associated with other functional polymorphisms that were not assessed. Comparison of haplotype analysis may be complicated due to variation in alleles in the haplotype, different software used to generate the haplotype and determining haplotype frequency.

The VEGF -634CC genotype correlated with elevated levels of mRNA expression. Elevated VEGF mRNA expression correlated with tumor size >20 mm, lymph node involvement and presence of LVI. These observations are consistent with a previous study employing the RT-PCR technique. Gomez-Esquer *et al* demonstrated a correlation between VEGF mRNA expression higher than the 25th percentile and more aggressive features in 103 breast cancer patients (23).

Incorporation of bevacizumab, a humanized monoclonal antibody that targets VEGF in chemotherapy, is a rapidly evolving area in the treatment of breast cancer. An association study of VEGF polymorphisms in 180 advanced breast cancer patients treated with paclitaxel alone or with bevacizumab and 183 untreated patients, revealed that the VEGF -2578A/A genotype was associated with an improved median OS in the combination arm when compared with AC combined with the CC genotype. The VEGF -1154A/A genotype also demonstrated an improved median OS when compared with GG combined with the GA genotype in the combination arm (24). These observations indicate that selection methods to identify the patients suitable for anti-VEGF therapy must be established. -634G/C polymorphisms may identify populations at risk and predict the outcome of breast cancer. It is possible that genotyping of VEGF -634 polymorphisms in breast cancer patients may be used to select appropriate patients for anti-angiogenesis treatment.

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