Predictive value of intrahepatic hepatitis B virus covalently closed circular DNA and total DNA in patients with acute hepatitis B and patients with chronic hepatitis B receiving anti-viral treatment

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Abstract. This study aimed to investigate the persistence and predictive values of intrahepatic (IH) hepatitis B virus (HBV) covalently closed circular DNA (cccDNA) and total DNA (tDNA) in patients with acute hepatitis B (AHB) and patients with chronic hepatitis B (CHB) receiving anti-viral treatment. The levels of IH cccDNA and tDNA, serum HBV DNA, hepatitis B surface antigen (HBsAg), hepatitis B e antigen (HBeAg) and alanine aminotransferase (ALT) were detected in 11 patients with AHB and 46 patients with CHB who were receiving anti-viral treatment, among whom 21 had primary treatment failure, 11 achieved virological response (VR) and 15 achieved VR and HBsAg seroclearance. The median IH cccDNA and tDNA levels in the patients with AHB (0.002 copies/cell and 0.04 copies/cell, respectively) were significantly lower than those in the patients with CHB. In the patients with CHB, the median IH cccDNA level among individuals who achieved VR and HBsAg seroclearance (0.012 copies/cell) was significantly lower than that in those who had failed primary treatment (4.18 copies/cell, P<0.0001) but not that in those who achieved solely VR (0.039 copies/cell, P=0.169). The median IH tDNA level in patients with CHB who achevied VR and HBsAg seroclearance (0.096 copies/cell) was significantly lower than that in those who failed primary treatment (371 copies/cell, P<0.0001) and those who achieved solely VR (1.62 copies/cell, P=0.001). No significant difference

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was observed in the area under the receiver operating characteristic (ROC) curve, which was used to predict the likelihood of achieving VR and HBsAg seroclearance, between IH tDNA and IH cccDNA levels (0.96 and 0.88, respectively; P>0.10). IH cccDNA levels were shown to be positively correlated with serum ALT (P=0.024), HBeAg (P=0.001) and IH tDNA levels (P<0.0001), but not with serum HBV DNA (P=0.12) and HBsAg levels in either HBeAg-positive (P=0.84) or in HBeAg-negative (P=0.146) patients. In conclusion, IH cccDNA may persist in patients with AHB and patients with CHB who acheive VR and HBsAg seroclearance following anti-viral treatment. Furthermore, IH cccDNA and tDNA may have potential in predicting successful therapeutic response in patients with CHB who receive anti-viral treatment.

Introduction

Hepatitis B virus (HBV) infection is a predominant global health concern with >350 million individuals chronically infected worldwide. Approximately one million mortalities occur annually due to the long-term complications associated with HBV infection (1). Following infection of hepatoctyes, HBV relaxed-circle DNA (rcDNA) is transferred to the nucleus where it forms covalently closed circular DNA (cccDNA), which represents the intracellular template of the virus. Within infected cells, the pregenomic RNA (pgRNA) is then transcribed from the cccDNA and transported to the cytoplasm, where the mature capsids of the rcDNA are reverse transcribed and either secreted from the cells or returned to the nucleus to form the cccDNA pool. While HBV is a non-cytopathic virus, the immune response against virus-infected liver cells and the production of inflammatory cytokines have been proposed to be responsible for liver disease and viral clearance (2).

HBV infections are predominantly divided into either self-limited acute hepatitis B (AHB), inactive hepatitis B surface antigen (HBsAg) carriers, chronic hepatitis B (CHB) and occult HBV infection (OBI) (3). AHB is characterized by transient liver inflammation and virusemia. CHB consists of four phases: Immune-tolerant (IT), immune-clearance

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(IC), non/low-replicative (LR) and hepatitis B e antigen (HBeAg)-negative hepatitis (ENH) (4). Patients are classified as having the OBI form of the disease if they have undetectable HBsAg, but detectable HBV DNA in the sera or liver tissues (5). Anti-viral drug treatment for patients with CHB predominantly involves interferon- α (INF- α) and nucleoside analogues (NUC), including lamivudine, adefovir and entecavir. However, such drugs are not capable of directly eradicating cccDNA, which has been reported to have a significant role in HBV infection relapse (6), the pathogenesis of hepatocellular carcinoma and cirrhosis (5), and the infection of liver transplant cases (7). Monitoring intrahepatic (IH) cccDNA levels in patients with CHB may have potential as a predictor of therapeutic response or disease progression (8). However, at present, IH HBV cccDNA and total DNA (tDNA) persistence, and the correlation between IH cccDNA and other biochemical, virological and serological parameters in patients with AHB and those with CHB who are receiving anti-viral treatment have yet to be elucidated.

This study aimed to investigate the persistence of IH cccDNA and tDNA and their correlation in patients with AHB and those with CHB receiving anti-viral treatment. Furthermore, the present study aimed to obtain and compare the thresholds of IH cccDNA and tDNA levels in order to assess successful therapeutic outcome for patients with CHB, which may be beneficial in determining an anti-viral treatment endpoint.

Patients and methods

Patients and samples. Liver biopsy specimens were collected from a total of 60 patients, including 11 patients with AHB who achieved spontaneous virological seroclearance, 46 patients with CHB in phase III antiviral clinical trials receiving NUC (adefovir, 10 mg/day or lamivudine, 100 mg/day), either as a monotherapy or with pegylated-INF- α (100 µg/week) and three patients with autoimmune hepatitis (AIH) as negative controls. Patients who were coinfected with hepatitis D, hepatitis C or human immunodeficiency virus, or those with Wilson's disease, primary biliary cirrhosis or with a substantial daily alcohol intake (20 g/day for females; 30 g/day for males) were excluded from the study. Among the patients with CHB, 21 had failed primary treatment, 11 had achieved virological response (VR) and 14 had achieved VR and HBsAg seroclearance and had been relapse-free for >6 months. Each patient signed an informed consent document approved by the Ethics Committee of Shenzhen Third People's Hospital (Shenzhen, China). Biopsy specimens were frozen in liquid nitrogen and stored at -80°C until experimental analysis.

IH HBV cccDNA quantification. DNA was extracted from biopsy specimens using the QIAamp[®] DNA Mini kit (Qiagen, Hilden, Germany). To enhance the specificity of cccDNA detection, the Plasmid-Safe[™] ATP-Dependent DNase (PSAD; Epicentre Biotechnologies, Madison, WI, USA) was used to degrade rcDNA and single-stranded DNA (ssDNA) prior to quantitative polymerase chain reaction (qPCR) analysis, according to the manufacturer's instructions (9). IH cccDNA levels were measured using qPCR analysis as described previously (9). Two forward primers, [CCC1, 5'-GCGGWCTCC CCGTCTGTGCC-3'; diaphanous-related formin 1 (DRF1), 5'-GTCTGTGCCTTCTCATCTGC-3'], and one reverse primer, (CCC2, 5'-GTC ATGCCCCAAAGCCACC-3'), were used for cccDNA amplification using the LightCycler[®] system (Roche Diagnostics, Mannheim, Germany) and SYBR Premix Ex Taq (Takara Bio Inc., Otsu, Japan). Serial dilutions of a plasmid containing an HBV monomer (pHBVEcoRI) served as a quantification standard. β -globin DNA (house keeping gene) was detected using the LightCycler[®] Control kit DNA (Roche Diagnostics) in order to count the cell number in the biopsies and calculate copies/cell.

Serum and IH HBV tDNA quantification. Blood was collected on the day of the liver biopsy and serum was immediately stored at -20°C. DNA was extracted from 200 μ l serum using the QIAamp[®] DNA Blood Mini kit (Qiagen). Prior to degradation using PSAD, serum and IH tDNA levels were measured using the Cobas[®] TaqMan[®] test as described previously (Roche Diagnostics) (10).

Serum HBsAg and HBeAg quantification. Serum HBsAg and HBeAg levels were quantified by enzyme immunoassay using the Abbott ARCHITECT platform (Abbott Laboratories, Chicago, IL, USA) according to the manufacturer's instructions. HBsAg >0.05 IU/ml and HBeAg >1 IU/ml were considered to be positive results.

Serum alanine aminotransferase (ALT) quantification. Serum ALT levels were measured using blood chemistry analyses (DiaSys Diagnostic Systems, Holzheim, Germany), according to the manufacturer's instructions.

Statistical analysis. Statistical analyses were performed using the SPSS 13.0 statistical software (SPSS Inc., Chicago, IL, USA). Continuous variables are expressed as the mean with the range, and were analyzed using the Mann-Whitney rank sum test or one-way analysis of variance. Serum HBsAg and HBeAg (IU/ml) levels were logarithmically transformed prior to analysis. Categorical variables were compared using Pearson's χ^2 test. Correlations were analyzed using Pearson's correlation coefficient. Area under the receiver operating characteristic (ROC) curve analysis was performed to predict the likelihood of achieving VR in combination with HBsAg seroclearance in the patients with CHB. Two-sided P<0.05 was considered to indicate a statistically significant difference.

Results

Baseline characteristics. The clinical, biochemical and serological characteristics of the patient groups used in this study are shown in Table I. Among the 11 patients with AHB, all patients were HBeAg-negative (HBeAg), and one patient was also anti-hepatitis B core antibody-positive (anti-HBcAb⁺). Among the patients with CHB who had failed primary treatment, three patients were HBeAg⁺ and anti-HBcAb⁺, and the remaining 18 patients were HBeAg⁺ and anti-HBcAb⁺. In the group of patients with CHB who had achieved VR, six patients were HBeAg⁻, of whom two patients were anti-HBcAb⁺, and five patients were HBeAg⁺ and anti-HBcAb⁺. In the group

	Patient groups						
	CHB receiving anti-viral treatment						
Parameters	AHB	1	2	3	P-value		
Age (years)	28.18 (21-41)	32.86 (22-42)	37.18 (27-73)	36.93 (21-55)	0.062		
HBeAg ⁺ /HBeAg	0/11	18/3	5/6	3/11	< 0.0001		
Gender (M/F)	10/1	19/2	9/2	11/3	0.713		
ALT (IU/ml)	34.27 (12-83)	76.86 (29-262)	36.27 (14-102)	25.57 (15-36)	0.001		
HBeAg (log ₁₀ IU/ml)	-0.42 (-1-0.009)	1.42 (-1-3.10)	0.59 (-0.77-2.49)	-0.44 (-2-0.74)	< 0.0001		
HBsAg (log ₁₀ IU/ml)	0.61 (-0.19-3.45)	3.01 (1.49-4.76)	3.41 (1.54-5.06)	-1.43 (-2.52-0.10)	< 0.0001		
HBV DNA (copies/ml)	0	$1.33 \times 10^{7} (2.32 \times 10^{4} - 9.07 \times 10^{7})$	0	0	0.120		

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1, Patients with CHB who failed primary treatment; 2, patients with CHB who achieved VR; and 3, patients with CHB who achieved VR and HBsAg seroclearance. ALT, alanine aminotransferase; HBV, hepatitis B virus; HBsAg, hepatitis B virus surface antigen; HBeAg, hepatitis B e antigen; AHB, acute hepatitis B; CHB, chronic hepatitis B; VR, virological response; M, male; F, female.

of patients with CHB who had achieved VR and HBsAg seroclearance, 11 patients were HBeAg⁻ and anti-HBcAb⁻ and three patients were HBeAg⁺ and anti-HBcAb⁺ (P<0.0001).

IH cccDNA and tDNA quantification. The lower limits of detection for IH cccDNA and tDNA quantification were 0.00024 copies/cell and 0.0001 copies/cell, respectively. IH cccDNA levels were detectable in four patients with AHB, 21 patients with CHB who had failed primary treatment, 10 patients who had achieved VR and 11 patients who had achieved VR and HBsAg seroclearance. IH tDNA levels were detectable in 10 patients with AHB, 21 patients with CHB who had failed primary treatment, 11 patients who had achieved VR, 13 patients who had achieved VR and HBsAg seroclearance and one patient with AIH. The median IH cccDNA level in patients with AHB (0.002 copies/cell; range, <0.00024-0.013) was identified to be significantly lower than that in patients with CHB who had failed primary treatment (4.18 copies/cell; range, 0.078-42.4; P<0.0001) as well as that in patients with CHB who had achieved VR (0.039 copies/cell; range, <0.00024-0.17; P=0.005) but not that in patients with CHB who had achieved VR and HBsAg seroclearance (0.012 copies/cell; range, <0.00024-0.093; P=0.076). The median IH tDNA level in patients with AHB (0.04 copies/cell; range, <0.0001-0.15) was observed to be significantly lower than that in patients with CHB who had failed primary treatment (371 copies/cell; range, 3.94-2,940; P<0.0001), that in patients with CHB who had achieved VR (1.62 copies/cell; range, 0.0039 to 5.50; P=0.001) but not that in patients with CHB who had achieved VR and HBsAg seroclearance (0.096 copies/cell; range, <0.0001-0.38; P=0.443) (Fig. 1).

Among patients with CHB, the median IH cccDNA level in those who had achieved VR and HBsAg seroclearance was observed to be significantly lower than that in those who had failed primary treatment (P<0.0001), but not that in those who had achieved VR (P=0.169). Furthermore, the median IH tDNA level in patients with CHB who had achieved VR and HBsAg seroclearance was detected to be significantly lower than that in patients with CHB who had failed primary treatTable II. Correlation between IH HBV cccDNA and tDNA levels and biochemical, virological and serological parameters.

	IH HBV cccDNA		IH HBV tDNA	
Parameters	P-value	r	P-value	r
Serum ALT	0.024	0.294	0.005	0.369
Serum HBeAg	0.001	0.480	0.015	0.321
Serum HBsAg				
Serum HBeAg ⁺ patients	0.840	0.038	0.247	0.236
Serum HBeAg ⁻ patients	0.146	0.267	0.007	0.554
Serum HBV DNA	0.121	0.210	0.437	0.105

HBV, hepatitis B virus; IH, intrahepatic; cccDNA, covalently closed circular DNA; tDNA, total DNA; VR, virological response; HBsAg, hepatitis B surface antigen; HBeAg, hepatitis B e antigen; ALT, alanine aminotransferase.

ment (P<0.0001) and those who had achieved VR (P=0.001). As shown in Fig. 1, the median IH cccDNA and tDNA levels in patients with CHB who had achieved VR were significantly lower than those who had failed primary treatment (P<0.0001 and P<0.0001, respectively). Furthermore, Fig. 2 shows that the IH tDNA:cccDNA ratio in patients with CHB who had achieved VR and HBsAg seroclearance (20.33; range, 0-79.25) was significantly lower that in those who had failed primary treatment (380.29; range, 1.89-2,753.93; P<0.0001) and those who had achieved VR (155.52; range, 0-643.11; P=0.004). The median IH cccDNA level in the patients who were HBeAg⁻ (0.14 copies/cell; range, < 0.00024-2.08)was significantly lower than in those who were HBeAg⁺ (3.24 copies/cell; range, 0.00042-42.4; P<0.0001). As shown in Fig. 1, the median IH tDNA level (1.38 copies/cell; range, <0.0001-17.2) in the former was significantly lower than that in the latter (299 copies/cell; range, 0.070-2940; P<0.0001).

Correlation analysis. IH cccDNA and tDNA levels were positively correlated with serum ALT (P=0.024 and P=0.0048, respectively) and serum HBeAg levels (P=0.0001 and P=0.015, respectively); however, no correlation was observed



Figure 1. IH cccDNA and tDNA levels in patients with AHB and patients with CHB receiving anti-viral treatment: Group 1, patients with AHB; group 2, patients with CHB who failed primary treatment; group 3, patients with CHB who achieved VR; group 4, patients with CHB who achieved VR and HBsAg seroclearance. **P<0.01; ***P<0.001. AHB, acute hepatitis B; CHB; chronic hepatitis B; IH, intrahepatic; cccDNA, covalently closed circular DNA; tDNA, total DNA; VR, virological response; HBsAg, hepatitis B surface antigen; HBeAg, hepatitis B e antigen.



Figure 2. Ratio between IH tDNA and cccDNA in patients with CHB receiving anti-viral treatment: Group 1, patients with CHB who failed primary treatment; group 2, patients with CHB who achieved VR; group 3, patients with CHB who achieved VR and HBsAg seroclearance.**P<0.01; ***P<0.001. CHB; chronic hepatitis B; IH, intrahepatic; cccDNA, covalently closed circular DNA; tDNA, total DNA; VR, virological response; HBsAg, hepatitis B surface antigen.

with HBV DNA levels (P=0.121 and P=0.437, respectively). IH cccDNA levels were not observed to be correlated with serum HBsAg levels in either patients who were HBeAg⁺ (P=0.84) or in patients who were HBeAg⁻ (P=0.146). As shown in Table II, IH tDNA levels were positively correlated with serum HBsAg levels in patients who were HBeAg⁻ (P=0.007), but not in those who were HBeAg⁺ (P=0.247). IH cccDNA levels were observed to be positively correlated with IH tDNA levels (P<0.0001) but not with the IH tDNA:cccDNA ratio (P=0.701). However, as demonstrated in Fig. 3, IH tDNA

levels were positively correlated with the IH tDNA:cccDNA ratio (P=0.0037).

Prediction for achieving VR in combination with HBsAg seroclearance. Areas under the ROC curve were used to predict the likelihood of achieving VR and HBsAg seroclearance in patients with CHB who received anti-viral treatment. The areas for IH cccDNA and tDNA levels were observed to be 0.88 [95% confidence interval (CI): 0.78-0.98] and 0.96 (95% CI: 0.91-1.00), respectively. As shown in Fig. 4, the cutoff value for IH cccDNA levels was 0.015 copies/cell with a sensitivity of 81.3%, specificity of 85.7%, positive predictive value (PPV) of 85.71% and negative predictive value (NPV) of 88.37%. The cutoff value for IH tDNA levels was 0.23 copies/cell with a sensitivity of 93.8%, specificity of 85.7%, PPV of 85.71% and NPV of 95.35% (P>0.10).

Discussion

Approximately two billion individuals are exposed to HBV worldwide, of whom >350 million are chronically infected. It is well established that cellular immune responses, particularly T cells, contribute to HBV clearance. CD4⁺T cells are classified into type 1 and type 2 T helper cells (Th1 and Th2, respectively), which differ in their patterns of secreted cytokines (11). Host immune reactions that are biased towards Th1 have been associated with AHB, with Th1-type cytokines, including IFN- γ and tumor necrosis factor- β (TNF- β), potentially controlling HBV replication by activating CD8⁺T-cell, natural killer (NK)



Figure 3. Correlation analysis between (A) IH cccDNA and tDNA levels; (B) IH cccDNA levels and the IH tDNA:cccDNA ratio and (C) IH tDNA levels and the IH tDNA:cccDNA ratio. IH, intrahepatic; cccDNA, covalently closed circular DNA; tDNA, total DNA.

cell and macrophage responses, consequently stimulating a cascade of inflammatory cytokines and degrading HBV RNA directly (12). Inefficiencies in the innate and adaptive immune responses may result in CHB, in which high levels of transforming growth factor- β 1, predominantly secreted by regulatory T cells, may impair NK cell function by reducing natural killer group 2 member D/DNAX activation protein 10 and natural killer cell receptor 2B4/SLAM-associated protein expression, and suppress innate antiviral immunity by blocking the cell cycle in G1 (13) as well as inhibit the secretion of IFN- γ and TNF- α from HBV-specific T cells (12).

In the present study, the median IH cccDNA level in patients with AHB was observed to be significantly lower than that of



Figure 4. ROC curve of IH cccDNA and tDNA levels to predict the likelihood of achieving VR and HBsAg seroclearance in patients with CHB receiving anti-viral treatment. ROC, receiver operating characteristic; CHB, chronic hepatitis B; IH, intrahepatic; HBV, heptitis B virus; cccDNA, covalently closed circular DNA; tDNA, total DNA.

the patients with CHB who were receiving anti-viral treatment. This finding suggests that in patients with AHB, the immune system may be capable of reducing IH cccDNA to markedly low levels. In patients with CHB, the median IH cccDNA level in those who had achieved VR in combination with HBsAg seroclearance, was observed to be significantly lower than that in those who had failed primary treatment and those that had achieved VR. The outcome of anti-viral treatment in patients with CHB is highly variable, ranging from primary treatment failure to VR, serological response, biochemical response and virological breakthrough (14). While it has been hypothesized that clinical resolution of HBV infection occurs when HBV replication is completely controlled by the host's immune system (15), it has been suggested that achieving VR in combination with HBsAg seroclearance may be the most effective therapeutic response (16). Furthermore, the present study demonstrated that cccDNA production may be attenuated in patients with CHB who achieve VR and HBsAg seroclearance following anti-viral treatment.

To predict the likelihood of achieving VR and HBsAg seroclearance following anti-viral treatment in patients with CHB, the areas under the ROC curves of IH tDNA and cccDNA levels were compared. The area under the ROC curve of IH tDNA levels was observed to be higher than that of IH cccDNA levels (0.96 vs. 0.88); however, the difference was not statistically significant (P>0.10). This may be explained by changes in virion productivity, which was defined as the ratio between IH HBV tDNA and cccDNA levels and reflected the level of cccDNA transcription (17). In the present study, while the IH tDNA:cccDNA ratio was significantly lower in patients with CHB who achieved VR and HBsAg seroclearance, it was revealed to be positively correlated with IH tDNA, but not cccDNA levels. IH cccDNA transcription is tightly controlled by various intrinsic factors, including methylation and acetylation of H3 and H4 histones in cccDNA minichromosomes (18,19) and recruitment of transcription factors, including histone deacetylase 1, sirtuin 1, enhancer of zeste homolog 2 and yin yang 1 (20). Effective anti-viral treatment may be beneficial to

control cccDNA transcription and consequently reduce virion productivity. In addition, it has been reported that amplification of the cccDNA pool may be suppressed by a negative-feedback mechanism involving the viral large surface protein (15), and HBeAg loss may significantly impair virion productivity (21). Such reports are verified by the present study, in which the proportion of HBeAg⁻ patients was observed to be significantly higher in the group of patients with CHB who had achieved VR and HBsAg seroclearance. A previous report indicated that in patients with CHB who had achieved therapeutic response with adefovir, the reduction rate of IH tDNA levels was significantly higher than that of cccDNA levels (22). However, it has also been reported that IH tDNA levels may be as sensitive as IH cccDNA levels for predicting the likelihood of achieving sustained virological response (SVR) (8), although achieving VR in combination with HBsAg seroclearance is likely to be more effective than achieving SVR in anti-viral treatment. The findings of the present study reveal a potentially novel therapeutic strategy for suppressing virion productivity through reducing the size of the cccDNA pool rather than through eradicating IH cccDNA completely.

IH cccDNA was not detected in the negative controls employed in the present study, however tDNA persistence was detected in one patient, being diagnosed as OBI. This finding may be attributed to the complex composition of IH tDNA, which aside from rcDNA, consists of linear DNA, including double linear DNA (dIDNA), as well as single-stranded DNA (ssDNA). Certain linear DNA molecules, produced through illegitimate replication and deficient HBV transcription (23), may be present in hepatocytes but not secreted into the blood (24). Alternatively, IH tDNA may be detected by means of integrated HBV (25). The findings of the present study indicate that IH tDNA detection may be valuable in the diagnosis of OBI in patients with hepatitis without HBV virusemia.

While cryo-preserved liver tissue is not always available clinically, serum HBsAg has been regarded as a surrogate marker for assessing IH cccDNA levels (21). Previous reports observed that serum HBsAg levels were positively correlated with IH cccDNA levels in patients with CHB who were HBeAg⁺, but not in those who were HBeAg⁻ (26,27). However, in the present study, no significant correlation was observed between serum HBsAg and IH cccDNA levels in either HBeAg⁺ or HBeAg⁻ patients. This may be a consequence of HBsAg production, which is complex, abundant and far exceeding that required for virion assembly, being independent of HBV replication (28), or the occurrence of increased HBsAg translation in HBeAg patients as a compensatory mechanism to adapt to the increased immunological pressure (17). Alternatively, HBsAg may be produced from integrated HBV (26), or IH cccDNA copies may be reduced by anti-viral treatment (29).

It was observed in the present study that IH cccDNA levels were positively correlated with serum HBeAg levels and that IH cccDNA and tDNA levels were significantly lower in HBeAg⁻ patients than in HBeAg⁺ patients. HBeAg has been reported to have an important role in HBV persistence, and HBeAg seroconversion has been suggested to increase Th1 cytokine secretion to inhibit HBV replication (12). The findings of the present study are in accordance with a previous study that demonstrated that pre-core mRNA was solely transcribed from the cccDNA template and not the integrated HBV, and that serum HBeAg detection may serve as a cccDNA surrogate in anti-viral screening assays (30).

In the present study, IH cccDNA levels were observed to be positively correlated with IH tDNA but not serum HBV DNA levels. This poor correlation between IH cccDNA and serum HBV DNA levels may be partially attributed to the fact that a number of patients were virusemia negative and indicates that serum HBV DNA levels may not represent an ideal biomarker for evaluating IH HBV production. CHB is a necroinflammatory liver disease and it has been suggested that raised serum ALT levels may predict activation of the host immune system, hepatocyte damage and increased cell proliferation (31). The positive correlation between serum ALT levels and IH cccDNA and tDNA levels observed in the present study indicate that cccDNA replication may have a role in the progression of HBV inflammation.

In conclusion, the present study showed that extremely low levels of IH cccDNA may be present in patients with AHB and patients with CHB who achieve VR and HBsAg seroclearance following anti-viral treatment. IH cccDNA levels were identified to be positively correlated with IH tDNA, serum HBeAg and ALT levels, and not correlated with serum HBV DNA and HBsAg levels. Therefore, IH cccDNA and tDNA have potential value for the prediction of a successful therapeutic response in patients with CHB receiving anti-viral treatment, and the latter may also be beneficial in OBI diagnosis.

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