

Role of α -synuclein in cognitive dysfunction: Studies in *Drosophila melanogaster*

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Abstract. α -Synuclein (α -Syn) is hypothesized to have a critical role in sporadic and genetic cases of Parkinson's disease (PD) in which Lewy bodies, as the hallmark of PD, are formed from abnormal aggregates of α -Syn. To determine the role of α -Syn in the motor and cognitive dysfunction observed in PD, a *Drosophila melanogaster* model was established to investigate the electrophysiological and ethological changes caused by overexpression of α -Syn. The present data indicated that α -Syn overexpression reduced the synaptic transmission of cholinergic neurons by modulating the calcium channel currents in the projection neurons in the antennal lobe region of the *Drosophila* brain, as well as the learning and memory ability of the flies. However, the locomotor ability of the *Drosophila* remained unaffected. The present findings suggested that α -Syn may be associated with senile dementia in patients with PD.

Introduction

Parkinson's disease (PD) is one of the most common degenerative diseases of the nervous system. The major pathological change observed in PD is degeneration and necrosis of the dopaminergic neurons of the substantia nigra, which results in a decrease in dopamine synthesis (1). The low dopamine level disrupts the balance between the dopaminergic and cholinergic activity that is required for normal movement, which accounts for a series of clinical symptoms, including tremor, muscle stiffness and slowness of movement. Genetic and environmental factors contribute to this disease. While the majority of cases of PD are sporadic, certain cases occur as a result of genetic factors and are associated with certain gene mutations, including parkin (2), α -synuclein (α -Syn) and

leucine-rich repeat kinase 2 (3,4). Lewy bodies (LBs) are a common feature of hereditary and idiopathic PD. α -Syn, the main component of LBs, is encoded by SNCA. Point mutations in SNCA, such as A30P and A53T, may cause familial cases of PD (5,6). α -Syn misexpression has been experimentally demonstrated to mimic several aspects of PD in transgenic animals, including motor dysfunction, α -Syn aggregation and neurodegeneration (7-9). At present, several molecular mechanisms have been proposed to explain the causes of PD, including misfolding and aggregation of α -Syn, posttranslational modifications of α -Syn and oxidative stress (10).

In addition to motor dysfunction, numerous patients with PD have non-motor symptoms, including depression, sleep disorders, dementia, autonomic dysfunction, hyposmia and gastrointestinal symptoms (11). Previous investigations have demonstrated that a specific cholinergic neurotransmitter defect in patients with senile dementia contributed significant damage to cholinergic neurons. This observation suggests a close association between the cholinergic system and cognitive function in the human brain. As a critical cognitive function, memory has been used to demonstrate that synaptic plasticity in the *Drosophila melanogaster* mushroom body neurons is critical for learning and memory of olfactory stimuli (12). The projection neurons (PNs) in the antennal lobe of the mushroom body are cholinergic and neurotransmission at the synapse conducts messages in the form of electrical activity (13). Voltage-gated calcium channels (CAC) are located at the plasma membrane of nerve terminals and are crucial for the process of synaptic transmission and neuronal communication. Ca_v2 -type calcium channels are encoded by *cac* and regulate action potential (AP)-independent neurotransmitter release at cholinergic synapses in the adult *Drosophila* brain (14). While certain details have been determined concerning the molecular mechanisms of PD, direct evidence for the association between α -Syn and cholinergic transmission remains to be demonstrated. The aim of the present study was to use *Drosophila* as a model organism to observe whether α -Syn affects cholinergic neuronal transmission and consequentially alters their learning and memory capabilities.

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Materials and methods

***Drosophila* strains.** *Drosophila* stocks were reared on standard cornmeal agar medium supplemented with dry yeast for 24 h

at 60% relative humidity. α -Syn-expressing *Drosophila* were modified by microinjection and expression of A53T α -Syn transgenes was driven by the elav-GAL4 expression system. In brief, the α -Syn clone was constructed using the elav-GAL4 expression system, then this was injected into the eggs of white eye *Drosophila* (control flies). After hybridization of the red eye flies, α -Syn flies were created. Elav-GAL4 flies were crossed with UAS-A53T flies (provided by the Biochemistry and Cell Institute of Shanghai Life Science Research Institute of the Chinese Academy of Sciences, Shanghai, China), to produce the A53T flies.

Western blot analysis. The cerebral proteins of α -Syn *Drosophila* brains were extracted from tissue lysate with the protease inhibitor phenylmethylsulfonyl fluoride (Sigma-Aldrich, St. Louis, MO, USA). The solution was sonicated and ultracentrifuged for 30 min at 4695 x g at below 4°C. The resulting supernatant was boiled for 15 min in a water bath. Adult fly heads of the appropriate genotypes were prepared and analyzed using standard SDS-PAGE with 10% Tris-glycine gradient gels. The primary α -Syn antibody (cat. no. 610786) was developed by BD Biosciences (Mountain View, CA, USA) and was diluted to 1:1,000 prior to use. The goat anti-mouse secondary antibody (cat. no. 81-6511; Thermo Fisher Scientific, San Francisco, CA, USA) was then diluted to 1:10,000 and applied, and an X-ray film exposure was conducted.

Climbing assays. Flies were divided into random groups of 20-30 individuals per vial and then assessed for geotaxis as described previously (8). Briefly, groups of 10 flies were placed in a 95x27 mm empty vial. Flies were gently taped to the bottom of the vial and the number of flies crossing an 8 cm mark was recorded after 10 sec. Researchers were blinded to the corresponding genotype and condition of the flies.

T maze test. Hungry flies (80-100), which had been in the dark for 30 min prior to the experiment were placed in the maze. After 90 sec, smell A was released in one side of the T maze and at the same time, twelve 1.5 sec of the 70 volt DC electrical stimulation was applied over 60 sec. Flies were then allowed to breathe air for 45 sec. Subsequently, smell B was released for 60 sec in the other side of the T maze followed by 45 sec air. The flies were then allowed to move around the maze freely for 2 min, after which the numbers of flies at the sides of the maze with smells A and B were counted.

Immunofluorescence. The flies brains were stripped and placed on a slide. The slide was rinsed with 1% PBS (Sigma-Aldrich) three times, blocked and incubated in a blocking buffer (0.1 M PBS, 0.1% Triton X-100 and 1% BSA; Sigma-Aldrich) for 3 h at room temperature. The α -synuclein antibody (1:250; 610786; BD Biosciences) was then applied and washed by PBS for 5 min, three times. Subsequently, the rabbit anti-human IgG secondary antibody (BA1020; Boster, Wuhan, China) was applied. Following incubation, the brain was washed three times with PBS with 5 min intervals and fixed with a coverslip. The brains of flies were observed with an immunofluorescence laser scanning confocal microscope (Zeiss LSM710; Carl Zeiss, Oberkochen, Germany).



Figure 1. Western blot analysis of *Drosophila* α -synuclein. Protein of α -synuclein and A53T are 19 kDa.

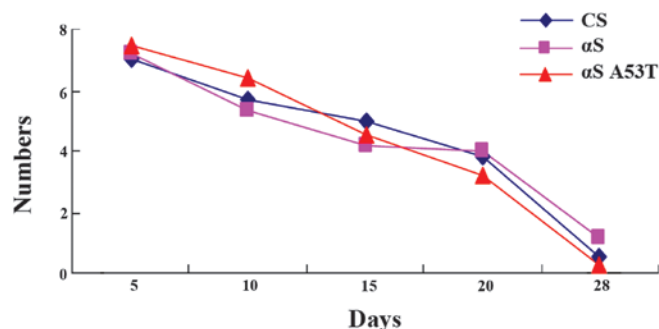


Figure 2. Climbing assays. No significant difference was identified in locomotor activity of α -synuclein and A53T flies compared with the CS group ($P > 0.05$). CS, control saline, α S, α -synuclein.

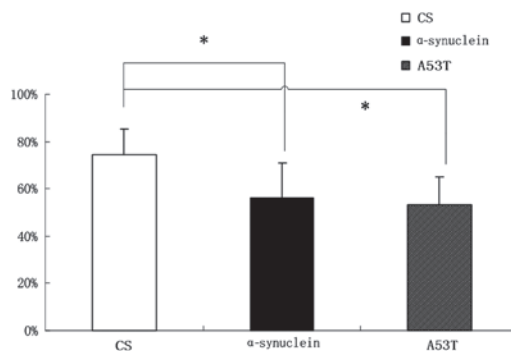


Figure 3. T maze results. α -synuclein and A53T flies' learning and memory ability reduced by 18% compared with CS flies ($P < 0.05$). CS, control saline.

Biocytin staining and immunohistochemistry images. In order to identify and confirm that the cells recorded were PNs, cells recorded were stained by biocytin (Sigma-Aldrich). While recording, cells were injected with biocytin through a recording pipette filled with internal solution containing biocytin in whole-cell recording mode for a minimum of 30 min. After recording, the brains were collected and fixed in 4% formaldehyde (Sigma-Aldrich) in phosphate-buffered saline (PBS; Sigma-Aldrich) at 4°C for 10 h. The brain was then rinsed in 1% PBS three times, blocked and incubated in a blocking buffer (0.1 M PBS, 0.1% Triton X-100, and 1% bovine serum albumin; Sigma-Aldrich) containing streptavidin-Cy3 (Molecular Devices Ltd., Wokingham, UK) for 3 h at room temperature. After incubation, the brain was washed three times with 5 min intervals in PBS. A BX51WI microscope

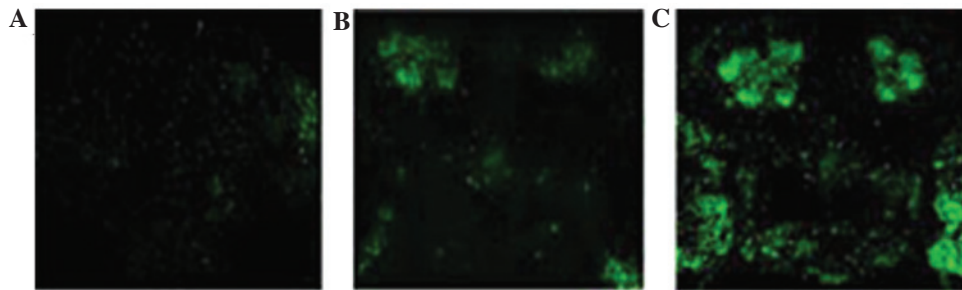


Figure 4. Laser scanning confocal microscope images of *Drosophila* brains. (A) CS *Drosophila*; (B) α -synuclein *Drosophila*; (C) A53T *Drosophila* (magnification $\times 63$). α -synuclein and A53T *Drosophila* exhibit green fluorescence, particularly in the mushroom body, while the CS *Drosophila* does not exhibit green fluorescence. CS, control saline.

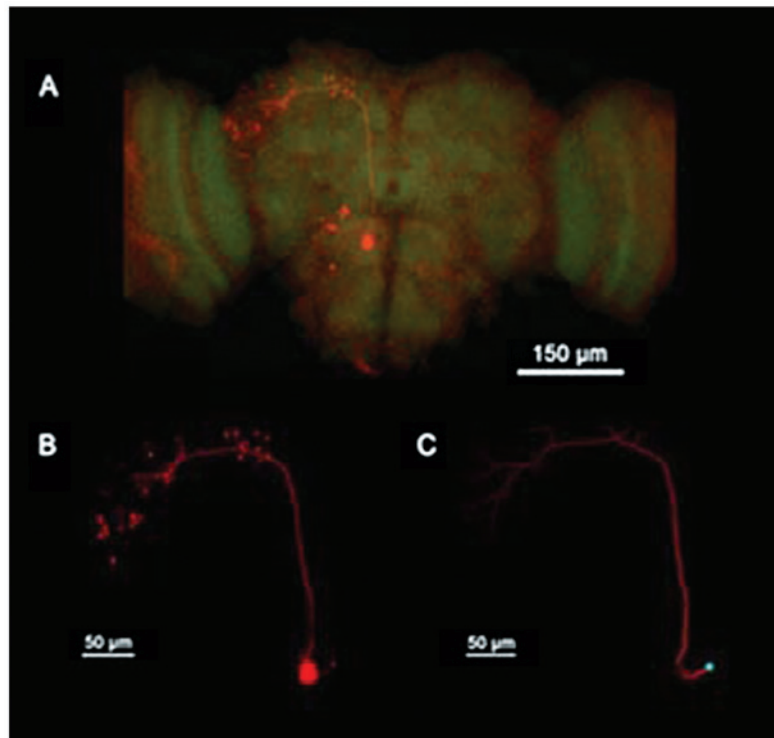


Figure 5. Image demonstrating the detailed morphology of the recorded projection neurons. (A) The morphology of the whole fly brain following injection of biocytin; (B) removal of the whole brain background; (C) the single projection neuron.

(Olympus, Tokyo, Japan) with a $\times 40$ objective and confocal camera was used to capture images of dendritic arborization of the PNs in the antennal lobe.

Immunohistochemistry. Adult *Drosophila* brains were fixed in 4% paraformaldehyde and embedded in paraffin. The paraffin-embedded brains were then cut into sections. The α -Syn antibody (1:250; BD Biosciences) was used to detect the expression of α -Syn.

Electrophysiological recordings from PNs in isolated whole brain. All brains were obtained from female flies two days prior to eclosion. The entire brain, including optic lobes, was removed from the head and prepared for recordings in standard external solution containing 20 U/ml papain (Sigma-Aldrich) with 1 mM l-cysteine. Subsequently, the dissected brains were mounted in an RC-26 perfusion

chamber (Sigma-Aldrich) containing the recording solution bubbled with 95% O_2 and 5% CO_2 (2 ml/min) throughout the experiments, with the ventral surface of the brain facing up. Pipettes were targeted to PNs in the dorsal neuron cluster in the antennal lobe. For measurements of cholinergic miniature excitatory postsynaptic currents (mEPSCs), tetrodotoxin (TTX; 1 μ M) was added to the external solution to block voltage-gated sodium currents and γ -aminobutyric (GABA)ergic synaptic currents, and picrotoxin (PTX; 10 μ M; Sigma-Aldrich) was added to block GABAergic synaptic currents. $CaCl_2$ was omitted and tetraethylammonium (TEA; 10 mM; Sigma-Aldrich) and 4-aminopyridine (4-AP; 1 mM) were added to the external solution to measure Ca^{2+} currents. For Ca^{2+} current measurements, the external solution consisted of 101 mM NaCl, 1.8 mM $CaCl_2$, 0.8 mM $MgCl_2$, 5.4 mM KCl, 5 mM glucose, 1.25 mM NaH_2PO_4 , 20.7 mM $NaHCO_3$, 1 μ M TTX, 10 mM TEA and 1 mM 4-AP.

Statistical analysis. Statistical analysis was conducted using SPSS software, version 11.5 (SPSS, Inc., Chicago, IL, USA) Comparison of the magnitude of the calcium currents prior to and following application of permethrin was made using an unpaired t-test. Comparison of the area below the baseline of sodium currents prior to and following application of permethrin was made using a paired t-test. Comparisons of more than two treatments were made with analysis of variance (ANOVA) and the Bonferroni post hoc correction.

Results

Expression of α -Syn protein. The pan-neuronal driver elav-GAL4 was used to drive expression of normal and mutant (A53T) human α -Syn throughout the nervous system. Western blot analysis was performed to verify the expression of α -Syn (Fig. 1).

No differences are present in locomotor ability of α -Syn *Drosophila*. Previous studies have demonstrated that a decrease in the ability of flies to climb up the wall of a plastic vial is associated with the progression of the pathology in PD (15,16). However, in the present study, no significant decrease in climbing ability was observed (Fig. 2).

Learning and memory skills are reduced in α -Syn *Drosophila*. Compared with CS flies, the learning and memory ability of α -Syn wild-type and A53T mutation flies decreased exhibiting a reduction of ~18% (Fig. 3) as elucidated through a T maze experiment.

Immunofluorescence analysis of α -Syn in *Drosophila*. The whole brain of the *Drosophila* were observed with an immunofluorescence laser scanning confocal microscopy, and it was identified that the levels of green fluorescence were greater in the α -Syn *Drosophila* compared with the control saline group. In addition, green fluorescence was particularly evident in the mushroom body (Fig. 4).

mEPSC frequency is decreased in α -Syn *Drosophila*. Previous studies have demonstrated that cac-encoded voltage-gated calcium channels are important in regulating the spontaneous release of neurotransmitter at excitatory cholinergic synapses and mEPSC frequency in the PNs of *Drosophila* (7,17,18). Recordings were performed at room temperature for a minimum of 1 min in the presence of TTX to block sodium channels and PTX to block GABA receptors. The dorsal antennal lobe glomeruli is the location of the PNs, with two main branches projecting to the lateral horn and the local mushroom body. The PNs were identified morphologically from biocytin fills and their specific electrical activities (Fig. 5). The PNs were selected for analysis as they were cholinergic and cholinceptive. PNs firing spontaneously were recorded in α -Syn *Drosophila* in the presence of TTX and PTX. The experiments indicated that the frequency of PNs was reduced. Sodium AP-independent mEPSCs in the antennal lobe PNs were monitored in the whole brain, which was isolated from female fly pupae two days prior to eclosion. mEPSCs were directly monitored by whole cell patch-clamp recordings from PNs. The average mEPSC frequency was 1.92 ± 0.3 Hz under

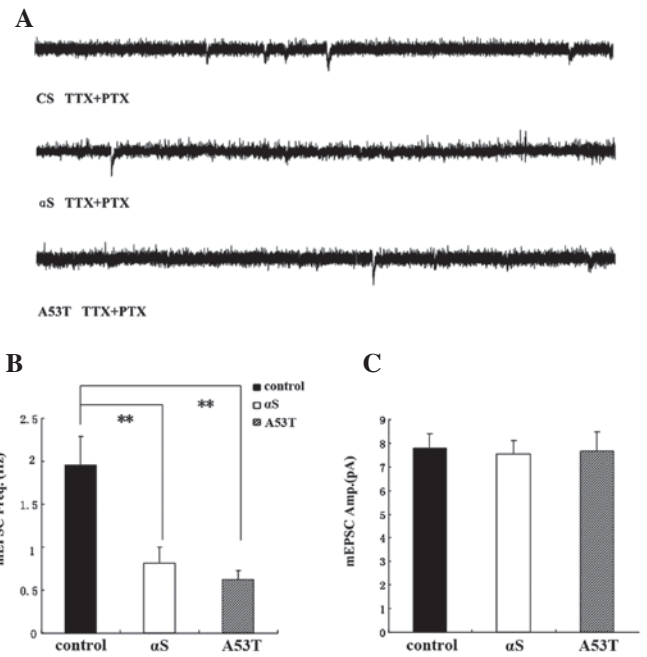


Figure 6. (A) α -synuclein modulated mEPSC frequency and amplitude in projection neurons of *Drosophila* brain. (B) mEPSC frequency of α -synuclein and A53T flies is reduced markedly (** $P < 0.01$), (C) α -synuclein and A53T do not exhibit a significant difference from that of CS ($P > 0.05$). CS, control saline, mEPSCs, miniature excitatory postsynaptic currents, α S, α -synuclein; TTX, tetrodotoxin; PTX, picrotoxin.

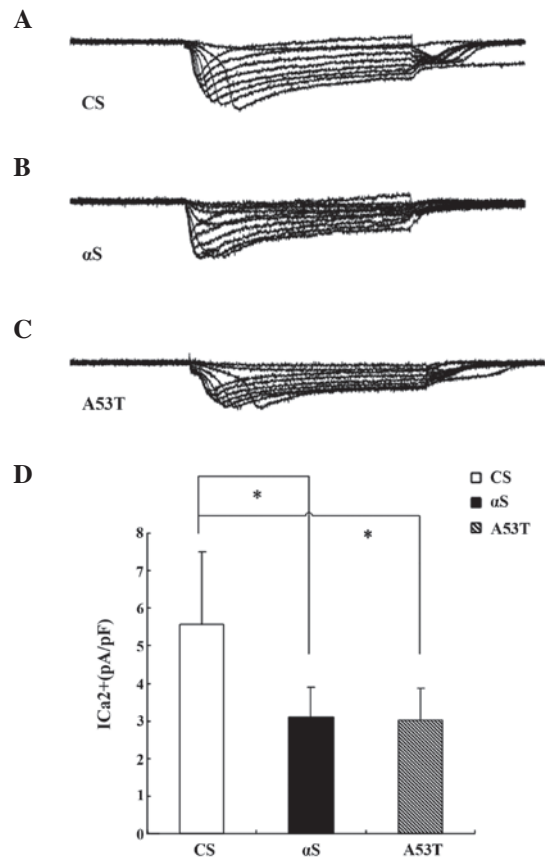


Figure 7. (A) Effect of α S on whole-cell calcium currents in a PN. (B) Typical calcium current family recorded from a PN in control saline. (C) Calcium current recorded in the PN with α S and A53T. (D) Density of calcium current was significantly reduced by α S compared with control saline ($P < 0.05$). α S, α -synuclein; PN, projection neuron; CS, control saline.

normal conditions. The average mEPSC frequency decreased significantly by 0.62 ± 0.11 Hz in the α -Syn *Drosophila*. The present results demonstrated that the mEPSC frequency in the α -Syn *Drosophila* was decreased (Fig. 6).

Cholinergic mEPSCs were recorded from single PNs following the addition of TTX and PTX to the external solution. The mean frequency of mEPSCs was significantly reduced by α -Syn (ANOVA, Bonferroni post hoc test; $P < 0.01$). No significant difference was identified between the mean mEPSC amplitude of the α -Syn and A53T groups and that of the CS group.

*Calcium currents of PNs in brains of α -Syn *Drosophila* pupae two days prior to eclosion.* Calcium currents have been revealed to have broad effects on neuronal function, particularly in presynaptic terminals for neurotransmitter release, electrical excitability and activity-dependent gene regulation (19,20). Calcium currents were directly calculated in antennal lobe PNs isolated from the whole brain of fly pupae two days prior to eclosion. Using whole cell voltage-clamp recordings, the inward calcium current was isolated in PNs as previously described (21). The average calcium current in normal PNs and α -Syn PNs were 5.57 ± 1.94 and 3.03 ± 0.85 pA/pF, respectively (Fig. 7).

Discussion

Drosophila are an ideal model for investigating the association between PD and Alzheimer's disease (AD) as transgenic techniques can be used to express human α -Syn in the CNS of *Drosophila*. The abundant expression of acetylcholine and GABA throughout the *Drosophila* CNS suggested that these classic neurotransmitters have a major role in mediating fast synaptic transmission (22). The PNs were selected for analysis as they are cholinergic and cholinceptive, receiving cholinergic synaptic input from olfactory receptor neurons and potentially lateral excitatory input (13). The preparations used in the current study and described in a previous study (23) allowed assessment of the cholinergic synaptic transmission in PNs in an isolated whole brain. Cholinergic currents were recorded using a whole-cell patch clamp at room temperature following treatment with TTX for at least 1 min to block sodium channels and PTX to block GABA receptors. The neurons were identified as cholinergic and mediated by nicotinic acetylcholine receptors based on the finding that the majority of the mEPSCs recorded could be inhibited by a competitive antagonist of the nicotinic acetylcholine receptor at the neuromuscular junction termed curare, which is classified as a long-duration (24), non-depolarizing neuromuscular blocking agent. A number of studies have revealed that the morbidity associated with senile dementia is more prevalent in PD patients and cholinergic neurons exhibit a critical role in AD. A number of studies had reported difficulties in recording cholinergic currents in the mammalian nervous system and as a result and in addition to their amenability to transgenic studies, *Drosophila* became a useful model for studying of the association between AD and PD.

Certain non-motor symptoms of PD, including cognitive impairment and sleep disorders often appear a number of years prior to the onset of motor symptoms. The main pathology

associated with PD involves dopaminergic degeneration, which is the main factor contributing to the motor symptoms. The motor symptoms observed in patients with PD are often accompanied by cognitive impairment, depression and other symptoms associated with degeneration of the forebrain. Mutations in α -Syn are associated with genetic and sporadic cases of PD. Previous studies have demonstrated that an α -Syn mutation in *Drosophila*, which leads to increased oligomerization, enhances the degree of neurotoxicity in the fruit fly (25). A previous study demonstrated that there is a long time period between the degeneration of dopaminergic neurons and the appearance of the typical motor symptoms of PD (26), which indicates that there is a compensatory mechanism in the central nervous system. It is hypothesized that the dynamic balance between dopamine and acetylcholine is important. Inhibiting the cholinergic system may cause a decline in learning and memory abilities and symptoms similar to senile amnesia (27). In addition, numerous patients with PD eventually develop cognitive deficits and dementia (28). The present experiments demonstrate that the non-motor symptoms in patients with PD may be associated with the cholinergic system.

The present results also demonstrate that learning and memory skills reduced in α -Syn wild-type and A53T mutant *Drosophila* by ~18%. Previous studies have demonstrated that the expression of human α -Syn may impair the motor capability of *Drosophila* (8,29,30). However, in the present study, no significant difference was identified in the motor function of the α -Syn and A53T *Drosophila*. It is possible that the flies in the present study were too young and the expression of α -Syn may only effect non-motor symptoms at this stage, without a marked impact on motor symptoms. α -Syn may reduce the frequency of mEPSCs in PNs and reduce the electrical signals of the calcium channel. Thus, it has been demonstrated that α -Syn expression in *Drosophila* may affect cholinergic synaptic transmission. The frequency of mEPSCs in PNs and the calcium current were reduced, which may be associated with the decline in the ability to learn and in memory in *Drosophila*. The present study demonstrated that mutations in α -Syn may be one of the reasons for the higher incidence of senile dementia in patients with PD. However, the exact mechanism requires further investigation.

References

1. Janezic S, Threlfell S, Dodson PD, *et al*: Deficits in dopaminergic transmission precede neuron loss and dysfunction in a new Parkinson model. *Proc Natl Acad Sci USA* 110: E4016-E1025, 2013.
2. Kitada T, Asakawa S, Hattori N, Matsumine H, Yamamura Y, Minoshima S, *et al*: Mutations in the parkin gene cause autosomal recessive juvenile parkinsonism. *Nature* 392: 605-608, 1998.
3. Paisán-Ruiz C, Jain S, Evans EW, Gilks WP, Simón J, van der Brug M, *et al*: Cloning of the gene containing mutations that cause PARK8-linked Parkinson's disease. *Neuron* 44: 595-600, 2004.
4. Zimprich A, Biskup S, Leitner P, Lichtner P, Farrer M, Lincoln S, *et al*: Mutations in LRRK2 cause autosomal-dominant parkinsonism with pleomorphic pathology. *Neuron* 44: 601-607, 2004.
5. Krüger R, Kuhn W, Müller T, *et al*: Ala30Pro mutation in the gene encoding α -synuclein in Parkinson's disease. *Nature Genetics* 18: 106-108, 1998.
6. Polymeropoulos MH, Lavedan C, Leroy E, *et al*: Mutation in the α -synuclein gene identified in families with Parkinson's disease. *Science* 276: 2045-2047, 1997.
7. Feany MB and Bender WW: A *Drosophila* model of Parkinson's disease. *Nature* 404: 394-398, 2000.

8. Lee MK, Stirling W, Xu Y *et al*: Human α -synuclein-harboring familial Parkinson's disease-linked Ala-53 \rightarrow Thr mutation causes neurodegenerative disease with α -synuclein aggregation in transgenic mice. *Proc Nat Acad Sci USA* 99: 8968-8973, 2002.
9. Masliah E, Rockenstein E, Veinbergs I, *et al*: Dopaminergic loss and inclusion body formation in α -synuclein mice: implications for neurodegenerative disorders. *Science* 287: 1265-1269, 2000.
10. Mizuno H, Fujikake N, Wada K, *et al*: α -Synuclein transgenic *Drosophila* as a model of Parkinson's disease and related synucleinopathies. *Parkinsons Dis* 2011: 212706, 2011.
11. Chaudhuri KR, Yates L and Martinez-Martin P: The non-motor symptom complex of Parkinson's disease: A comprehensive assessment is essential. *Curr Neurol Neurosci Rep* 5: 275-283, 2005.
12. Heisenberg M: Mushroom body memoir: from maps to models. *Nat Rev Neurosci* 4: 266-275, 2003.
13. Kazama H and Wilson RI: Homeostatic matching and nonlinear amplification at identified central synapses. *Neuron* 58: 401-413, 2008.
14. Bosboom JL, Stoffers D and Wolters ECh: Cognitive dysfunction and dementia in Parkinson's disease. *J Neural Transm* 111: 1303-1315, 2004.
15. Rieckhof GE, Yoshihara M, Guan Z and Littleton JT: Presynaptic N-type calcium channels regulate synaptic growth. *J Biol Chem* 278: 41099-41108, 2003.
16. Pendleton RG, Parvez F, Sayed M and Hillman R: Effects of pharmacological agents upon a transgenic model of Parkinson's disease in *Drosophila melanogaster*. *J Pharmacol Exp Ther* 300: 91-96, 2002.
17. Stone E, Haario H and Lawrence JJ: A kinetic model for the frequency dependence of cholinergic modulation at hippocampal GABAergic synapses. *Math Biosci* 258: 162-175, 2014.
18. Zhaowei L, Yongling X, Jiajia Y and Zhuo Y: The reduction of EPSC amplitude in CA1 pyramidal neurons by the peroxynitrite donor SIN-1 requires Ca²⁺ influx via postsynaptic non-L-type voltage gated calcium channels. *Neurochem Res* 39: 361-371, 2014.
19. Gu H, Jiang SA, Campusano JM, Iniguez J, Su H, Hoang AA, *et al*: Ca_v2-type calcium channels encoded by cac regulate AP-independent neurotransmitter release at cholinergic synapses in adult *Drosophila* brain. *J Neurophysiol* 101: 42-53, 2009.
20. Kawasaki F, Zou B, Xu X and Ordway RW: Active zone localization of presynaptic calcium channels encoded by the cacophony locus of *Drosophila*. *J Neurosci* 24: 282-285, 2004.
21. Catterall WA: Structure and regulation of voltage-gated Ca²⁺ channels. *Annu Rev Cell Dev Biol* 16: 521-555, 2000.
22. Karpinar DP, Balija MBG, Kügler S, *et al*: Pre-fibrillar α -synuclein variants with impaired β -structure increase neurotoxicity in Parkinson's disease models. *EMBO J* 28: 3256-3268, 2009.
23. Rascol O, Payoux P, Ory F, *et al*: Limitations of current Parkinson's disease therapy. *Ann Neuro* 53: s3-s15, 2003: discussions 12-5.
24. Restifo LL and White K: Molecular and genetic approaches to neurotransmitter and neuromodulator systems in *Drosophila*. *Adv Insect Physiol* 22: 115-219, 1990.
25. Gu H and O'Dowd DK: Cholinergic synaptic transmission in adult *Drosophila* Kenyon cells in situ. *J Neurosci* 26: 265-272, 2006.
26. Thompson MA: Muscle relaxant drugs. *Br J Hosp Med* 23: 153, 1980.
27. Rinne JO, Myllykylä T, Lönnberg P and Marjamäki P: A post-mortem study of brain nicotinic receptors in Parkinson's and Alzheimer's disease. *Brain Res* 547: 167-170, 1991.
28. Foltynie T, Brayne CE, Robbins TW and Barker RA: The cognitive ability of an incident cohort of Parkinson's patients in the UK. The CamPaIGN study. *Brain* 127: 550-560, 2004.
29. Chen AY, Wilburn P, Hao X, Tully T: Walking deficits and centrophobism in an α -synuclein fly model of Parkinson's disease. *Genes Brain Behav* 13: 812-820, 2014.
30. Gajula Balija MB, Griesinger C, Herzig A, Zweckstetter M, Jäckle H: Pre-fibrillar α -synuclein mutants cause Parkinson's disease-like non-motor symptoms in *Drosophila*. *PLoS One* 6: e24701, 2011.