

Upregulation of retinoic acid receptor- β reverses drug resistance in cholangiocarcinoma cells by enhancing susceptibility to apoptosis

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Abstract. Retinoic acid receptor β (RAR β), a known tumor suppressor gene, is frequently silenced in numerous malignant types of tumor. Recent reports have demonstrated that loss of RAR β expression may be responsible, in part, for the drug resistance observed in clinical trials. However, little is known about the role of RAR β in regulating drug sensitivity in patients with cholangiocarcinoma (CCA) with a high risk of mortality and poor outcomes. In the present study, low RAR β expression was observed in the majority of CCA tissues investigated (28/33, 84.8%). In addition, the CCA cell line QBC939, which exhibits low RAR β expression, was found to be significantly resistant to chemotherapeutic agents compared with SK-ChA-1, MZ-ChA-1 and Hccc9810 CCA cell lines, which exhibit high RAR β expression. Furthermore, upregulation of RAR β significantly enhanced the sensitivity of QBC939 cells to common chemotherapeutic agents both *in vitro* and *in vivo*. Upregulation of RAR β was shown to increase the expression of proapoptotic genes *bax*, *bak* and *bim*, in addition to caspase-3 activity, and decrease the expression of antiapoptotic genes *bcl-2*, *bcl-xL* and *mcl-1*. As a result, CCA cells were more susceptible to caspase-dependent apoptosis. Taken together, these data suggest that RAR β upregulation rendered CCA cells more sensitive to chemotherapeutic agents by increasing the susceptibility of cells to caspase-dependent apoptosis. These results support the hypothesis that RAR β

may be an ideal chemosensitization target for the treatment of patients with drug-resistant CCA.

Introduction

Altered expression of nuclear retinoid receptors is associated with the malignant transformation of human cells (1). Retinoic acid receptor β (RAR β) belongs to the RAR superfamily and is frequently silenced during epithelial cell carcinogenesis, including those of the breast, lung, esophagus, pancreas, cervix and prostate (2-7). Thus, RAR β is believed to play a role as a tumor suppressor gene in human tumorigenesis (8). Of particular note, esophageal, lung and breast cancer cell lines that do not express RAR β have been found to be resistant to retinoid treatment (9,10). This has led to the hypothesis that RAR β expression may be responsible, in part, for mediating the efficacy of chemopreventative agents in clinical trials, such as inhibitors of the epidermal growth factor receptor and β -cryptoxanthin. An increasing number of studies suggest that restoration of RAR β expression in tumor cells may enhance their response to chemotherapeutic agents (11).

Cholangiocarcinoma (CCA) is the second most common primary liver malignancy, which is highly resistant to available chemotherapeutic agents and confers a 5-year relative survival rate of less than 5% (12). Surgery is the only curative treatment for CCA, however only a very small percentage of tumors are resectable. This is primarily due to the majority of patients presenting with late-stage disease, such as local advanced or metastatic disease (13). Therefore, the identification of new and effective therapies against CCA is critical. To date, clinical strategies to enhance drug sensitivity have included efforts that aim to initiate apoptotic signaling pathways, activate specific tumor suppressors and determine effective combination regimens (14). However, expression of the RAR β tumor suppressor in CCA, together with its role in regulating sensitivity to chemotherapeutic agents, are currently unknown.

In the present study, the expression of RAR β in CCA tissues was investigated and the potential mechanisms by which RAR β confers sensitivity to chemotherapeutic agents within CCA cells *in vitro* and *in vivo* was explored.

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Materials and methods

Reagents. 5-fluorouracil (5-FU), vincristine sulfate (VCR), cisplatin (CDDP), mitomycin C (MMC), 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT), caspase-3 inhibitor (z-VAD-fmk), G418, BMS453 and CoCl₂ were all purchased from Sigma-Aldrich (St. Louis, MO, USA). The RPMI-1640 and fetal bovine serum (FBS) were all purchased from Gibco BRL (Grand Island, NY). The 100 U/ml penicillin and bicinchoninic acid protein assay reagent were purchased from Wuhan Boster (Wuhan, China). Glyceraldehyde 3-phosphate dehydrogenase (GAPDH; ab9485) and RAR β (ab198557) antibodies were purchased from Abcam (Cambridge, UK). The annexin V-fluorescein isothiocyanate (FITC)/propidium iodide (PI) double staining apoptosis detection kit and TdT-mediated dUTP nick end labeling (TUNEL) kit were obtained from Roche Diagnostics (Basel, Switzerland). Lipofectamine 2000 was purchased from Invitrogen (Thermo Fisher Scientific, Inc., Waltham, MA., USA). The Caspase-3 assay kit was from R&D Systems, Inc. (Minneapolis, MN, USA). All of other chemicals and reagents were the highest quality and purchased from standard commercial sources.

Patients and tumor specimens. Analysis of RAR β expression in clinical CCA tissues was performed in agreement with the ethical guidelines of the 1975 Declaration of Helsinki, and was approved by the Institute Research Ethics Committee at The First Affiliated Hospital of Xiamen University. Between 2009-2013, 33 CCA samples were collected from patients without metastatic disease that had not received pre-operative treatment. Tumor tissues were fixed in 10% formalin and then paraffin-embedded for immunohistochemical (IHC) analysis and routine histological characterization.

Cell culture and stable transfection. The human CCA cell line QBC939 was a kind gift from Professor Shu-Guang Wang from Southwest Hospital, the Third Military Medical University (Chongqing, China). The CCA cell lines Sk-ChA-1, MZ-ChA-1 and Hccc9810 were a kind gift from Professor Chun-Dong Yu laboratory of Xiamen University (Xiamen, China). HIBEpIC human intrahepatic biliary epithelial cells were purchased from Cell Bank of the Type Culture Collection of Chinese Academy of Sciences (Shanghai, China). All four human CCA cell lines and HIBEpIC cells were all cultured in RPMI-1640 supplemented with 10% FBS and 100 U/ml penicillin in a humidified chamber at 37°C in 5% CO₂. RAR β cDNA was cloned into the expression vector pBoBi as described previously (15). The RAR β expression vector (4 μ g) was stably transfected into QBC939 cells (1x10⁶) to produce pBoBi-RAR β using 10 μ l Lipofectamine 2000, according to the manufacturer's instructions. Positive selection of stable transfectants was achieved by supplementing complete medium with 400 mg/ml of G418. Stable control vector QBC939 transfectants are defined as pBoBi-Ctrl from herein.

Cell proliferation assay. To determine cell viability in response to treatment, an MTT assay was used. Briefly, QBC939 cells were seeded onto 96-well culture plates at a density of 5x10³ cells/well and grown in complete RPMI-1640 culture

medium. Following overnight incubation, cells were treated with a variety of agents (60 μ M) including 5-FU, CDDP, VCR and MMC for 48 h. MTT (5 mg/ml) was added to each well and incubated at 37°C for 4 h, before the resulting formazan crystals were dissolved in dimethyl sulfoxide. Absorbance was read at 490 nm using a microplate reader (Model 680; Bio-Rad Laboratories, Inc., Hercules, CA, USA).

Hypoxic growth conditions. Cells were cultured in a 35 mm dish until 70-80% confluence was reached. They were then transferred to a hypoxic incubator with a humidified atmosphere that was flushed with $\leq 0.1\%$ O₂, 5% CO₂ and 95% N₂, followed by incubation at 37°C for 12 h.

Serum starvation. Cells were seeded onto 6-well culture plates at a density of 2x10⁵ cells/well and grown in complete RPMI-1640 culture medium overnight. Then the medium was removed and washed with PBS, and added the RPMI 1640 medium without serum. The duration of serum starvation was determined according to the experimental requirements.

Tumor xenograft study. *In vivo* drug sensitivity experiments were divided into two groups: RAR β (pBoBi-RAR β) and control (pBoBi-Ctrl). Female specific pathogen-free BALB/c nude mice (Shanghai Laboratory Animal Center, Shanghai, China) (age, 4-5 weeks) were injected subcutaneously with 100 μ l cells (2x10⁶). Mice were treated with 70 mg/kg/d 5-FU at day 8 post-transplantation. Tumor volumes were determined using the following formula: (AxB²)/2, where A is the largest diameter and B is the perpendicular diameter. After 35 days from injection, mice were sacrificed by inhalation of carbon dioxide. All animal procedures were approved by the Animal Care and Use Committee of the First Affiliated Hospital of Xiamen University.

Gene and protein expression analyses. mRNA expression levels were determined by reverse transcription-quantitative polymerase chain reaction (RT-qPCR). Protein expression was measured using a western blotting assay. Both analyses were performed as described previously (15).

Immunohistochemical (IHC) staining. IHC was performed to detect RAR β expression in CCA tissues as described previously (16). Human CCA tissue sections were immunostained with an antibody against RAR β (1:100). IHC scoring was conducted by two independent pathologists. The number of positively-stained cells per 1,000 cells in randomly selected fields was recorded, and the mean number from a minimum of five slides was calculated. Staining intensity was categorized as low (<50%) or high (>50%).

Analysis of apoptosis. Apoptosis was evaluated using an annexin V-FITC/PI double-staining assay and a TUNEL assay. The annexin V-FITC/PI double-staining assay was performed according to the manufacturer's instructions (Roche Diagnostics) and analyzed using a flow cytometer (FACSCalibur; BD Biosciences, Franklin Lakes, CA, USA). TUNEL staining was performed using an *in situ* cell death detection kit according to the manufacturer's instructions (Roche Diagnostics).

Analysis of caspase-3 activity. Caspase-3 activity was determined using the caspase-3 assay kit (R&D Systems Inc., Minneapolis, MN, USA) according to the manufacturer's instructions. Briefly, cells were lysed and centrifuged at 10,000 x g for 20 min at 4°C to obtain supernatants. The protein concentration of sample supernatants was determined using a bicinchoninic acid protein assay reagent before they were added to a dithiothreitol and caspase-3 substrate reaction mixture and incubated for 2 h at 37°C. Absorbance was measured at 405 nm using a microplate reader (Model 680; Bio-Rad Laboratories, Inc.).

Statistical analysis. Data are presented as the mean ± standard error from a minimum of three independent experiments. Student's *t*-test or one-way analysis of variance was conducted using the SPSS 13.0 software package (SPSS, Inc., Chicago, IL, USA). Dunn test was used if there was statistical significance after one-way analysis. *P*<0.05 was considered to indicate a statistically significant difference.

Results

Low expression of RARβ is partially responsible for the drug resistance of CCA. IHC was performed to investigate RARβ protein expression in a set of 33 paraffin-embedded human CCA tissues. A total of 28/33 (84.8%) tissues were found to exhibit low RARβ protein expression (Fig. 1A), only 5 tissues exhibited high RARβ protein expression (Fig. 1B). In addition, RARβ mRNA and protein expression levels in four CCA cell lines (QBC939, SK-ChA-1, MZ-ChA-1 and Hccc9810 cells) were found to be significantly lower than the HBd Epi normal bile duct epithelial cell line (Fig. 2A). RARβ mRNA and protein levels were markedly reduced within QBC939 cells. Notably, the proportion of surviving QBC939 cells following exposure to three commonly used chemotherapeutic agents including 5-FU, VCR and MMC, was found to be significantly higher compared with the remaining three CCA cell lines (Fig. 2B). These results give rise to the hypothesis that the observed resistance of CCA cells to common therapeutic agents, may be associated with silenced RARβ expression.

Upregulation of RARβ enhances the sensitivity of CCA cells to chemotherapeutic agents in vitro. In order to investigate the role of RARβ in the therapeutic response of CCA cells, RARβ expression vectors were transfected into QBC939 cells prior to treatment with chemotherapeutic agents. As shown in Fig. 3A, an increase in RARβ mRNA and protein levels was observed within stably RARβ-transfected cells (pBoBi-QBC939) compared with control (pBoBi-Ctrl) cells. Cell survival analysis results indicated that the sensitivity of pBoBi-QBC939 cells to common chemotherapeutic agents, including 5-FU, CDDP, VCR and MMC, was significantly enhanced following RARβ upregulation compared with pBoBi-Ctrl cells (Fig. 3B). Furthermore, the proportion of apoptotic pBoBi-QBC939 cells generated in response to 5-FU, CDDP, VCR and MMC, was found to be significantly higher than pBoBi-Ctrl cells (Fig. 3C). These *in vitro* findings suggest that upregulation of RARβ enhances the sensitivity of CCA cells to chemotherapeutic agents.

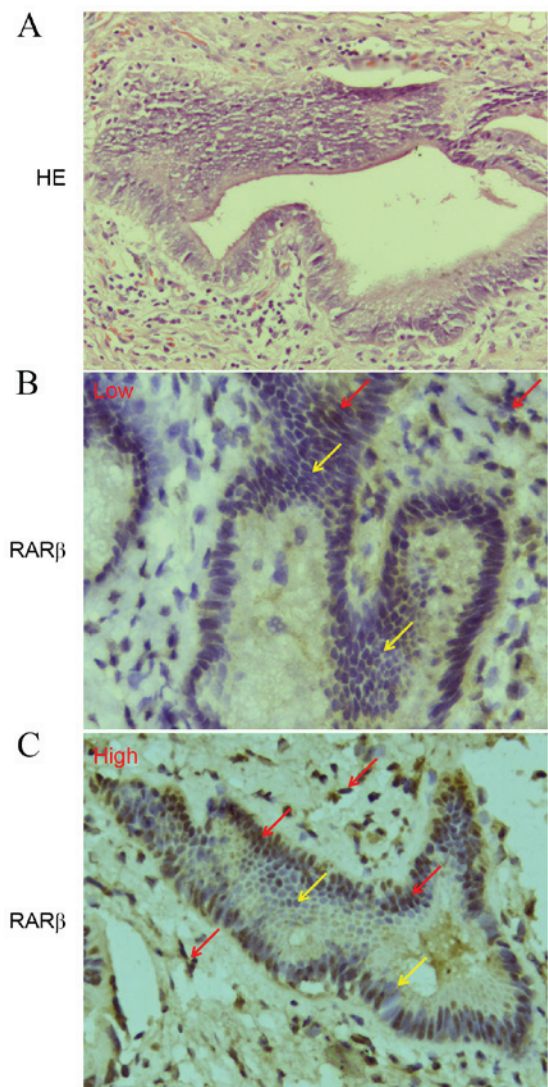


Figure 1. Expression of RARβ protein detected by IHC in human CCA tissues. (A) HE staining. (B and C) Expression of RARβ (magnification, x400). Yellow arrows indicate negative RARβ expression and red arrows indicate positive expression. The staining intensities of RARβ were categorized as low and high expression in 33 CCA tissues. RARβ, retinoic acid receptor-β; IHC, immunohistochemical staining; CCA, cholangiocarcinoma; HE, hematoxylin-eosin.

Upregulation of RARβ increases the cytotoxic effect of 5-FU on xenografted CCA tumors. 5-FU is a common chemotherapeutic agent used for the treatment of hepatobiliary tumors. Therefore, this agent was selected to investigate the contribution of RARβ upregulation in the sensitivity to chemotherapeutic treatment in the xenografted QBC939 CCA tumors. As shown in Fig. 4A, the rate of tumor growth inhibition in xenografted pBoBi-RARβ tumors following treatment with 5-FU, was significantly higher than in control pBoBi-Ctrl tumors. A significant difference in tumor growth inhibition was apparent from day 20 and increased further during late tumor growth (Fig. 4B). Moreover, analysis of apoptosis in xenografted CCA tumor tissues using the TUNEL assay was consistent with the results of the *in vitro* assays, and indicated that the number of apoptotic cells induced by 5-FU in xenografted pBoBi-RARβ tumors was significantly higher than control pBoBi-Ctrl tumors (*P*=0.0035; Fig. 4C). These *in vivo* findings provide

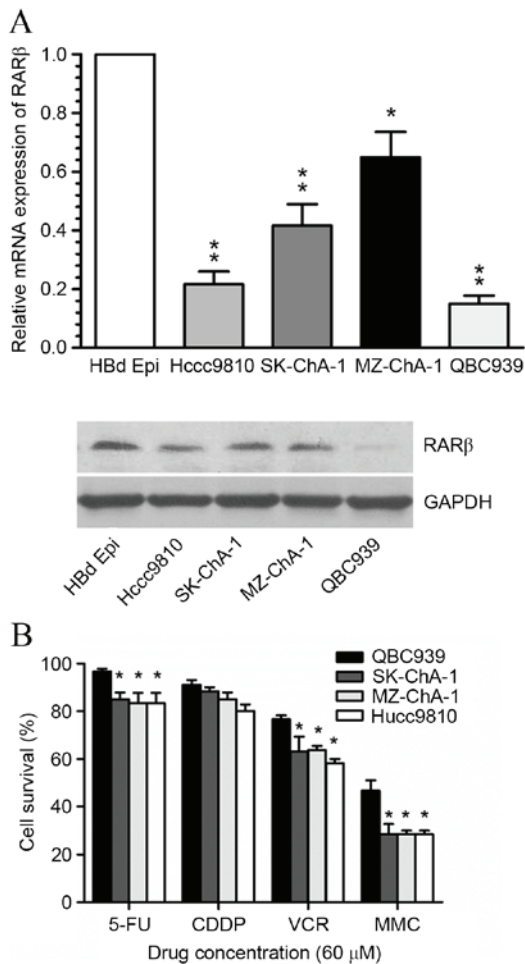


Figure 2. Expression of RAR β and drug sensitivity in human CCA cell lines. (A) Relative mRNA expression levels of RAR β in the normal bile duct cells and 4 CCA cell lines, as detected by reverse transcription-quantitative polymerase chain reaction. * $P < 0.05$, ** $P < 0.001$ vs. HBd Epi cells. Protein expression detected by western blotting analysis. (B) Drug sensitivities of four CCA cell lines in response to 5-FU, CDDP, VCR and MMC chemotherapeutic agents as determined using the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide assay. * $P < 0.05$, ** $P < 0.001$ vs. QBC939 cells. RAR β , retinoic acid receptor- β ; CCA, cholangiocarcinoma; 5-FU, 5-fluorouracil; CDDP, cisplatin; VCR, vincristine; MMC, mitomycin C.

additional evidence that RAR β upregulation enhances the sensitivity of CCA cells to chemotherapeutic agents.

High RAR β expression renders CCA cells more susceptible to caspase-dependent apoptosis. Taking the above results into consideration, the observed increase in the sensitivity of CCA cells to chemotherapeutic agents following RAR β upregulation may render CCA cells more susceptible to apoptosis. Hence, the expression levels of genes associated with apoptosis following RAR β upregulation were investigated. Results from RT-qPCR assays showed that the expression of proapoptotic genes, including *bax*, *bak* and *bim*, were increased >4 fold in pBoBi-RAR β QBC939 cells, while the expression of antiapoptotic genes, including *bcl-2*, *bcl-xL* and *mcl-1*, were all significantly decreased compared with pBoBi-Ctrl cells (Fig. 5A). To investigate these findings further, the association between RAR β expression and caspase-3 activity was explored. Treatment of pBoBi-RAR β cells with the RAR β -activator, BMS453,

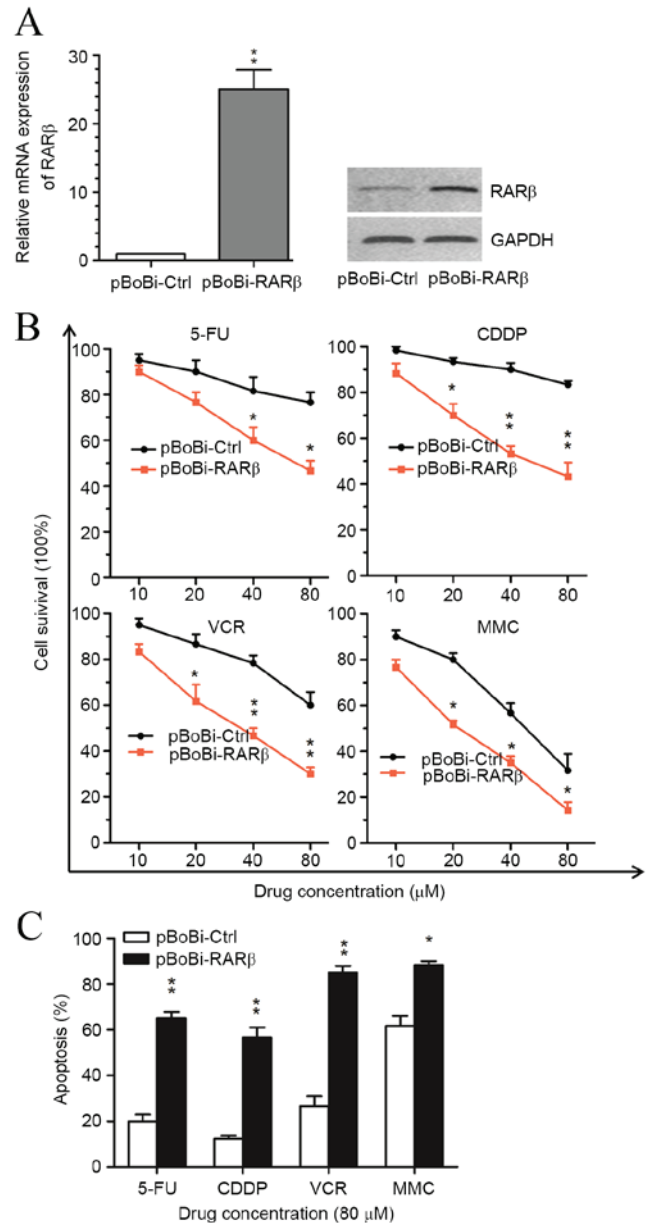


Figure 3. Sensitivity of QBC939 cells to chemotherapeutic agents after RAR β upregulation. Expression of (A) RAR β mRNA and protein in QBC939 cells following stable transfection with RAR β (pBoBi-RAR β) or control (pBoBi-Ctrl) expression vectors. (B) Cell survival analysis of pBoBi-Ctrl and pBoBi-RAR β cells following treatment with 5-FU, CDDP, VCR and MMC chemotherapeutic agents for 48 h. * $P < 0.05$, ** $P < 0.001$ vs. pBoBi-Ctrl. (C) The proportion of apoptotic pBoBi-Ctrl and pBoBi-RAR β QBC939 cells induced by each chemotherapeutic agent. * $P < 0.05$, ** $P < 0.01$ vs. pBoBi-Ctrl. RAR β , retinoic acid receptor- β ; 5-FU, 5-fluorouracil; CDDP, cisplatin; VCR, vincristine; MMC, mitomycin C.

resulted in a 3.5-fold increase in caspase-3 activity compared with pBoBi-Ctrl cells, which exhibited a 1.8-fold increase. The observed increase in caspase-3 activity was further enhanced when cells were cultured under hypoxic conditions or following serum starvation. By contrast, the activity of caspase-3 was significantly decreased in both pBoBi-RAR β and pBoBi-Ctrl cells after treatment with the caspase-3 inhibitor z-VAD-fmk, even when cells were cultured under hypoxic conditions or following serum starvation (Fig. 5B). Furthermore, apoptosis induced by 5-FU, hypoxia or serum

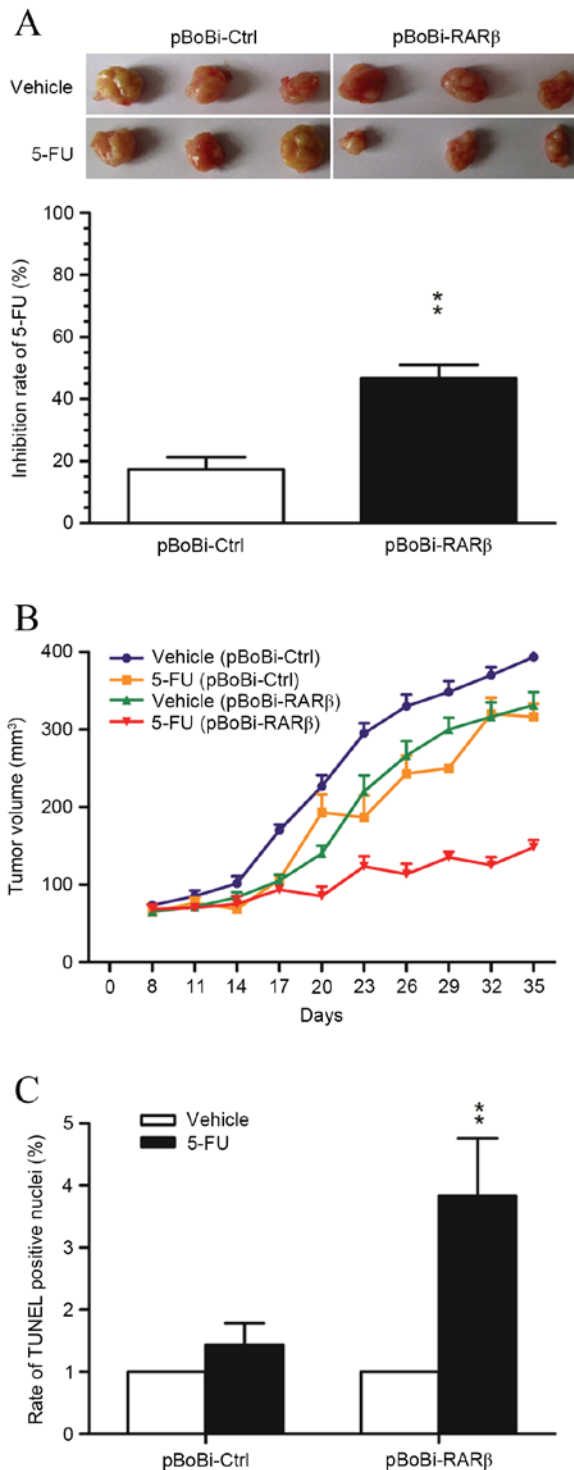


Figure 4. RARβ upregulation contributes to the cytotoxic effect of 5-FU *in vivo*. (A) Tumor images and quantitative analysis of tumor weight show the rate of growth inhibition of CCA xenograft tumors following 5-FU treatment. *P<0.05, **P<0.001 vs. pBoBi-Ctrl. (B) Tumor volume (mm³) of CCA xenograft tumors at 8-35 days post-injection. Tumor latency was considered to be the time taken for the tumor to grow to a size of 80 mm³. (C) The number of apoptotic cells in CCA xenograft tumors was detected using the TUNEL assay. *P<0.05, **P<0.01 vs. pBoBi-Ctrl. RARβ, retinoic acid receptor-β; CCA, cholangiocarcinoma; 5-FU, 5-fluorouracil; TUNEL, TdT-mediated dUTP nick end labeling.

starvation was enhanced upon upregulation or activation of RARβ (Fig. 5C). These results indicate that the susceptibility

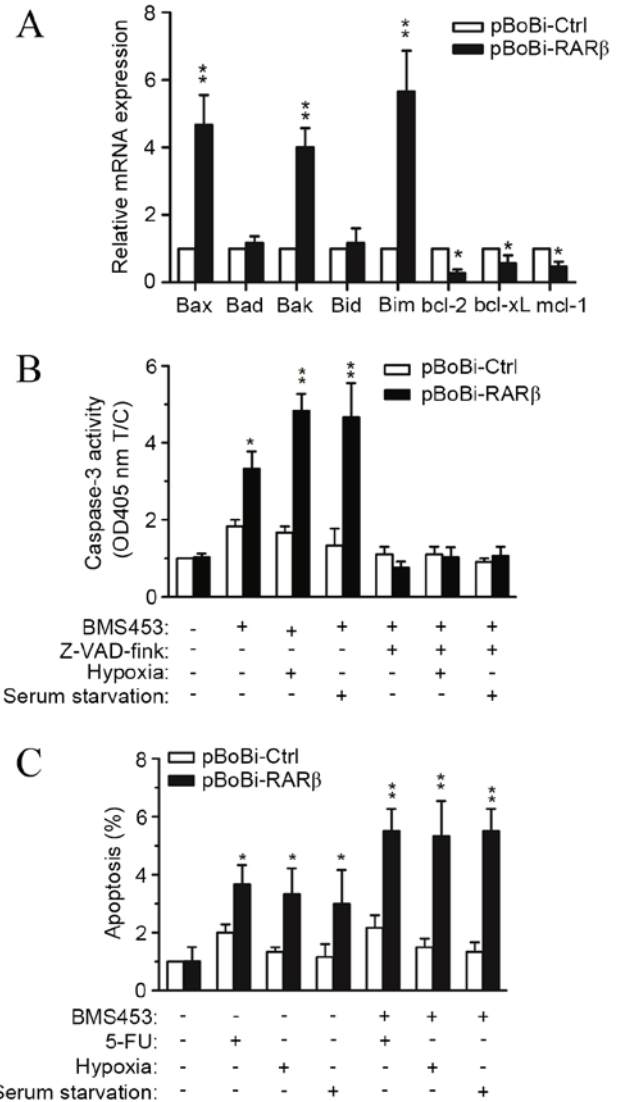


Figure 5. Efficacy of RARβ upregulation on cholangiocarcinoma apoptosis induction. (A) Analysis of apoptosis-associated gene expression by reverse transcription-quantitative polymerase chain reaction in pBoBi-RARβ and pBoBi-Ctrl QBC939 cells. *P<0.05, **P<0.001 vs. pBoBi-Ctrl. (B) Caspase-3 activity within pBoBi-RARβ and pBoBi-Ctrl QBC939 cells following treatment with the RARβ-activator BMS453 (10 μM), the caspase-3 inhibitor Z-VAD-fmk (20 μM), or following hypoxia or serum starvation for 6 h. *P<0.05, **P<0.001 vs. pBoBi-Ctrl. (C) The proportion of apoptotic pBoBi-RARβ and pBoBi-Ctrl QBC939 cells following treatment with BMS453 (10 μM), 5-FU (60 μM) or following hypoxia or serum starvation for 12 h. *P<0.05, **P<0.01 vs. pBoBi-Ctrl. RARβ, retinoic acid receptor-β; 5-FU, 5-fluorouracil.

of CCA cells to caspase-dependent apoptosis may be enhanced by the upregulation of RARβ.

Discussion

The results of the present study indicate the role of RARβ upregulation on the sensitivity of CCA cells to chemotherapeutic agents. Of note, RARβ upregulation was found to increase the sensitivity of CCA cells and tumors to various chemotherapeutic agents currently used for the treatment of CCA. In addition, RARβ upregulation was observed to be associated with an increase in the Bax/Bcl-2 ratio and caspase-3

activity, and rendered CCA cells and xenograft tumors more susceptible to apoptosis. These findings provide evidence to suggest that the upregulation of RAR β may represent a beneficial adjunct therapeutic strategy for the treatment of CCA.

Previous studies have shown that loss of RAR β is a common event in the development of malignant tumors, and the induction of RAR β expression suppresses cancer development (2-7,17). Increased expression of RAR β has been found to correlate with the clinical response to various retinoids (1,18); however, the molecular mechanism of RAR β in regulating sensitivity to chemotherapeutic agents is little known. A major problem in the treatment of human cancer is the lack of response of many tumors to chemotherapeutic agents. Following exposure to anti-cancer agents, tumor cells can alter the expression profile of a specific set of genes to activate and/or suppress signaling networks, which de-sensitizes cells to drug-induced death signals and confers cell survival and chemoresistance (19,20).

Similar to other types of human carcinoma, the results of the present study indicate that CCA, a chemoresistant bile duct carcinoma with a poor prognosis (21), demonstrates loss of RAR β expression. Upregulation of RAR β within the drug-resistant CCA cell line QBC939, which exhibits loss of RAR β expression, was observed to increase the susceptibility of cells to apoptosis induced by chemotherapeutic agents *in vitro* and *in vivo*. Hypoxia and nutrient deficiency are common adverse microenvironments encountered by tumor cells. Based on the presence of specific intracellular signaling pathways, tumor cells can either become more aggressive or undergo apoptosis in response to these microenvironmental stress conditions. Differentially expressed genes, including tumor suppressor genes and oncogenes, are involved in the processes of adaptation. Once upregulated, tumor suppressor genes can initiate apoptosis (22). Consistent with these observations, upregulation of RAR β in the present study was found to induce CCA cells to undergo apoptosis in response to hypoxic and starvation conditions.

It is possible that RAR β upregulation may initiate the process of apoptosis. It is known that the Bcl-2 family of proteins regulates the apoptotic pathway by balancing the expression of proapoptotic (Bax) and antiapoptotic (Bcl-2) factors. In this regard, the Bax/Bcl-2 ratio is suggested to be a useful predictor of apoptotic cell death (23). In the present study, RAR β upregulation was found to induce changes in the expression of genes belonging to the Bcl-2 family, which resulted in an increase in the Bax/Bcl-2 ratio. Caspase-3 functions as an executor of apoptosis by activating DNA fragmentation (24). During the apoptotic processes, cytochrome c is released from the mitochondria and into the cytosol. Cytoplasmic cytochrome c then activates caspase-9, which in turn triggers the activation of caspase-3 and leads to cell death. Increased caspase-3 activity is associated with an increase in the Bax/Bcl-2 ratio (25). Despite the fact that the expression of cytochrome c and caspase-9 was not investigated in the present study, apoptosis induced by RAR β upregulation was observed to be associated with the activation of caspase-3, which led to an increase in the Bax/Bcl-2 ratio.

In conclusion, the data presented in the present study demonstrate a clear functional role for RAR β in sensitizing

CCA cells to chemotherapy-induced cell death. Taken in conjunction with previous studies, it is proposed that the use of adjunct therapies that confer RAR β upregulation should be explored for the treatment of chemoresistant tumors, such as CCA, with the aim of overcoming drug resistance and/or increasing the susceptibility of tumor cells to initiate intrinsic cell death pathways.

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