

Annexin A2 functions downstream of c-Jun N-terminal kinase to promote skin fibroblast cell migration

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Abstract. Delayed healing of skin wounds is one of the outcomes of diabetes mellitus (DM), a condition that affects a significant number of patients worldwide. However, the underlying mechanisms remain unknown. In order to examine proteome alterations in DM, a rat model of type 1 diabetes was developed using streptozotocin injections. The proteomic responses of normal and DM rat skin were analyzed by two-dimensional electrophoresis, and differentially expressed proteins were identified using a liquid chromatography/mass spectrometry system. DM induced 36 and repressed 41 differentially expressed proteins, respectively. Altered proteins were involved in a number of biological processes, including RNA and protein metabolism, the tricarboxylic acid cycle, glycolysis, cytoskeleton regulation, hydrogen detoxification and calcium-mediated signal transduction. In addition, overexpression of annexin A2, one of the signaling proteins altered by DM, accelerated the rate of human skin fibroblast cell migration. Application of SP600125, an inhibitor of a key regulator of cell migration c-Jun N-terminal kinase (JNK), inhibited the migration of normal cells. By contrast, SP600125 treatment did not inhibit the migration of annexin A2-overexpressed cells, indicating that annexin A2 may function downstream of JNK. In conclusion, the results of the present study reveal the potential proteomic responses to DM in skin tissues, and

demonstrate a positive functional role of annexin A2 in fibroblast cell migration.

Introduction

Diabetes mellitus (DM) is one of the most severe metabolic diseases, which affects >170 million people worldwide (1). A major symptom of DM is an increased blood sugar level, which results in a number of additional complications, such as skin ulcers, which affect ~15% of DM patients (1). In addition, DM affects angiogenesis, which leads to the delay of wound healing (2); however, the underlying mechanisms remain unclear.

Wound healing is a complex process that requires the cooperation of keratinocytes, fibroblasts, endothelial cells, macrophages and platelets (3). Fibroblast cell proliferation and migration is known to serve an important role in the wound repair process, as well as in collagen deposition and remodeling, wound contraction, angiogenesis and the formation of extracellular matrix (ECM) (3). It is possible that poor DM-associated wound healing may be due to altered protein and lipid metabolism and the abnormal formation of granulation tissue (4). Increased blood sugar levels alter glycosylation enzymes and induce covalent binding of aldose sugars to a protein or lipid. Binding of aldose sugar to a protein or lipid produces a stable product, termed advanced glycation end-product, which accumulate on the surface of the cell membrane and ECM proteins leading to delayed turnover rates (4). Reactive oxygen species (ROS) accumulate in high quantities in the skin of diabetic rats (5). In addition, nitric oxide (NO) is a known signaling molecule that stimulates fibroblast cell proliferation, maturation, differentiation and collagen production during the wound healing process (6). In fibroblasts from diabetic ulcers, the cells often possess a dilated endoplasmic reticulum, an increased number of vesicular bodies and defected micro tubular structures (7). These subcellular alterations may be induced by activated protein synthesis and turnover, as well as defects in microtubule structure thus inhibiting protein secretion via autonomous trafficking through vesicles and induction of microtubules to release their products into the extracellular space in diabetic ulcer fibroblasts (7,8). Diabetic ulcers exhibit defective fibroblast proliferation, which may be due to reduced ECM protein production, and delayed wound contraction and

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healing (8). However, a limited number of biochemical studies have been performed to date, that have investigated the mechanisms underlying DM-mediated delay of wound healing (5-8).

Fibroblast cell migration is an important step during the wound closure process, and the phosphoinositide 3-kinase (PI3K)-Ras-related C3 botulinum toxin substrate 1-c-Jun N-terminal kinase (JNK) signaling pathway is important in the promotion of cell migration (9). In a recent study, RNA-Seq analysis was performed using high-glucose stimulated human foreskin fibroblasts, in order to examine the response of a large population of transcripts to high glucose-induced stress (9). The results demonstrated that the inflammatory response and Wnt signaling pathways generated negative and positive responses to high glucose-induced stress, respectively (10). Xuan *et al* (10) revealed that nuclear factor- κ B functions independently of PI3K signaling to activate JNK, resulting in accelerated fibroblast cell migration.

The present study investigated DM-induced alterations in a large number of proteins in diabetic rat skin using two-dimensional (2-D) gel electrophoresis. Proteomic analysis and liquid chromatography/mass spectrometry (LC/MS) were performed to identify the differentially expressed proteins. In addition, the effect of annexin A2 on human foreskin fibroblast cell migration was analyzed, as well as the potential association between annexin A2 and JNK, a key regulator of cell migration. The present study provides useful proteomic information that may enhance what is currently known regarding the mechanisms underlying skin wound closure.

Materials and methods

Generation of skin wounds in diabetic rats. A rat model of type 1 diabetes was generated using a previously described method (11). A total of 12 male Sprague-Dawley rats (age, 8 weeks; weight, 150 to 200 g) were housed in cages (12 h light/12 h dark cycle; free access to water and food) at 25°C and 30% humidity, divided into 2 groups (n=6/group) and were injected intraperitoneally with *streptozotocin* (STZ; Sigma-Aldrich; Merck KGaA, Darmstadt, Germany; 50 mg/kg) which was dissolved in sodium citrate buffer (pH 4.5) (12). STZ-injected rats with blood glucose levels >16.7 mmol/l were considered to have developed DM. To generate the control group, the same volume of sodium citrate without STZ was injected into the male rats. To generate skin wounds, the rats were anesthetized with pentobarbital (Sigma-Aldrich; Merck KGaA; 45 mg/ml), and two circular wounds (~200 mm²) that were 1 cm deep were created on the lower back of each rat. Ethical approval was obtained from the ethics committee of Wenzhou Medical University (Wenzhou, China).

Analysis of wound healing. Following 8 weeks of DM onset, the wound area was evaluated every 4 days for 16 days. The skin wounds from 3 diabetic and 3 control rats were photographed with rulers, and the area of the unhealed regions was calculated using these images together with TINA 2.0 software (DesignSoft, Inc., Budapest, Hungary; <http://www.designsoftware.com/>) (11).

Protein preparation and 2-D SDS-PAGE analysis. Following anesthesia with pentobarbital (45 mg/ml; Sigma-Aldrich;

Merck KGaA), the dorsal area of diabetic rats was totally depilated with Na₂S (8.0%, w/v; Sigma-Aldrich; Merck KGaA) and two full-thickness circular wounds (~300 mm² each) were created on the lower back of each rat using a pair of sharp scissors and a scalpel. Skin tissues (2 g) that were 1-cm deep were collected from the diabetic and control rats for protein extraction 2 weeks following the establishment of DM. The following reagents were used for protein extraction: PBS, 9.5 M urea, 0.1% (w/v) DTT, 2% (w/v) CHAPS and 0.8% (w/v) pharmalyte (Sigma-Aldrich; Merck KGaA). The extracts were dissolved in immobilized pH gradient (IPG) rehydration buffer [8 M urea, 2% (w/v) 3-[(3-cholamidopropyl) dimethylammonio]-1-propanesulfonate hydrate, 2% (w/v) IPG buffer, 0.04 M dithiothreitol, 1X nuclease solution and 0.1% (w/v) of bromophenol blue] prior to isoelectric focusing electrophoresis. The following reagents were used for 2-D gel analysis: 1% DTT, 50 mM Tris-HCl, pH 8.8, 6 M urea, 30% (v/v) glycerol, 2% (w/v) SDS, 0.1% (w/v) bromophenol blue, equilibration buffer (2.5% (w/v) iodoacetamide (Sigma-Aldrich; Merck KGaA). The MS/MS spectra were processed using Proteome Discoverer software (version 1.3; Thermo Fisher Scientific, Inc., Waltham, MA, USA) and the database search was performed using the Mascot search engine (Matrix Science Mascot version 2.3; <http://www.matrixscience.com/>) against a concatenated forward-decoy approach (13).

In-gel digestion. Protein spots were visualized using ImageMaster 2D Platinum version 6.0 (GE Healthcare Bio-Sciences, Pittsburgh, PA, USA). A total of 77 protein spots of interested were excised and digested using a 1% protease cocktail blue (Sigma-Aldrich; Merck KGaA), which were further analysed using MS. In-gel digestion was performed in 3 steps according to the procedure described by Russell *et al* (14). The following reagents were used for analysis: 50% (v/v) acetonitrile (ACN), 10 mM DTT containing 100 mM ammonium bicarbonate and 1% trypsin solution (Sigma-Aldrich; Merck KGaA).

LC-MS/MS analysis and bioinformatics analysis. The proteins extracted from the gel sections were digested using 1% trypsin solution (Sigma-Aldrich; Merck KGaA) and 4 μ l of samples were submitted to online nanoflow liquid chromatography using the easy-nano LC system (Proxeon Biosystems; Thermo Fisher Scientific, Inc.) with 10 cm capillary columns of an internal diameter of 75- μ m, filled with 3- μ m Reprosil-Pur C18-A2 resin (Dr. Maisch GmbH, Ammerbuch-Entringen, Germany) (14). The gradient consisted of 10-30% (v/v) ACN in 0.1% (v/v) formic acid at a flow rate of 200 nl/min for 45 min, 30-100% (v/v) ACN in 0.1% (v/v) formic acid at a flow rate of 200 nl/min for 1 min and 100% CAN in 0.1% formic acid at a flow rate of 200 nl/min for 10 min. The elution was electrosprayed through a Proxeon nanoelectrospray ion source by (electrospray ionization) ESI-MS/MS analysis on a Thermo Fisher LTQ Velos Pro (Thermo Fisher Scientific, Inc.) using full ion scan mode over the m/z range 200-1800. Collision-induced dissociation (CID) was performed in the linear ion trap using a 4.0-Th isolation width and 35% normalized collision energy with helium as the collision gas. Five independent MS/MS scans were performed on each ion using dynamic exclusion. The precursor ion that was selected for

CID was dynamically excluded from further MS/MS analysis for 30 sec. Further corresponding protein identification was performed using the Swiss-Prot protein sequence database (version 54.5; http://web.expasy.org/docs/swiss-prot_guide-line.html). LC-MS/MS and bioinformatics analyses were performed by LC Bio, Inc. (Hangzhou, China).

Western blot analysis. Skin tissues (100 mg) were ground using liquid nitrogen and an ice-cold lysis solution (7 M urea, 2 M thiourea, 2% CHAPS, 40 mM Tris base, 40 mM dithiothreitol, and 1% protease inhibitor; Sigma-Aldrich; Merck KGaA) was added to procure whole cell extracts. Total proteins (20 μ g) from skin samples with or without induced diabetes 2 weeks prior, were separated on a 12% SDS-PAGE gel and electro-transferred to Immobilon®-P PVDF Transfer Membranes (EMD Millipore, Billerica, MA, USA). The membranes were blocked in 1X Tris-buffered saline containing 5% skim milk and 0.05% Tween-20 for 2 h. All of the reagents for SDS gel electrophoresis and membrane blocking were purchased the Sigma-Aldrich; Merck KGaA. To analyse protein levels in the samples the membranes were incubated with the following primary antibodies at room temperature (25°C) for 2 h: Anti-eNOS antibody (cat. no. ab76198; 1:2,000, Abcam, Cambridge, MA, USA), anti-cleaved (c)-caspase-3 antibody (cat. no. ab2302; 1:2,000, Abcam), anti-plasminogen activator inhibitor (PAI) -1 antibody (cat. no. ab125687; 1:2,000; Abcam), anti-cluster of differentiation (CD) 34 antibody (cat. no. ab81289; 1:2,000; Abcam), anti-collagen1 antibody (cat. no. ab34710; 1:2,000; Abcam), anti-annexin A2 antibody (cat. no. ab41803; 1:1,000; Abcam), anti-JNK antibody (cat. no. ab179461; 1:2,000; Abcam), anti-phosphorylated (p) -JNK antibody (cat. no. ab4821; 1:2,000; Abcam) and anti-GAPDH antibody (cat. no. ab8245; 1:2,000; Abcam). The membranes were then incubated with either the anti-mouse or anti-rabbit horseradish peroxidase-conjugated secondary antibody (cat. no. 7074; 1:2,000; Cell Signaling Technology, Inc., Danvers, MA, USA) at room temperature (25°C) for 2 h. Antigen-antibody complexes were visualized using an electrochemiluminescence kit (BioTrand, Crystal Lake, Illinois, USA). Protein levels were normalized against GAPDH and protein expression was analyzed using Image J2 version 2.0 software (13).

Human foreskin fibroblast cell culture. Human fibroblast cell culture was performed as previously described (5). Human foreskin tissues were collected from 3 patients diagnosed with redundant prepuce (all three patients were healthy without DM or other disease diagnoses; ages, 26, 30 and 32 years old) who were admitted in January 2014 to the Department of Dermatology at the First Affiliated Hospital of Wenzhou Medical University (Wenzhou, China). The present study was approved by the ethics committee of Wenzhou Medical University, and written informed consent was obtained from all patients involved. The fat was first removed from the tissue samples, when were subsequently cut into 3x2-mm strips and incubated overnight at 4°C in 0.05% Dispase I (Sigma-Aldrich; Merck KGaA). The epidermis was then removed and the dermis was placed into 25 cm² flasks pre-treated with foetal bovine serum (FBS; HyClone; GE Healthcare Life Sciences, Logan, UT, USA). The flasks were positioned horizontally for 1 h, and then vertically for 3 h

in a culture chamber at 37°C with 5% CO₂. The cells were cultured in Dulbecco's modified Eagle's medium (DMEM; HyClone; GE Healthcare Life Sciences) containing 5.5 mM glucose (Sigma-Aldrich; Merck KGaA), 10% FBS and 1% penicillin-streptomycin (Gibco; Thermo Fisher Scientific, Inc.); the medium was refreshed every 3 days. When cell confluence had reached 70 to 80% the cells were digested and passaged using 0.25% trypsin (Gibco; Thermo Fisher Scientific, Inc.). Cells were cultured for 3 days in 5.5 mM glucose medium, before they were transferred to media with or without 25 μ M SP600125 (Cell Signaling Technology, Inc.) for up to 1 day prior to the migration assay, or for 1 h prior to western blot analysis.

Overexpression of annexin A2 in fibroblast cells. For over-expression of annexin A2, the open reading frame (ORF) sequences of the annexin A2 gene (ANXA2; GenBank Accession no. NM_001002858.2; National Center for Biotechnology Information, <https://www.ncbi.nlm.nih.gov/>) was amplified for 28 cycles of 95°C for 30 sec, 60°C for 30 sec and 72°C for 90 sec using the following primers: Forward, 5'-ATCGTAGGATCCATGGGCCGCCAGCTAG-3', and reverse, 5'-AGCTATTCTAGATCAGTCATCTCCACCACA CAGGTA-3'. The amplified polymerase chain reaction (PCR) fragments were then cloned into the *Bacillus amyloliquefaciens* and *Xanthomonas badrii* restriction enzyme sites of the pcDNA3 expression vector (Invitrogen; Thermo Fisher Scientific, Inc.). Primer design and sequencing were performed by Sangon Biotech Co., Ltd. (Shanghai, China; <http://www.sangon.com/>). A total of 2 or 3 μ g pcDNA3-ANXA2 plasmid was transformed into human foreskin primary fibroblasts from the 3 patients using a Lipofectamine 2000 kit (Invitrogen; Thermo Fisher Scientific, Inc.) according the manufacturer's instructions. Empty vector pcDNA3 using Lipofectamine 2000 transformed cells was used as control. The 2 and 3 μ g of pcDNA3-ANXA2 plasmid transformed cells were termed OX1 and OX2, respectively. Following 2 days of transformation, annexin A2 levels in the fibroblast cells were examined by western blotting, and cell migration tests were performed.

Cell migration assay. Primary fibroblast cells were seeded onto 6-well plates at 80-90% confluency and incubated at 37°C overnight in DMEM containing 0.5% FBS and 5 μ g/ml mitomycin-C (Sigma-Aldrich; Merck KGaA). Linear scratch wounds were subsequently created in the confluent fibroblast monolayer using a sterile 200 μ l pipette tip (Ningbo MFLab Medical Instruments Co., Ltd., Ningbo, Zhejiang, China). The medium was immediately replaced with prewarmed (37°C) fresh DMEM containing 0.5 % FBS and 5 μ g/ml mitomycin-C. At 0, 12 and 24 h following wound establishment, images were captured using a light microscope (IX70; Olympus Corporation, Tokyo, Japan) equipped with a CCD camera (CoolSNAP HQ; Nippon Roper, Toyko, Japan), which was controlled by MetaMorph 7.1 software (Molecular Devices, LLC, Sunnyvale, CA, USA). To quantify cell migration, 10 cells on the border of the wound area were randomly selected from each well and the migration distance was measured using ImageJ version 14.8 software (<https://imagej.nih.gov/ij/>) at the indicated time points.

Statistical analysis. Statistical analyses were performed using GraphPad Prism software (version 5.0; GraphPad Software, Inc., La Jolla, CA, USA). Comparisons between 2 groups were performed using a Student's *t*-test, while significant differences among 3 groups were analysed using one-way analysis of variance, followed by Bonferroni's multiple comparison test. $P < 0.05$ was considered to indicate a statistically significant difference.

Results

DM impairs skin wound healing. Diabetes-induced skin wound healing was analyzed in a rat model of diabetes. STZ, an inducer of type 1 diabetes was used to induce diabetes in rats, and wound healing was assessed by monitoring and comparing the size of ~200 mm² circular wounds created on the waist of each rat at 8 weeks following the onset of diabetes (8). Wound repair was then analysed over the course of 16 days, and the results indicated that the rate of wound healing was significantly reduced in diabetic rats when compared with non-diabetic normal rats (Fig. 1).

Proteomic analysis of rat skin tissues under DM stress. In order to gain an improved understanding of the molecular mechanisms underlying DM-mediated impairment of skin wound healing, comparative proteomic assays were performed to identify differentially expressed proteins in the skin of DM rats and normal rats using 2-D gel electrophoresis. Total skin protein extracts were first subjected to 2-D gel analysis followed by identification by LC/MS. This led to the detection of proteins altered by DM. Proteins extracted from the normal rat skin were used as a control (Fig. 2). Optimization of the 2-D gels and image processing identified 36 and 41 protein spots (representing differentially expressed proteins) that were induced and repressed by DM, respectively (Fig. 2). The experiments were repeated 3 times and only the reproducible alterations were taken into account. To identify differentially expressed proteins, the spots were excised from the 2-D gels and identified by LC/MS and trypsin digestion. The results presented in Table I demonstrate that 72 proteins were identified among the 77 spots observed in the 2-D gel (Fig. 2). The major group of proteins identified, either overexpressed or repressed, were observed to be involved in regulation of the cytoskeleton (tubulin and actin formation), carbohydrate metabolism (glycolysis and the tricarboxylic acid cycle) and protein folding. In the present study, the calcium binding proteins, calmodulin and calreticulin, were induced in DM (Table I). Formation of the ECM is an important step in the wound healing process (3). DM reduced the levels of collagen $\alpha 1$ and dermatopontin, two ECM-associated proteins. In addition to the major group of proteins, RNA metabolism, adenosine triphosphate (ATP) synthesis and cell migration-associated protein levels were altered (Table I).

Protein expression was altered in DM rat skin. To verify the results identified by proteomic assay, western blot analysis was performed using specific antibodies to detect select marker proteins in the pathways identified. The results revealed that the expression levels of CD34, collagen-1 and endothelial NO synthase were decreased (NO synthesis), whereas PAI-1

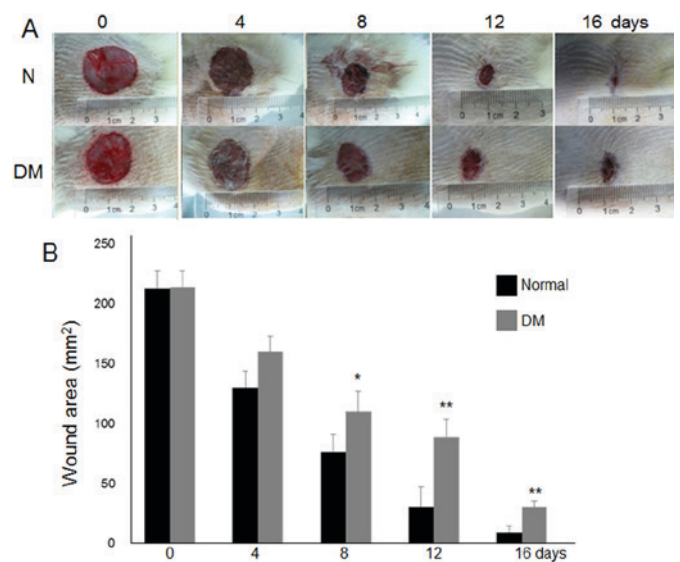


Figure 1. Effect of DM on the rate of skin wound repair in rats. (A) Representative images of skin wounds from normal and DM rats. (B) The wound area in normal and DM rats at each time point. Wound areas were measured using TINA 2.0 software. * $P < 0.05$ and ** $P < 0.05$ vs. control. DM, diabetes mellitus; N, normal group.

(inflammation), c-caspase3 (apoptosis) and annexin A2 were increased in DM rat skin when compared with normal rat skin (Fig. 3). These results were similar to those identified by the proteomic analysis.

Overexpression of annexin A2 promotes human foreskin fibroblast cell migration. Annexin A2 was induced in DM rat skin (Fig. 3), and the transcript of its orthologue gene in humans was previously demonstrated to be induced by high-glucose treatment of foreskin fibroblast cells (9). Therefore, the function of annexin A2 in fibroblast cells was examined further, by forced overexpression of annexin A2. Human *ANXA2* (GenBank Accession no. NM_001002858.2) ORF sequences were amplified by PCR, and the PCR products were then cloned into the pcDNA3 expression vector. Following sequencing, 2 and 3 μ g pcDNA3-*ANXA2* plasmid were transformed into human foreskin primary fibroblast cells using the Lipofectamine 2000 system to generate two independent transformants, termed OX1 and OX2, respectively. Compared with the control cells (untransformed), OX1 and OX2 cells exhibited a significantly increased cell migration rate following wound generation (Fig. 4A and B). To verify the expression levels of annexin A2, western blot analysis was performed. The results demonstrated that the level of annexin A2 was increased in OX1 and OX2 when compared with the controls (Fig. 4C). In addition, annexin A2 overexpression maintained an increased cell migration rate when compared with the control cells when they were cultured in high-glucose containing medium (data not shown). However, the level of p-JNK, a key regulator of cell migration, was not altered by the overexpression of annexin A2 (Fig. 4C).

Annexin A2 functions downstream of JNK. As annexin A2 overexpression was not observed to alter p-JNK protein

Table I. Differentially expressed proteins identified by liquid chromatography/mass spectrometry analysis.

A, Proteins highly expressed in DM skin

Spot no.	UniProt ID	Protein	Function
1	P69897	Tubulin β -5 chain	Cytoskeleton
8	P85108	Tubulin β -2A chain	Cytoskeleton
9	Q4QRB4	Tubulin β	Cytoskeleton
13	P48675	Desmin	Cytoskeleton
16	O35763	Moesin	Cytoskeleton
17	O35763	Moesin	Cytoskeleton
2	P18418	Calreticulin	Ca ²⁺ binding
4	P18418	Calreticulin	Ca ²⁺ binding
12	P43742	Reticulocalbin-3	Ca ²⁺ binding
34	P62161	Calmodulin (inflammation and apoptosis)	Ca ²⁺ binding
3	P06761	78 kDa glucose-regulated protein	Protein folding and oligomerization
7	P04785	Protein disulfide-isomerase	Protein folding
19	P11598	Protein disulfide-isomerase A3	Protein folding
36	P11598	Protein disulfide-isomerase A3	Protein folding
24	P34064	Proteasome subunit α type-5	Protein degradation
14	G3V9R8	Heterogeneous nuclear ribonucleoprotein C1/C2	RNA metabolism
6	P61980	Heterogeneous nuclear ribonucleoprotein K	RNA metabolism
22	B5DEN5	Elongation factor 1- β	RNA metabolism
33	P02401	60S acidic ribosomal protein P2	RNA metabolism
35	P63324	40S ribosomal protein S12	RNA metabolism
20	P62630	Elongation factor 1- α 1	RNA metabolism
5	P33840	Antithrombin-III	Plasma protease inhibitors
10	P10719	ATP synthase subunit β	ATP synthesis
11	P31000	Vimentin	Signaling
23	P20332	14-3-3 protein σ	Signaling
25	Q79342	14-3-3 protein σ	Signaling
15	Q9WTT6	Guanine deaminase	Purine metabolism
18	P04764	α -enolase	Carbon metabolism
21	P02650	Apolipoprotein E	Carbon metabolism (inflammation)
29	P04797	Glyceraldehyde-3-phosphate dehydrogenase	Carbon metabolism
30	P04797	Glyceraldehyde-3-phosphate dehydrogenase	Carbon metabolism
31	P04797	Glyceraldehyde-3-phosphate dehydrogenase	Carbon metabolism
26	Q07936	Annexin A2	Apoptosis and inflammation
27	P63029	Translationally-controlled tumor protein	Tumor protein
28	P23764	Glutathione peroxidase 3	Superoxide
32	Q04374	Ig κ chain V-II region 26-10	Immune system

B, Proteins with reduced expression in DM skin

37	P02454	Collagen α -1	Tissue connection
68	B2RZ77	Dermatopontin	ECM matrix
38	P04785	Protein disulfide-isomerase	Protein folding
40	P04785	Protein disulfide-isomerase	Protein folding
77	P73842	Peptidyl-prolylcis-trans isomerase A	Protein refolding
65	P40112	Proteasome subunit β type-3	Protein degradation
42	P39423	Lamin-A/C	Gene regulation
47	P70564	Serpin B5	Gene regulation
61	Q9WUH4	Four and a half LIM domains protein 1	Signaling
44	P60711	Actin, cytoplasmic 1	Cytoskeleton

Table I. Continued.

B, Proteins with reduced expression in DM skin			
Spot no.	UniProt ID	Protein	Function
48	Q4FZT0	Stomatin-like protein 2	Cytoskeleton
49	Q6AYC4	Macrophage-capping protein	Cytoskeleton
71	Q4KLF8	Actin-related protein 2/3 complex subunit 5	Cytoskeleton
66	P02600	Myosin light chain 1/3	Cytoskeleton
67	P13413	Troponin I	Cytoskeleton
72	P45592	Cofilin-1	Cytoskeleton
73	P04466	Myosin regulatory light chain 2	Cytoskeleton
74	P16409	Myosin regulatory light chain	Cytoskeleton
45	P15429	β -enolase	Carbon metabolism
46	P05065	Fructose-bisphosphate aldolase A	Carbon metabolism
43	P04764	α -enolase	Carbon metabolism
50	P12007	Isovaleryl-CoA dehydrogenase	Carbon metabolism
51	P16617	Phosphoglycerate kinase 1	Carbon metabolism
52	P41562	Isocitrate dehydrogenase	Carbon metabolism
53	P17764	Acetyl-CoA acetyltransferase	Carbon metabolism
55	P05065	Fructose-bisphosphate aldolase A	Carbon metabolism
56	P07943	Aldose reductase	Carbon metabolism
57	P04797	Glyceraldehyde-3-phosphate dehydrogenase	Carbon metabolism
58	P04636	Malate dehydrogenase	Carbon metabolism
62	P15429	β -enolase	Carbon metabolism
69	P39069	Adenylate kinase isoenzyme 1	ATP synthesis
59	A7VJC2	Heterogeneous nuclear ribonucleoproteins A2/B1	RNA metabolism
60	P55260	Annexin A4	Apoptosis and inflammation
63	P63029	Translationally-controlled tumor protein	Tumor protein
64	A48593	Glutathione S-transferase P1	Superoxide
75	P02625	Parvalbumin α	Ca ²⁺ -binding

Differentially expressed proteins were identified following optimization of two-dimensional gels and image processing. The spot number refers to the corresponding protein on the gel images presented in Fig. 2. DM, diabetes mellitus; ID, identification number; ATP, adenosine triphosphate, ECM, extracellular matrix.

expression levels, SP600125, an inhibitor of JNK was used to analyze its effect on annexin A2 expression. SP600125 significantly inhibited fibroblast cell migration (Fig. 5A and B), which is consistent with results presented in a previous study (11). In addition, SP600125 suppressed p-JNK expression while maintaining normal total JNK expression levels (Fig. 5C). SP600125 treatment marginally repressed annexin A2 protein expression levels (Fig. 5C).

Due to the observed inhibitory effects of SP600125 on cell migration and annexin A2 expression levels, its effects were further examined in OX1 fibroblast cells over-expressing *ANXA2*. SP600125+OX1 was associated with a significant increase in cell migration when compared with SP600125-treated controls at 12 and 24 h. In addition, SP600125 treatment did not alter the expression levels of annexin A2 in OX1 transformed cells (Fig. 6A and B). Western blotting results demonstrated that SP600125 markedly suppressed p-JNK levels in OX1 cells (Fig. 6C). Collectively, these results indicate that annexin A2 may

function downstream of JNK to promote fibroblast cell migration.

Discussion

One of the strategies employed by living organisms to adapt to environmental alterations, is the rapid reprogramming of transcriptional and translational regulation via cell signalling mechanisms. Therefore, the analysis of proteomic alterations following exposure to specific stressors is a suitable strategy to understand its regulatory basis (9,10). In the present study, proteomic analysis was performed on DM rat skin to identify global alterations in the proteome. The proteomic results demonstrated that the expression of a number of important proteins involved in metabolic processes was significantly altered.

Cell migration, a key process in wound healing, is multi-step process. During the migration process a protrusion extends from one cell to attach to a nearby leading edge

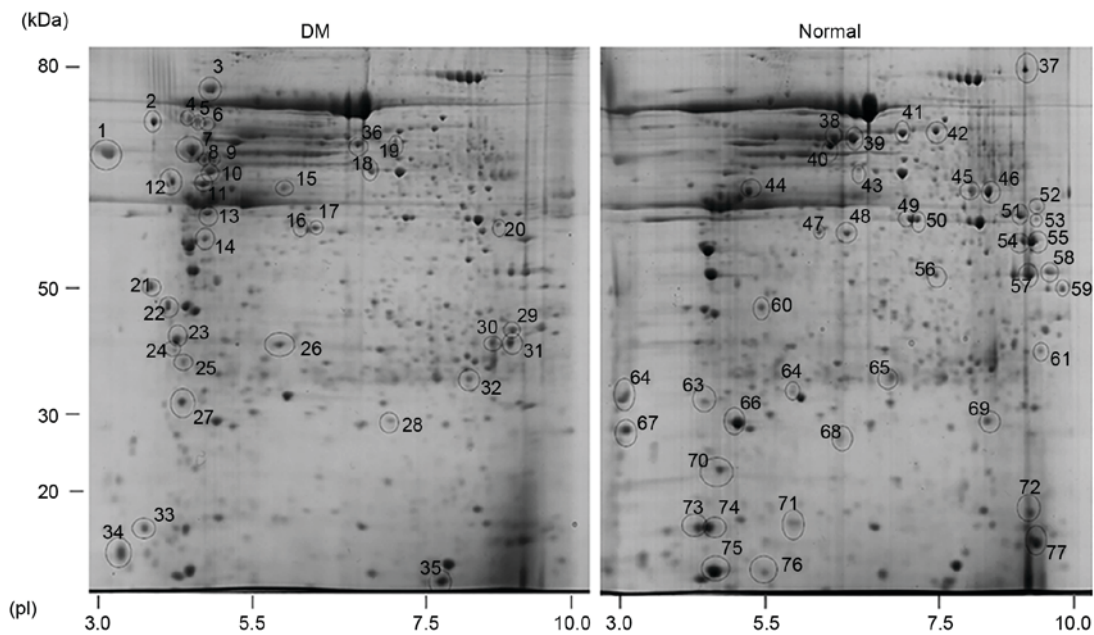


Figure 2. Comparison of 2-D gel images of the total protein extracts from normal and DM rat skins. Total protein extracted from the normal rat (right panel) and DM rat (left panel) skins were separated using a 2-D gel system and detected by silver staining. The 77 differentially expressed proteins identified in the normal and DM rat skins are indicated by circles and a corresponding number. 2-D, two-dimensional; DM, diabetes mellitus; pl, isoelectric point.

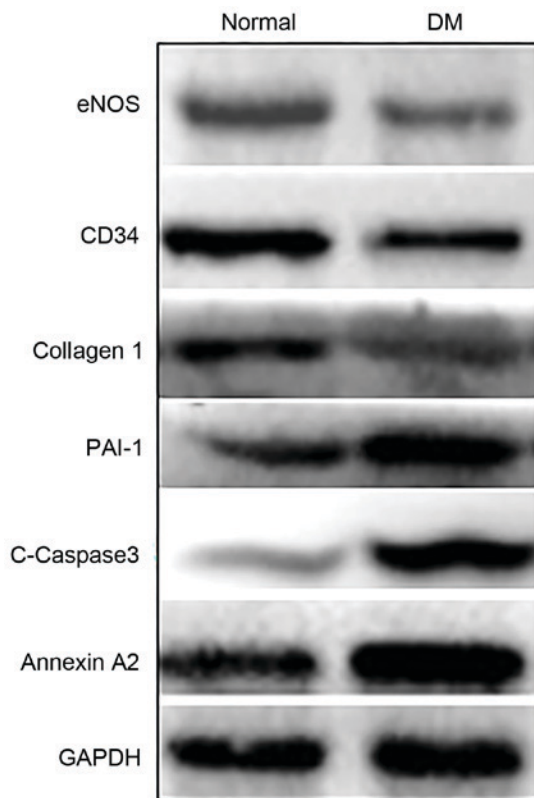


Figure 3. Western blot analysis of protein marker expression. Proteins extracted from the skin of normal and DM rats were used to analyse the expression of eNOS, CD34, annexin A2, collagen1, c-caspase-3 and PAI-1. GAPDH was used as a loading control. DM, diabetes mellitus; eNOS, endothelial nitric oxide synthase; CD34, cluster of differentiation 34; c-caspase-3, cleaved caspase-3; PAI-1, plasminogen activator inhibitor.

of another cell protrusion (3). The cells then migrate forward and release their adhesions and retractions at the rear of the

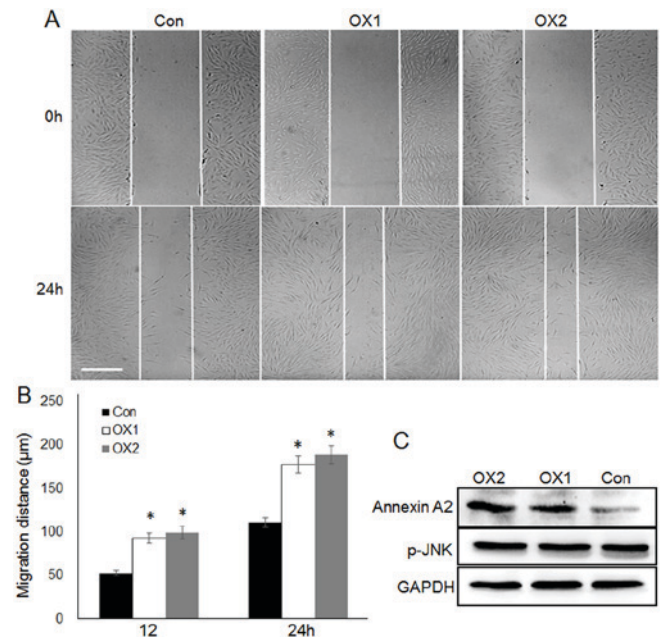


Figure 4. Effect of annexin A2 overexpression on human fibroblast cell migration. (A) A wound healing assay was performed to analyse the effects of annexin A2 overexpression on fibroblast cell migration (scale bar, 500 μ m). OX1 and OX2 represent human fibroblast cells transformed with 2 and 3 μ g pcDNA3-ANXA2 plasmids, respectively. (B) Quantification of cell migration distance. Data represent the mean \pm standard error of 10 replicates. Significant differences between non-transformed and annexin A2-overexpressed cells are indicated; * $P < 0.01$ vs. control. (C) Annexin A2 and p-JNK levels were analysed by western blot analysis. GAPDH was used as a loading control. ANXA2, annexin A2 gene; p-JNK, phosphorylated-c-Jun N-terminal kinase; Con, untransformed control cells.

cell (15-17). The actin-mediated Wiskott-Aldrich syndrome protein-family verprolin homologous protein complex is important in the formation of the lamellipodial protrusions at

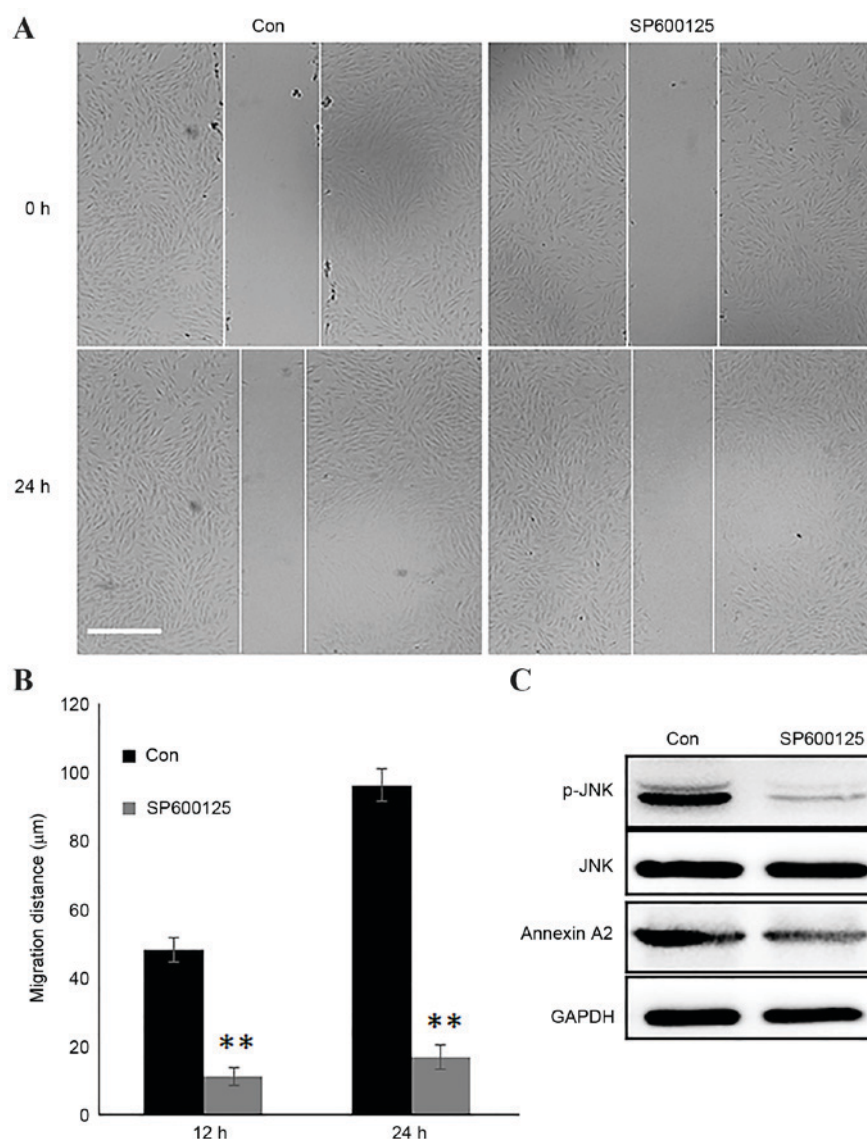


Figure 5. Effect of SP600125 on fibroblast migration and annexin A2 levels. (A) A wound healing assay was performed to analyse the effects of SP600125, an inhibitor of p-JNK, on fibroblast migration (scale bar, 500 μ m). (B) Quantification of cell migration distance. (A) Data represent the mean \pm standard error of 10 replicates. The significant differences between SP600125-treated and untreated cells are indicated; ** $P < 0.001$ vs. control. (C) p-JNK, JNK and annexin A2 levels were analysed by western blot analysis. GAPDH was used as a loading control. p-JNK, phosphorylated-c-Jun N-terminal kinase; Con, untreated control cells.

the leading edge of the migrating cells (18), and the Rho family guanosine triphosphatase protein, cell division control protein 42, reportedly regulates the polarity of migrating cells (19). In the present study, cytoskeleton-associated proteins were part of a major group of proteins regulated by DM in rat skin. Actin and the actin-associated protein complex were repressed by DM, as were tubulin, myosin, troponin and additional cytoskeleton-associated proteins. The ECM serves important roles in cell adhesion, communication between nearby cells, and during differentiation induced by the deposition of extracellular molecules secreted by cells (20). In the present study, collagen $\alpha 1$ and dermatopontin, an ECM-associated protein, were repressed in DM while moesin, a protein involved in cell-to-cell recognition, was induced. A previous study identified protein alterations induced by DM that involved the superoxide detoxification process (glutathione peroxidase 3 and glutathione S-transferase P1) and ROS accumulation in

rat skin (5), indicating that DM-mediated ROS accumulation may be induced via inhibition of the superoxide detoxification pathway. In addition, DM regulation in carbon metabolism and the ATP synthesis process may affect the supply of carbon skeletons and energy to the cell, which may in turn inhibit wound closure (21).

Annexin A2, a member of the annexin protein family, has been reported to induce cell migration and neoangiogenesis in breast tumors (22). In addition, annexin A2 reportedly regulates intestinal epithelial cell spreading and wound closure (23). In the present study, annexin A2 was overexpressed while annexin A4 was repressed in DM. Western blot analysis confirmed the increased levels of annexin A2 in DM rat skin when compared with normal controls. Similarly, a previous transcriptomics study revealed the induction of *ANXA2* by high-glucose in human foreskin fibroblast cells (9). Overexpression of *ANXA2* in human foreskin primary

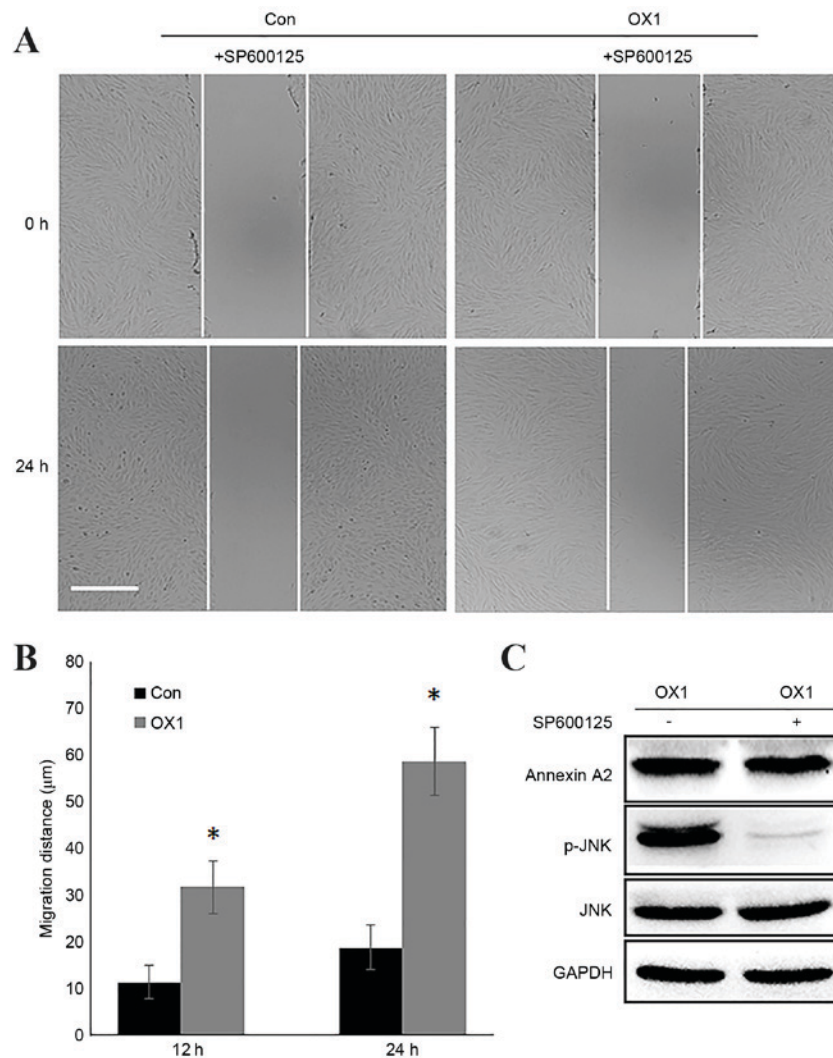


Figure 6. Effect of SP600125 on migration of cells overexpressing annexin A2. (A) A wound healing assay was performed to analyse the effects of SP600125, an inhibitor of p-JNK, on control and cells overexpressing annexin A2 (scale bar, 500 μ m). (B) Quantification of cell migration distance. Data represent the mean \pm standard error of 10 replicates. Significant differences between SP600125-treated and untreated cells are indicated; * $P < 0.05$ vs. control. (C) Annexin A2, p-JNK and JNK levels were analysed by western blot analysis. GAPDH was used as a loading control. p-JNK, phosphorylated-c-Jun N-terminal kinase; Con, untreated control cells; OX1, human fibroblast cells overexpressing annexin A2.

fibroblast cells in the present study resulted in a marked increase in the rate of cell migration. Furthermore, inhibition of JNK by treating cells with SP600125, delayed cell migration and reduced annexin A2 levels. However, a significant difference in cell migration was identified when comparing the SP600126-treated OX1 cells with the SP600126-treated control cells (Fig. 6), indicating that annexin A2 may function downstream of JNK during cell migration. A previous study demonstrated that high-glucose stress increased the nitrosative modification of annexin A2 in human foreskin fibroblasts, and that regulation occurred downstream of JNK regulation (5). These results indicated that JNK may regulate ANXA2 expression and its post-translational modifications, which may be important for wound repair. In the present study, the level of annexin A2 was increased in rat DM skin when compared with normal skin. This may be due to feed-back activation of annexin A2 in an attempt to try to repair the wound in DM rat skin.

In conclusion, 2-D gel electrophoresis and LC/MS proteomics were performed to gain insight into the intricate

mechanisms underlying the response to DM-mediated delay of wound healing in rats. This approach resulted in the identification of a number of differentially expressed proteins in DM that were associated with major processes, including carbohydrate metabolism, protein folding and cell redox homeostasis. The majority of proteomic studies conducted to date, that have used stress models, have been performed in rat skin against diabetes. Overexpression of annexin A2 accelerated the rate of fibroblast migration, and JNK-annexin A2 was identified as a potential pathway that promotes cell migration during wound healing. It would be of interest to analyze skin wound repair in an annexin A2 overexpressed animal model in future studies.

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