Alcohol consumption and corresponding factors: A novel perspective on the risk factors of esophageal cancer (Review)

QIAO PENG 1 , HUI CHEN 2 and JI-RONG HUO 1

¹Department of Gastroenterology, The Second Xiangya Hospital of Central South University, Changsha, Hunan 410011; ²Department of Gastroenterology, People's Hospital of Taizhou, Taizhou, Jiangsu 225300, P.R. China

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Abstract. Esophageal cancer is the eighth most common type of cancer in the world, and the sixth most common cause of mortality from cancer. Alcohol consumption is the major risk factor for esophageal cancer, due to the worldwide prevalence and high carcinogenicity of the ethanol metabolite. In epidemiological studies, the efficiency of alcohol intake to enhance the risk of esophageal cancer is altered by daily ethanol consumption, type of alcoholic beverages ingested, time since quitting drinking, age of drinking initiation, differences in population and subtypes of esophageal cancer. Corresponding factors, including gene polymorphisms, tobacco smoking, oral microorganisms and folate deficiency, reveal a synergistic effect in concurrent alcohol users that may lead to an increased risk of developing esophageal cancer. Consequently, esophageal cancer prevention involves multiple aspects, including quitting drinking and smoking, maintaining an adequate oral health and ingesting adequate quantities of folate, particularly in genetically high-risk populations.

Contents

- 1. Introduction
- 2. Association between alcohol consumption and the risk of esophageal cancer
- 3. Interactions between alcohol consumption and the corresponding factors
- 4. Conclusion

Correspondence to: Professor Ji-Rong Huo, Department of Gastroenterology, The Second Xiangya Hospital of Central South University, 139 Renminzhong Road, Changsha, Hunan 410011, P.R. China

E-mail: huojirong2014@outlook.com

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1. Introduction

Esophageal cancer is the eighth most common type of cancer in the world and the sixth most common cause of mortality from cancer, according to the results of the GLOBOCAN project, published by the International Agency for Research on Cancer in 2012 (1). Alcohol consumption has been demonstrated to be a major risk factor for developing esophageal cancer, particularly esophageal squamous cell carcinoma (ESCC) in men (2-4). Globally, prevailing alcohol intake has been an increasingly dire health problem due to the carcinogenicity of ethanol (5,6). Chronic ethanol ingestion leads to nutritional deficiencies and generation of reactive oxygen species (ROS). In addition, ethanol acts as a solvent of carcinogens (7). Acetaldehyde, the primary metabolite of ethanol, is highly mutagenic, due to its ability to form exocyclic DNA adducts (8). The diversity of carcinogenetic mechanisms may reflect the wide interaction between ethanol and cofactors from the inner and outer environment (9). The present study focuses on how alcohol consumption associates with the risk of esophageal cancer and interacts with corresponding factors, mainly from an epidemiological aspect.

2. Association between alcohol consumption and the risk of esophageal cancer

Daily ethanol consumption. The risk of developing esophageal cancer has been indicated to increase with an increase in alcohol intake (10-18). Fan et al (18) demonstrated a positive association between the total amount of ethanol intake during lifetime and the risk of esophageal cancer. In another study, heavy ethanol intake (>53.3 g/day) was significantly associated with the risk of esophageal cancer, even in a relatively short duration (≤20 years) (12). Castellsagué et al (19) demonstrated that, compared with individuals with a decreased daily alcohol consumption for numerous years, those drinking large amounts of alcohol for a shorter period of time tend to carry an increased risk of developing esophageal cancer. Therefore, compared with a long duration of alcohol consumption, an increased daily amount of alcohol consumption may be a more effective risk factor of esophageal cancer.

Type of alcoholic beverages. As the major component of alcoholic beverages, ethanol is the determinant of the risk of

esophageal cancer (20). Acetaldehyde, though present in trace amounts in beverages, may be another risk factor, due to its strong carcinogenicity (21). The most prevalent beverage in a region tends to have the greatest relative risk (22). Baijiu (a type of hard liquor with a high alcohol content) in China, wine in Italy, calvados in France and spirits in South America enhance the development of esophageal cancer, due to their high ethanol content and great popularity in each particular region (12,19,22,23). In contrast, wine consumption was observed to reduce the risk of esophageal cancer in a previous cohort study (17). Despite the regional variations in drinking habits, certain antitumor substances, including flavonoids, that are contained in wine may explain the inconsistencies reported across different studies (24). In previous studies, beer had a relatively mild effect on the risk of esophageal cancer, even with large consumption, compared with other beverages, due to its low ethanol content (17-19,22).

Years since quitting drinking. There are controversies regarding the association between the time since quitting drinking and the risk of developing esophageal cancer. Castellsagué et al (19) and Wu et al (25) demonstrated a virtually negative association in men. The conclusions of the study by Zambon et al (14) were more complex. Compared with persistent drinkers, drinking cessation was associated with certain increased risk in the first 10 subsequent years and with a non-significant risk reduction thereafter (14). These differences may be due to variations in reference and population stratification in case-control studies.

Age of drinking initiation. This variant has an uncertain effect on the risk of esophageal cancer. In the study by Castellsagué et al (19), along with increasing daily alcohol intake, there was a greater tendency for individuals that started drinking at an older age to develop esophageal cancer. However, Zambon et al (14) showed no association between the age of drinking initiation and the risk of developing esophageal cancer.

Differences in population. Different populations involved in various studies affect the strength of the association between alcohol and esophageal cancer. Alcohol consumption may be a great risk factor for esophageal cancer in Caucasian populations (14,19,26); however, the effect of alcohol consumption on the risk of developing esophageal cancer appears to be much weaker in Asian populations (25,27), particularly in certain regions with high incidence of esophageal cancer, including Linxian (China), where alcohol consumption is not a major risk factor for esophageal cancer (28). This weak association may be attributed to strong confounding factors, including gene polymorphism and other carcinogens that dilute the effect of alcohol (11,28). In studies concerning gender, the association between alcohol consumption on the risk of developing esophageal cancer was weaker in women compared with men, which may be partly explained by a short history of alcohol exposure and low alcohol prevalence among women (19,25).

Subtypes of esophageal cancer. A large number of studies have indicated a strong positive association between alcohol intake and ESCC (11,12,15,17,18,29). However, the association

between alcohol intake and esophageal adenocarcinoma (EAC) is attenuated. The two cohort studies by Steevens *et al* (17,30) revealed that alcohol consumption did not promote the risk of developing EAC or Barrett's esophagus (precancerous lesions of EAC). Other analyses reached similar conclusions (29,31,32). The study by Akiyama *et al* (33) showed a moderate increase in the risk of Barrett's esophagus in Japanese male drinkers. In the meta-analysis conducted by Lubin *et al* (15), odd ratios (ORs) with drinking-years exhibited an inverse association with alcohol consumption in <5 drinks/day consumers, and no association in heavier consumers. The distinct outcome of EAC and ESCC may be explained by the different pathogenesis of these two subtypes of esophageal cancer (34).

3. Interactions between alcohol consumption and the corresponding factors

Gene polymorphism. Since numerous genes participate in the catabolism of ethanol directly or indirectly, gene polymorphism greatly affects the carcinogenicity of ethanol in various populations (35). In several epidemiological studies, even genes that are barely associated with ethanol metabolism revealed similar effects (Table I).

Alcohol dehydrogenases (ADH) and aldehyde dehydrogenases (ALDH) are the major enzymes that participate in the metabolism of ethanol. Ethanol is oxidated to acetaldehyde by ADH, and the subsequent oxidation of acetaldehyde to acetate is catalyzed by ALDH. Polymorphism of these gene families caused by point mutations alters enzymatic activity, resulting in potential individual variations in acetaldehyde exposure (36-38).

The ADH1B*2 (Arg48His) allele encodes a more active subunit of ADH1B, compared with the ADH1B*1 allele, the homodimer of which has a ~40-fold greater maximum velocity than the ADH1B*1/2*1 form of ADH1B (39,40). The enzyme encoded by the ADH1C*1 (Ile350Val) allele has a 2.5-times faster speed of acetaldehyde production than the enzyme encoded by the ADH1C*2 allele (39,40). ADH1B*1 has been previously demonstrated to enhance the risk of esophageal cancer in Asian populations (41-45). By contrast, the association between ADH1C and the risk of esophageal cancer is contradictory. Yokoyama et al (41) and Muto et al (46) demonstrated that ADH1C*2 increased the risk of developing esophageal cancer; however, the study by Wu et al (45) showed no enhanced risk. Furthermore, ADH1C*1, but not ADH1C*2, enhanced the risk of esophageal cancer in certain studies conducted in western countries (47,48). This contradiction may be explained by different linkage disequilibrium patterns among various populations (40,48). ADH7 is mainly expressed in the upper gastrointestinal tract (49), and certain studies showed that single nucleotide polymorphisms of ADH7 were associated with esophageal cancer in alcohol drinkers (50-52).

The ALDH2*2 (Glu487Lys) allele, which encodes an inactive subunit of ALDH, occurs most frequently in Asian countries, including China, Japan, Korea, Mongolia and Indochina (53). As the major enzyme affecting blood acetaldehyde concentration, the ALDH2*1*2 and ALDH2*2*2 forms of ALDH produced a 6- and 19-fold increased acetaldehyde concentrations, respectively, compared with the ALDH2*1*1

form (54). The enhancing effect of the ALDH2*2 allele on the risk of ESCC in Japanese men was demonstrated in previous reports by Yokoyama et al (55,56). A previous meta-analysis including seven case-control studies in Asia revealed a positive association between the risk of developing esophageal cancer and the level of alcohol consumption in subjects carrying the ALDH2*1*2 genotype (57). The ALDH2*2 and ADH1B*1 alleles acted in a multiplicative manner to enhance the risk of esophageal cancer (41,42). Notably, although the ALDH2*1*2 genotype increased the risk of developing esophageal cancer, the ALDH2*2*2 genotype reduced the risk in a previous study (42). It may be hypothesized that increased blood levels of acetaldehyde due to the ALDH2*2 homodimer results in the 'alcohol flushing response', which includes facial flushing, nausea and tachycardia, preventing people from heavy drinking, thus decreasing the possibility of esophageal cancer (58,59). Individuals that possessed the ALDH2*1/2*2 genotype and also carried ADH2*1/2*1 did not exhibit flushing following drinking. These individuals tended to be heavy drinkers and had the greatest risk of developing esophageal cancer (58).

Cytochrome P450 2E1 (CYP2E1) is the major enzyme in the microsomal ethanol oxidation system (60). Induced by ethanol consumption, CYP2E1 metabolically activates procarcinogens and produces noxious ROS during ethanol oxidation (61). Compared with CYP2E1*c1 (Pst I-/Rsa I+), the mutant CYP2E1*c2 (Pst I+/Rsa I-) is considered to display a decreased activity (38). Individuals with the CYP2E1*c1*c1 genotype possessed a much greater risk of developing ESCC compared with those carrying the CYP2E1*c2 allele among Chinese drinkers; Furthermore, CYP2E1*c1*c1 exhibits a synergistic interaction with ALDH2*1*2 and methylenetetrahydrofolate reductase (MTHFR)677 (C/T+T/T) (62,63). However, in several cases, particularly in Caucasians, gene polymorphisms of CYP2E1 have no association with esophageal cancer (64-67).

Polymorphisms of the enzymes involved in the folate metabolic pathways also impact the development of esophageal cancer. MTHFR, a key enzyme in folate metabolism, is important for DNA methylation (68,69). Altered activity of MTHFR may cause DNA hypomethylation, a process associated with carcinogenesis (70,71). The most commonly studied MTHFR mutant is the change from C to T at nucleotide 677 of the MTHFR gene, which results in an alanine to valine substitution in the MTHFR enzyme (72). The homozygote and heterozygote of 667T exhibit a 30 and 65% activity, respectively, compared with the 100% activity exhibited by the 677CC genotype (73). Previous case-control studies and meta-analyses have revealed that the MTHFR C677T allele increased the risk of developing ESCC (74-78). In coordination with ADLH2*2, individuals with C677T demonstrated elevated ORs (74). The MTHFR 677T allele collaborated with alcohol consumption to increase the risk of esophageal cancer in former, moderate and heavy drinkers (75). However, the interaction between MTHFR 677T and alcohol intake is not consistent in certain meta-analyses (77,78). A previous study conducted in Japan reached the opposite conclusion, and reported that the MTHFR 677TT genotype significantly decreased esophageal cancer risk in heavy drinkers (79). This inconsistency may be due to the regional variations in folate consumption among different populations, which would suggest a significant gene-nutrient interaction between folate consumption and MTHFR genotype (78-80).

Certain gene polymorphisms with little association with the metabolism of alcohol somewhat alter the risk of esophageal cancer. According to the review by Hiyama et al (81), carcinogen metabolism-associated genes [such as cytochrome P450 family 1 subfamily A member 1 (CYP1A1), glutathione S-transferases (GSTs) and natural antisense transcripts], DNA repair genes (such as X-ray repair complementing defective repair in Chinese hamster cells 1 and xeroderma pigmentosum group D), cell cycle control genes (such as tumor protein p53 and cyclin D1) and oncogenes (such as v-myc avian myelocytomatosis viral oncogene lung carcinoma derived homolog) showed varying degrees of association with esophageal cancer. However, inconsistencies exist in various studies, and certain studies showed that the aforementioned genes interacted with alcohol consumption in the development of esophageal cancer. The GSTM1 deletion, GSTP1 341C/T+341T/T and CYP1A1 Val/Val genotypes were indicated to possess much greater ORs in alcohol drinkers compared with non-drinkers in two case-control studies (82,83).

Tobacco smoking. Similarly to alcohol beverages, tobacco has spread all over the world, with >1 billion current smokers (84). The majority of alcohol drinkers are also tobacco smokers (85). Epidemiological studies involving the combined effects of tobacco and alcohol have consistently revealed the existence of a positive synergistic effect between these two factors on the risk of developing esophageal cancer (14,17-19,25,86-88) (Fig. 1). The two variants usually exhibit a mutual dose-response association (14,17,19), and a combined OR reached a value of 130 in an Italian study (14). However, the effect of this combination appeared to be much weaker in Asian populations, with a combined OR of <10 at the highest alcohol and tobacco use (18,25). Similarly, women had a less combined OR compared with men (19,25). The variation between populations may be attributed to regional and gender differences in terms of tobacco prevalence (25,89). Reportedly, ~66.9% of men but only 4.2% of women are tobacco smokers in China, whereas the prevalence of smoking among men and women was estimated to be 35 and 22%, respectively, in developed countries, and 50 and 9%, respectively, in developing countries (25,89).

Tobacco smoke contains >60 carcinogens, including tobacco-specific nitrosamines such as 4-(methylnitrosamino) -1-(3-pyridyl)-1-butanone and N'-nitrosonornicotine, and polycyclic aromatic hydrocarbons (PAHs) such as benzo[a]pyrene and acetaldehyde (84,90,91). Tobacco smoke interacts with ethanol in the initiation of cancer in several ways: i) Ethanol activates the CYP members that metabolize tobacco procarcinogens to carcinogens (91); ii) ethanol increases cellular membrane permeability and acts as a solvent to facilitate the penetration of molecules like PAHs into the intracellular domain of mucosal epithelial cells (91); and iii) tobacco smoke, as a direct source of acetaldehyde or as a regulator of the population of oral bacteria, cooperatively elevates acetaldehyde exposure in a direct or indirect way by 7-fold, compared with alcohol drinking alone (92).

Oral microorganisms. Epidemiological evidence of the associations between salivary acetaldehyde and esophageal cancer remains limited, but several experiments conducted in animals may aid to elucidate the local carcinogenic effects of acetaldehyde on the mucosa of the upper digestive tract. Previous studies suggested that long-term alcohol consumption may induce increased cell proliferation in the oral and esophageal mucosa of rats (93,94). As a more direct model for reflecting the effect of salivary acetaldehyde, rats that drank water with an increased concentration of acetaldehyde showed hyperplasia and hyperproliferation in the epithelia of their upper gastrointestinal tract (95).

Oral microbes and prolonged ethanol use are two major factors in the generation of salivary acetaldehyde. This hypothesis has been previously demonstrated *in vivo* by Homann *et al* (96). In this study, moderate ethanol ingestion resulted in carcinogenic amounts of acetaldehyde in the saliva; whereas using an antiseptic mouthwash with chlorhexidine prior to ethanol exposure, *in vivo* acetaldehyde production decreased by >50%, with a parallel evident decrease in bacterial counts. Yokoyama *et al* (97) showed that following 3 weeks of abstinence, the microorganism count and salivary acetaldehyde production decreased in alcoholics. These results indicate a certain mutual effect between ethanol and oral microorganisms. Chronic alcohol consumption may increase bacterial concentrations through affecting salivary gland morphology and decreasing salivary flow (98,99).

As another promoter of microbial acetaldehyde production, tobacco smoking may exhibit a strong association with increased salivary acetaldehyde during alcohol drinking. Smokers that smoke while drinking have 7-fold increased salivary acetaldehyde levels compared with non-smokers (100). With the exception of the direct contribution of acetaldehyde by tobacco smoke, the alteration of oral microorganisms by smoking is also a major source of the increased concentration of salivary acetaldehyde (99). As previously reported, increased yeast infections and conversions from Gram-negative to Gram-positive bacteria has been demonstrated in smokers (99,101,102). However, oral bacteria may activate the nitrosamines from tobacco smoking to carcinogenic adducts by forming hydroxylated products (103-105).

Although>700 bacterial species inhabit the oral cavity (105), only the prevalence of several aerobic Gram-positive bacteria and yeast have been proved to play a leading role in acetaldehyde production (106). This phenomenon may due to the increased ADH activity, which has been confirmed in Streptococcus gordonii V2016 (107), Neisseria (108) and Streptococcus salivarius (109), the prevalence of aerobic Gram-positive bacteria and yeast are important in acetaldehyde production (109), due to their increased ADH activity. Other species, including hemolytic Streptococcus viridans var., Stomatococcus sp. and Corynebacterium sp., are not highly efficient in producing acetaldehyde; however, these species were significantly associated with increased acetaldehyde levels in saliva (110). Yeasts such as Candida albicans were indicated to have great capacity to produce carcinogenic acetaldehyde (111).

Folate deficiency. In previous studies, people ingesting a greater quantity and variety of fresh vegetable and fruits

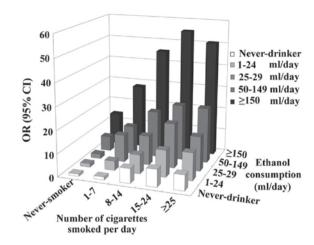


Figure 1. Combined exposure to alcohol consumption and cigarette smoking, and risk of developing esophageal cancer, as reported by Castellsagué *et al* (19). OR, odds ratio; CI, confidence interval.

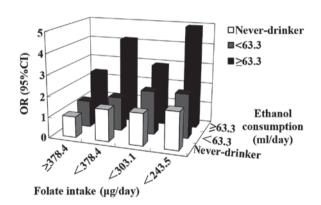


Figure 2. Combined exposure to alcohol consumption and folate intake, and risk of developing oral and pharyngeal cancer, as reported by Matsuo *et al* (122). OR, odds ratio; CI, confidence interval.

were less likely to develop esophageal cancer (112-115). Among numerous anticarcinogenic nutrients contained in plant foods, folate intake has been widely demonstrated to be closely associated with cancer of the brain, lung, esophagus, pancreas, colorectum, breast, cervix and breast in previous epidemiological studies (116). Three meta-analyses involving worldwide case-control studies conducted between 1988 and 2011 reached a consensus that folate intake may effectively protect individuals from ESCC and EAC, with a pooled OR and relative risk (RR) between 0.50 and 0.66 (78,117,118). In a large cohort study, a decreased intake of folate compared with the daily median intake (405 μ g) demonstrated an inverse association with the risk of developing ESCC, but increased intake showed no association; furthermore, there were no significant associations between dietary folate and risk of EAC (119). One hypothesis proposed that folate intake was not linearly associated with cancer risk, with a protective effect only in moderate folate intake but no protection or even tumor promotion in low or excessive ingestion (120). In that study, the cohort had a relatively high median of folate consumption, which may explain this inconsistency with other studies.

A significant interaction has been observed between folate and alcohol intake in upper digestive tract cancer. In

Table I. Interaction between alcohol consumption and gene polymorphisms in the risk of esophageal cancer.

Genes	Odds ratio/relative risk (95% confidence interval)				
	Gene polymorphisms ^a	Alcohol consumption ^b	Synergism ^c	First author, year	Refs.
ADH1B*1	4.25 (0.41-43.80)	7.01 (3.77-13.00)	38.60 (13.3-113.00)	Yokoyama, 2002	(41)
	2.37 (1.40-4.01)	6.21 (2.39-16.30)	25.20 (10.90-53.40)	Zhang, 2010	(42)
	0.97 (0.21-3.62)	2.72 (0.55-79.60)	3.70 (0.34-282.00)	Wang, 2011	(44)
	1.39 (0.84-2.31)	1.67 (1.18-2.37)	3.58 (2.20-5.84)	Wu, 2013	(45)
ADH1C*2	0.81 (0.17-3.99)	6.64 (3.66-12.10)	23.80 (7.67-74.10)	Yokoyama, 2002	(41)
ALDH2*2	1.44 (0.22-9.54)	10.40 (2.85-37.80)	88.90 (24.0-74.10)	Yokoyama, 2002	(41)
	1.70 (1.05-2.75)	4.22 (2.03-8.77)	21.50 (6.44-71.60)	Zhang, 2010	(42)
	1.59 (0.87-3.71)	1.03 (0.07-27.50)	7.05 (0.48-331.00)	Wang, 2011	(44)
	0.74 (0.53-1.02)	1.35 (0.99-1.85)	2.34 (1.52-3.61)	Wu, 2013	(45)
CYP2E1*C1	0.56 (0.20-1.59)	1.93 (0.43-2.41)	7.64 (2.82-11.30)	Guo, 2008	(62)
	2.70 (1.31-5.57)	1.94 (0.82-4.60)	7.10 (3.44-14.70)	Qin, 2008	(63)
MTHFR C677T	2.19 (1.03-4.73)	3.42 (1.28-8.93)	5.77 (2.11-15.70)	Zhao, 2011	(75)
GSTM1 deletion	1.56 (0.80-3.04)	2.74 (1.01-7.55)	6.27 (2.30-17.70)	Wang, 2004	(82)
CYP1A1 Val/Val	2.01 (0.94-3.43)	3.02 (1.31-7.03)	7.71 (2.39-32.20)	Wang, 2004	(82)

^aIndividuals who are homozygotes or heterozygotes for one of the listed genes but not alcohol-drinkers. ^bIndividuals who are heavy drinkers but non-carriers of one of the listed genes, or heterozygotes for that gene. ^cSynergism between gene polymorphisms and alcohol consumption in individuals who are heavy alcohol-drinkers and homozygotes or heterozygotes for one of the listed genes. ADH, alcohol dehydrogenase; ALDH, aldehyde dehydrogenase; CYP, cytochrome P450; MTHFR, methylenetetrahydrofolate reductase; GST, glutathione S-transferase.

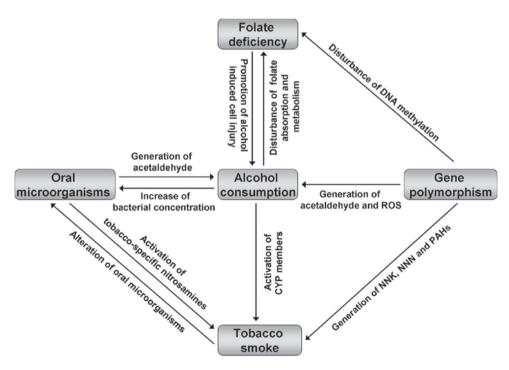


Figure 3. Interactions between alcohol consumption and its corresponding factors. ROS, reactive oxygen species; CYP, cytochrome P450; PAHs, polycyclic aromatic hydrocarbons; NNK, 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone; NNN, N-nitrosonornicotine.

the case-control study by Ibiebele *et al* (121), individuals with increased alcohol and decreased folate intake demonstrated a 3-fold OR of ESCC compared with individuals with decreased alcohol and increased folate intake. Matsuo *et al* (122) reported a similar association in oral and pharyngeal cancer (Fig. 2).

Another large prospective cohort study additionally confirmed the antagonistic interaction between alcohol and folate intake in oral cancer (123).

Folate is important in DNA metabolism, since it mediates the synthesis of S-adenosylmethionine, a methyl

donor used in biological methylation reactions and de novo deoxynucleoside triphosphate synthesis (116). Folate depletion may be oncogenic through altered DNA/RNA methylation, disruption of DNA integrity and disruption of DNA repair (116). Alcohol ingestion is a primary cause of folate deficiency (124,125). In addition to dietary inadequacy, alcohol may decrease internal folate levels through intestinal malabsorption, decreased hepatic storage and increased renal excretion (124,125). Folate homeostasis depends on transporter proteins, including reduced folate carrier, proton-coupled folate transporter, folate-binding protein, mitochondrial folate transporter and enzymes such as folylpolyglutamate synthetase (126-129). Chronic ethanol exposure may downregulate gene expression, thus impairing the transportation of folate across membranes (126-129). With the exception of causing folate deficiency, ethanol intake interferes widely in folate-dependent intermediary metabolism by inhibiting enzymes in the one-carbon metabolism, particularly methionine synthase and its associated products of metabolism, thus disturbing the synthesis of nucleotides (130). By contrast, folate deficiency facilitates the adverse effects of alcohol in methionine metabolism and promotes alcohol-induced oxidative cell injury (124).

4. Conclusion

Alcohol consumption significantly increases the risk of esophageal cancer, exhibiting a dose-response association with daily intake and an altered efficiency in various beverage types, populations and cancer subtypes. Gene polymorphisms, tobacco smoking, oral microorganisms and folate deficiency act as collaborators with concurrent alcohol use (Fig. 3). Current evidence suggests that, rather than mere alcohol consumption, the synergy of alcohol consumption and corresponding factors is important for the development of esophageal cancer. Therefore, quitting alcohol drinking and tobacco smoking, maintaining an adequate oral hygiene and ingesting adequate levels of plant foods may effectively protect high-risk individuals from developing esophageal cancer.

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