

Experimental and clinicopathological analysis of HOXB9 in gastric cancer

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Abstract. The association between homeobox (HOX)B9 expression and tumor malignancy was identified recently. It was reported that HOXB9 induced tumor angiogenesis, and associated with poor prognosis in patients with breast and colon cancer. On the other hand, regional lymph nodes are the most common site of tumor spread, and lymph node metastasis is a major prognostic factor in gastric cancer. It was hypothesized that HOXB9 promotes tumor lymphangiogenesis and induces tumor progression, invasion and metastasis in gastric cancer. The aim of the present study was to evaluate the correlation between HOXB9 expression, prognosis and clinicopathologic factors in patients with gastric cancer, and to assess the contribution of HOXB9 expression to tumor cell lymphangiogenesis *in vitro*. HOXB9 expression was evaluated by immunohistochemistry in resected tumor tissues from 58 patients with gastric cancer, and the association between prognosis and clinicopathologic factors was determined. HOXB9 gene was overexpressed in human gastric cancer TMK-1 cells and the effect of HOXB9 overexpression on the expression of vascular endothelial growth factor (VEGF)-C, VEGF-D and VEGF receptor (R)-3 was determined. It was demonstrated that the depth of tumor invasion, the number of node metastases, lymphatic invasion and vascular invasion were significantly associated with HOXB9 expression. Overall survival was decreased in patients with HOXB9 expression. The mRNA expression of VEGF-D but not of VEGF-C and VEGFR-3 was increased in HOXB9-overexpressing TMK-1 cells compared with control cells. In conclusion, HOXB9 expression was positively correlated with gastric cancer progression and lymphangiogenesis marker expression. HOXB9 may be associated with lymphogenic metastasis.

Introduction

Surgical resection is the most important treatment for patients with gastric cancer. The prognosis of patients with early gastric cancer after gastrectomy is favorable, with a 5-year survival rate of ~90% (1,2). The prognosis of patients with advanced gastric cancer remains poor. Even if patients are asymptomatic, the rate of lymph node metastasis and risk of early recurrence is high (3).

Prognostic clinicopathologic factors, including depth of invasion, lymph node status and distant metastasis are important to assess the malignant potential of gastric cancer. Recently, gene expression profiling was developed as a method for defining the phenotypes of malignant tumors at the molecular level. Therefore, indexes based on gene expression may indicate the benefit of systemic therapy and serve as predictive, prognostic and therapeutic biomarkers in gastric cancer (4).

The homeobox (HOX) gene family consists of 39 members with a shared and highly conserved 61 amino acid homeodomain motif. These genes are associated with embryologic regulation, and their roles in neoplastic transformation and tumor progression are now being recognized (5). HOXB9 is a direct transcriptional target of WNT/TCF-4, whose expression is normally restricted to embryogenesis (5,6).

The relationship between HOXB9 expression and tumor malignancy was identified previously. Nguyen *et al* (7) reported that WNT/TCF signaling through lymphoid enhancer-binding factor 1 (LEF1) and HOXB9 mediated lung adenocarcinoma metastasis. It was reported that elevated HOXB9 expression correlated with high tumor grade in breast cancer patients, and that HOXB9 expression was an independent prognostic factor for disease-free survival (8,9). Fang *et al* (10) clarified that higher levels of HOXB9 expression were significantly associated with advanced clinical stage in patients with glioma. HOXB9 overexpression also stimulated the proliferation, migration and sphere formation of glioma cells *in vitro*, and induced the tumorigenicity of glioma cells *in vivo*.

Induction of HOXB9 in breast cancer cells may increase the concentrations of several angiogenic factors, including vascular endothelial growth factor (VEGF), basic fibroblast growth factor (bFGF) and interleukin (IL)-8. In addition, HOXB9 promotes the formation of large, well-vascularized breast cancer tumors *in vivo* (8). HOXB9 also induces angiogenesis and tumor proliferation of colon cancer cells *in vitro*

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resulting in high tumorigenicity *in vivo* and poor overall survival (11). Bevacizumab, an anti-VEGF antibody, remarkably suppresses tumor proliferation by inhibiting angiogenesis in HOXB9-overexpressing xenografts, improving overall survival and providing prolonged progression-free survival in HOXB9-overexpressing patients (11). These data suggest that HOXB9 has a significant association with tumor progression and therefore may be a prognostic factor in clinical outcomes.

Breast and colon cancer often spread through the blood stream to lung, liver, brain or bones. Regional lymph nodes are the most common site of tumor spread, and lymph node metastasis is a major prognostic factor in gastric cancer (12). Thus, understanding the mechanism of lymphatic metastasis may contribute to the identification of a new therapeutic target for the treatment of gastric cancer. VEGF and its receptors (VEGFRs) have crucial roles in physiological and pathological vasculogenesis. Among VEGFs, VEGF-C and VEGF-D, which bind only to VEGFR-3, are known to regulate lymphangiogenesis (13). These ligands and receptors are often used as lymphangiogenic markers.

In the present study it was hypothesized that, HOXB9 promotes tumor lymphangiogenesis and induces tumor progression, invasion and metastasis in gastric cancer. The aim of this present study was to evaluate the correlation between HOXB9 expression, prognosis and clinicopathologic factors in patients with gastric cancer, and to assess the role of HOXB9 on tumor cell lymphangiogenesis *in vitro*.

Materials and methods

Patients. A panel of 58 patients, 42 male and 16 female, with gastric cancer were recruited between June 2008 and February 2012 and were selected from the database from the Department of Surgery of Keio University Hospital (Tokyo, Japan). Their age ranged between 32 and 88, and the median was 69 years old. The clinical staging was described based on the Union for International Cancer Control TMN Classification of Malignant Tumors (7th edition) (14). The study was approved by the Ethics Committee of Keio University School of Medicine (Tokyo, Japan). Written informed consent was obtained from each patient.

Immunohistochemical staining of HOXB9. Specimens were cut into 4 μ m sections, mounted onto glass slides, dewaxed in xylene and dehydrated in alcohol. For antigen retrieval, these sections were autoclaved in citric acid solution (Ph 6.0) at 121°C for 10 min. Endogenous peroxidase activity was blocked with a peroxidase blocking reagent (0.03% hydrogen peroxide containing sodium azide; Dako; Agilent Technologies, Inc., Santa Clara, CA, USA) for 10 min at 4°C. Samples were incubated with a rabbit polyclonal anti-human HOXB9-specific antibody (1:50; cat. no., sc-133671; Santa Cruz Biotechnology, Inc., Dallas, TX, USA) at 4°C for 12 h, followed by incubation using a horseradish peroxidase-labeled secondary antibody (cat. no., K4003; Dako; Agilent Technologies, Inc.) at room temperature for 30 min. The slides were stained with 3,3'-diaminobenzidine (Dako; Agilent Technologies, Inc.) at room temperature for 2 min, according to manufacturer's protocol. Slides were rinsed in PBS. Sections were counterstained with hematoxylin, dehydrated and mounted. HOXB9

Table I. Immunoreactive scores of HOXB9.

Variables	Scores
Percentage of positive cells	
<33%	0
33-66%	1
>66%	2
Intensity of reaction	
No reaction	0
Weak reaction	1
Intense reaction	2

0-1 scores = HOXB9 negative, 2-4 scores = HOXB9 positive. HOX, homeobox.

Table II. Patient characteristics (n=58).

Parameters	Number (%)
Gender	
Male	42 (72.4)
Female	16 (27.6)
Median age in years (range)	69 (32-88)
Pathological depth of tumor invasion (14)	
T1	14 (24.1)
T2	9 (15.6)
T3	21 (36.2)
T4	14 (24.1)
Pathological nodal stage	
N0	28 (48.3)
N1	8 (13.8)
N2	8 (13.8)
N3	14 (24.1)
Pathological stage	
I	16 (27.6)
II	21 (36.2)
III	19 (32.8)
IV	2 (3.4)

Pathological tumor stage was described based on the Union for International Cancer Control TMN Classification of Malignant Tumors (7th edition) (14).

expression was evaluated independently by two authors, both blinded to the clinicopathological parameters. A light microscope was used at a magnification of x200. The percentage of positive cells and staining intensity were evaluated (Table I).

Reverse transcription-quantitative polymerase chain reaction (RT-qPCR). The expression levels of HOXB9 mRNA were determined in four human gastric cancer cell lines (TMK-1, MKN-45, MKN-74 and KATO-III) using RT-qPCR. RNA was extracted from the cells using the RNeasy kit (Qiagen, Inc.,

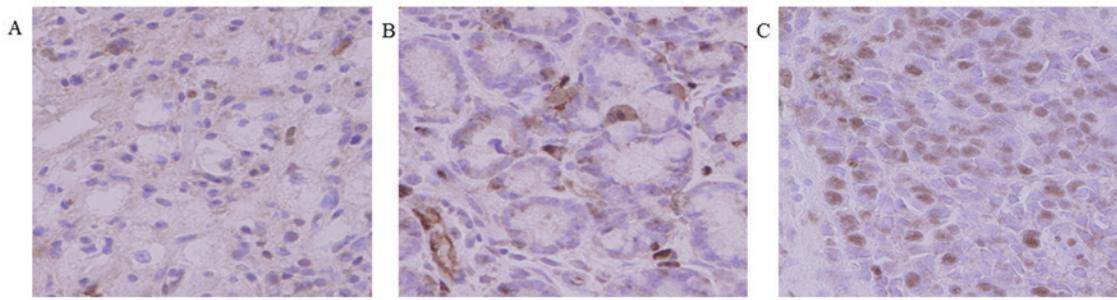


Figure 1. Immunohistochemical staining for homeobox B9 expression in human gastric cancer tissues. Staining intensity was graded as (A) no reaction, (B) weak reaction and (C) intense reaction (original magnification, x400).

Valencia, CA, USA), according to manufacturer's protocol. The concentration of total RNA was determined using a NanoDrop ND-1000 Spectrophotometer (Thermo Fisher Scientific, Inc., Waltham, MA, USA). Total RNA was reverse transcribed into cDNA using Transcriptor High Fidelity cDNA Synthesis kit (Roche Diagnostics, Rotkreuz, Switzerland) according manufacturer's protocol. The quality and quantity of the cDNA samples were assessed by standard electrophoresis and NanoDrop ND-1000 spectrophotometer. The following primer pairs were used for the qPCR: HOXB9: Sense, 5'-CCG GCTACGGGGACAATAA-3' and antisense, 5'-GGTGTA GGGACAGCGCTTTTT-3'; GAPDH, sense, 5'-ATCATCCCT GCCTCTACTGG-3' and antisense, 5'-TTTCTAGACGGC AGGTCAGGT-3'; and β -actin: Sense, 5'-CTCTTCCAGCCT TCCTTCCT-3' and antisense, 5'-AGCACTGTGTTGGCG TACAG-3'. The following thermocycling conditions were used: Initial denaturation at 95°C for 2 min; 25 cycles of 95°C for 30 sec; 56°C for 30 sec and 72°C for 60 sec; and a final extension at 72°C for 10 min. Each experiment was performed using triplicates. qPCR was subsequently performed using a Light Cycler 480 Real-Time PCR System and SYBR Green 1 Master Mix (Roche Diagnostics, Basel, Switzerland) which contained Taq DNA polymerase. The following thermocycling conditions were used for PCR: Pre-denaturation at 95°C for 5 min, followed by 45 cycles of denaturation at 95°C for 10 sec, annealing at 57°C for 10 sec and extension at 72°C for 10 sec. Subsequently, melting curve analysis was performed. The gene expressions were quantified using $2^{-\Delta\Delta C_q}$ method (15).

Transfection of HOXB9 gene. TMK-1 cells with low HOXB9 expression were infected with HOXB9-expressing lentiviral vector using pLenti6.3/V5-DEST Gateway Vector kit (Invitrogen; Thermo Fisher Scientific, Inc.), according to the manufacturer's protocols. The time interval between transfection and subsequent experimentation was >1 week.

Gel electrophoresis of PCR products. KOD FX Neo (Toyobo Life Science, Osaka, Japan) was used for PCR. The following primer pairs were used: VEGF-C, sense, 5'-AAAGAACCT GCCCAGAAAT-3' and antisense, 5'-TGGTGGTGGAACT TTCTTCC-3'; VEGF-D, sense, 5'-TCCAGGAACCAGCTC TCTGT-3' and antisense, 5'-TTTTTGGGGTGCTGGATT AG-3'; and VEGFR-3, sense, 5'-GAGACAAGGACAGCG AGGAC-3' and antisense, 5'-CTGTGTCGTTGGCATGTA CC-3'. The following thermocycling conditions were used: Initial denaturation at 94°C for 2 min, followed by 30 cycles

at 98°C for 10 sec and at 68°C 30 sec. Bands were analyzed by electrophoresis in 2% gel and visualized under UV light. The expression levels of lymphangiogenic factors in HOXB9 transfected cells were compared with that in LacZ transfected cells (control).

RT-qPCR for lymphangiogenic markers. RT-qPCR was performed to quantify the difference of expression of lymphangiogenic markers between transfected cells. cDNA purified from the cultured cell lines (TMK-1, TMK-1/LacZ, and TMK-1/HOXB9) were amplified. The thermocycling conditions and fluorophore were the same as described above. The following primer pairs were used: VEGF-D, sense, 5'-GAACAC CAGCACCTCGTACA-3' and antisense; and 5'-TTCTTCAGG GATCTGGATGG-3'; VEGFR-3, sense, 5'-CTGCAAGAACGT GCATCTGT-3' and antisense, 5'-TTCTTGTGGCAGTGCTTG TC-3'. The aforementioned primers were used for VEGF-C. The gene expressions were quantified using $2^{-\Delta\Delta C_q}$ method (15).

Statistical analysis. The relevant data were presented as mean \pm standard deviation (n=3). HOXB9 mRNA expression levels in each cell line were compared using the Kruskal-Wallis test. Dunn's multiple comparisons test was used for pairwise post-hoc comparisons. The proportions were compared using Fisher's exact test. Overall survival curves were created by Kaplan-Meier estimates and compared by log-rank tests. Statistical analyses were performed using SPSS (version 19; IBM Corp., Armonk, NY, USA). $P < 0.05$ was considered to indicate a statistically significant difference.

Results

Patient characteristics. Patient characteristics are shown in Table II. The majority of the 58 patients were male (72.4%) with a median age of 69 years. The median follow-up period was 59.2 months. The majority of the patients (75.9%) exhibited advanced gastric cancer that had invaded beyond the submucosal layer, and 51.7% were positive for lymph node metastasis.

HOXB9 expression in patients with gastric cancer. The expression of HOXB9 was evaluated in 58 tumor specimens. Immunohistochemical staining for HOXB9 (Fig. 1) was performed. Staining intensity was graded as none, weak and intense. According to the classification criteria (Table I), a total of 30 tumors (51.7%) were positive for HOXB9 expression.

Table III. Relation between HOXB9 expression and clinicopathologic factors in 58 gastric cancer patients.

Characteristics	No. of patients	HOXB9 staining		P-value
		Negative (n=28)	Positive (n=30)	
Gender				0.77
Male	42	21	21	
Female	16	7	9	
Age				0.19
<70	29	17	12	
≥70	29	11	18	
T feature				0.015
T1-2	23	16	7	
T3-4	35	12	23	
LN metastasis				0.008
Negative	28	19	9	
Positive	30	9	21	
Lymphatic invasion				0.001
Negative	17	14	3	
Positive	41	14	27	
Vascular invasion				0.023
Negative	18	13	5	
Positive	40	15	25	

T, pathological depth of tumor invasion; LN: lymph node; HOX, homeobox.

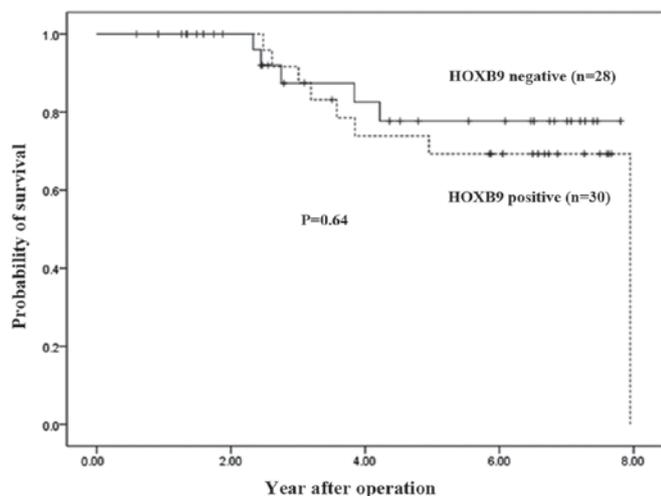


Figure 2. Kaplan-Meier plots of overall survival in HOXB9-positive and HOXB9-negative patients with gastric cancer. The P value was calculated by the log-rank test. HOX, homeobox.

HOXB9 staining was observed in the nucleus of gastric cancer cells. Association between HOXB9 expression and clinicopathologic variables is shown in Table III.

In addition, overall survival was decreased in patients with positive HOXB9 expression ($P=0.64$; Fig. 2). However, this did not reach a statistically significant difference. The depth of tumor invasion ($P=0.015$), the number of node metastases ($P=0.008$), lymphatic invasion ($P=0.001$) and vascular

invasion ($P=0.023$) were significantly associated with HOXB9 expression (Table III).

Association between HOXB9 and lymphangiogenic factors. TMK-1 cells expressed the lowest amount of HOXB9 mRNA expression levels among the four cell lines tested (Fig. 3). Using a viral vector, HOXB9 mRNA expression level was increased by 3,377-fold in TMK-1/HOXB9 cells compared with that in TMK-1/LacZ cells; this difference reached statistical significance ($P=0.027$; Fig. 4). Gel electrophoresis of PCR products showed that the expression levels of VEGF-D mRNA were increased in TMK-1/HOXB9 cells compared with TMK-1/LacZ cells (Fig. 5). The relative mRNA level of VEGF-D was increased by 6.06-fold in TMK-1/HOXB9 compared with that in TMK-1/LacZ cells ($P=0.066$; Fig. 6). VEGF-C and VEGFR-3 levels were unchanged (Figs. 5 and 6).

Discussion

HOXB9 is considered to be a transcription factor. Transcription factors are proteins, which bind to DNA sequences to control the expression of genes (16). They may have a dynamic behavior depending on intracellular/extracellular signals. In the present study, the expression of HOXB9 and clinicopathological factors were investigated in patients with gastric cancer at various pathological stages, in order to compare the association between HOXB9 expression and gastric cancer progression. Immunohistochemistry revealed that 51.7% of resected gastric cancer tissues were positive for HOXB9 expression. In a previous

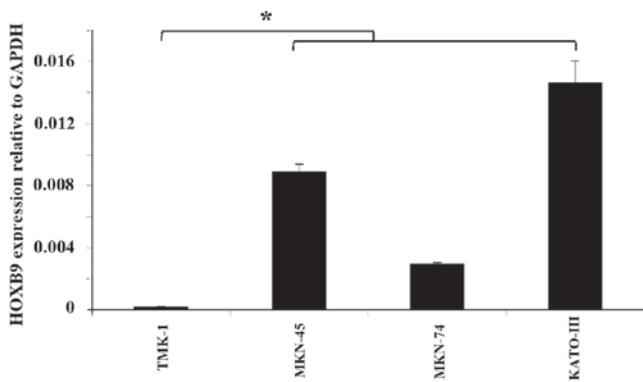


Figure 3. Comparison of HOXB9 mRNA expression levels in TMK-1, MKN-45, MKN-74 and KATO-III cell lines. TMK-1 cells exhibited the lowest HOXB9 expression level among all cell lines. *P=0.083 with comparisons as indicated by lines. HOX, homeobox.

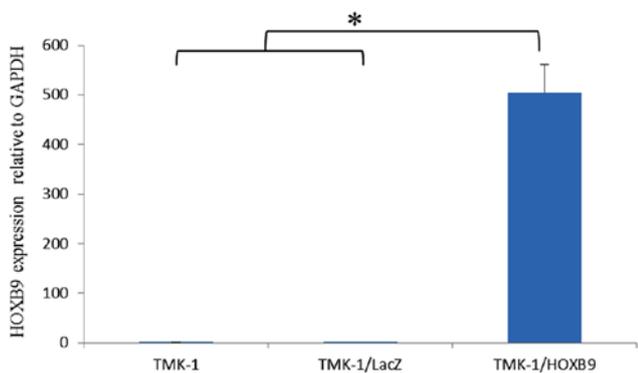


Figure 4. Comparison of HOXB9 mRNA expression levels in TMK-1, TMK-1/LacZ and TMK-1/HOXB9 in cell lines. Overexpression of HOXB9 mRNA in TMK-1 cells. *P<0.05 with comparisons indicated by lines. HOX, homeobox.

study, it was showed that HOXB9 was expressed in 49% of resected breast cancer tissues and in 67% of resected colorectal cancer tissues (9,11). Although the methods used to evaluate HOXB9 expression were different in each study, the positive rates were comparable. In addition, it was indicated that HOXB9 expression was localized to the nucleus of gastric cancer cells, which is in accordance to a previous study on breast cancer (9).

In the present study, HOXB9-positive patients with gastric cancer tended to exhibit a poorer prognosis compared with those in the HOXB9-negative group. Although this difference did not reach statistical significance, it is possible that additional data may clarify the potential association between HOXB9 expression and survival of patients with gastric cancer. HOXB9 expression was the only independent prognostic factor for disease-free survival in breast cancer patients (hazard ratio=15.532; P=0.009) (9), and increased HOXB9 expression in colorectal cancer was significantly associated with poorer overall survival (11). However, Sha *et al* (17) reported that decreased expression of HOXB9 was associated with a poor overall survival in Chinese patients with gastric cancer. The reason for this discrepancy in the association between survival and HOXB9 expression are unclear, but may be related to differences in the subjects and methodology.

In our previous study, HOXB9 positivity was significantly associated with tumor virulence in breast cancer patients

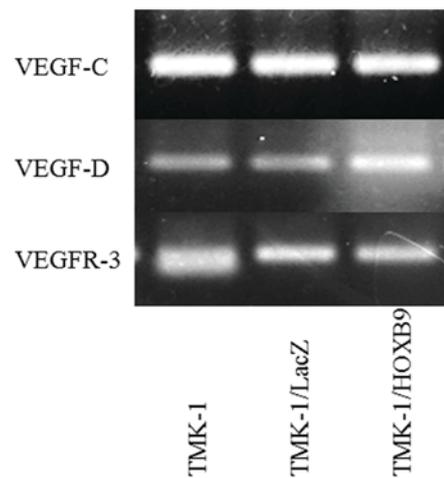


Figure 5. Expression levels of VEGF-C, VEGF-D and VEGFR-3 mRNA in TMK-1, TMK-1/LacZ and TMK-1/HOXB9 cells. The expression of VEGF-D mRNA was increased in TMK-1/HOXB9 compared with TMK-1/LacZ. VEGF-C and VEGFR-3 levels were unchanged. HOX, homeobox; VEGF, vascular endothelial growth factor; VEGFR, VEGF receptor.

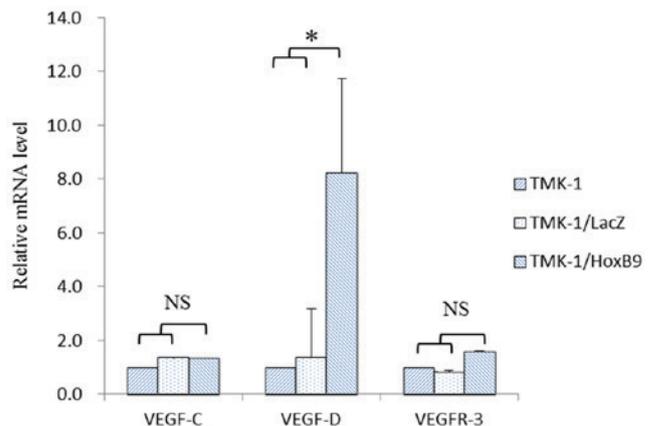


Figure 6. Quantification of expression levels of lymphangiogenic markers. mRNA expression levels of lymphangiogenic factors in TMK-1, TMK-1/LacZ and TMK-1/HOXB9 cell lines were compared. The relative mRNA level of VEGF-D was increased by 6.06-fold in TMK-1/HOXB9 cells compared with that in TMK-1/LacZ cells. *P=0.066 with comparisons indicated by lines. HOX, homeobox; VEGF, vascular endothelial growth factor; VEGFR, VEGF receptor; NS, not significant.

(tumor size, nuclear grade and lymph node metastasis) (9). In the present study, it was found that HOXB9 expression was significantly associated with the depth of invasion, lymph node metastasis, lymphatic invasion and vascular invasion in patients with gastric cancer, suggesting a role of this transcription factor in gastric cancer.

Lymph node metastasis and lymphatic invasion are related to lymphogenic metastasis, a specific phenomenon of gastric cancer progression (12). A previous study reported that HOXB9 induces the expression of several angiogenic factors (epidermal growth factor (EGF), bFGF, IL-8 and angiopoietin-like 2 (ANGPTL-2)), as well as ErbB (amphiregulin, epiregulin and neuregulins) and transforming growth factor-β (TGF-β) in patients with breast cancer (8). These factors activate their respective pathways, leading to increased cell motility and the acquisition of mesenchymal phenotypes (8). Additionally, a

study on colon cancer reported that HOXB9 induced angiogenesis and tumor proliferation *in vitro*, consistent with high tumorigenicity *in vivo* (11). The results of the present study demonstrated that there was no association between HOXB9 expression and the angiogenic factors VEGF-A, bFGF, IL8, ANGPTL2, TGF- β 1, and TGF- β 2 in patients with gastric cancer (data not shown). However, the lymphangiogenic factor VEGF-D, but not VEGF-C and VEGFR-3, was elevated in TMK-1 cells transfected with the HOXB9 gene. To the best of our knowledge this is the first study suggesting that HOXB9 promotes lymphangiogenesis. Therefore, HOXB9 may be associated with lymphogenic metastasis.

Overall, HOXB9 expression positively associated with gastric cancer progression and may be associated with poor prognosis. These results suggest the potential clinical utility of HOXB9 as a prognostic factor or therapeutic target in gastric cancer.

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Availability of data and materials

The datasets used and/or analyzed during the present study are available from the corresponding author on reasonable request.

Authors' contributions

FK, NW, TH, and YK conceived the study. RN, TT, HK, and HT were the major contributors in analyzing and interpreting the patient data. FK and KF performed the experiments. FK and NW evaluated immunostained sections. FK wrote the manuscript. YK supervised the study, and performed the critical review. All authors read and approved the final manuscript.

Ethics approval and consent to participate

The study was approved by the Ethics Committee of Keio University School of Medicine (approval no. 20080189). Written informed consent was obtained from each patient.

Patient consent for publication

The patients consented to publication.

Competing interests

The authors declare that they have no competing interests.

References

1. Sano T, Sasako M, Kinoshita T and Maruyama K: Recurrence of early gastric cancer. Follow-up of 1475 patients and review of the Japanese literature. *Cancer* 72: 3174-3178, 1993.
2. Japanese Gastric Cancer Association Registration Committee, Maruyama K, Kaminishi M, Hayashi K, Isobe Y, Honda I, Katai H, Arai K, Kodera Y and Nashimoto A: Gastric cancer treated in 1991 in Japan: Data analysis of nationwide registry. *Gastric Cancer* 9: 51-66, 2006.
3. Gotoda T, Yanagisawa A, Sasako M, Ono H, Nakanishi Y, Shimoda T and Kato Y: Incidence of lymph node metastasis from early gastric cancer: Estimation with a large number of cases at two large centers. *Gastric Cancer* 3: 219-225, 2000.
4. Bang YJ, Van Cutsem E, Feyereislova A, Chung HC, Shen L, Sawaki A, Lordick F, Ohtsu A, Omuro Y, Satoh T, *et al*: Trastuzumab in combination with chemotherapy versus chemotherapy alone for treatment of HER2-positive advanced gastric or gastro-oesophageal junction cancer (ToGA): A phase 3, open-label, randomised controlled trial. *Lancet* 376: 687-697, 2010.
5. Abate-Shen C: Deregulated homeobox gene expression in cancer: Cause or consequence? *Nat Rev Cancer* 2: 777-785, 2002.
6. Hatzis P, van der Flier LG, van Driel MA, Guryev V, Nielsen F, Denissov S, Nijman IJ, Koster J, Santo EE, Welboren W, *et al*: Genome-wide pattern of TCF7L2/TCF4 chromatin occupancy in colorectal cancer cells. *Mol Cell Biol* 28: 2732-2744, 2008.
7. Nguyen DX, Chiang AC, Zhang XH, Kim JY, Kris MG, Ladanyi M, Gerald WL and Massagué J: WNT/TCF signaling through LEF1 and HOXB9 mediates lung adenocarcinoma metastasis. *Cell* 138: 51-62, 2009.
8. Hayashida T, Takahashi F, Chiba N, Brachtel E, Takahashi M, Godin-Heymann N, Gross KW, Vivanco Md, Wijendran V, Shioda T, *et al*: HOXB9, a gene overexpressed in breast cancer, promotes tumorigenicity and lung metastasis. *Proc Natl Acad Sci USA* 107: 1100-1105, 2010.
9. Seki H, Hayashida T, Jinno H, Hirose S, Sakata M, Takahashi M, Maheswaran S, Mukai M and Kitagawa Y: HOXB9 expression promoting tumor cell proliferation and angiogenesis is associated with clinical outcomes in breast cancer patients. *Ann Surg Oncol* 19: 1831-1840, 2012.
10. Fang L, Xu Y and Zou L: Overexpressed homeobox B9 regulates oncogenic activities by transforming growth factor- β 1 in gliomas. *Biochem Biophys Res Commun* 446: 272-279, 2014.
11. Hoshino Y, Hayashida T, Hirata A, Takahashi H, Chiba N, Ohmura M, Wakui M, Jinno H, Hasegawa H, Maheswaran S, *et al*: Bevacizumab terminates homeobox B9-induced tumor proliferation by silencing microenvironmental communication. *Mol Cancer* 13: 102, 2014.
12. Pak KH, Jo A, Choi HJ, Choi Y, Kim H, and Cheong JH: The different role of intratumoral and peritumoral lymphangiogenesis in gastric cancer progression and prognosis. *BMC Cancer* 15: 498, 2015.
13. Takahashi S: Vascular endothelial growth factor (VEGF), VEGF receptors and their inhibitors for antiangiogenic tumor therapy. *Biol Pharm Bull* 34: 1785-1788, 2011.
14. Sobin LH, Gospodarowicz MK and Wittekind C: TNM classification of malignant tumours. 7th edition. Wiley-Blackwell, Hoboken, Oxford, 2009.
15. Livak KJ and Schmittgen TD: Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta C(T)) method. *Methods* 25: 402-408, 2001.
16. Bouhlel MA, Lambert M and David-Cordonnier MH: Targeting transcription factor binding to DNA by competing with DNA binders as an approach for controlling gene expression. *Curr Top Med Chem* 15: 1323-1358, 2015.
17. Sha S, Gu Y, Xu B, Hu H, Yang Y, Kong X and Wu K: Decreased expression of HOXB9 is related to poor overall survival in patients with gastric carcinoma. *Dig Liver Dis* 45: 422-429, 2013.

