

Comparative integromics on Ephrin family

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Abstract. EFNA1, EFNA2, EFNA3, EFNA4, EFNA5, EFNB1, EFNB2 and EFNB3 are EFN family ligands for EPH family receptors. EFN/EPH signaling pathway networks with the WNT signaling pathway during embryogenesis, tissue regeneration, and carcinogenesis. Comparative genomics analyses on *EFNB1*, *EFNB2* and *EFNB3* were performed by using bioinformatics and human intelligence (humint). *EFNB1* mRNA was expressed in human embryonic stem (ES) cells, neural tissues, diffuse type gastric cancer, pancreatic cancer, colon cancer, brain tumors and esophageal cancer, *EFNB2* mRNA in human ES cells, neural tissues and colon cancer, *EFNB3* mRNA in human ES cells, neural tissues, brain tumors, pancreatic cancer and colon cancer. Because triple TCF/LEF-binding sites were identified within the 5'-promoter region of human *EFNB3* gene, comparative genomics analyses on *EFNB3* orthologs were further performed. Chimpanzee *EFNB3* gene, consisting of five exons, was identified within AC164921.3 genome sequence. AY421228.1 was not a correct coding sequence for chimpanzee EFNB3. Chimpanzee *EFNB3* gene was found to encode a 340-amino-acid protein showing 99.4% and 96.6% total-amino-acid identity with human EFNB3 and mouse *Efnb3*, respectively. Three TCF/LEF-binding sites within human *EFNB3* promoter were conserved in chimpanzee *EFNB3* promoter, and the second TCF/LEF-binding site in rodent *Efnb3* promoters. CpG hypermethylation of *EFNB3* promoter with 63.2% GC content as well as deletion of *EFNB3* gene closely linked to *TP53* tumor suppressor gene at human chromosome 17p13.1 should be investigated to elucidate the mechanism of infrequent *EFNB3* upregulation in human colorectal cancer. *EFNB3*, identified as potential transcriptional target of WNT/ β -catenin signaling pathway, is a pharmacogenomics target in the fields of regenerative medicine and oncology.

Introduction

EFNA1, EFNA2, EFNA3, EFNA4, EFNA5, EFNB1, EFNB2 and EFNB3 are EPHRIN (EFN) family ligands for EPH family receptors (1-15). EFNA1, EFNA2, EFNA3, EFNA4 and EFNA5 are GPI-anchored cell-surface proteins with EPH-binding domain, while EFNB1, EFNB2 and EFNB3 are transmembrane proteins with extracellular EPH-binding domain and cytoplasmic PDZ-binding motif. EPH family members share common domain architecture, consisting of extracellular EFN-binding domain, cysteine-rich domain, two fibronectin type III repeats as well as cytoplasmic tyrosine kinase domain and C-terminal SAM motif. EPHA1, EPHA2, EPHA3, EPHA4, EPHA5, EPHA6, EPHA8 and EPHA10 are classified into EPHA subfamily, while EPHB1, EPHB2, EPHB3, EPHB4 and EPHB6 are classified into EPHB subfamily. EFN/EPH signaling pathway is implicated in a variety of processes, including axon guidance, angiogenesis, and gastrointestinal morphogenesis.

Canonical WNT signaling activation leads to transcriptional activation of *DKK1*, *DKK4*, *FGF18*, *FGF20*, etc. depending on the transcriptional complex consisting of TCF/LEF, β -catenin, BCL9/BCL9L and PYGO1/PYGO2 (16-32). WNT/ β -catenin signaling pathway is implicated in the cell fate determination.

Mouse *Efnb1* is expressed in intestinal differentiated cells, while *Efnb2* and *Efnb3* in intestinal proliferating cells depending on the WNT/ β -catenin signaling pathway (33,34). EFN/EPH and WNT signaling pathways network together during embryogenesis, tissue regeneration and carcinogenesis; however, direct transcriptional regulation of EFN family members by the WNT/ β -catenin signaling pathway remains unclear. Comparative genomics analyses on *EFNB1*, *EFNB2* and *EFNB3* were performed, and *EFNB3* was identified as potential target gene of the WNT/ β -catenin signaling pathway.

Materials and methods

WNT target gene screening. Genome sequences corresponding to human *EFNB1*, *EFNB2* and *EFNB3* genes were searched for with BLAST programs (<http://www.ncbi.nlm.nih.gov>) as described previously (35-39). TCF/LEF-binding sites within the 5'-flanking promoter region of the above genes were searched for based on bioinformatics and manual inspection as described previously (28-32).

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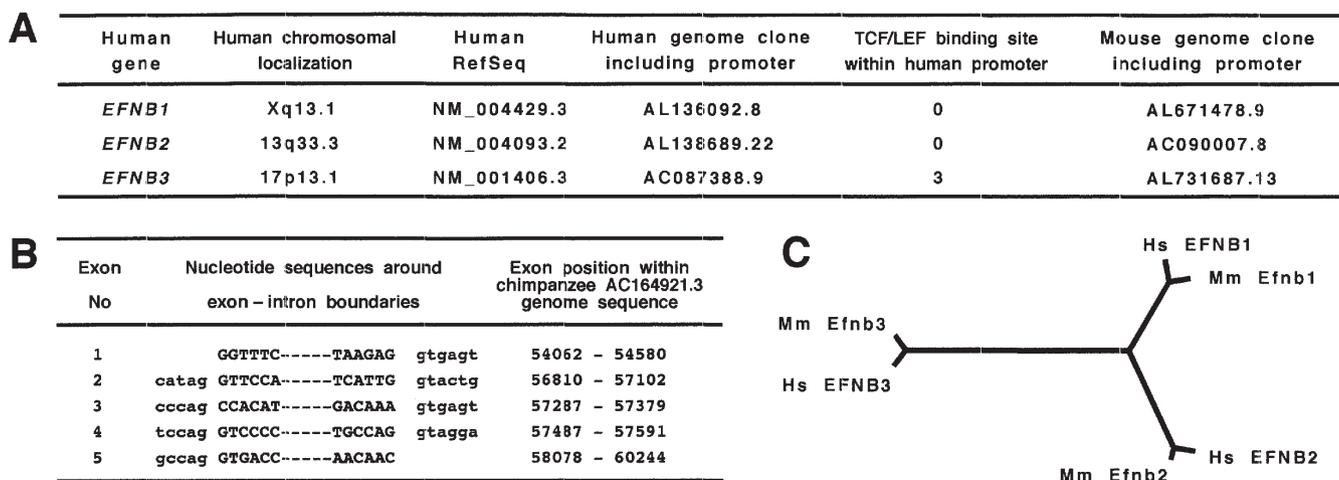


Figure 1. (A), *EFNB* gene family. Three TCF/LEF-binding sites exist within the *EFNB3* promoter. (B), Exon-intron structure of the chimpanzee *EFNB3* gene. Nucleotide sequences around exon-intron boundaries are shown by upper-case letters (exon) and by lower-case letters (intron). (C), Phylogenetic analyses on the *EFNB* family.

Identification of the chimpanzee *EFNB3* orthologs. Chimpanzee genome sequences homologous to human *EFNB3* were searched for with BLAST programs as described previously (40-43). Exon-intron boundaries were determined based on the consensus sequence of exon-intron junctions ('gt ... ag' rule of intronic sequence) and codon usage within the coding region as described previously (44-47). Coding sequence of chimpanzee *EFNB3* was determined by assembling exonic regions.

Comparative proteomics analysis. Phylogenetic analyses on mammalian *EFNB* family members were performed by using the CLUSTALW program.

Comparative genomics analyses. Promoter region of mammalian *EFNB3* orthologs were aligned by using the Genetyx program and manual curation. TCF/LEF-binding sites within the promoter region were determined as mentioned above.

In silico expression analysis. Expressed sequence tags (ESTs) derived from human *FENB1*, *EFNB2*, and *EFNB3* genes were searched for by using the BLAST programs. The sources of *FENB1*, *EFNB2*, and *EFNB3* ESTs were listed up for *in silico* expression analysis.

Results

Screening of TCF/LEF-binding site within promoter region of *EFNB* family genes. Human *EFNB1* RefSeq (NM_004429.3), *EFNB2* RefSeq (NM_004093.2) and *EFNB3* RefSeq (NM_001406.3) were used as query sequences for the BLAST programs to identify genome clones corresponding to *EFNB* family genes. The 5'-flanking promoter region of human *EFNB1*, *EFNB2* and *EFNB3* genes were identified within AL136092.8, AL138689.22 and AC087388.9 genome sequences, respectively (Fig. 1A). TCF/LEF-binding sites within the 5'-promoter region of human *EFNB1*, *EFNB2* and *EFNB3* genes were then searched for based on manual inspection. Triple TCF/LEF-binding sites were identified within human *EFNB3* promoter (Fig. 1A).

Identification of the chimpanzee *EFNB3* gene. BLAST programs using human *EFNB3* RefSeq revealed that chimpanzee *EFNB3* gene was located within AC164921.3 genome sequence. Exon-intron boundaries of chimpanzee *EFNB3* gene were determined based on the consensus sequence of exon-intron junctions. Chimpanzee *EFNB3* gene was found consisting of five exons (Fig. 1B).

Because the chimpanzee AY421228.1 predicted sequence accompanied by sequence gaps within exons 2, 4 and 5 was not the correct chimpanzee *EFNB3* sequence, complete coding sequence (CDS) of chimpanzee *EFNB3* was determined in this study by assembling nucleotide sequences of five exons (Fig. 2).

Genetyx program revealed that nucleotide position 398-1420 was the coding region of chimpanzee *EFNB3* complete CDS (Fig. 2). Chimpanzee *EFNB3* gene was found to encode a 340-amino-acid protein showing 99.4% and 96.6% total-amino-acid identity with human *EFNB3* and mouse *Efnb3*, respectively.

Comparative proteomics analysis on mammalian *EFNB3* family members. Phylogenetic analysis revealed that *EFNB1* orthologs and *EFNB2* orthologs were more related to each other than to *EFNB3* orthologs (Fig. 1C). Extracellular EPH-binding domain as well as the C-terminal cytoplasmic region with five tyrosine residues and PZD-binding motif were well conserved among *EFNB* family members (Fig. 2B).

Expression of human *EFNB1*, *EFNB2* and *EFNB3* mRNAs. *In silico* expression analyses were performed to investigate the expression profile of *EFNB* family members. *EFNB1* mRNA was expressed in human embryonic stem (ES) cells, diffuse type gastric cancer, pancreatic cancer, colon cancer, brain tumors and esophageal cancer. *EFNB2* mRNA was expressed in human ES cells, neural tissues and colon cancer. *EFNB3* mRNA was expressed in human ES cells, neural tissues, brain tumors, pancreatic cancer and colon cancer.

Comparative genomics analyses on *EFNB3* promoters. Human *EFNB3* promoter and chimpanzee *EFNB3* promoter were

located within AC087388.9 and AC164921.3 genome sequences, respectively, as mentioned above. BLAST programs revealed that mouse and rat *Efnb3* promoters were located within AL731687.13 and AC134317.3 genome sequences, respectively. Promoter alignment revealed that 5'-promoter region of human, chimpanzee, mouse and rat *EFNB3* orthologs were well conserved (Fig. 3).

GC content of human *EFNB3* promoter was 63.2%, that of chimpanzee *EFNB3* promoter was 63.1%, that of mouse *Efnb3* promoter was 55.5%, and that of rat *Efnb3* promoter was 56.3%. GC contents of primate *EFNB3* promoters were higher than the rodent *Efnb3* promoters.

Triple TCF/LEF-binding sites within human *EFNB3* promoters were located about 1100, 1000, and 400 bp upstream of the transcription start site (Fig. 3). Three TCF/LEF-binding sites within human *EFNB3* promoter were conserved in the chimpanzee *EFNB3* promoter. The second TCF/LEF-binding site within human *EFNB3* promoter was conserved in rodents *Efnb3* promoters.

Discussion

TCF/LEF-binding sites within *EFNB1*, *EFNB2*, *EFNB3* promoters were searched for to identify the WNT/ β -catenin target gene among the *EFNB* family in this study. The 5'-flanking promoter region of human *EFNB1*, *EFNB2* and *EFNB3* genes were identified within AL136092.8, AL138689.22 and AC087388.9 genome sequences, respectively. Because triple TCF/LEF-binding sites were identified within the 5'-promoter region of human *EFNB3* gene (Fig. 1A), comparative genomics analyses on *EFNB3* orthologs were further performed.

Chimpanzee *EFNB3* gene, consisting of five exons, was identified within the AC164921.3 genome sequence (Fig. 1B). AY421228.1 was not the correct coding sequence for chimpanzee *EFNB3*, and complete CDS of chimpanzee *EFNB3* was determined in this study (Fig. 2). Chimpanzee *EFNB3* gene was found to encode a 340-amino-acid protein showing 99.4% and 96.6% total-amino-acid identity with human *EFNB3* and mouse *Efnb3*, respectively.

Three TCF/LEF-binding sites within human *EFNB3* promoter were conserved in chimpanzee *EFNB3* promoter, while the only second TCF/LEF-binding site within human *EFNB3* promoter was conserved in rodent *Efnb3* promoters (Fig. 3). GC contents of primate *EFNB3* promoters were higher than the rodent *Efnb3* promoters. Although mammalian *EFNB3* promoters were relatively well conserved, TCF/LEF-binding sites were triplicated in primate *EFNB3* promoters compared with the rodent *Efnb3* promoters due to nucleotide changes during mammalian evolution.

Expression of human *EFNB1*, *EFNB2* and *EFNB3* mRNAs was investigated by using the *in silico* expression analyses. *EFNB1*, *EFNB2* and *EFNB3* mRNAs were expressed in human ES cells and neural tissues. In addition, *EFNB1* mRNA was expressed in a variety of tumors, such as gastric cancer, pancreatic cancer, colon cancer, brain tumors and esophageal cancer.

Mouse *Efnb3*, interacting with *Epha4* on the axons, is implicated in axon repulsion during embryogenesis as well as inhibition of neurite outgrowth after traumatic spinal cord injury (48-50). Repression of *EFNB3* transcription could contribute to the acceleration of neurite outgrowth after traumatic spinal cord injury.

EFNB3 expression in colorectal cancer was relatively infrequent, although WNT/ β -catenin signaling pathway is frequently activated in colorectal cancer. GC content of *EFNB3* promoter was 63.2%, and *EFNB3* gene is closely linked to *TP53* tumor suppressor gene at human chromosome 17p13.1. CpG hypermethylation of *EFNB3* promoter as well as deletion of *EFNB3* gene might explain the relatively infrequent expression of *EFNB3* mRNA in colorectal cancer. Epigenetic changes and genetic alterations of *EFNB3* gene in colorectal cancer should be investigated in the future.

EFNB3 was identified as potential transcriptional target of WNT/ β -catenin signaling pathway in this study. *EFNB3* is a pharmacogenomics target in the fields of regenerative medicine and oncology.

References

- Davis S, Gale NW, Aldrich TH, *et al*: Ligands for EPH-related receptor tyrosine kinases that require membrane attachment or clustering for activity. *Science* 266: 816-819, 1994.
- Beckmann MP, Cerretti DP, Baum P, *et al*: Molecular characterization of a family of ligands for eph-related tyrosine kinase receptors. *EMBO J* 13: 3757-3762, 1994.
- Fletcher FA, Huebner K, Shaffer LG, *et al*: Assignment of the gene (EPLG2) encoding a high-affinity binding protein for the receptor tyrosine kinase *elk* to a 200-kilobasepair region in human chromosome Xq12. *Genomics* 25: 334-335, 1995.
- Bennett BD, Zeigler FC, Gu Q, *et al*: Molecular cloning of a ligand for the EPH-related receptor protein-tyrosine kinase *Htk*. *Proc Natl Acad Sci USA* 92: 1866-1870, 1995.
- Pandey A, Lindberg RA and Dixit VM: Cell signalling. Receptor orphans find a family. *Curr Biol* 5: 986-989, 1995.
- Cerretti DP, Lyman SD, Kozlosky CJ, *et al*: The genes encoding the eph-related receptor tyrosine kinase ligands LERK-1 (EPLG1, Epl1), LERK-3 (EPLG3, Epl3), and LERK-4 (EPLG4, Epl4) are clustered on human chromosome 1 and mouse chromosome 3. *Genomics* 33: 277-282, 1996.
- Cerretti DP, Copeland NG, Gilbert DJ, *et al*: The gene encoding LERK-7 (EPLG7, Epl7), a ligand for the Eph-related receptor tyrosine kinases, maps to human chromosome 5 at band q21 and to mouse chromosome 17. *Genomics* 35: 376-379, 1996.
- Gale NW, Flenniken A, Compton DC, *et al*: *Elk-L3*, a novel transmembrane ligand for the Eph family of receptor tyrosine kinases, expressed in embryonic floor plate, roof plate and hind-brain segments. *Oncogene* 13: 1343-1352, 1996.
- Gale NW, Holland SJ, Valenzuela DM, *et al*: Eph receptors and ligands comprise two major specificity subclasses and are reciprocally compartmentalized during embryogenesis. *Neuron* 17: 9-19, 1996.
- Aasheim HC, Pedeutour F, Grosgeorge J and Logtenberg T: Cloning, chromosomal mapping, and tissue expression of the gene encoding the human Eph-family kinase ligand ephrin-A2. *Biochem Biophys Res Commun* 252: 378-382, 1998.
- Flanagan JG and Vanderhaeghen P: The ephrins and Eph receptors in neural development. *Annu Rev Neurosci* 21: 309-345, 1998.
- Holder N and Klein R: Eph receptors and ephrins: effectors of morphogenesis. *Development* 126: 2033-2044, 1999.
- Frisen J, Holmberg J and Barbacid M: Ephrins and their multi-talented directors of embryonic development. *EMBO J* 18: 5159-5165, 1999.
- Wilkinson DG: Multiple roles of Eph receptors and ephrinin neural development. *Nat Rev Neurosci* 2: 155-164, 2001.
- Mancia F and Shapiro L: ADAM and Eph: how Ephrin-signaling cells become detached. *Cell* 123: 185-187, 2005.
- Katoh M: *WNT* and *FGF* gene clusters. *Int J Oncol* 21: 1269-1273, 2002.
- Katoh M: Regulation of WNT signaling molecules by retinoic acid during neuronal differentiation in NT2 cells: threshold model of WNT action. *Int J Mol Med* 10: 683-687, 2002.
- Heller RS, Klein T, Ling Z, Heimberg H, Katoh M, Madsen OD and Serup P: Expression of *WNT*, *Frizzled*, *sFRP*, and *DKK* genes in adult human pancreas. *Gene Expr* 11: 141-147, 2003.
- Katoh M: *WNT2* and human gastrointestinal cancer. *Int J Mol Med* 12: 811-816, 2003.

 SPANDIDOS M and Katoh M: Identification and characterization of *BCL9L* gene and mouse *Bcl9l* gene *in silico*. Int J Mol Med 12: 643-649, 2003.

21. Garciadiego-Cazares D, Rosales C, Katoh M and Chimal-Monroy J: Coordination of chondrocyte differentiation and joint formation by $\alpha 5\beta 1$ integrin in the developing appendicular skeleton. Development 131: 4735-4742, 2004.
22. Katoh M and Katoh M: Comparative genomics on *WNT8A* and *WNT8B* genes. Int J Oncol 26: 1129-1133, 2005.
23. Katoh M: Molecular evolution of *WNT2B* orthologs. Int J Oncol 26: 1135-1139, 2005.
24. Katoh M: Comparative genomics on *WNT3-WNT9B* gene cluster. Int J Mol Med 15: 743-747, 2005.
25. Swain RK, Katoh M, Medina A and Steinbeisser H: *Xenopus* frizzled-4S, a splicing variant of Xfz4, is a context-dependent activator and inhibitor of Wnt/ β -catenin signaling. Cell Commun Signal 3: 12, 2005.
26. Katoh M: Epithelial-mesenchymal transition in gastric cancer. Int J Oncol 27: 1677-1683, 2005.
27. Katoh M: WNT2B: comparative integromics and clinical application. Int J Mol Med 16: 1103-1108, 2005.
28. Katoh Y and Katoh M: Comparative genomics on *DKK1* orthologs. Int J Oncol 27: 275-279, 2005.
29. Katoh Y and Katoh M: Comparative genomics on *DKK2* and *DKK4* orthologs. Int J Mol Med 16: 477-481, 2005.
30. Katoh M and Katoh M: Comparative genomics on *FGF20* orthologs. Oncol Rep 14: 287-290, 2005.
31. Katoh M and Katoh M: Comparative genomics on *FGF8*, *FGF17*, and *FGF18* orthologs. Int J Mol Med 16: 493-496, 2005.
32. Katoh Y and Katoh M: Comparative genomics on *FGF16* orthologs. Int J Mol Med 16: 959-963, 2005.
33. Battle E, Henderson JT, Begthel H, *et al*: β -catenin and TCF mediate cell positioning in the intestinal epithelium by controlling the expression of EphB/EphrinB. Cell 111: 251-263, 2002.
34. Battle E, Bacani J, Begthel H, *et al*: EphB receptor activity suppresses colorectal cancer progression. Nature 435: 1126-1130, 2005.
35. Katoh M: Paradigm shift in gene-finding method: from bench-top approach to desk-top approach. Int J Mol Med 10: 677-682, 2002.
36. Katoh M and Katoh M: Evolutionary conservation of *CCND1-ORAOV1-FGF19-FGF4* locus from zebrafish to human. Int J Mol Med 12: 45-50, 2003.
37. Katoh M and Katoh M: *CLDN23* gene, frequently down-regulated in intestinal-type gastric cancer, is a novel member of *CLAUDIN* gene family. Int J Mol Med 11: 683-689, 2003.
38. Katoh M and Katoh M: Identification and characterization of human *MPP7* gene and mouse *Mpp7* gene *in silico*. Int J Mol Med 13: 333-338, 2004.
39. Katoh M and Katoh M: Identification and characterization of *Crumbs* homolog 2 gene at human chromosome 9q33.3. Int J Oncol 24: 743-749, 2004.
40. Katoh M and Katoh M: Identification and characterization of human *HES2*, *HES3*, and *HES5* genes *in silico*. Int J Oncol 25: 529-534, 2004.
41. Katoh M and Katoh M: Identification and characterization of human *HESL*, rat *Hesl* and rainbow trout *hesl* genes *in silico*. Int J Mol Med 14: 747-751, 2004.
42. Katoh M and Katoh M: Comparative genomics on *WNT5A* and *WNT5B* genes. Int J Mol Med 15: 749-753, 2005.
43. Katoh Y and Katoh M: Comparative genomics on *WNT11* gene. Int J Mol Med 15: 879-883, 2005.
44. Katoh Y and Katoh M: Comparative genomics on *VANGL1* and *VANGL2* genes. Int J Oncol 26: 1435-1440, 2005.
45. Katoh Y and Katoh M: Comparative genomics on *SFRP1* orthologs. Int J Oncol 27: 861-865, 2005.
46. Katoh Y and Katoh M: WNT antagonist, SFRP1, is Hedgehog signaling target. Int J Mol Med 17: 171-175, 2006.
47. Katoh Y and Katoh M: Comparative genomics on HHIP family orthologs. Int J Mol Med 17: 391-395, 2006.
48. Kullander K, Croll SD, Zimmer M, *et al*: Ephrin-B3 is the midline barrier that prevents corticospinal tract axons from recrossing, allowing for unilateral motor control. Genes Dev 15: 877-888, 2001.
49. Kullander K, Butt SJ, Lebreton JM, *et al*: Role of EphA4 and EphrinB3 in local neuronal circuits that control walking. Science 299: 1889-1892, 2003.
50. Benson MD, Romero MI, Lush ME, *et al*: Ephrin-B3 is a myelin-based inhibitor of neurite outgrowth. Proc Natl Acad Sci USA 102: 10694-10699, 2005.