

Expression of Myc target gene *mina53* in subtypes of human lymphoma

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Abstract. *Mina53* (*mina*) was identified as a gene, which is directly induced by the oncogene *c-myc*. Elevated expression of *Mina53* protein was found in >80% of colon cancer and esophageal squamous cell carcinoma (ESCC). Patients with high expression of *Mina53* had shorter survival, suggesting the prognostic usefulness of *Mina53*. We studied *Mina53* expression in lymphoma subtypes to examine its diagnostic significance and its possible role in lymphoma-genesis. Surgical cases of 28 lymphoma and 4 non-neoplastic tissues were stained immunochemically using anti-*Mina53* monoclonal antibody. *Mina53* expression correlated well with *c-Myc* expression in lymphoma, suggesting that *c-Myc* is a controlling factor for *mina53* expression also in lymphomas. Although the expression of *Mina53* as well as *c-Myc* was less frequent in lymphoma compared with those of colon and ESCC, increased expression of *Mina53* was found in Burkitt-like lymphoma (1/1), Hodgkin's lymphoma (3/5), diffuse large B cell lymphoma (DLBCL) (5/13), lymphomas with a transition from follicular to DLBCL (1/2), with none in follicular (0/4) and T cell lymphoma (0/3). Analyses of the data suggested that *Mina53* was frequently expressed in aggressive types of B cell lymphoma. To get more information about the expression of *Mina53* in DLBCL, which most frequently occurs among lymphomas, we analyzed the expression of *Mina53* in another 21 DLBCL specimens, which were in more advanced stages than those

described above. The expression level of *Mina53* correlated to the international prognostic index (IPI) values with statistical significance ($r=0.477$, $P=0.0275$). Notably, in this group, *Mina53* expression did not correlate with *c-Myc* expression, suggesting that other factor(s) besides *c-Myc* largely affect the expression of *Mina53* in advanced DLBCL. These results suggest that although *Mina53* expression is not prominent in lymphoma in general, it may be related to tumor progression of B cell lymphoma.

Introduction

The *c-myc* proto-oncogene belongs to a family of related genes, which include *L-myc* and *N-myc* (1). Deregulated expression of *c-myc* is common in cancer and the activation of *c-myc* by chromosomal translocation, gene amplification and proviral insertion was described in a variety of tumors from several species, including humans (2-5). Constitutive *c-myc* expression inhibits exit from the cell cycle and abolishes differentiation (1,6). *c-Myc* activity is sufficient to drive quiescent cells into the cell cycle in the absence of growth factors (7). Studies in rodent model systems have shown that overexpression of *c-Myc* can cause malignant transformation and that sustained tumor growth depends on its continued expression (8-11). Thus, *c-Myc* is a potent and critical promoter of tumor cell proliferation.

Although the precise mechanism remains poorly understood, *c-Myc* is thought to exert its biological effects by regulating the expression of target genes (12,13). To date, many Myc target genes have been identified. While many of them are related to cell proliferation, they have a variety of functions and their modes of contribution to tumorigenesis are different to each other. A novel gene, *mina53*, whose expression was demonstrated through experimental evidence to be directly induced by *c-myc* has been identified (14). The *mina53* gene encodes a protein with a molecular weight of 53 kDa that is localized in the nucleus. Expression of *mina53* is increased during cell proliferation and specific inhibition of *mina53* expression by an RNA interference (RNAi) method

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suppressed cell proliferation in certain cultured cell lines (14-16). Using a specific monoclonal antibody against human Mina53 protein, it was found that expression of Mina53 is frequently increased in human colon cancer (15) and esophageal squamous cell carcinoma (ESCC) (16), but not in non-neoplastic well proliferating cells. In the case of ESCC, patients with high expression of Mina53 had shorter survival (16). These results suggest that Mina53 is a cancer related protein, which may play a role in c-Myc-induced carcinogenesis.

It is difficult to predict the prognosis of lymphoma. Although the International Prognostic Index (IPI) is useful in predicting the outcome of Diffuse large B cell lymphoma (DLBCL), patients with identical IPI still exhibit a marked variety in survival and additional molecular markers should help the diagnosis of the disease (17). In human lymphoma, translocation of the *c-myc* gene to the heavy or light chain of the immunoglobulin locus is seen in virtually every case of Burkitt's lymphoma and in certain cases of lymphomas that have progressed from indolent ones (18-21). The presence of *c-myc* rearrangements in higher-grade tumors may suggest a role for c-Myc in the clonal evolution of low-grade tumors into more aggressive lymphomas (22). Transcription of the translocated *c-myc* gene is greater than that seen in resting B cells (20,23). A subgroup of patients with dual translocation of *c-myc* and *bcl-2* appear to have an extremely poor outcome despite high-dose chemotherapy (24). c-Myc induces lymphoma in the bursa of Fabricius in bird (25,26). Transgenic mice bearing *c-myc* gene developed clonal B cell malignancies and constitutive expression of c-Myc caused human B lymphoblastic cells that were immortalized by the Epstein-Barr virus to be tumorigenic in immunodeficient mice (8,27,28). These results suggest that c-Myc plays a causal role in lymphomagenesis and may be associated with tumor progression.

We studied the expression of Mina53 in several subtypes of lymphoma using immunohistochemical methods to examine a possible role of Mina53 in lymphoma-genesis. We showed that Mina53 is markedly expressed in certain subtypes of lymphoma.

Materials and methods

Antibodies. Anti-human Mina53 mouse monoclonal antibody M532 was used in this study. The specificity of M532 to human Mina53 was demonstrated in a previous study (15). Rabbit anti-c-Myc polyclonal antibody (sc-764) (Santa Cruz Biotechnology, Santa Cruz, CA), mouse anti-Ki-67 monoclonal antibody (clone MIB-1) (Dako A/S, Glostrup, Denmark), peroxidase-labeled goat anti-mouse IgG Fab' (Nichirei, Tokyo, Japan) and peroxidase-labeled goat anti-rabbit IgG Fab' (Nichirei) were purchased.

Tissues for immunostaining. Routinely processed formalin-fixed and paraffin-embedded specimens from 28 patients with lymphoma were obtained from the Department of Otolaryngology, Kurume University Hospital and St. Mary's Hospital, Kurume. The sections included B cell lymphomas (n=25) and T cell lymphomas (n=3). The sections for B cell lymphomas were classified into the major subtypes of

lymphoma as follicular lymphoma (n=4), diffuse large B cell lymphoma (n=13), Hodgkin's lymphoma (n=5) and Burkitt-like lymphoma (n=1). There were also 2 cases in which there was a transition from follicular to diffuse large B cell lymphoma. Four apparently non-neoplastic lymphoid tissues were also used in this study. The characteristics of the tissues are outlined in Table I. Additional routinely processed formalin-fixed and paraffin-embedded specimens from 21 patients with diffuse large B cell lymphoma were obtained from the Department of Internal Medicine, Saga University Hospital.

Immunostaining of lymphoma tissues. Immunostaining was performed essentially as previously described (15) with minor modifications. Briefly, thin sections of formalin-fixed, paraffin-embedded tissue specimens were mounted on glass slides, deparaffinized, hydrated and autoclaved for 20 min in 10 mM sodium citrate buffer, pH 6.0, for antigen retrieval. After pretreatment with 3% H₂O₂ in PBS and then with 1% skim milk in PBS, sections were incubated with the primary antibodies overnight at 4°C and then with peroxidase-labeled goat anti-mouse IgG Fab' or peroxidase-labeled goat anti-rabbit IgG Fab' (Nichirei). Color was developed with 3,3'-diaminobenzidine and H₂O₂. After light counterstaining with haematoxylin, the slides were dehydrated, coverslipped and observed with an Olympus AX80 microscope (Olympus Optical, Tokyo, Japan).

Evaluation of immunostaining. The staining intensity of each section was scored on a scale from 0 to 4 by visual observation. The section with the highest staining intensity was scored as 4, the lowest as 1 and no staining at all as 0. The percentage of tumor cells stained by the antibodies was also estimated as 0, 0-50, 50, 50-100 and 100%. The staining index was calculated as the staining intensity x percentage of cells stained. For calculation of the staining index, the percentage of cells stained was estimated as 0, 25, 50, 75 and 100%. Sections were defined as positive or increased expression when the staining intensity was higher than the average staining intensity of the non-neoplastic tissues. The correlations between the staining intensities of the proteins were examined by using the Spearman rank correlation coefficient.

Results

Mina53 expression in lymphoma. Anti-Mina53 monoclonal antibody was used to examine the expression of Mina53 in lymphoma tissues. Initially, Mina53 expression was investigated in 4 apparently non-neoplastic lymphoid tissues. As shown in Table I, Mina53 was only weakly expressed in 3 cases, while no expression was detected in the other case. These results were consistent with our previous results (15,16).

The section shown in Fig. 1a contained diffuse large B cell lymphoma (DLBCL) and Mina53 was markedly expressed in the tumor cells. When the expression of Mina53 was analyzed in 13 cases of DLBCL, 5 cases (38.5%) contained elevated amounts of Mina53, compared to lymphocytes in the non-neoplastic tissues (Table II). The section shown in Fig. 1b contained follicular lymphoma, in which Mina53 expression

Table I. Characteristics of lymphoma tissues and summary of immunohistochemistry.

No.	Age	Sex	Histological type	Staining intensity			% Tumor cells stained			Staining index		
				Mina53	Myc	Ki-67	Mina53	Myc	Ki-67	Mina53	Myc	Ki-67
1	67	F	DLBCL	0	1.5	3	0	0-50	0-50	0	37.5	75
2	71	M	DLBCL	1	3	3	100	100	100	100	300	300
3	45	M	DLBCL	1	2	4	0-50	0-50	50-100	25	50	300
4	63	F	DLBCL	1	2	4	50	50-100	50-100	50	150	300
5	80	M	DLBCL	1	2	3	50-100	50-100	50-100	75	150	225
6	72	F	DLBCL	1	1.5	3	50	0-50	50-100	50	37.5	225
7	57	M	DLBCL	1	3	3	100	50	50-100	100	150	225
8	80	M	DLBCL	1	3	3	50-100	100	50-100	75	300	225
9	76	M	DLBCL	1.5	3	3	100	100	50-100	150	300	225
10	63	M	DLBCL	2	2	3	100	100	100	200	200	300
11	54	M	DLBCL	2	3	4	100	100	100	200	300	400
12	60	M	DLBCL	2	3	3	100	100	50-100	200	300	225
13	43	M	DLBCL	3	3	4	100	100	50-100	300	300	300
14	52	F	FL	0	1	3	0	25	0-50	0	25	225
15	53	M	FL	1	1	3	50	0-50	100	50	25	300
16	44	M	FL	1	2	3	100	100	0-50	100	200	75
17	56	M	FL	0	1	3	0	10-50	0-50	0	25	75
18	50	M	FL>DLBCL	1	1	3	50-100	50-100	0-50	75	75	75
19	47	F	FL>DLBCL	1.5	3	3	100	100	50-100	150	300	225
20	18	M	LRCHL	1	1	3	50-100	50	100	75	50	300
21	74	M	NSHL	0	2	3	0	100	100	0	200	300
22	34	M	NSHL	1.5	1	3	50-100	10	50-100	112.5	25	150
23	18	M	NSHL	2	2	3	100	100	50-100	200	200	225
24	65	M	MCHL	3	3	3	100	100	100	300	300	300
25	75	F	TCL	0	1	3	0	0-50	0-50	0	25	75
26	68	F	TCL	1	2	3	50-100	50	50-100	75	100	225
27	30	M	TCL	0	1	3	0	0-50	50-100	0	25	225
28	55	M	BL	4	3	4	100	100	100	300	400	300
29	19	F	N	1	2	3	50	0-50	100	50	50	300
30	79	F	N ^a	1.5	2	3	0-50	0-50	100	37.5	50	300
			^b	1.5	2	3	0-50	0-50	10	37.5	50	30
31	56	M	N	0	2	3	0	0-50	100	0	50	300
32	60	M	N	1	2	3	100	0-50	50-100	100	50	225

DLBCL, Diffuse large B cell lymphoma; FL, follicular lymphoma; FL>DLBCL, contained follicular and diffuse large B cell lymphoma; LRCHL, Lymphocyte-rich classical Hodgkin's lymphoma; NSHL, Nodular sclerosis Hodgkin's lymphoma; MC Hodgkin's lymphoma, Mixed cellularity Hodgkin's lymphoma; TCL, T cell lymphoma; BLL, Burkitt-like lymphoma; N, non-neoplastic. ^aGerminal center B cells. ^bNon-germinal B cells.

was hardly detected. As demonstrated in Table II, all cases of follicular lymphoma did not show increased expression of Mina53. There were two cases (#18 and #19) with a transition from follicular to diffuse large B cell lymphoma in which Mina53 expression was slightly increased in 1 case (50%) (Table II). It was shown that diffuse large B cell lymphoma is more aggressive than follicular B cell lymphoma and some proportion of follicular B cell lymphomas progress to diffuse large B cell lymphoma (29,30).

The section shown in Fig. 1c contained a mixed cellularity Hodgkin's lymphoma (MCHL). Tumor cells were markedly stained by anti-Mina53 antibody (Table II) whereas most non-neoplastic cells showed little staining (Fig. 1c). Elevated expression of Mina53 was found in 2 out of 3 cases of nodular sclerosis Hodgkin's lymphoma (NSHL) (Tables I and II), but

not in the 1 case of lymphocyte-rich classical Hodgkin's lymphoma (LRCHL) (Tables I and II). It was revealed that MCHLs are more aggressive than NSHLs and that LCHLs show far less aggressive clinical behavior (31-33). Therefore, while Mina53 expression was increased in 60% of all cases of Hodgkin's lymphoma (Table II), aggressive forms of Hodgkin's lymphoma tend to express Mina53 highly and frequently (Tables I and II).

We investigated one case of Burkitt-like lymphoma, a very aggressive lymphoma and it was found that Mina53 was highly expressed in the tumor cells of the lymphoma (Fig. 1d). All cases of T cell type lymphoma (3/3) showed very little or no expression of Mina53 (Tables I and II). Thus, our results above suggest that increased expression of Mina53 is associated with high-grade B cell lymphoma.

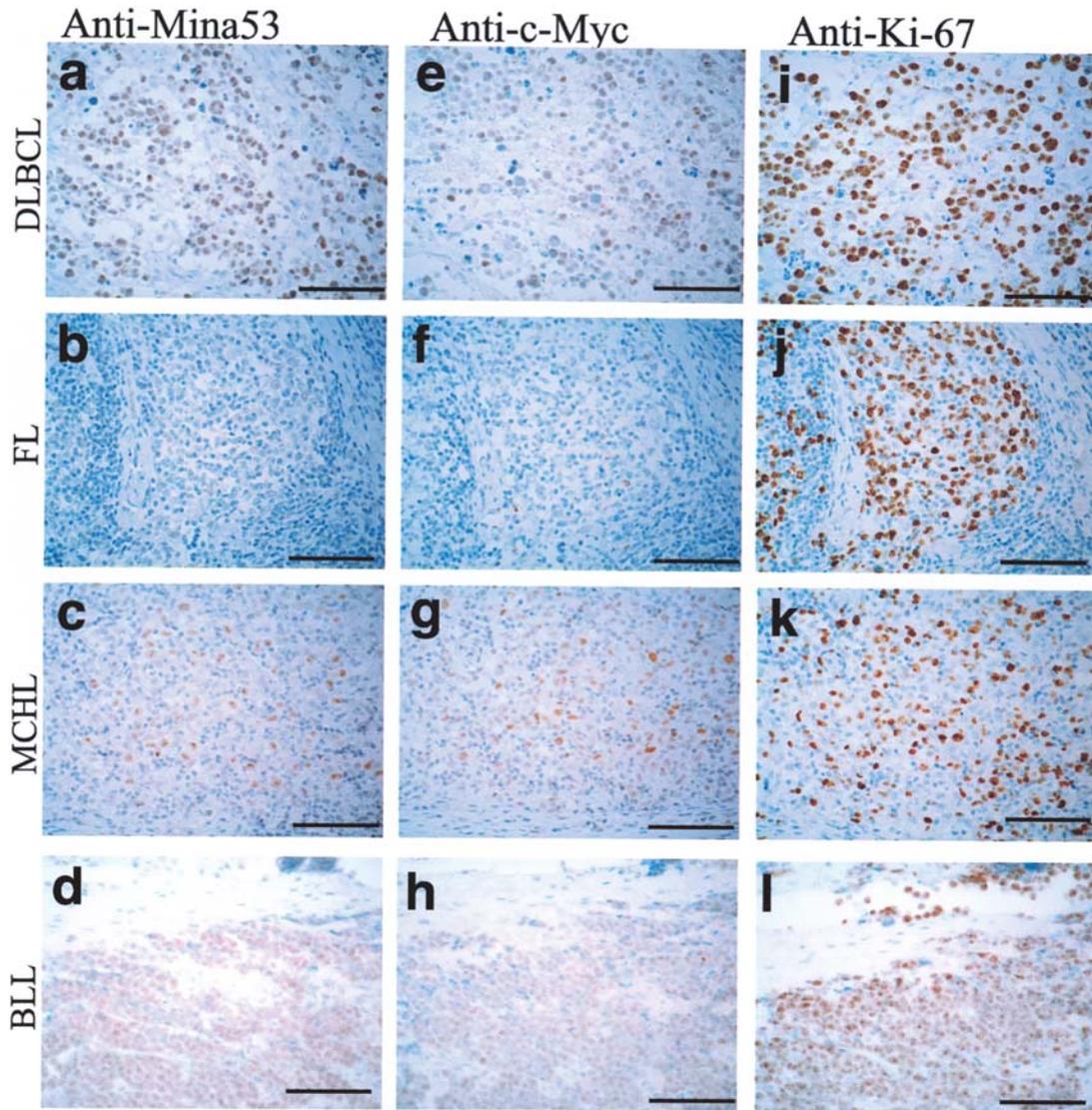


Figure 1. Immunohistochemical analysis of Mina53, Myc and Ki-67 proteins in lymphoma tissues. Serial sections were stained by anti-Mina53 (left), anti-c-Myc (middle), or anti-Ki-67 (right) antibody. Staining of diffuse large B cell lymphoma for Mina53 (a), Myc (e), or Ki-67 (i). Section (a) showed marked expression of Mina53 in tumor cells. Staining of follicular lymphoma for Mina53 (b), c-Myc (f) or Ki-67 (j). Section (b) showed lack of Mina53 expression. Staining of a mixed cellularity Hodgkin's lymphoma for Mina53 (c), c-Myc (g), or Ki-67 (k). Section (c) showed marked Mina53 expression. Staining of Burkitt-lymphoma for Mina53 (d), c-Myc (h), or Ki-67 (l). Section (d) showed high expression of Mina53. Scale bars, 75 μ m.

Table II. Summary of Mina53 staining in lymphoma by histological type.

Histological Type ^a	Cases	Mina53 staining intensity						Positive rate (%)	
		-	+	+1.5	+2	+3	+4	Mina53	Myc
DLBCL	13	1	7	1	3	1	0	38.5	53.8
FL	4	2	2	0	0	0	0	0	0
FL>DLBCL	2	0	1	1	0	0	0	50	50
LRCHL	1	0	1	0	0	0	0	0	0
NSHL	3	1	0	1	1	0	0	66.7	0
MCHL	1	0	0	0	1	1	0	100	100
BLL	1	0	0	0	0	1	1	100	100
TCL	3	1	2	0	0	0	0	0	0

^aDLBCL, Diffuse large B cell lymphoma; FL, follicular lymphoma; FL>DLBCL, contained follicular and diffuse large B cell lymphoma; LRCHL, Lymphocyte-rich classical Hodgkin's lymphoma; NSHL, Nodular sclerosis Hodgkin's lymphoma; MCHL, Mixed cellularity Hodgkin's lymphoma; TCL, T cell lymphoma; BLL, Burkitt-like lymphoma.

Table III. Summary of Mina53 and c-Myc expression in a subset of DLBCL lymphoma.

Patient No.	Age	Gender	Subtype	Staining intensity		Staining index		IPI
				Mina53	Myc	Mina53	Myc	
33	38	F	NonGCB DLBL	2	1	150.0	25.0	3
34	75	F	GCB DLBL	1	1	50.0	25.0	1
35	59	F	NonGCB DLBL	2.5	1	250.0	25.0	3
36	63	F	GCB DLBL	1.5	1	112.5	25.0	3
37	67	M	NonGCB DLBL	1	2	50.0	150.0	1
38	80	F	NonGCB DLBL	2	3	200.0	225.0	2
39	56	M	NonGCB DLBL	1.5	1	112.5	25.0	4
40	79	F	GCB DLBL	2.5	1	250.0	25.0	5
41	32	F	GCB DLBL	2	1.5	200.0	37.5	3
42	47	M	NonGCB DLBL	2	3	200.0	225.0	2
43	69	M	NonGCB DLBL	3	2	300.0	150.0	5
44	65	M	GCB DLBL	2	2.5	200.0	187.5	1
45	63	M	GCB DLBL	3	2	300.0	150.0	3
46	73	F	NonGCB DLBL	3	1	225.0	25.0	2
47	72	F	GCB DLBL	2	1	150.0	25.0	4
48	65	M	NonGCB DLBL	2	3	200.0	75.0	3
49	41	M	NonGCB DLBL	2	1	200.0	25.0	1
50	78	F	GCB DLBL	2.5	2	250.0	100.0	4
51	62	F	GCB DLBL	3	2	300.0	150.0	4
52	75	M	GCB DLBL	4	1	400.0	25.0	3
53	62	M	NonGCB DLBL	1	2	100.0	50.0	1
Average						200.0	83.3	2.8

Table IV. Correlation between Mina53 expression and other factors.

Factors	Cases	r	95% CI	P
All lymphoma cases				
Mina53, Myc	28	0.775	0.565-0.891	<0.0001
Mina53, Ki-67	28	0.427	0.064-0.690	0.0225
Diffuse large B cell lymphoma with low average IPI value				
Mina53, Myc	13	0.724	0.288-0.911	0.0038
Mina53, Ki-67	13	0.490	-0.084-0.820	0.0900
Mina53, IPI	12	-0.402	-0.793-0.223	0.2007
Myc, IPI	12	0.093	-0.508-0.633	0.7803
Diffuse large B cell lymphoma with high average IPI value				
Mina53, Myc	21	0.157	-0.295-0.551	0.503
Mina53, IPI	21	0.477	-0.058-0.754	0.0275
Myc, IPI	21	-0.149	-0.508-0.633	0.5230

CI, confidence interval. ^aFor diffuse large B cell lymphoma subgroup.

Expression of Mina53 in diffuse large B cell lymphomas with more advanced stages. The results above suggest that increased expression of Mina53 may be associated with advanced stages in some cases of B cell lymphomas. However, when staining index of Mina53 was compared with IPI values (international prognostic index), no correlation was observed (Table IV). This may be due to the fact that there are few

progressed cancer specimens examined. In the specimens in Table I, the lymphomas have relatively lower IPI values (the average of the IPI value was 1.7), because most of the specimens were obtained from the Department of Otolaryngology, where patients with early stages of the lymphomas usually receive surgical treatment. To clarify this point, the expression of Mina53 was further examined in another 21 cases

of DLBCL with more advanced stages (the average of the IPI value was 2.8), which were obtained from the Department of internal medicine. The results are shown in Table III. When the staining index of *Mina53* was compared to IPI values, a positive correlation was found between them with statistical significance ($r=0.477$, $P=0.0275$). These results suggest that *Mina53* is highly expressed in aggressive DLBCL.

c-Myc was also stained in these specimens. Notably, the staining level of *c-Myc* was not similar to that of *Mina53* ($r=0.157$, $p=0.5030$) (Tables III and IV). In this group, the averages of staining indices for *c-Myc* and *Mina53* were 83.3 and 200, respectively (Table III), while those for *c-Myc* and *Mina53* were 198 and 117, respectively, in the group of DLBCL with lower IPI values (Table I). Although these values are not directly compared between two groups, which were obtained from different hospitals, the results suggest that the expression of *Mina53* is higher in the group of patients with the high average of the IPI, while that of *c-Myc* is higher in the group of patients with the low average of the IPI.

Comparison of expression of Mina53, Myc and Ki-67 in lymphoid tissues. Serial sections from the above specimens were stained with anti-*c-Myc* antibody. *c-Myc* was detected in certain lymphoma tissues, including diffuse large B cell lymphoma (Fig. 1e-h). The staining level of *c-Myc* in each section was quite similar to that of *Mina53* ($r=0.775$, $p<0.0001$) (Table IV and Fig. 1). These results suggest that *Mina53* expression is generally closely related to *c-Myc* expression in lymphoid cells. However, as mentioned above, expression of *Mina53* did not correlate with that of *c-Myc* in the specimens with advanced DLBCL (Tables III and IV).

The expression of *Ki-67*, a well-used cell proliferation marker, was also investigated in serial sections in all cases shown in Table I. As reported before (34,35), anti-*Ki-67* antibody stained lymphoma tissues intensely (Fig. 1i-l). In all cases, an antibody against *Ki-67* stained tumor cells strongly and frequently. *Ki-67* staining was also detected in follicular lymphoma lacking *Mina53* and *c-Myc* staining [Fig. 1b (*Mina53*), f (*Myc*) and j (*Ki-67*)]. The staining levels of *Mina53* showed a statistically significant correlation with those of *Ki-67* ($r=0.427$, $p=0.0225$) (Table IV and Fig. 1), suggesting that *Mina53* is related to cell proliferation in lymphoma. However, *Ki-67* was also highly expressed in nearly all cells in the non-neoplastic germinal center, in which *Mina53* was faintly expressed (Table I; #30). Thus *Mina53* is not always expressed in proliferating cells.

Discussion

The elevation of Mina53 expression is frequently found in some aggressive types of lymphomas. In our previous studies we demonstrated that *Mina53* is highly expressed in >80% of ESCC and colon cancer cases and it was suggested that increased expression of *Mina53* is a hallmark in these cancers (15,16). In this study we found that although the expression of *Mina53* was less frequent in lymphoma, *Mina53* is markedly expressed in one third of all lymphoma cases (Table I).

Mina53 is poorly expressed in non-neoplastic lymphoid tissues including germinal centers. Burkitt-like lymphoma, an aggressive form of lymphoma, showed very high expression of

Mina53. We also found very high expression of *Mina53* in Burkitt's lymphoma cell line Daudi (data not shown). Among Hodgkin's lymphomas, *Mina53* expression was strongest and most frequent in MCHL, which is the most aggressive, followed by NSHL, which is intermediate grade, with none in the indolent LRCHL (Table I). *Mina53* expression was found frequently in DLBCL but not in follicular B cell lymphomas. In a group of DLBCL with relatively advanced stages (Table III), there was positive correlation between them with statistical significance ($r=0.477$, $P=0.0275$, Table IV). These results suggest that *Mina53* expression was found more often in aggressive DLBCL. Together, these results suggest that *Mina53* may be activated during the transformation from indolent to more aggressive forms of the disease.

Mina53 expression is cell type-specific rather than cell proliferation-specific. The expression of *Mina53* was compared with that of *Ki-67*, a widely used biomarker for cell proliferation. We noted strong and frequent expression of *Ki-67* in lymphoma (Fig. 1 and Table I) and that *Ki-67* was also strongly expressed in non-neoplastic lymphoid tissues. On the other hand, *Mina53* expression was confined specifically to tumor cells, suggesting that *Mina53* is dispensable for cell proliferation of non-neoplastic lymphocyte. Therefore, *Mina53* may not directly be involved in basic mechanisms of cell proliferation but plays a role in the pathogenesis of certain B cell lymphoma.

There is precedence that the expression of *Mina53* is not associated with cell proliferation. In mouse testis there is a prominently high expression of *Mina53* in some oval-shaped cells in the periphery of the seminiferous epithelium of normal adult testis, which appears to be well-proliferating type A spermatogonia. However, the expression of *Mina53* was also detected in Sertoli cells and even in the Leydig cells, which are highly differentiated and actively functioning but no longer dividing (36). These results suggest that the expression of *Mina53* is cell type-specific but not cell proliferation-specific.

Control of Mina53 expression. In this study we found that *Mina53* expression correlated well with that of *c-Myc* ($r=0.724$, $P=0.0038$) in the specimen with relatively low IPI values. These results suggest that *c-Myc* largely determined *Mina53* expression in lymphoma at the early stages. However, in another 21 specimens of DLBCL with relatively high IPI values, the staining indices of *c-Myc* were different from those of *Mina53* ($r=0.157$, $p=0.5030$) (Table IV). Therefore, in advanced stages of DLBCL *Mina53* expression increased without high expression of *c-Myc*. As mentioned above, the expression of *Mina53* was detected in Sertoli cells and the Leydig cells, which were shown not to express *c-myc* (37). Thus these results suggest that there should be other controlling molecule(s) for *mina53* expression besides *c-myc* in testicular somatic cells and advanced DLBCL.

Lymphomagenesis and c-Myc. *c-Myc* translocations are present in virtually all cases of Burkitt's lymphoma (18-21). In our study, the expression of *c-Myc* was also high in this disease (Fig. 1 and Table I). Aggressive forms of Hodgkin's lymphoma tend to express *c-Myc* as well as *Mina53* highly and frequently (Tables I and II). These results suggest that in

these two types of B cell lymphomas, the elevated expression of c-Myc appears to be correlated to aggressiveness.

In DLBCL, c-Myc level did not correlate with IPI especially in the group of aggressive DLBCL (Tables III and IV). This suggests that although the high level expression of c-Myc may contribute to tumorigenesis in the early stages of DLBCL, it may be dispensable in the advanced stage. A similar observation was previously reported. Results from gene expression profile analysis performed before and after follicular lymphomas were transformed to DLBCL showed that expression levels of c-Myc can be increased, decreased, or unchanged during the transformation (38). Large scale analysis of gene expression during *myc*-induced lymphomagenesis in the bursa of Fabricius revealed that while overexpression of *myc* results in transformation, expression profiles of late metastatic tumors showed a large variation in *myc* overexpression levels and some showed minimal *myc* overexpression (26). These results together with the results described in this report suggest that overexpression of *myc* may be important for the early induction of these lymphomas than the maintenance of late-stage metastases.

Since c-Myc controls the expression of many genes that are related to cell proliferation, the change of only one gene *c-myc* can largely contribute to cell proliferation. Indeed, c-Myc activation induced proliferation and tumorigenic ability of pancreatic cells without additional genetic changes when apoptosis was suppressed (39). However, during the progression of the disease, many mutations are accumulated, many of which favor cell proliferation. At that stage c-Myc expression becomes dispensable and/or reduction of c-Myc expression would give an advantage for carcinogenesis since one Myc function, the ability inducing apoptosis, is unfavorable for cell proliferation.

In conclusion, we found that the elevated expression of Mina53 is associated with some specific subtypes of lymphoma, most plausibly in subtypes of high-grade B cell lymphoma, suggesting that Mina53 staining may allow patient selection for systematic therapies.

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