

Detection of an elevated HER2 expression in MCF-7 breast cancer cells overexpressing estrogen receptor β 1

CLAUS LATTRICH, INGOLF JUHASZ-BOESS, OLAF ORTMANN and OLIVER TREECK

Department of Obstetrics and Gynecology, University of Regensburg, D-93053 Regensburg, Germany

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Abstract. The estrogen receptor (ER) expression and HER2 amplification are important factors in determining the prognosis and therapy of breast cancer. Interactions between the two signaling pathways for example resulted in ER α -dependent regulation of HER2 expression in breast cancer cells. In this study, we investigated to what extent ER β is able to affect the HER2 expression. For this purpose, we analyzed HER2 levels in ER β 1-overexpressing clones of the breast cancer cell lines MCF-7 and SK-BR-3 and of the ovarian cancer cell lines SK-OV-3 and OVCAR-3 by both RT-PCR and Western blot analysis. Treatment with ligand 17- β estradiol diminished the HER2 expression in MCF-7 wild-type cells, an effect partially inhibited by treatment with 4-OH tamoxifen. MCF-7 breast cancer cells stably overexpressing ER β 1 exhibited elevated >5-fold HER2 mRNA levels and elevated >3-fold HER2 protein levels even in the absence of estradiol. In contrast, ER β 1 overexpression did not affect HER2 protein levels in the ER α -positive OVCAR-3 ovarian cancer cells and in the HER2 overexpressing, hormone-independent SK-BR-3 and SK-OV-3 cells. By demonstrating the elevated HER2 expression in a hormone-dependent breast cancer cell line overexpressing ER β 1, our data suggest the presence of cross-talk between the two receptors. This is one of the molecular mechanisms underlying the significant ER β /HER2 co-expression observed in recent clinical studies.

Introduction

The growth of hormone-dependent breast and ovarian cancer cells can be regulated by estrogen receptors (ER) and receptor tyrosine kinases such as HER2. Estrogen receptors ER α and - β are expressed in normal breast and ovary cancer cells and in malignancies derived from these tissues (1-3). Although the

role of ER β in breast and ovary is not fully elucidated, the interaction of ER α and - β is important for the normal development and function of these tissues. In animal studies, while ER α has been shown to be essential for normal mammary gland development, ER β effects are more subtle, with roles in terminal differentiation (4). ER β is the dominant estrogen receptor in normal breast tissues (5-7), but its expression declines during tumorigenesis (8,9) suggesting a role for ER β as a tumor suppressor in breast tissues (10). However, ~55-72% of primary breast cancer cases show a lower or higher expression of ER β (11-14). Our recent studies and others suggest that ER β has similar functions in ovarian carcinogenesis (15,16). Malignant ovarian tumors originating from the epithelial surface constitute ~90% of ovarian cancers and express low levels of ER β , compared to normal tissues. In addition, the restoration of ER β in ovarian cancer cells, leads to a strong inhibition of their proliferation and motility, while apoptosis is enhanced (15).

The receptor tyrosine kinase HER2 is overexpressed in breast and ovarian cancer tissues. In 20% of all ovarian epithelial cancer, HER2 is overexpressed (17-19) and may be associated with chemoresistance (20,21). The amplification or overexpression of the HER2 gene is found in 20-25% of human breast cancers (22) and is known as a poor prognostic marker and therapeutic target (23,24).

Recent studies demonstrated an interaction between the ER α and HER2 signal transduction (25,26). For example estradiol is able to reduce HER2 expression in an ER α -dependent manner (27). This may be a reason why the coexpression of the two receptors, which is correlated with a poor prognosis, is infrequent (28-30). In contrast, a significant positive correlation between HER2 overexpression and ER β expression was reported (11,13).

In this study, we elucidated the molecular mechanism underlying this co-expression of ER β and HER2. For this purpose, we examined the effect of the ER β overexpression on HER2 expression in breast and ovarian cancer cell lines.

Materials and methods

Materials. DMEM/F12 culture medium was obtained from Invitrogen (Karlsruhe, Germany), fetal calf serum (FCS) was purchased from PAA (Pasching, Austria) and 17- β estradiol, 4-OH tamoxifen and serum replacement 2 (SR2) were obtained from Sigma (Deisenhofen, Germany). M-MLV-P

Correspondence to: Dr Oliver Treeck, Klinik für Frauenheilkunde und Geburtshilfe, Universität Regensburg, Caritas Krankenhaus St. Josef, Landshuter Str. 65, D-93053 Regensburg, Germany
E-mail: otreeck@caritasstjosef.de

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Table I. Primer sequences used for RT-PCR amplification.

Target	Oligonucleotides	Sequences 5'-3'
ER α	ER α 1	TGATGAAAGGTGGGATACGA
	ER α 2	AAGGTTGGCAGCTCTCATGT
ER β 1	ER β 1	CAAGGCCGGTGTGTTTATCT
	ER β 2	GGCGTCACTGAGACTGTGG
HER2	HER2-1	AACTGCACCCACTCCTGTGT
	HER2-2	TGATGAGGATCCCAAAGACC
β -actin	actin-2573	CTGTGGCATCCACGAAACTA
	actin-2876	CGCTCAGGAGGAGCAATG

reverse transcriptase was purchased from Promega (Mannheim, Germany). The RNeasy mini kit, RNase-free DNase set and Quantitect SYBR-Green PCR kit were obtained from Qiagen (Hilden, Germany). PCR primers were synthesized at Metabion (Planegg-Martinsried, Germany). Platinum Pfx polymerase was purchased at Invitrogen (Karlsruhe).

Cell culture. MCF-7, SK-BR-3, OVCAR-3 and SK-OV-3 breast and ovarian cancer cells were obtained from the American Type Culture Collection (Manassas, USA). Cells were maintained in phenol red-free DMEM/F12 medium supplemented with 10% FCS. Cells were cultured with 5% CO₂ at 37°C in a humidified incubator. Before treatment with E2 or 4-OH tamoxifen, FCS was replaced by 1x serum replacement 2 (SR2). Stimulation was carried out the following day using 3 nM E2 alone or in combination with 100 nM 4-OH tamoxifen for 48 h prior to RNA and protein extraction.

Reverse transcription and PCR. Total RNA was isolated by means of the RNeasy kit (Qiagen) according to the manufacturer's instructions. From 1 μ g total RNA, cDNA was synthesized using 100 U M-MLV-P reverse transcriptase (Promega), 2.5 mM dNTP mixture and 50 pM random primers (Invitrogen). For real-time PCR detection of ER α , ER β 1 or HER2 (primer sequences in Table I), 2 μ l cDNA was amplified using the Quantitect SYBR-Green PCR kit (Qiagen) and the LightCycler PCR device (Roche Diagnostics, Mannheim, Germany). The PCR program was 95°C for 15 min, followed by 35 PCR cycles (95°C for 10 sec, 56°C for 30 sec and 72°C for 30 sec) and a final extension for 5 min at 72°C, followed by a standard melting curve analysis. In all the RT-PCR experiments, a 190 bp β -actin fragment was amplified as a reference gene using intron-spanning primers actin-2573 and -2876. After performing dilution experiments with a sample cDNA over a 100-fold range confirming the PCR efficiencies of all primer pairs to be approximately equal (31), data were analyzed using the comparative $\Delta\Delta C_T$ method (32) by calculating the difference between the threshold cycle (C_T) values of the target and reference gene of each sample and then comparing the resulting ΔC_T values between different samples. In these experiments, mRNA not subjected to reverse transcription was used as a negative control to distinguish cDNA and vector or genomic DNA amplification.

Antibodies and Western blot analysis. Cells were lysed in RIPA buffer 1% (v/v) Igepal CA-630, 0.5% (w/v) sodium deoxycholate, 0.1% (w/v) sodium dodecyl sulphate (SDS) in phosphate-buffered solution (PBS) containing aprotinin and sodium orthovanadate. Aliquots containing 15 μ g protein were resolved by 10% (w/v) SDS-polyacrylamide gel electrophoresis, followed by electrotransfer to a PVDF hybrid (Amersham, UK) membrane. Immunodetection was carried out using the HER2 antibody NEU (C-18) (Santa Cruz Biotechnology, Santa Cruz, USA) or β -actin antibody (8226, ABCAM, Germany) diluted at 1:5000 in PBS containing 5% skim milk (w/v) followed by horseradish peroxidase conjugated secondary antibody (diluted at 1:50000), which was detected using the chemiluminescence (ECL) system (Amersham, Buckinghamshire, UK).

Results

Expression of ER α , ER β and HER2 in breast and ovarian cancer cell lines. To characterize the employed cancer cell lines, we first examined the expression of HER2, ER α and ER β in the cells on the mRNA level. The real-time RT-PCR analysis revealed a strong expression of HER2 in the SK-BR-3 breast and SK-OV-3 ovarian cancer cells, which are known to be HER2-overexpressing and estrogen-unresponsive. In contrast, the estrogen-responsive MCF-7 breast and OVCAR-3 ovarian cancer cells exhibited only relatively low HER2 mRNA levels (Fig. 1). All four cell lines tested expressed detectable levels of ER β 1 mRNA, which were highest in the ER α -positive, estrogen-responsive MCF-7 and OVCAR-3 cells (Fig. 2). As expected ER β 1 mRNA levels were shown to be strongly elevated in the ER β 1-overexpressing (ER β ⁺) clones of these cell lines as previously described (15,33). The levels were generated by stable transfection with an ER β 1 expression plasmid (Fig. 2). Real-time RT-PCR analysis of the ER α expression confirmed the previously described positive ER α status of MCF-7 and OVCAR-3 cells and the absence of functional ER α in the SK-BR-3 and SK-OV-3 cells (data not shown).

Effect of ER β overexpression on HER2 expression in breast and ovarian cancer cell lines. We analyzed the expression of the HER2 gene in the ER β 1-overexpressing breast and

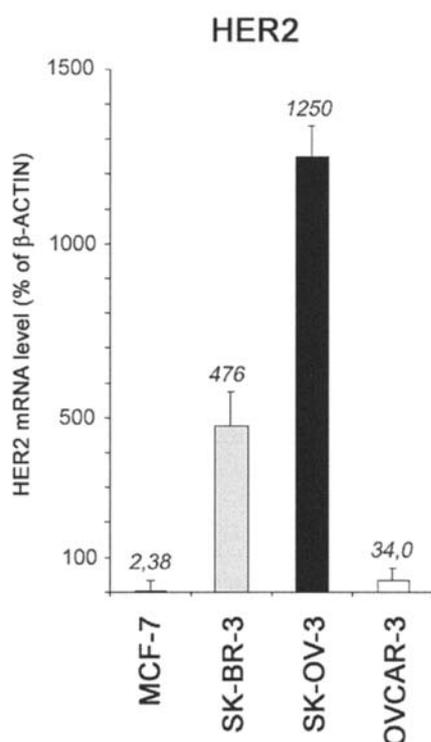


Figure 1. HER2 mRNA levels in breast and ovarian cancer cell lines. Total RNA isolated from the indicated cancer cell lines was subjected to real-time RT-PCR. Relative HER2 mRNA levels are expressed in percent of β -actin control (n=4).

ovarian cancer cell lines on the mRNA and protein levels. HER2 mRNA levels were elevated >5-fold in the MCF-7/ER β ⁺ cells (Fig. 3a) and elevated ~4-fold in the SK-OV-3/ER β ⁺ cells (Fig. 4a) cultured in serum- and steroid hormone-free medium when compared to wild-type cells. The addition of 17- β estradiol (E2) significantly decreased the HER2 mRNA levels in the hormone-dependent, ER α -positive breast cancer cell line MCF-7 down to 59.7% (Fig. 3a), but not in the hormone-independent ovarian cancer cell line SK-OV-3. This effect could be partially inhibited in the MCF-7 cells by treatment with 4-OH tamoxifen and was also present in the MCF-7/ER β ⁺ cells. In contrast, the HER2 mRNA levels were not altered in the SK-BR-3/ER β ⁺ breast cancer and OVCAR-3/ER β ⁺ ovarian cancer cells.

The HER2 protein levels as assessed by Western blot analysis were elevated >3-fold in the MCF-7 cells overexpressing ER β 1 (Fig. 3b and c). This effect was not dependent on the presence of E2 and was not affected by the addition of 4-OH tamoxifen. In contrast, in the SK-OV-3/ER β ⁺ cells exhibiting increased amounts of HER2 transcripts, we detected only a marginal increase of the HER2 protein level which was not significant (Fig. 4b and c). SK-BR-3 and OVCAR-3 cells overexpressing ER β 1 did not exhibit changes in the HER2 expression on the protein level.

Discussion

In this report we demonstrated that elevated ER β receptor levels can trigger a significant increase of HER2 expression in hormone-dependent breast cancer cells. Our findings are

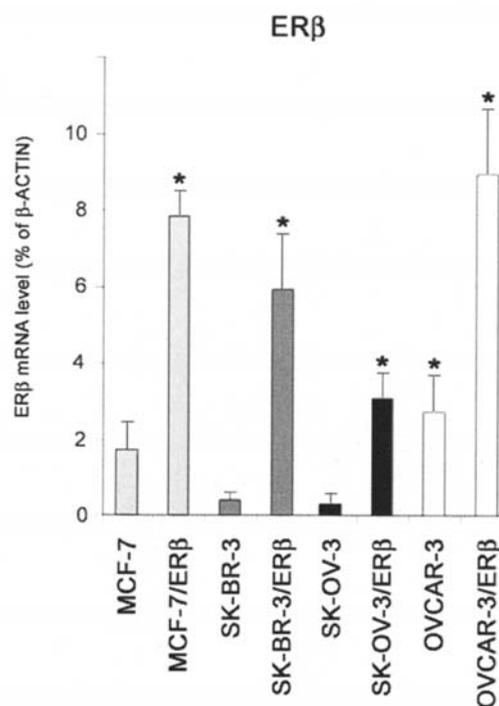


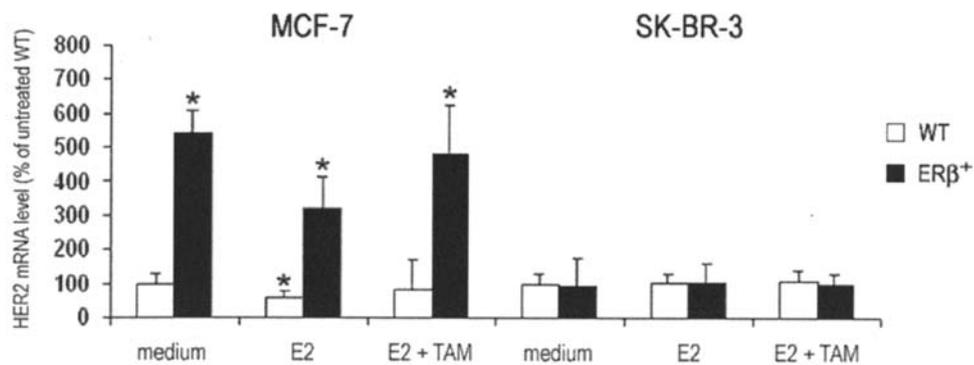
Figure 2. Estrogen receptor β mRNA levels in wild-type and stably transfected ER β ⁺ cell lines. Total RNA isolated from the indicated cancer cell lines and clones was subjected to real-time RT-PCR. Relative ER β mRNA levels are expressed in percent of β -actin control (n=4); *p<0.01 vs. wild-type.

one explanation for the recently published observation that ER β and HER2 expression is strongly and positively correlated in primary breast cancer (11,13).

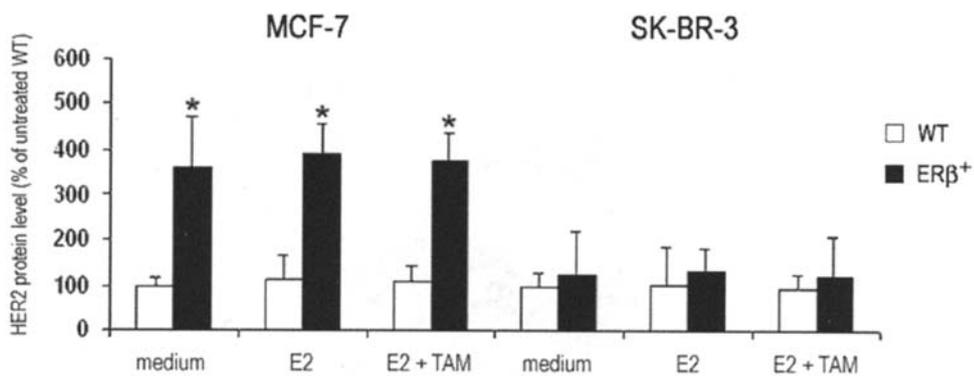
The interactions between the ER α -mediated cellular estrogen response and HER2 signal transduction have previously been reported. The activity of ER α and estrogen-related receptor α 1 is modulated by the HER2 signaling pathway (25,26). On the other hand, the HER2 expression is known to be suppressed by ER α -dependent signaling mechanisms (27). We also examined whether cross-talk exists between ER β and HER2 in breast and ovarian cancer cells.

For this purpose, we used a set of four hormone-dependent and -independent breast and ovarian cancer cell lines exhibiting different levels of ER α including the HER2-overexpressing cell lines SK-BR-3 and SK-OV-3. We compared the HER2 expression level of these wild-type cells to the expression of this receptor tyrosine kinase in ER β 1-overexpressing clones of these cell lines generated by stable transfection with an ER β 1-expression plasmid as previously described (15,33). The ER β 1-overexpressing clones used in this study exhibited elevated ER β mRNA levels which are still in a physiological range resembling the ER β expression level of tumor cell lines such as MDA-MD-231 (34) and are characterized by a significantly slowed proliferation, increased basal apoptosis and impaired motility (15,33). These characteristics of our ER β ⁺ clones are in line with reports from other groups demonstrating that ER β may negatively regulate cellular proliferation, promote apoptosis and thus may have a protective role in hormone-dependent tissues such as breast and prostate (35-37). Similar observations

a)



b)



c)

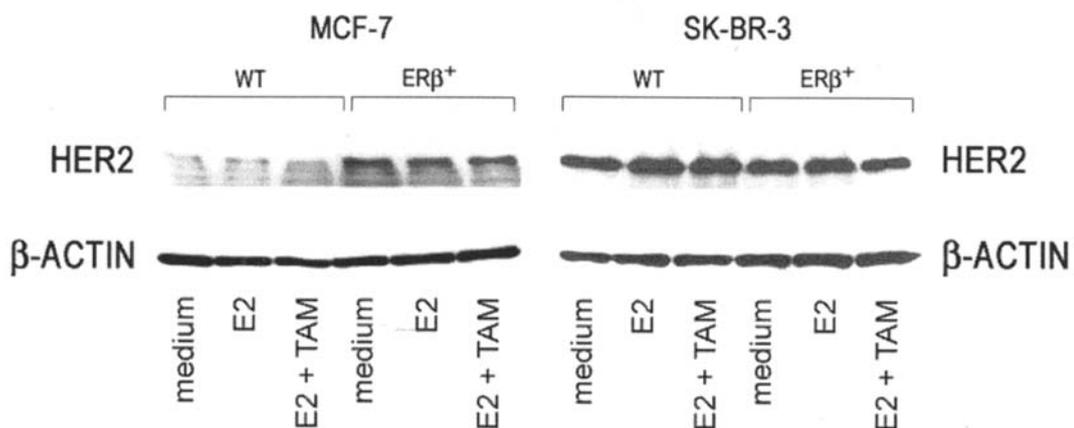
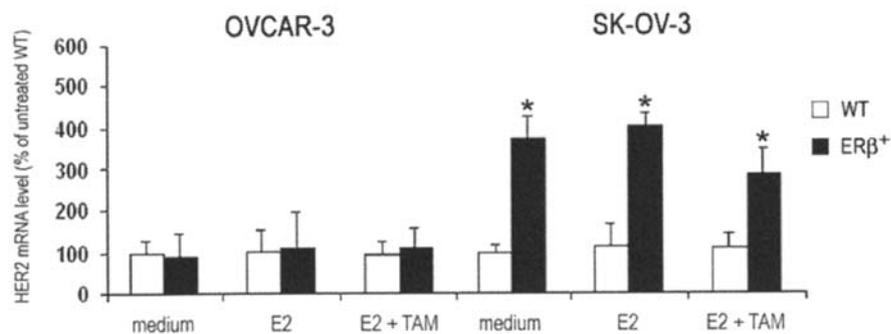


Figure 3. HER2 expression in wild-type and ER β -transfected breast cancer cell lines. The indicated cell lines and clones were cultured in steroid-hormone-free medium and treated with 3 nM 17- β estradiol (E2) alone or in combination with 100 nM 4-OH tamoxifen (TAM) for 48 h prior to RNA and protein preparation. (a) HER2 mRNA levels expressed in percent of the β -actin control. (b) HER2 Protein level in percent of the untreated wild-type control. (c) Representative Western blot (n=4); *p<0.01 vs. wild-type.

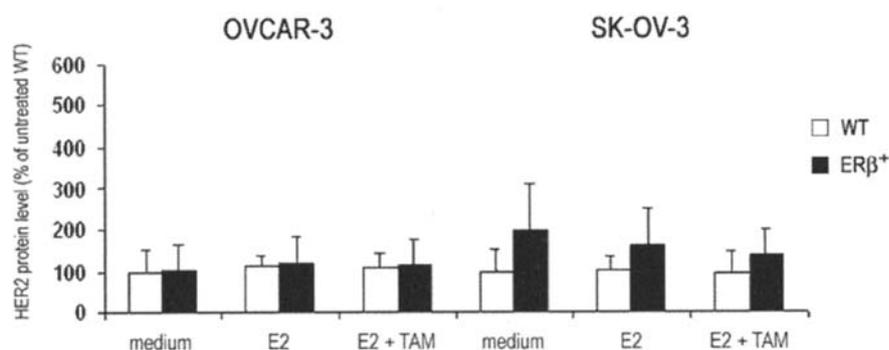
were made in studies analyzing the function of ER β in ovarian cancer cells (15). These observations taken together with the fact that the ER β expression is reported to decrease during tumorigenesis suggest that ER β has a tumor-suppressor

function in hormone-dependent tissues. In contrast, the HER-2 overexpression and/or amplification in breast cancer with positive hormone receptor status is correlated with an aggressive tumor phenotype (38). Thus, our findings, which

a)



b)



c)

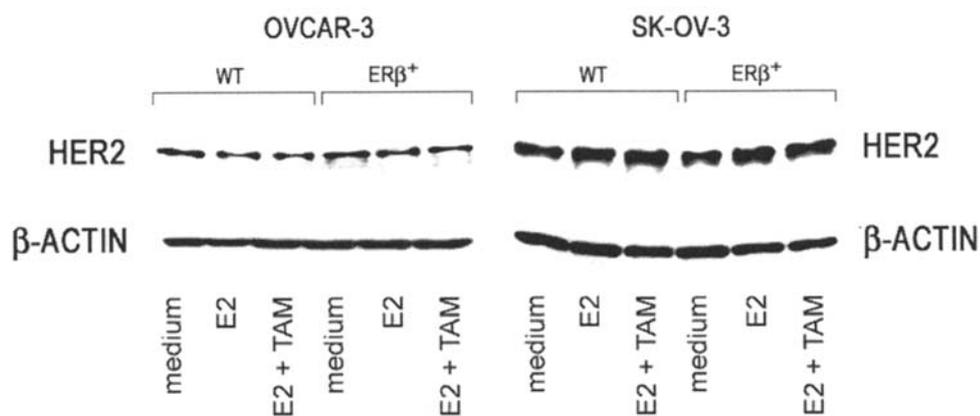


Figure 4. HER2 expression in wild-type and ERβ-transfected ovarian cancer cell lines. The indicated cell lines and clones were cultured in steroid-hormone-free medium and treated with 3 nM 17-β estradiol (E2) alone or in combination with 100 nM 4-OH tamoxifen (TAM) for 48 h prior to RNA and protein preparation. (a) HER2 mRNA levels expressed in percent of the β-actin control. (b) HER2 Protein level in percent of the untreated wild-type control. (c) Representative Western blot (n=4); *p<0.01 vs. wild-type.

demonstrate that an overexpression of ERβ leads to the elevation of HER2 expression in ERα-positive MCF-7 breast cancer cells, are noteworthy. However, results from our previous studies demonstrating a diminished proliferation of the MCF-7/ERβ⁺ clone when compared to wild-type MCF-7 cells clearly suggest that the up-regulation of HER2 is not sufficient to trigger increased cell growth.

We confirmed the results of previous studies by showing that treatment with E2 is able to diminish the HER2 expression in MCF-7 wild-type cells expressing ERα (27). Given that the overexpression of ERβ1 was able to increase the HER2 expression in this cell line, it is tempting to speculate that the ERα-triggered down-regulation of HER2 was counteracted by a high expression of ERβ1 in a ligand-independent

manner. This interpretation of our observations would be in line with previous reports demonstrating that ER β acts as an ER α antagonist in specific settings (39,40).

The fact that we did not observe any significant effect of ER β 1 overexpression on HER2 protein levels of SK-OV-3 and SK-BR-3 cells could simply be explained by the strong HER2 overexpression already present in the two cell lines which did not allow for the detection of possible ER β actions (41,42). The second reason for the observed unchanged HER2 expression in these cell lines was the known loss of functional ER α in SK-OV-3 and SK-BR-3 cells which does not allow for the observation of antagonistic ER β effects (39,40).

In this study, we reported the up-regulation of the HER2 receptor tyrosine kinase expression in estrogen-responsive MCF-7 breast cancer cells overexpressing ER β 1. This effect could be one of the reasons for the significantly correlated co-expression of the two receptors observed in recent clinical reports.

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