





Figure S2. Map of pLKO.1-TRC cloning vector. TRC, The RNAi Consortium.

Figure S3. Lentivirus containing IDO-modifying plasmids was successfully constructed and transfected into 293FT cells. The IDO (A) overexpression or (B) knockdown recombinant plasmids were transformed into active *Escherichia coli* and amplified. The positive colonies were identified by bacterial PCR. Lanes 1-4 were all PCR products from individual colonies of *Escherichia coli* transformed with IDO-overexpressing recombinant plasmid (1,224 bp). Lanes 5-8 were the PCR products from individual colonies of *Escherichia coli* transformed with IDO-knockdown recombinant plasmid (258 bp). IDO, indoleamine 2,3-dioxygenase; sh, short hairpin RNA.



Figure S4. Sequence alignment and transfection 293FT cells. (A) Sequences inserted into vector were confirmed by comparing with the theoretical sequence of *shIDO*. (B) In the presence of liposome transfection reagent, the confirmed EGFP-IDO overexpression construct and two packaging plasmids were co-transfected into 293FT cells to produce effective lentivirus particles. The transfection efficiency in 293FT cells was observed under a fluorescence microscope (magnification, x10; scale bar, 230.5 μ m). EGFP, enhanced green fluorescent protein; IDO, indoleamine 2,3-dioxygenase; shRNA/sh, short hairpin RNA.



293FT after lentivirus transfection

Figure S5. Chromatogram demonstration of Trp and Kyn in genetically modified DCs. Detection of Trp and Kyn concentrations in the culture medium samples of (A) IDO^{oe}DCs or (B) IDO^{kd}DCs with their controls and standard was performed by high-performance liquid chromatography. DC, dendritic cell; IDO, indoleamine 2,3-dioxygenase; Kyn, kynurenine; Trp, tryptophan; Vector^{Ctrl}DCs, DCs infected with control vector of pLJM1-EGFP; IDO^{oe}DCs, IDO-overexpressing DCs; vector^{ctrl}DCs, DCs infected with control vector of pLKO.1; IDO^{kd}DCs, IDO-knockdown DCs.



Figure S6. Effects of IDO on the survival and migration of DCs. (A) Flow cytometric analyses of viable cells stained with FITC-Annexin V and PI to detect the effect of IDO on the viability of DCs. The cells negative for both Annexin V and PI were considered alive. Representative scatter plots (left) and statistics (right) are shown. (B) After CFSE staining (37°C for 10 min), both 10x10⁶ vector^{ctrl}DCs and IDO^{kd}DCs were subcutaneously injected into mice (n=3 mice per group). After 24 h, the popliteal lymph nodes of mice were extracted to make a single cell suspension. The CFSE+DCs were detected by flow cytometry. Representative scatter plots (left) and statistics (right) are shown. The results are presented as the mean \pm SEM (n=3). DC, dendritic cell; CFSE, carboxyfluorescein diacetate succinimidyl ester; IDO, indoleamine 2,3-dioxygenase; ns, not significant; FSC-A, forward scatter area; vector^{ctrl}DCs, DCs infected with control vector of pLKO.1; IDO^{kd}DCs, IDO-knockdown DCs.

