

p38 mitogen-activated protein kinase/activator protein-1 involved in serum deprivation-induced human alkaline ceramidase 2 upregulation

ZHAOQUAN HUANG¹, GUOJIN HUANG², QUANZHONG LI³ and JUNFEI JIN^{2,4}

¹Department of Pathology, Guilin Medical University; ²Laboratory of Hepatobiliary and Pancreatic Surgery;

³Department of Cardiology, Affiliated Hospital of Guilin Medical University;

⁴Guangxi Key Laboratory of Molecular Medicine in Liver Injury and Repair, Guilin Medical University, Guilin, Guangxi 541001, P.R. China

Received October 15, 2014; Accepted November 14, 2014

DOI: 10.3892/br.2014.394

Abstract. Our previous study revealed that serum deprivation upregulated human alkaline ceramidase 2 (haCER2) activity and mRNA in HeLa cells, but the mechanism remains unknown. In the present study, serum deprivation also upregulated haCER2 activity in HepG2 human hepatoma cell line cells due to an increase in *haCER2* mRNA, in which mRNA transcription, not mRNA stability, is involved. Furthermore, p38 mitogen-activated protein kinase (MAPK)/activator protein-1 (AP-1) signaling pathway is involved in *haCER2* mRNA upregulation by serum deprivation, and this mechanism may explain why *haCER2* is upregulated in human liver cancer. In conclusion, p38 MAPK, AP-1 or haCER2 may be used as targets in liver cancer therapy.

Introduction

Ceramide, sphingosine and sphingosine-1-phosphate (S1P) are three major metabolites of the sphingolipid signaling pathway. In response to various stresses, ceramide has been shown to mediate differentiation, growth arrest and apoptosis (1-5); sphingosine acts similarly to ceramide (6), and S1P has the opposite effects (7). Therefore, the balance between the intracellular levels of ceramide/sphingosine and S1P may determine the fate of a cell (8). It is well known that ceramide

can be metabolized by ceramidases to generate sphingosine, which in turn can be phosphorylated by sphingosine kinases to form S1P. In addition to sphingosine kinases, ceramidases may play a pivotal role in the regulation of the intracellular ratio between ceramide, sphingosine and S1P (6). Based on optimal pHs and structures, ceramidase are classified into acid, neutral and alkaline ceramidase. Thus far, three alkaline ceramidases, such as alkaline ceramidase 1, 2 and 3, have been identified in humans (9). Human alkaline ceramidase 2 (haCER2), cloned by us (7), regulates sphingosine and S1P by controlling ceramides hydrolysis. Different expression levels of haCER2 have opposite effects; high expression of haCER2 caused sphingosine-induced growth arrest and low expression of haCER2 promoted S1P-mediated cell proliferation (7). Thus, regulation of haCER2 may affect the fate of a cell by controlling the balance of ceramide/sphingosine and S1P. In our previous study, serum deprivation upregulated *haCER2* mRNA and activity, but the mechanism remains unknown. In the present study, the HepG2 human hepatoma cell line was used as a cell model and it was found that p38/activator protein-1 (AP-1) signaling is involved in serum deprivation-induced *haCER2* transcriptional activation and subsequently caused an increase in haCER2 enzymatic activity.

Materials and methods

Cells and materials. HepG2 cells were cultured in Dulbecco's modified Eagle's medium (HyClone Corp., Logan, UT, USA) supplemented with 10% heat-inactivated fetal bovine serum (FBS; HyClone Corp.), 100 U/ml penicillin and 100 µg/ml streptomycin in a humidified atmosphere at 37°C and 5% CO₂.

o-Phthalaldehyde was from Fluka (Milwaukee, WI, USA). D-e-C24:1-ceramide and D-e-C17-sphingosine were purchased from Avanti Polar Lipids, Inc., (Pelham, AL, USA). Actinomycin D and 12-*O*-tetradecanoylphorbol-13-acetate (TPA; an activator of AP-1) were purchased from Sigma-Aldrich (St. Louis, MO, USA). PD-98059 [inhibitor of the extracellular signal-regulated kinase (ERK) pathway], SP-600125 [inhibitor of the c-Jun NH₂-terminal kinase (JNK) pathway], SB-203580 [inhibitor of p38 mitogen-activated

Correspondence to: Dr Zhaoquan Huang, Department of Pathology, Guilin Medical University, 109 Huan Cheng Bei Er Road, Guilin, Guangxi 541004, P.R. China
E-mail: gxlzzq@163.com

Dr Junfei Jin, Guangxi Key Laboratory of Molecular Medicine in Liver Injury and Repair, Guilin Medical University, 15 Lequn Road, Guilin, Guangxi 541001, P.R. China
E-mail: changliangzjin@163.com

Key words: serum deprivation, HepG2 cells, human alkaline ceramidase 2, gene regulation, activator protein-1, mitogen-activated protein kinases, p38

protein kinase (MAPK) pathway] and Bay117085 [inhibitor of nuclear factor- κ B (NF- κ B) pathway] were from Calbiochem/EMD Biosciences, Inc., (San Diego, CA, USA). AP-1 and NF- κ B Signal Reporter were purchased from Qiagen (Valencia, CA, USA). *Renilla* luciferase reporter plasmid pRL-TK and the Dual-Luciferase Reporter Assay system were from Promega Corp., (Madison, WI, USA). SR11302 (AP-1 specific inhibitor) were purchased from Tocris Bioscience (Ellisville, MO, USA). Human p38 small interfering RNA (siRNA) (sc-29433), p38 primers for quantitative polymerase chain reaction (qPCR) (sc-29433-PR) and p38 antibody (sc-7972) were purchased from Santa Cruz Biotechnology, Inc., (Santa Cruz, CA, USA). The RNeasy Mini kit was from Qiagen (Santa Clarita, CA, USA) and the iScript cDNA Synthesis kit was purchased from Bio-Rad Laboratories, Inc., (Hercules, CA, USA). Other unlisted chemicals were purchased from Sigma-Aldrich Co., LLC.

haCER2 activity assays. *haCER2* activity was determined by the release of sphingosine from ceramide as described previously (10). In brief, D-e-C24:1-ceramide was selected as a substrate dispersed into A buffer (pH 9.0), including 25 mmol/l glycine-NaOH, 5 mmol/l CaCl_2 and 0.3% Triton X-100, by water bath sonication. The microsomes containing *haCER2* from HepG2 cells were suspended in B buffer (pH 9.0), including 25 mmol/l glycine-NaOH and 5 mmol/l CaCl_2 . Incubation of the microsomes with ceramide substrate at 37°C for 20 min initiated the enzymatic reactions, which were stopped by boiling the mixture. After Bligh-Dye extraction, sphingosine was assayed by high-performance liquid chromatography analysis with D-e-C17-sphingosine as an internal standard. HPLC was conducted using the Agilent 1050-HPLC model fitted with an eclipse XDB-C18 column (Agilent Technologies, Palo Alto, CA, USA). The solvent was methanol-potassium phosphate buffer (90:10 v/v) and the flow rate was 0.7 ml/min. A HP1046 fluorescence detector with an excitation at 345 nm and emission at 455 nm was used.

Reverse transcription-qPCR (RT-qPCR). Total RNA was isolated from HepG2 cells using the RNeasy Mini kit according to the manufacturer's instructions. RNA was reverse-transcribed into first-strand cDNA with the iScript cDNA Synthesis kit using 20 μ l of the reaction mixture containing 1 μ l iScript reverse transcriptase, 4 μ l 5X iScript reaction mixture and 0.5 μ g total RNA. The complete reaction was cycled for 5 min at 25°C, 30 min at 42°C and 5 min at 85°C using a PTC-200 DNA Engine (MJ Research Inc., Waltham, MA, USA) (11). cDNA was subjected to RT-qPCR analysis, which was performed on an iCycler system (Bio-Rad Laboratories, Inc., Hercules, CA, USA). The initial PCR step was 3 min at 95°C, followed by 40 cycles of a 10-sec melting at 95°C and a 45-sec annealing/extension at 60°C. The final step was 1-min incubation at 60°C. All the reactions were performed in triplicate. Data are expressed as the mean normalized expression, which is directly proportional to the amount of *haCER2* mRNA relative to the amount of β -actin mRNA. The primers used were forward, 5'-AGTGTCTGTCTGCGTTACG-3'; and reverse, 5'-TGTTGTTGATGGCAGGCTTGAC-3' for *haCER2*; and forward, 5'-CAATGTTCTGGTGCAATTCAGAG-3' and reverse, 5'-GGATCCCATTCCTACCACTGTG-3' for β -actin (9).

mRNA stability analysis. HepG2 cells were plated in 12-well plates at a density of 0.4×10^6 cells/well overnight and treated with serum deprivation, followed by the addition of 10 μ g/ml actinomycin D. HepG2 cells were harvested 2 h after the addition of actinomycin D and *haCER2* mRNA was quantified using RT-qPCR as above.

Transfection and luciferase activity assay. HepG2 cells were transiently transfected with 1 μ g AP-1 or NF- κ B Signal Reporter for 24 h using FuGENE HD as the transfection reagent. The cells were cotransfected with the *Renilla* luciferase reporter plasmid pRL-TK (50 ng/10⁶ cells) as an internal control. The cells were subsequently treated with serum deprivation for 8 h. Following the treatment, the cells were rinsed with cold PBS and lysed with the buffer from the Dual-Luciferase Reporter Assay system. Firefly and *renilla* luciferase levels were measured in a luminometer using the dual-luciferase reporter assay reagents according to the manufacturer's instructions. The firefly luciferase levels were normalized to the *renilla* luciferase levels.

Treatment of cells with the inhibitors of the signaling pathways. HepG2 cells were treated with serum deprivation in the absence or presence of 10 μ M PD-98059, SP-600125, SB-203580 for 8 h. Following the treatment, *haCER2* mRNA was quantified using qPCR.

Transfection of siRNA. In order to silence the *p38 MAPK* gene expression, the siRNA approach was used. The HepG2 cells were cultured for 24 h before the transfection was processed in 24-well plates with 40 nM siRNAs using Lipofectamine[®]2000 (Invitrogen Corp., Carlsbad, CA, USA). After a 24-h incubation with *p38* siRNA or Con-siRNA, the HepG2 cells were cultured with or without serum deprivation for an additional 8 h and subsequently the cells were collected and *haCER2* mRNA was quantified using RT-qPCR. *p38 MAPK* knock-down was confirmed by western blot analysis (12) and qPCR.

Statistical analysis. Data are expressed as the mean \pm standard deviation, with a minimum of three independent experiments analyzed by Student's t-test or analysis of variance. $P < 0.05$ was considered to indicate a statistically significant difference.

Results

haCER2 is upregulated by serum deprivation in HepG2 cells. Our previous study revealed that *haCER2* is upregulated by serum deprivation in HeLa cells; this prompted the investigation of whether the expression of the endogenous *haCER2* was upregulated upon serum deprivation in HepG2 cells. *In vitro* activity assays showed that serum deprivation markedly increased alkaline ceramidase activity on D-e-C24:1-ceramide in a time-dependent manner, reaching a peak at 12 h (Fig. 1A). RT-qPCR analysis demonstrated that *haCER2* mRNA was significantly upregulated in response to serum deprivation in a time-dependent manner and the increase plateaued at 8 h (Fig. 1B). These results indicate that serum deprivation upregulates *haCER2* activity and mRNA. Comparing the kinetics of *haCER2* activity and mRNA, it is indicated that the serum deprivation induced-*haCER2* activity increase in HepG2 happens at mRNA level. Based on this kinetics study,

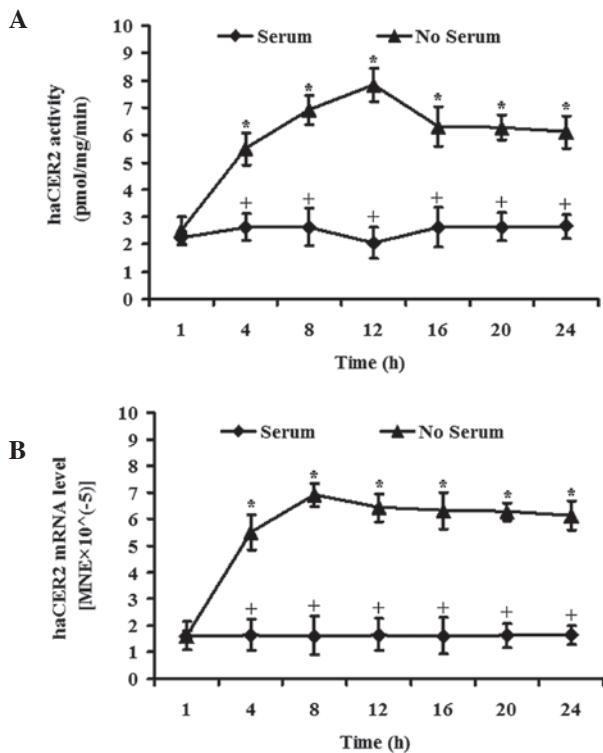


Figure 1. Human alkaline ceramidase 2 (haCER2) expression is upregulated by serum deprivation. HepG2 cells were grown to 80% confluence in serum-containing Dulbecco's modified Eagle's medium (DMEM) prior to growth in the same medium or in serum-free DMEM. The cells were harvested at different time points following the medium change and subjected to (A) *in vitro* activity assays for haCER2 or total RNA was extracted from another portion of the above cells and subjected to (B) reverse transcription-quantitative PCR analysis. Data represent the mean value \pm standard deviation of three independent experiments performed in duplicate. At the same time point, * $P < 0.05$. MNE, mean normalized expression.

haCER2 mRNA was measured at 8 h and haCER2 activity at 12 h in the subsequent studies.

Transcription, but not mRNA stability, is involved in haCER2 upregulation in HepG2 cells by serum deprivation. The data above showed that haCER2 mRNA was upregulated by serum deprivation and it is well known that gene expression can be regulated at the transcriptional level or the post-transcriptional level. In the last years, it became evident that mRNA stability/turnover provides an important mechanism for post-transcriptional control of gene expression. Therefore, whether mRNA stability was involved in the regulation of haCER2 expression by serum deprivation was determined. In the present study, the inhibitory effect of actinomycin D, an inhibitor of transcription, on haCER2 transcription was confirmed by the finding that the treatment of cells with actinomycin D for 2 h reduced the haCER2 mRNA level by 92% in medium containing 10% FBS (Fig. 2). Furthermore, a result showed that in the presence of actinomycin D for 2 h, haCER2 mRNA level was observed to reduce by 88% in HepG2 cells treated with serum starvation (Fig. 2). Comparing the mRNA level in HepG2 cells exposure to actinomycin D for 2 h between serum starvation and 10% FBS treatment, it was indicated that serum deprivation-stimulated haCER2 upregulation was due to the transcriptional regulation and not mRNA stability.

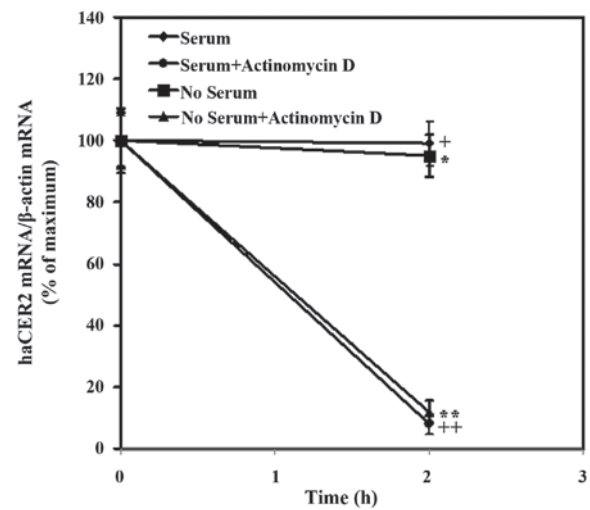


Figure 2. mRNA stability is not involved in serum deprivation-stimulated human alkaline ceramidase 2 (haCER2) upregulation. HepG2 cells were treated with serum deprivation for 6 h, followed by the addition of 10 μ g/ml actinomycin D. The cells were harvested 2 h after the addition of actinomycin D and haCER2 mRNA was quantified using reverse transcription-quantitative PCR. The data presented (means \pm standard deviation) were from 1 of 2 experiments with similar results. +, +, +, * $P < 0.05$.

AP-1 signaling is involved in haCER2 upregulation in HepG2 cells by serum deprivation. A previous study revealed that transcriptional factor AP-1 signaling regulated human neutral ceramidase gene transcription and the degree of decrease in ceramide mass was significantly greater compared to the reduction of human neutral ceramidase transcription, raising the possibility that AP-1 may regulate other ceramide metabolizing enzymes, including haCER2 (13). Therefore, in the present study, whether AP-1 regulated serum deprivation-induced haCER2 upregulation or not was investigated. In the first experiment, HepG2 cells were transfected with the luciferase reporter vectors constructed with either AP-1 or NF- κ B-binding element in the promoter. The results showed that serum deprivation stimulated AP-1 but not NF- κ B activity at 12 h (Fig. 3A). Consistent with the above results, haCER2 mRNA could be upregulated by serum deprivation in HepG2 cells transfected with AP-1 or NF- κ B (Fig. 3B). In addition, the AP-1 specific inhibitor SR11302 (14,15) completely blocked the serum deprivation-induced haCER2 mRNA upregulation (Fig. 3C), and TPA, as an activator of AP-1 (16), has an additive effect on serum deprivation-induced haCER2 mRNA upregulation (Fig. 3D). Notably, in 10% FBS medium, TPA also stimulates haCER2 mRNA expression (Fig. 3D).

p38 MAPK is involved in haCER2 upregulation in HepG2 cells by serum deprivation. AP-1 transcriptional activity can be regulated by MAPK, including ERK, JNK and p38 MAPK signal transduction pathways (17). Thus, which one was involved in haCER2 upregulation by serum deprivation in HepG2 cells was determined. In the study, HepG2 cells were treated with serum deprivation in the presence of the pharmacological inhibitors of the MAPK pathways. The results showed that haCER2 upregulation by serum deprivation were inhibited by SB-203580, an inhibitor for the p38 MAPK pathway, but not by SP-600125 (an inhibitor for JNK) and PD-98059 (an inhibitor for ERK)

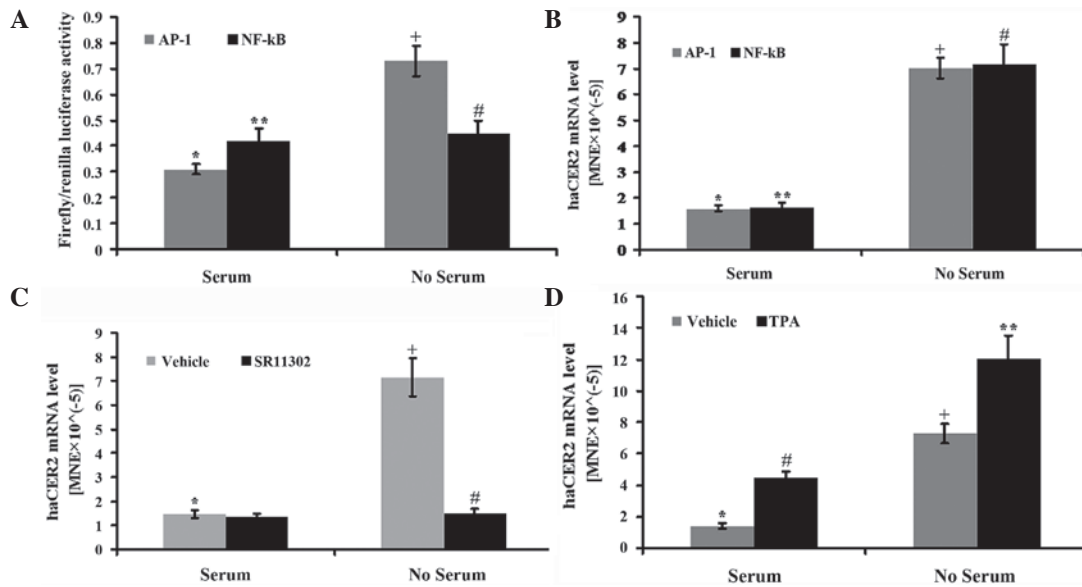


Figure 3. Activator protein-1 (AP-1) signaling is involved in human alkaline ceramidase 2 (*haCER2*) upregulation in HepG2 cells by serum deprivation. HepG2 cells were transfected with the DNA vectors constructed with AP-1 or the nuclear factor-κB (NF-κB)-binding element in the presence of FuGENE HD as a transfection reagent for 24 h. After the transfection, the cells were treated with serum deprivation for 8 h (for *haCER2* mRNA) or 12 h (for luciferase activity) and collected for further analysis. (A) The AP-1 or NF-κB activity was presented as the ratio of firefly luciferase vs. *renilla* luciferase activity (*[†]P<0.05; **[#]P>0.05) and (B) *haCER2* mRNA was quantified using reverse transcription-quantitative PCR (RT-qPCR), (*[†]P<0.05; **[#]P<0.05). HepG2 cells were treated with (C) 1 μM SR11302 (*[†]P<0.05; [†]#P<0.05) or (D) 10 ng/ml 12-*O*-tetradecanoylphorbol-13-acetate (TPA) (*[†]P<0.05; [†]#P<0.05; **[#]P<0.05; [†]#P<0.05) in serum deprived medium or 10% fetal bovine serum medium for 8 h and *haCER2* mRNA was measured by RT-qPCR. MNE, mean normalized expression.

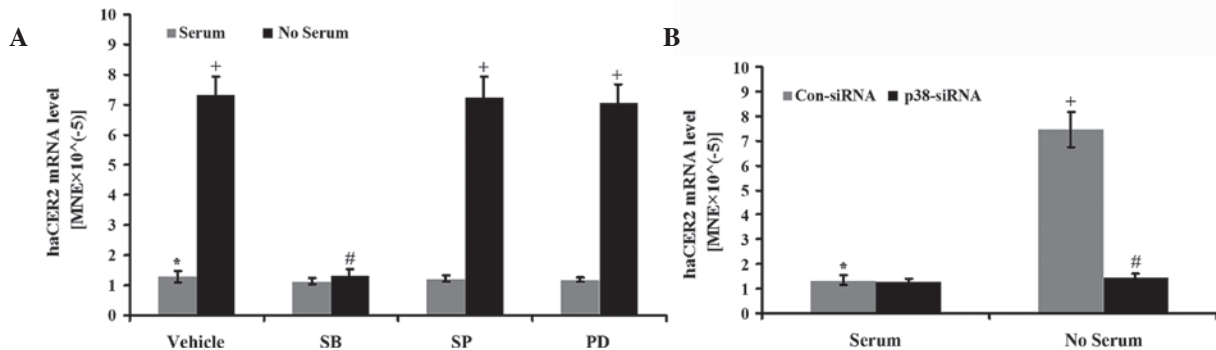


Figure 4. p38 mitogen-activated protein kinase (MAPK) may regulate human alkaline ceramidase 2 (*haCER2*) upregulation in HepG2 cells by serum deprivation. (A) HepG2 cells were treated with or without serum deprivation in the absence or presence of 10 μM SB-203580 (SB), an inhibitor for the p38 MAPK pathway; 10 μM SP-600125 (SP), an inhibitor for the JNK pathway; or 10 μM PD-98059 (PD), an inhibitor for the ERK pathway for 8 h. Subsequently the cells were collected and *haCER2* mRNA was measured by reverse transcription-quantitative PCR (RT-qPCR). (B) HepG2 cells were transfected with 40 nM p38 or scrambled small interfering RNA (Con-siRNA) for 24 h. The transfected cells were subsequently treated with serum deprivation for 8 h and collected for *haCER2* mRNA measurement by RT-qPCR. (*[†]P<0.05; [†]#P<0.05. MNE, mean normalized expression; JNK, c-Jun NH2-terminal kinase; ERK, extracellular signal-regulated kinase.

(Fig. 4A). Consistent with this result, downregulation of *p38* by specific siRNA completely inhibited *haCER2* upregulation by serum deprivation (Fig. 4B). *p38* MAPK knockdown was confirmed by western blot analysis and qPCR (data not shown). These data indicate that the *p38* MAPK pathway is involved in *haCER2* upregulation by serum deprivation.

Discussion

haCER2 plays an important role in cellular responses by regulating the hydrolysis of ceramides in cells (18). High and low ectopic expression of *haCER2* leads to different cell fates through changing the ratio of ceramide/sphingosine and SIP (7). Therefore, the regulation of *haCER2* expression by physiological or pathological stimuli is extremely important, but

little is known regarding it. In the present study, firstly, serum deprivation was found to cause *haCER2* activation in HepG2 cells in a time-dependent manner, which is in agreement with our previous finding conducted in HeLa cells (7). Comparing the kinetics of *haCER2* activity and mRNA expression, it is indicated that *haCER2* activity elevation was caused by increasing *haCER2* mRNA. The level of an mRNA within a cell depends on its rate of synthesis and rate of decay (19). mRNA stability assay showed that actinomycin D, an inhibitor of transcription, completely inhibited the *haCER2* mRNA increase caused by serum deprivation. Taking the above results together, it was indicated that serum deprivation-stimulated *haCER2* mRNA elevation was due to the transcriptional regulation.

In response to a plethora of physiological and pathological stimuli, numerous gene regulation is mediated by AP-1 (20). A

previous study suggested that human neutral ceramidase gene transcription is regulated by AP-1 signaling, it also raised the possibility that other ceramidases may be regulated by AP-1 as the degree of decrease in ceramide mass was significantly greater than the reduction of human neutral ceramidase transcription (13). Therefore, whether *haCER2* gene transcription by serum deprivation is regulated by AP-1 signaling pathway was investigated. The results showed that serum deprivation stimulated AP-1 luciferase activity, whereas NF- κ B activity was not stimulated by serum deprivation. Notably, SR11302 (a specific inhibitor of AP-1) blocked serum deprivation-induced *haCER2* mRNA increase, whereas TPA (an activator of AP-1) additively increased *haCER2* mRNA with serum deprivation. All these data confirmed that transcription factor AP-1 is involved in *haCER2* gene transcription by serum deprivation.

Three different types of MAPKs, including ERK, JNK and p38, contribute to the induction of AP-1 activity in response to a diverse array of extracellular stimuli (21). In the present study, the pharmacological inhibitors experiment revealed that SB-203580 blocked *haCER2* transcription induced by serum deprivation, whereas SP-600125 and PD-98059 could not. These indicated that the p38 MAPK, and not JNK and ERK, pathway is involved in serum deprivation-induced *haCER2* transcription. The involvement of the p38 MAPK pathway in *haCER2* transcription mediated by serum starvation was further confirmed by p38-specific siRNA downregulation.

Notably, AP-1 is composed of the Jun protein family (c-Jun, Jun B and Jun-D) and the Fos protein family (such as c-Fos, FosB, Fra-1 and Fra-2) (20), and the present study does not identify which subunit(s) of AP-1 play a pivotal role in *haCER2* transcription mediated by serum starvation and this is an area for future investigation.

A low-nutrient environment is commonly found in the central region of solid tumor, such as liver cancer (22,23). Serum starvation mimics the tumor growth environment *in vivo* (24). In HepG2 cells, serum deprivation upregulated *haCER2* expression in which the p38 MAPK/AP-1 signaling pathway is involved, and this mechanism may explain why *haCER2* is upregulated in solid liver cancer (7,25).

The regulation of mRNA stability is an important factor in modulating gene expression, and the p38 MAPK pathway has been indicated in the regulation of the mRNA half-lives of a number of genes (26). Based on the aforementioned studies, we hypothesized that mRNA stability may be involved in serum starvation-induced *haCER2* mRNA increase, but mRNA decay experiments exclude this possibility and future research is required to explore this reason.

In conclusion, the present study conducted in the human HepG2 hepatoma cell line indicates that serum deprivation affects the p38 MAPK signaling pathway leading transcription factor AP-1 activation and subsequently regulates *haCER2* mRNA expression. This mechanism may interpret why *haCER2* is upregulated in liver cancer.

Acknowledgements

The present study was supported in part by the National Natural Science Foundation of China (grant no. 81360309), 'Sphingolipids and Related Diseases' Program for Innovative Research Team of Guilin Medical University, and Hundred

Talents Program of Universities and Colleges Directly under the Guangxi Zhuang Autonomous Region.

References

1. Spiegel S and Merrill AH Jr: Sphingolipid metabolism and cell growth regulation. *FASEB J* 10: 1388-1397, 1996.
2. Merrill AH Jr, Schmelz EM, Wang E, *et al*: Importance of sphingolipids and inhibitors of sphingolipid metabolism as components of animal diets. *J Nutr* 127 (Suppl 5): S830-S833, 1997.
3. Hannun YA and Obeid LM: The Ceramide-centric universe of lipid-mediated cell regulation: stress encounters of the lipid kind. *J Biol Chem* 277: 25847-25850, 2002.
4. Levade T, Malagarie-Cazenave S, Gouaze V, *et al*: Ceramide in apoptosis: a revisited role. *Neurochem Res* 27: 601-607, 2002.
5. Kolesnick R and Fuks Z: Radiation and ceramide-induced apoptosis. *Oncogene* 22: 5897-5906, 2003.
6. Cuvillier O: Sphingosine in apoptosis signaling. *Biochim Biophys Acta* 1585: 153-162, 2002.
7. Xu R, Jin J, Hu W, *et al*: Golgi alkaline ceramidase regulates cell proliferation and survival by controlling levels of sphingosine and S1P. *FASEB J* 20: 1813-1825, 2006.
8. Cuvillier O, Pirianov G, Kleuser B, *et al*: Suppression of ceramide-mediated programmed cell death by sphingosine-1-phosphate. *Nature* 381: 800-803, 1996.
9. Xu R, Sun W, Jin J, Obeid LM and Mao C: RoleA of alkaline ceramidases in the generation of sphingosine and its phosphate in erythrocytes. *FASEB J* 24: 2507-2515, 2010.
10. Sun W, Hu W, Xu R, *et al*: Alkaline ceramidase 2 regulates beta1 integrin maturation and cell adhesion. *FASEB J* 23: 656-666, 2009.
11. Jin J, Zhang X, Lu Z, *et al*: Acid sphingomyelinase plays a key role in palmitic acid-amplified inflammatory signaling triggered by lipopolysaccharide at low concentrations in macrophages. *Am J Physiol Endocrinol Metab* 305: E853-E867, 2013.
12. Zhu Q, Lin L, Cheng Q, *et al*: The role of acid sphingomyelinase and caspase 5 in hypoxia-induced HuR cleavage and subsequent apoptosis in hepatocytes. *Biochim Biophys Acta* 1821: 1453-1461, 2012.
13. O'Neill SM, Houck KL, Yun JK, Fox TE and Kester M: AP-1 binding transcriptionally regulates human neutral ceramidase. *Arch Biochem Biophys* 511: 31-39, 2011.
14. Huang C, Ma WY, Dawson MI, Rincon M, Flavell RA and Dong Z: Blocking activator protein-1 activity, but not activating retinoic acid response element, is required for the antitumor promotion effect of retinoic acid. *Proc Natl Acad Sci USA* 94: 5826-5830, 1997.
15. Li JJ, Westergaard C, Ghosh P and Colburn NH: Inhibitors of both nuclear factor-kappaB and activator protein-1 activation block the neoplastic transformation response. *Cancer Res* 57: 3569-3576, 1997.
16. Fisch TM, Prywes R and Roeder RG: An AP1-binding site in the c-fos gene can mediate induction by epidermal growth factor and 12-O-tetradecanoyl phorbol-13-acetate. *Mol Cell Biol* 9: 1327-1331, 1989.
17. Whitmarsh AJ and Davis RJ: Transcription factor AP-1 regulation by mitogen-activated protein kinase signal transduction pathways. *J Mol Med (Berl)* 74: 589-607, 1996.
18. Sun W, Jin J, Xu R, *et al*: Substrate specificity, membrane topology, and activity regulation of human alkaline ceramidase 2 (ACER2). *J Biol Chem* 285: 8995-9007, 2010.
19. Bellofatto V and Wilusz J: Transcription and mRNA stability: parental guidance suggested. *Cell* 147: 1438-1439, 2011.
20. Hess J, Angel P and Schorpp-Kistner M: AP-1 subunits: quarrel and harmony among siblings. *J Cell Sci* 117: 5965-5973, 2004.
21. Karin M: The regulation of AP-1 activity by mitogen-activated protein kinases. *J Biol Chem* 270: 16483-16486, 1995.
22. Harrington EA, Fanidi A and Evan GI: Oncogenes and cell death. *Curr Opin Genet Dev* 4: 120-129, 1994.
23. Dang CV and Semenza GL: Oncogenic alterations of metabolism. *Trends Biochem Sci* 24: 68-72, 1999.
24. Xiong Y, Fang JH, Yun JP, *et al*: Effects of microRNA-29 on apoptosis, tumorigenicity, and prognosis of hepatocellular carcinoma. *Hepatology* 51: 836-845, 2010.
25. Graveel CR, Jatke T, Madore SJ, Holt AL and Farnham PJ: Expression profiling and identification of novel genes in hepatocellular carcinomas. *Oncogene* 20: 2704-2712, 2001.
26. Frevel MA, Bakheet T, Silva AM, Hissong JG, Khabar KS and Williams BR: p38 Mitogen-activated protein kinase-dependent and -independent signaling of mRNA stability of AU-rich element-containing transcripts. *Mol Cell Biol* 23: 425-436, 2003.