

Association of polymorphisms and haplotypes in the cytochrome P450 1B1 gene with uterine leiomyoma: A case control study

SAEEDAH SALIMI^{1,2}, MARYAM KHODAMIAN², MEHRNAZ NAROOIE-NEJAD^{3,4}, AZAM HAJIZADEH², KIMIA FAZELI⁵, LIDA NAMAZI⁶ and MINOO YAGHMAEI⁶

¹Cellular and Molecular Research Center, ²Department of Clinical Biochemistry, School of Medicine, ³Department of Genetics, ⁴Genetics of Non-Communicable Disease Research Center, Zahedan University of Medical Sciences, Zahedan; ⁵Faculty of Medicine, Mashhad University of Medical Sciences, Mashhad 9137913316; ⁶Department of Obstetrics and Gynecology, School of Medicine, Zahedan University of Medical Sciences, Zahedan 9816743175, Iran

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Abstract. Uterine leiomyoma (UL) is an estrogen-dependent neoplasm of the uterus and estrogen metabolizing enzymes affect its promotion and progression. The aim of the present study was to evaluate the association between four single-nucleotide polymorphisms (SNPs) of the cytochrome P450 1B1 (*CYP1B1*) gene and UL risk. Four SNPs of the *CYP1B1* gene in 105 UL patients and 112 unrelated healthy controls were genotyped using a direct sequencing method. Haplotype analyses were performed with UNPHASED software and linkage disequilibrium (LD) was assessed by Haploview software. There were no associations between Leu432Val (rs1056836), Asp449Asp (rs1056837) and Asn453Ser (rs1800440) polymorphisms of the *CYP1B1* gene and UL. Although the genotypic frequencies of the Arg368His (rs79204362) polymorphism did not differ between the two groups, the frequency of A (His) allele was significantly higher in UL females ($P=0.02$). In addition, the frequency of GTAA haplotype was significantly higher in the controls and played a protective role in UL susceptibility. A strong LD between the three common SNPs (rs1056836, rs1056837 and rs1800440) in the *CYP1B1* gene was observed in the population. In conclusion, a higher frequency of the *CYP1B1* 368His (A) allele was observed in UL females. The frequency of the GTAA haplotype was significantly higher in healthy females and this haplotype played a protective role in UL susceptibility.

Introduction

Uterine leiomyoma (UL) is a benign monoclonal tumor of the smooth muscle cells, which arises more specifically from the underlying myometrial tissue in the uterus (1). These tumors contain high amounts of extracellular matrix components, such as collagen, fibronectin and proteoglycans (2). UL has been diagnosed in 30-40% of females at a reproductive age and is the leading indication for hysterectomy (1). Although this complication is often asymptomatic, it may result in symptoms including abdominal discomfort to uterine bleeding, menometrorrhagia, severe dysmenorrhea, pelvic pain and infertility (3). Despite the high prevalence of UL, the associated pathophysiology and proliferative pathway remains unknown. Different risk factors have been suggested for UL, including age, endogenous hormone levels (estrogen and progesterone), certain enzymes, obesity, nulliparity and genetic factors (2,3). The genetic liability of UL is supported by the fact that hysterectomy and hospitalization due to UL is twice as high in monozygote female twins compared to dizygotes (4,5).

UL is an estrogen-dependent tumor and is more prevalent during the reproductive age. The estrogen hormone stimulates the progression of cell proliferation in the G₁ phase of the cell cycle in various target tissues, including the uterus and breast (2,6). Evidence has shown that leiomyoma tissue is more sensitive to estrogen compared to myometrium and the concentration of estrogen in leiomyoma tissue is higher compared to the normal myometrium. Additionally, the leiomyoma growth is regulated by serum estrogen and estrogen in the tumor (7).

Estrogen exerts its biological effect through the high affinity binding to estrogen receptors (ESR α and ESR β) and muscle cells with leiomyoma express α and β ESRs (8,9).

The P450 cytochrome system (CYP450) contains a group of enzymes that are involved in steroid hormone biosynthesis and metabolic activation of carcinogens (10,11). Five different enzymes have been identified in this system, which are involved in estrogen synthesis and degradation. Cytochrome

Correspondence to: Miss. Maryam Khodamian, Department of Clinical Biochemistry, School of Medicine, Zahedan University of Medical Sciences, Khalij-e Fars High Way, Zahedan 9816743175, Iran
E-mail: maryam.khodamian@yahoo.com

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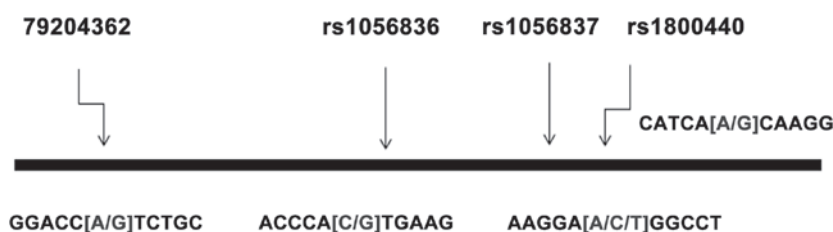


Figure 1. Schematic diagram of the distribution of four single-nucleotide polymorphism (SNP) loci in the cytochrome P450 1B1 (*CYP1B1*) gene.

P450 17a (CYP17) and cytochrome P450 19 (CYP19 or aromatase) control the early and terminal steps of the estrogen biosynthesis pathway respectively (12).

Cytochrome P450 1A1 (CYP1A1) and cytochrome P450 1B1 (CYP1B1) are the main enzymes of the CYP450 system in estrogen catabolism that catalyze the formation of 2-hydroxy and 4-hydroxy estrogen metabolites. Catechol-O-methyl transferase catalyzes the excretion of carcinogenic 4-hydroxy estrogen metabolites (12). Certain evidence has shown that human endometrial epithelium produce cytochrome P450 (CYP P450) metabolic enzymes that generate genotoxic intermediates and affect ESRs. Therefore, these enzymes are associated with carcinogenesis in extrahepatic tissues (13,14). The study by Shastry (15) found high 4-hydroxylase activities in human uterine myometrium and benign UL. This evidence strongly proposes that estrogens and their metabolic products affect the occurrence and development of UL. Therefore, *CYP1A1* and *CYP1B1* variants are expected to affect estrogen synthesis or degradation (16).

The association between polymorphisms of estrogen-metabolizing enzyme genes and UL susceptibility have been studied in different populations, however, their results were inconsistent (14,17). Therefore, in the present study the association between Arg368His (rs79204362), Leu432Val (rs1056836), Asp449Asp (rs1056837) and Asn453Ser (rs1800440) polymorphisms of the *CYP1B1* gene and UL susceptibility was examined in southeast Iran.

Materials and methods

Subject characteristics. The present case control study was performed on 105 females in their pre-menopausal stage that had undergone myomectomy or hysterectomy in Ali Ibn Abitaleb Hospital, Zahedan, southeast Iran, and were enrolled between 2011 and 2013. All the participants were selected using the convenient sampling method. In all the participants, leiomyoma had been confirmed pathologically.

In total, 112 healthy females in their premenopausal stage were selected as the control group among females referring for a routine annual check-up and performing the Pap smear test. The groups were matched with respect to demographic factors, such as age and ethnicity (Fars or Balouch). Physical and ultrasonographic evaluations were performed for females in the control group to eliminate UL. The Pap smear test was also negative.

The exclusion criteria were existence of systemic diseases and history of malignancy. All the participants were Iranian and provided their informed consent prior to participating

in the study. The protocol of the study was approved by the Ethics Committee of Zahedan University of Medical Sciences.

Genotype analysis. Genomic DNA was extracted from 2 ml peripheral blood leukocytes using the salting out phenol chloroform method and stored at -20°C until analysis. The analysis of the *CYP1B1* polymorphisms was performed using the sequencing method.

The reference sequence was the human *CYP1B1* gene (GeneBank accession number U56438). The fragment containing all the polymorphisms was amplified using the forward and reverse primers of 5'-TCACTTGCTTTTCTCTCTCC-3' and 5'-AATTTTCAGCTTGCTCTTG-3', respectively (18).

The total volume of the PCR mixture was 25 μl and contained 300 ng genomic DNA, 25 pM of each primers, 0.1 mM dNTP, 1.5 mM MgCl_2 , 2.5 μl PCR buffer 10X and 1 unit of Taq polymerase.

Amplification was performed in a Bio-Rad thermal cycler (Bio-Rad, Hercules, CA, USA) using a thermal profile of initial denaturation at 96°C for 6 min, followed by 30 cycles at 96°C for 30 sec, annealing at 61°C for 40-base pair insertion/deletion polymorphism and 60°C for T309G polymorphism for 30 sec, primer extension at 72°C for 60 sec, and a final extension step at 72°C for 6 min.

The PCR products were sequenced in a forward direction with the same primers as used in the PCR, using ABI Big Dye terminator chemistry and an ABI PRISM 3730/3730xl instrument (Applied Biosystems, Foster City, CA, USA).

A schematic diagram of Arg368His (rs79204362), Leu432Val (rs1056836), Asp449Asp (rs1056837) and Asn453Ser (rs1800440) polymorphisms of the *CYP1B1* gene is presented in Fig. 1.

Statistical analysis. Statistical analyses were performed using SPSS software, version 20 (IBM Corp, Armonk, NY, USA). The frequency of the genotypes and alleles were compared between two groups using the χ^2 or Fisher's exact tests. Quantitative variables were compared using the Student's t-test. Odds ratios (OR) and 95% confidence intervals (CI) were accounted to the significant allelic and genotypic associations. $P < 0.05$ was considered to indicate a statistically significant difference.

The frequency of haplotypes was calculated using PHASE version 2.1 software (19,20). Haplotypes with estimated frequency $< 5\%$ were excluded from the analysis. Logistic regression analysis was used to assess the independent effect of each risk polymorphism and haplotypes on UL. Bonferroni's correction was applied to confirm the association of haplotypes with the disease.

The computation of linkage disequilibrium (LD) between single-nucleotide polymorphisms (SNPs) was estimated using the normalized measure of allelic association D' and the characterization of these patterns was determined using Haploview 4.2 software (<http://www.borad.mit.edu/mpg/haploview>).

Results

Subject characteristics. The demographic and clinical characteristics of the UL patients and controls are shown in Table I. No significant differences in the age, body mass index and marital status between the two groups were observed. There were no significant differences in menstrual histories, including duration of menstruations, age of menarche and menstrual cycle between the two groups. Additionally, the frequency of bleeding and pain were significantly higher in the UL females ($P < 0.0001$).

Genotyping of CYP1B1 SNPs. Four SNPs of the *CYP1B1* gene were successfully genotyped in 105 UL females and 112 healthy controls. The frequency of alleles and genotypes of *CYP1B1* polymorphisms are shown in Table II. All the loci conformed to the Hardy-Weinberg equilibrium in UL females and control groups ($P > 0.05$). The frequency of the *CYP1B1* 432Val (G) allele in females with UL and controls were 38 and 30%, respectively, and the difference was not statistically significant. The genotype frequency of the *CYP1B1* Leu432Val polymorphism was not significantly different between the two groups. Although the frequency of the *CYP1B1* 449Asp (T) allele was higher in the UL group compared to controls (38 vs. 31%), the difference was not significant. The genotype frequency of the *CYP1B1* Asp449Asp (T/C) polymorphism was not significantly different between the two groups.

Genotype frequencies of polymorphisms. Although there was no significant difference in the genotypes frequency of the Arg368His (G/A) polymorphism between UL females and controls, a higher frequency of the A allele was observed in UL females compared to healthy females (13 vs. 6%) and the risk of UL was increased to 2.2-fold in the presence of the A allele (OR, 2.2; 95% CI, 1.1-4.4; $P = 0.02$). There was no association between *CYP1B1*, Asn453Ser (A/G) polymorphism and UL. The *CYP1B1* Arg368His (rs79204362) polymorphism has not been found in HapMap. Therefore, the LD patterns of the three *CYP1B1* SNPs were analyzed, as shown in Fig. 2. There was a strong LD between the three common SNPs in *CYP1B1* gene.

The frequency of five common haplotypes of four SNPs in the *CYP1B1* gene [Leu432Val (C/G), Asp449Asp (T/C), Asn453Ser (A/G) and Arg368His (G/A)] are shown in Table III.

Although the frequency of the CCGG haplotype was higher in UL females compared to the controls (24 vs. 18%), the difference was not statistically significant ($P = 0.3$). The frequency of the GTAA haplotype in controls was significantly higher than that of the cases (12.7 vs. 6%, $P = 0.039$) and the haplotype-based association analysis revealed that the GTAA haplotype may have potential to protect against UL in females (OR, 0.8; 95% CI, 0.7-1; $P = 0.039$). This association was also statistically significant following Bonferroni's correction.

Table I. Demographic and clinical characteristics of the UL females and control group.

	UL females (n=105)	Controls (n=112)	P-value
Age, years	38.1±9.8	36.7±5.7	0.23
Marriage status, n (%)	100 (95)	109 (97)	0.49
BMI, kg/m ²	25.1±5.5	25.4±4.7	0.70
Age of menarche, years	13.5±1.5	13.1±1.6	0.07
Duration of menses, days	6.1±1.6	5.8±1.5	0.07
Menstrual cycle, days	28.3±3.3	28.7±2.1	0.30
Bleeding, n (%)	60 (57)	4 (4)	<0.0001
Pain, n (%)	30 (29)	6 (5)	<0.0001

Data are mean ± standard deviation. UL, uterine leiomyoma; BMI, body mass index.

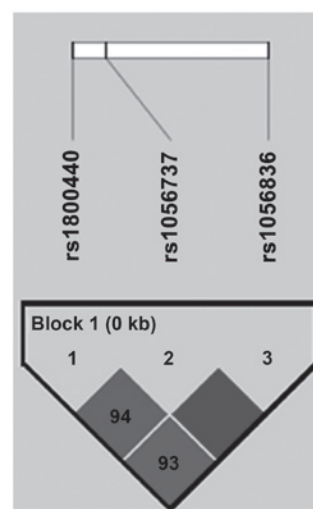


Figure 2. Linkage disequilibrium pattern of three single-nucleotide polymorphisms (SNPs) in the cytochrome P450 1B1 (*CYP1B1*) gene.

Discussion

In the present study, there was no association between the Leu432Val, Asp449Asp and Asn453Ser polymorphisms of the *CYP1B1* gene and SLE. However, a higher frequency of the A allele of Arg368His (G/A) polymorphism was observed in UL females compared to healthy females and according to the current findings, the presence of the A allele increased the SLE risk. In addition, the frequency of the GTAA haplotype in the controls was significantly higher compared to the patients; therefore, the GTAA haplotype played a protective role in SLE susceptibility. A strong LD between the three common SNPs in the *CYP1B1* gene was observed in the population of the present study.

UL is a common neoplasm associated with an interaction between various genes, cytokines, growth factors, hormones and environmental factors. Whether ovarian steroid hormones play an essential role in the growth of leiomyoma is unknown. The mitogenic effects of steroid hormones are induced through their receptors or by regulation of growth factor expression (7,8).

Table II. Allelic and genotypic frequency of the *CYP1B1* gene polymorphisms in UL females and healthy controls.

<i>CYP1B1</i>	UL females (n=105)	Controls (n=112)	P-value	Non-adjusted OR (95% CI)	P-value	Adjusted OR (95% CI) ^a
Leu432Val (C/G) rs1056836						
CC, n (%)	42 (40)	56 (50)		1		1
CG, n (%)	47 (45)	45 (40)	0.30	1.4 (0.8-2.5)	0.30	1.4 (0.8-2.4)
GG, n (%)	16 (15)	11 (10)	0.13	1.3 (0.8-2.0)	0.20	1.4 (0.9-2.1)
CG+GG, n (%)	63 (60)	56 (50)	0.14	1.5 (0.9-2.6)	0.20	1.5 (0.8-2.5)
C, n (%)	131 (62)	157 (70)		1		-
G, n (%)	79 (38)	67 (30)	0.09	1.4 (1.0-2.1)		-
Asp449Asp (T/C) rs1056837						
CC, n (%)	42 (40)	53 (47)		1		1
CT, n (%)	47 (45)	48 (43)	0.50	1.2 (0.7-2.2)	0.50	1.2 (0.7-2.2)
TT, n (%)	16 (15)	11 (10)	0.20	1.4 (0.9-2.1)	0.30	1.3 (0.8-2.0)
CT+TT, n (%)	63 (60)	59 (53)	0.30	1.4 (0.8-2.3)	0.30	1.3 (0.8-2.3)
C, n (%)	131 (62)	154 (69)		1		
T, n (%)	79 (38)	70 (31)	0.19	1.3 (0.9-2.0)		
Asn453Ser (A/G) rs1800440						
AA	71 (68)	64 (57)		1		
AG	29 (27)	42 (37)	0.11	0.6 (0.4-1.1)	0.12	0.6 (0.4-1.1)
GG	5 (5)	6 (6)	0.70	0.9 (0.5-1.6)	0.70	0.9 (0.5-1.6)
AG+GG	34 (32)	48 (43)	0.11	0.6 (0.4-1.1)	0.12	0.7 (0.4-1.1)
Allele						
A	171 (82)	170 (76)				
G	39 (18)	54 (24)	0.16	0.7 (0.5-1.1)		
Arg368His (G/A) rs79204362						
GG	81 (77)	98 (88)		1		
GA	21 (20)	14 (13)	0.11	1.8 (0.9-3.8)	0.11	1.8 (0.9-3.8)
AA	3 (3)	0 (0)	-	-	-	-
GA+AA	24 (23)	14 (13)	0.11	1.8 (0.9-3.8)	0.11	1.8 (0.9-3.8)
Allele						
G	183 (87)	210 (94)				
A	27 (13)	14 (6)	0.02	2.2 (1.1-4.4)		

^aAdjusted for age and ethnicity. *CYP1B1*, cytochrome P450 1B1; UL, uterine leiomyoma; OR, odds ratios; CI, confidence intervals.

Table III. Haplotypes frequency of the *CYP1B1* gene polymorphisms in UL patients and controls.

<i>CYP1B1</i> SNPs				Leiomyoma (n=105)	Control (n=112)	P-value	OR (95% CI)
rs1056836	rs1056837	rs1800440	rs79204362				
C	C	A	G	0.446518	0.445568		1
C	C	G	G	0.240873	0.177685	0.30	1.3 (0.8-2.2)
C	T	A	G	0.013393	0	0.10	-
G	T	A	G	0.236670	0.240574	0.95	1.0 (0.9-1.2)
G	T	A	A	0.062302	0.127683	0.039 ^a	0.8 (0.7-1)

^aP<0.05. *CYP1B1*, cytochrome P450 1B1; SNP, single-nucleotide polymorphism; UL, uterine leiomyoma; OR, odds ratios; CI, confidence intervals.

The UL development possibly occurs in response to a certain type of injury to the myometrium, such as hypoxia (21).

CYP1B1 is known as a key enzyme that metabolizes a variety of putative human carcinogens and estrogens. Certain evidence stated that this enzyme may play an essential role in tumor development and progression (22). Therefore, polymorphisms in the *CYP1B1* gene may affect the metabolism of environmental estrogens and hormone-like substances. The polymorphisms influenced the inter-individual risk of developing a tumor (22). The CYP1B1 enzyme catalyzes the conversion of estrogens to 4-hydroxy estrogen metabolites and recent products can increase the quantity of estrogen within the cells by activating the ESR. Certain evidence has shown that the CYP1B1 variants are more efficient in the conversion and accumulation of carcinogenic estrogens (23,24).

Thus, inherited variations in the CYP1B1 activity lead to differences in estrogen metabolism and explain inter-individual differences in the UL risk in association with estrogen-mediated tumorigenesis (25).

Several studies have addressed the association between the genetic polymorphisms of enzymes involved in the estrogen metabolism and the occurrence of UL (14,17,26,27). In 2006, Herr *et al* (26) reported a significant correlation between *CYP2A13* and *CYP1A1* SNPs and UL in a Caucasian population. The genetic distribution of Ile462Val and T6235C polymorphisms of the *CYP1A1* gene and Arg48Gly, Ala119Ser, Leu432Val and Asp449Asp polymorphisms of the *CYP1B1* gene were studied by Ye *et al* (17) in Chinese females. The studies reported that only the *CYP1A1* Ile462Val polymorphism was associated with the increased risk of UL. However, similar to the findings of the present study, there was no association between polymorphisms of the *CYP1B1* gene and UL susceptibility in China. Despite the results of Ye *et al* (17) in Chinese females, El-Shennawy *et al* (14) observed an elevated risk of UL in females with the CC genotype of the *CYP1B1* Leu432Val (C/G) polymorphism and according to their results this genotype can predict the susceptibility to UL in Egyptian females. Shen *et al* (27,28) studied the association between the Ala119Ser, Leu432Val and Asp449Asp polymorphisms of the *CYP1B1* gene and UL in Han Chinese females. Consistent with the results of the present study, an association between the Asp449Asp (T/C) polymorphism gene and UL was not observed, however, the studies found that the Ala119Ser polymorphism was a risk factor and the Leu432Val polymorphism was a protective factor for UL development. The present study findings showed a strong LD between the three common SNPs in the *CYP1B1* gene in the population. Similarly, Bethke *et al* (29) observed a strong LD between the five common SNPs in the *CYP1B1* gene in UK.

To the best of our knowledge, this is the first study regarding the effect of *CYP1B1* haplotypes (four SNPs) on UL susceptibility, therefore, further studies in different populations are required to evaluate the association between *CYP1B1* haplotypes and UL.

In conclusion, there was an association between the A (His) allele of the *CYP1B1* Arg368His (rs79204362) polymorphism and UL. In addition, the GTAA haplotype may play a protective role in SLE susceptibility. There was a strong LD between the three common SNPs (rs1056836, rs1056837 and rs1800440) of the *CYP1B1* gene in the southeast Iranian population.

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