

Anti-allergic effect of the naturally-occurring conjugated linolenic acid isomer, jacaric acid, on the activated human mast cell line-1

WAI NAM LIU and KWOK NAM LEUNG

Biochemistry Programme, School of Life Sciences, The Chinese University of Hong Kong, Shatin, Hong Kong, SAR, P.R. China

Received June 24, 2015; Accepted September 9, 2015

DOI: 10.3892/br.2015.517

Abstract. The present study aimed to investigate the immunomodulatory effect of jacaric acid, a naturally-occurring conjugated linolenic acid isomer that can be found in jacaranda seed oil, on the activated human mast cell line-1 (HMC-1). Our previous studies have demonstrated that jacaric acid only exerted minimal, if any, cytotoxicity on normal murine cells. In the present study, jacaric acid at concentrations $\leq 100 \mu\text{M}$ did not exhibit direct cytotoxicity on human peripheral blood mononuclear cells after 72 h of incubation, as determined by the MTT reduction assay. By contrast, jacaric acid could alleviate the calcium ionophore A23187 and phorbol 12-myristate 13-acetate-triggered allergic response in the HMC-1 cells at concentrations that were non-cytotoxic to the HMC-1 cells. Following pre-treatment with jacaric acid, the secretion of two inflammatory mediators, β -N-acetylglucosaminidase and tryptase, as well as the T helper 2 cytokines [interleukin (IL)-4 and IL-13] was significantly reduced in HMC-1 cells. The alleviation of allergic response was accompanied by downregulation of the matrix metalloproteinase-2 and -9 proteins and upregulation of the tissue inhibitor of metalloproteinase-1 protein. Collectively, the results indicated that the naturally-occurring jacaric acid exhibits a suppressive effect on the allergic response in activated human mast cells *in vitro*, and this could not be attributed to the direct cytotoxicity of jacaric acid on the treated cells.

Introduction

Conjugated fatty acids (CFAs) refer to the positional and geometric isomers of polyunsaturated FAs with conjugated

double bonds, in which 2 carbon-carbon double bonds are separated solely by one carbon-carbon single bond (1). Common CFA isomers, such as conjugated linoleic acids (CLAs) and conjugated linolenic acids (CLNAs), can be found naturally in meat or dairy products of ruminant animals (2) and different plant seed oils (3), respectively. CLAs have been studied most extensively due to their diverse metabolic and physiological effects (4,5). In recent years, CLNAs have received increasing attention as their relative abundance in certain plant seed oils (30-70% of total lipids) was much higher compared to that of CLAs (<1% of total lipids) (3,6). Previous *in vitro* and *in vivo* studies have demonstrated that CLNAs exhibit pleiotropic physiological and pharmacological activities, including anti-carcinogenic, anti-inflammatory, anti-obese, antioxidative and immunomodulatory properties (7,8). An earlier study by Ike *et al* (9) showed that α -eleostearic acid (9Z, 11E, 13E-CLNA isomer) isolated from bitter melon could induce interferon- γ production in mice treated with heat-inactivated *Propionibacterium acnes*, suggesting that T helper 1 (Th1) cellular immunity can be activated by α -eleostearic acid, which is responsible for the defense against intracellular parasitic infection. By contrast, Yamasaki *et al* (10) reported that consumption of pomegranate seed oil that is rich in punicic acid (9Z, 11E, 13Z-CLNA isomer) could ameliorate the function of B cells, which have a key role in the humoral immune response. These studies started to explain the immunostimulatory activities of CLNAs, however, the anti-allergic activity of CLNAs and their modulatory effects on mast cells have not yet been investigated. In the present study, jacaric acid (8Z, 10E, 12Z-CLNA isomer; Fig. 1) alleviated the allergic response in calcium ionophore A23187 (Iono) and phorbol 12-myristate 13-acetate (PMA)-activated human mast cells by reducing the release of inflammatory mediators and Th2 cytokines, and by modulating the protein expression levels of matrix metalloproteinases (MMPs).

Materials and methods

Chemicals and reagents. Jacaric acid (8Z, 10E, 12Z-CLNA isomer) used in the study, with an estimated purity >97%, was purchased from Larodan Fine Chemicals AB (Limhamn, Sweden). The stock solution (0.2 M) was prepared by dissolving the powder in sterile, cell culture-tested ethanol

Correspondence to: Professor Kwok Nam Leung, Biochemistry Programme, School of Life Sciences, The Chinese University of Hong Kong, 12 Chak Cheung Street, Shatin, Hong Kong, SAR, P.R. China

E-mail: knleung@cuhk.edu.hk

Key words: anti-allergic effect, β -N-acetylglucosaminidase, conjugated linolenic acids, interleukins, jacaric acid, mast cells, matrix metalloproteinases, tryptases

(Sigma-Aldrich, St. Louis, MO, USA). All other chemicals, unless otherwise stated, were purchased from Sigma-Aldrich.

Culture of cell line. Human mast cell line-1 (HMC-1) was a generous gift from Professor C.K. Wong of the Department of Chemical Pathology, The Chinese University of Hong Kong (Shatin, Hong Kong, SAR, China) (11). The cells were maintained in RPMI-1640 medium (Gibco, Thermo Fisher Scientific, Grand Island, NY, USA) supplemented with 10% fetal bovine serum (Gibco, Thermo Fisher Scientific) and 1% antibiotics (100 U/ml penicillin G, 100 µg/ml streptomycin sulfate and 0.25 µg/ml amphotericin B in 0.85% saline from Gibco, Thermo Fisher Scientific) in a humidified incubator containing 5% CO₂ in air at 37°C.

The procedures for the stimulation of HMC-1 cells were modified from a previously described method (12). In brief, HMC-1 cells (3x10⁵ cells/ml) were pre-treated with different concentrations of jacaric acid at 37°C for 72 h in 175-cm² tissue culture flasks. Following incubation, cells were centrifuged at 400 x g for 5 min, washed with phosphate-buffered saline three times and 3x10⁶ cells were transferred to a flat-bottomed 6-well microtiter plate in 1 ml complete RPMI medium, and were activated with 2 µM Iono and 40 nM PMA at 37°C for 6 h for the further assays.

Cytotoxicity assay. Human peripheral blood mononuclear cells (PBMCs) were prepared from the fresh human buffy coats supplied by the Hong Kong Red Cross Blood Transfusion Service (Kowloon, Hong Kong, SAR, China). The PBMCs used were a generous gift from Professor C.K. Wong of the Department of Chemical Pathology, The Chinese University of Hong Kong (13). Briefly, the viability of jacaric acid-treated HMC-1 cells and PBMCs was measured by the colorimetric MTT reduction assay, as described previously (14). Briefly, HMC-1 cells (3x10⁵/ml) and PBMCs (3x10⁶/ml) seeded in a flat-bottomed 96-well microtiter plate were incubated at 37°C with different concentrations of jacaric acid for various periods of time, and the cell viability was recorded by a Benchmark microplate reader (Bio-Rad Laboratories, Inc., Hercules, CA, USA). To further confirm the viability of jacaric acid-treated HMC-1 cells followed by Iono- and PMA-activation, the cells were counted by the trypan blue exclusion assay and the percentages of necrotic cells in the samples were also determined by the annexin V-green fluorescent protein (GFP)/propidium iodide (PI) dual staining assay using the FACSCanto™ flow cytometer (BD Biosciences, Franklin Lakes, NJ, USA) as described previously (14).

Determination of β-N-acetylglucosaminidase (NAG) release in HMC-1 cells. The measurement was performed according to the manufacturer's instructions in the NAG assay kit (Sigma-Aldrich) with slight modifications. Briefly, HMC-1 cells (3x10⁵ cells/ml) were incubated with different concentrations of jacaric acid at 37°C for 72 h in 175-cm² tissue culture flasks and the treated cells (3x10⁶) were further stimulated by 2 µM Iono and 40 nM PMA at 37°C for 6 h in a 6-well plate. Following incubation, the stimulated cells were centrifuged at 400 x g for 5 min, and 50 µl cell-free supernatant or cell lysate was transferred to the well of a flat-bottomed 96-well plate. A total of 50 µl of 0.09 M sodium citrate solution (pH 4.7)

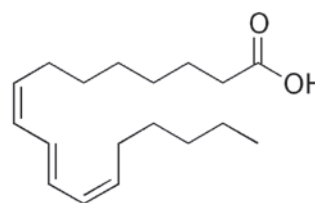


Figure 1. Chemical structure of jacaric acid (8Z, 10E, 12Z-conjugated linolenic acid isomer).

containing 1 mg/ml 4-nitrophenyl-*N*-acetyl-β-D-glucosaminide was added to the wells and incubated at 37°C for 30 min for color development. The enzymatic reaction was terminated by adding 200 µl of sodium carbonate solution [0.4 M (pH 11.6)] and the absorbance at 405 nm was measured by the Benchmark microplate reader. The concentrations of NAG in the samples were expressed in U/ml, in which 1 unit of NAG will hydrolyze 1 nanomole of 4-nitrophenyl-*N*-acetyl-β-D-glucosaminide to *p*-nitrophenol and *N*-acetyl-β-D-glucosaminide per min at pH 4.7 and 37°C. The percentage of NAG release was calculated as follows: % NAG release = (concentration of NAG in supernatant)/(concentration of NAG in supernatant and whole cell lysate) x100.

Measurement of tryptase release in HMC-1 cells. The assessment was performed according to the manufacturer's instructions in the Mast Cell Degranulation Assay kit (Millipore Corp., Billerica, MA, USA). HMC-1 cells (3x10⁵ cells/ml) were incubated with different concentrations of jacaric acid at 37°C for 72 h in 175-cm² tissue culture flasks and the treated cells (3x10⁶) were further stimulated by Iono and PMA at 37°C for 6 h in a 6-well plate. Following incubation, the stimulated cells were centrifuged at 400 x g for 5 min, and 180 µl cell-free supernatant was transferred to the well of a flat-bottomed 96-well plate. A total of 20 µl of tryptase substrate was added to each well and the samples were incubated at 37°C for 2 h. The absorbance at 405 nm was measured by the Benchmark microplate reader and the concentrations of tryptase in the samples were quantified with reference to a series of tryptase positive control solutions with concentrations ranging from 0 to 10,000 ng/ml.

Assessment of cytokine secretion by ELISA. To determine the secretion of interleukin (IL)-4 and IL-13, the corresponding ELISA kit was used (ExCell Biology, Inc., Shanghai, China). Briefly, HMC-1 cells (3x10⁵ cells/ml) were incubated with different concentrations of jacaric acid at 37°C for 72 h in 175-cm² tissue culture flasks and the treated cells (3x10⁶) were further stimulated by 2 µM Iono and 40 nM PMA at 37°C for 6 h in a 6-well plate. The cell-free supernatant was transferred to another 96-well plate provided in the ELISA kit. Subsequently, the absorbance at 405 nm was measured by the Benchmark microplate reader.

Western blotting. Protein expression levels were determined by the western blotting technique with the aid of a panel of specific antibodies. HMC-1 cells (3x10⁵ cells/ml) were incubated with different concentrations of jacaric acid at 37°C for 72 h in 175-cm² tissue culture flasks and the treated HMC-1 cells were

transferred to a 6-well plate and sensitized in the presence of 2 μ M Iono and 40 nM PMA at 37°C for 6 h. After incubation, the cell pellet was collected and total protein was extracted by the cell lysis buffer. Protein concentration was measured by the Bradford reagent and the protein sample was resolved on 12% polyacrylamide gels and transferred to polyvinylidene difluoride membranes. The membrane was first incubated with the following primary antibodies: Rabbit anti-MMP-2 (CST-4022S), anti-MMP-9 (CST-3852S), anti-tissue inhibitor of metalloproteinase-1 (TIMP-1) (CST-8946S) (Cell Signaling Technology, Inc., Danvers, MA, USA) and mouse anti- β -actin antibody (SC-A5316) (Santa Cruz Biotechnology, Inc., Dallas, TX, USA), followed by incubation with mouse IgG horseradish peroxidase-conjugated secondary antibody (GE-NA931) or rabbit IgG horseradish peroxidase-conjugated secondary antibody (GE-NA934) (GE Healthcare, Buckinghamshire, UK and finally developed with the enhanced chemiluminescence reagent (Santa Cruz Biotechnology, Inc.).

Statistical analysis. Each experiment was repeated at least three times and only the results of the most representative experiments are shown. The data are expressed as the arithmetic mean \pm standard error. One-way analysis of variance was used for statistical analysis, and $P < 0.05$ was considered to indicate a statistically significant difference.

Results

Jacaric acid exhibits no direct cytotoxicity on HMC-1 cells and PBMCs. To investigate whether jacaric acid was cytotoxic to the HMC-1 cells or PBMCs, the colorimetric MTT reduction assay was employed. Fig. 2A shows that jacaric acid at concentrations ≤ 8 μ M exhibited no significant cytotoxicity to the HMC-1 cells ($>90\%$ cell viability) after 72 h of incubation when compared with the solvent control. Additionally, the viability of jacaric acid-treated HMC-1 cells following Iono- and PMA-activation was examined by the trypan blue exclusion assay and by the annexin V-GFP/PI dual staining assay. As shown in Fig. 2B and C, the cell viability of HMC-1 cells and the percentage of the necrotic cells respectively did not differ significantly between the jacaric acid-treated group and the solvent-treated control. Notably, the viability of the PBMCs remained $>90\%$ when the cells were incubated with ≤ 100 μ M jacaric acid for 6, 24, 48 and 72 h (Fig. 3), suggesting that jacaric acid was neither cytotoxic to the HMC-1 cells, nor to the PBMCs under the prescribed experimental conditions.

Jacaric acid suppresses the release of NAG and tryptase from HMC-1 cells. Mast cells have a well-known central role in allergic response by releasing pharmacologically active inflammatory mediators, which cause the symptoms of allergic inflammatory reaction (15). Common inflammatory mediators, including NAG and tryptase, are potent markers of allergic disease (16). In the present study, the release of NAG and tryptase in HMC-1 cells was increased by 2-fold upon activation with Iono and PMA (data not shown), suggesting that the combined use of Iono and PMA can elicit an allergic response. To investigate whether jacaric acid can exhibit an anti-allergic response, jacaric acid-treated HMC-1 cells were stimulated by 2 μ M Iono and 40 nM PMA, and the release

Table I. Jacaric acid suppresses β -N-acetylglucosaminidase and tryptase release in HMC-1 cells.

Release of inflammatory mediator	Concentration of jacaric acid, μ M		
	0	2	4
NAG, %	31.80 \pm 3.82	25.85 \pm 2.02	16.32 \pm 1.02 ^a
Tryptase, U/ml	503.93 \pm 25.73	496.38 \pm 26.46	408.85 \pm 20.38 ^a

Percentage of β -N-acetylglucosaminidase (NAG) release was calculated as follows: % NAG release = (concentration of NAG in supernatant)/(concentration of NAG in supernatant and whole cell lysate) \times 100. ^a $P < 0.001$. HMC-1, human mast cell line-1.

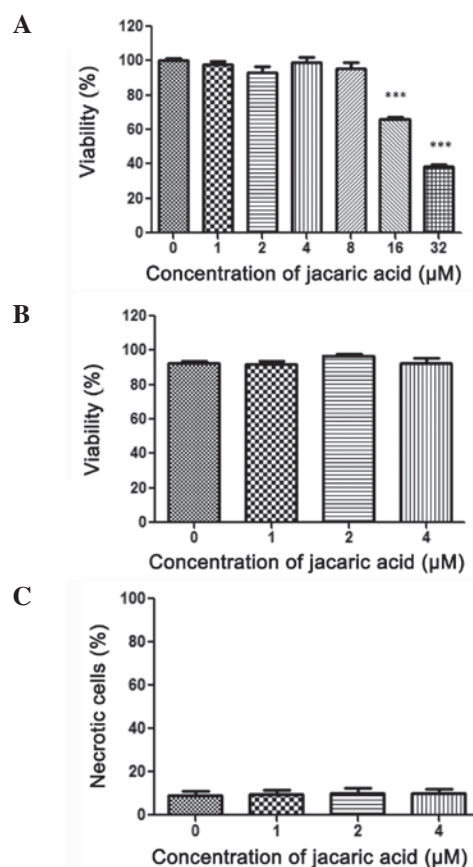


Figure 2. Effect of jacaric acid on the viability of the human mast cell line-1 (HMC-1) cells. (A) HMC-1 cells were incubated with different concentrations of jacaric acid at 37°C for 72 h. Cells treated with 0.02% ethanol acted as the control. The cell viability was determined by the MTT reduction assay. The results are expressed as the mean percentage of cell viability \pm standard error (SE) of quadruplicate cultures; *** $P < 0.001$. (B and C) HMC-1 cells were pre-treated with either different concentrations of jacaric acid or 0.02% ethanol at 37°C for 72 h, followed by Iono- and phorbol 12-myristate 13-acetate-sensitization at 37°C for 6 h. (B) Cell viability was determined by the trypan blue exclusion assay, and the results are expressed as mean percentage of cell viability \pm SE. (C) Following incubation, the cells were stained with annexin V-green fluorescent protein and propidium iodide, and the fluorescence intensity was measured by FACSanto™ flow cytometer. The percentage of necrotic cells was quantified and expressed as mean \pm SE.

of NAG and tryptase from HMC-1 cells was determined. As shown in Table I, pre-treatment of HMC-1 cells with jacaric acid at 4 μ M could significantly suppress the secretion of NAG

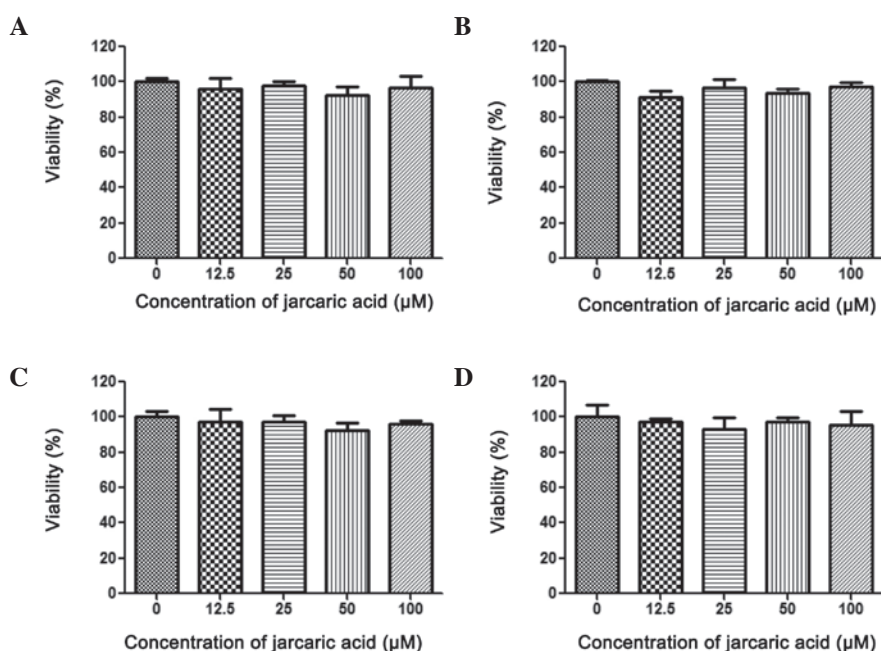


Figure 3. Jacaric acid does not exhibit cytotoxicity on the human peripheral blood mononuclear cells. Human peripheral blood mononuclear cells were incubated with different concentrations of jacaric acid at 37°C for (A) 6, (B) 24, (C) 48 or (D) 72 h. Cells treated with 0.02% ethanol acted as the control. Cell viability was determined by the MTT reduction assay. The results are expressed as the mean percentage of cell viability \pm standard error of quadruplicate cultures.

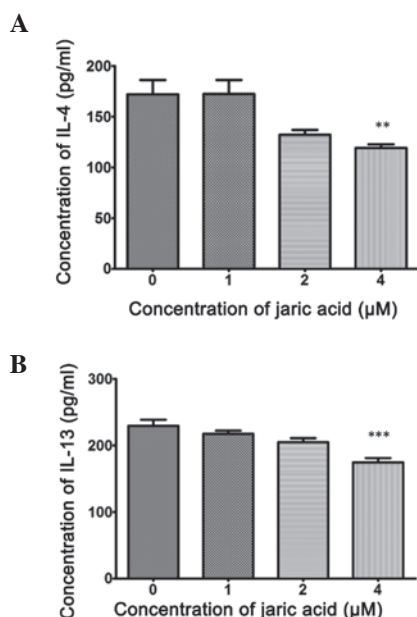


Figure 4. Jacaric acid reduces the secretion of interleukin (IL)-4 and IL-13 in human mast cell line-1 (HMC-1) cells. HMC-1 cells were incubated with different concentrations of jacaric acid at 37°C for 72 h. Cells treated with 0.02% ethanol acted as the control. The treated cells were further sensitized with Iono and phorbol 12-myristate 13-acetate at 37°C for 6 h. The concentrations of (A) IL-4 and (B) IL-13 were quantified by ELISA kit according to manufacturer's instructions, and the results are expressed as mean \pm standard error. ** $P < 0.01$; *** $P < 0.001$.

from sensitized HMC-1 cells. To confirm the anti-allergic effect of jacaric acid, the release of tryptase in HMC-1 cells was assessed by the Mast Cell Degranulation Assay kit. Jacaric acid could also inhibit the release of tryptase from sensitized HMC-1 cells (Table I). These results suggest that jacaric acid could alleviate the allergic response in sensitized HMC-1 cells.

Jacaric acid suppresses the secretion of Th2 cytokines in HMC-1 cells. Apart from the release of NAG and tryptase, a previous study demonstrated that the secretion of two Th2 cytokines, IL-4 and IL-13, is also a marker of allergic response upon stimulation by Iono and PMA (12). The present results showed that the secretion of IL-4 (Fig. 4A) and IL-13 (Fig. 4B) in activated HMC-1 cells was significantly reduced when the cells were pre-treated with 4 μ M jacaric acid.

Jacaric acid modulates the expression levels of MMP-2, MMP-9 and TIMP-1 proteins in HMC-1 cells. The recruitment and migration of inflammatory cells require degradation of extracellular matrix proteins by secreted MMP. MMP-2 and MMP-9 are considered to have a key role in tissue remodeling and repair through degradation of type IV collagen, whereas the activation of MMP is inhibited by TIMPs (17). To unravel the molecular action mechanisms underlying the anti-allergic response, the effects of jacaric acid on modulating the expression levels of MMP-2, MMP-9 and TIMP-1 proteins were examined by Western blotting (Fig. 5A). Following pre-treatment with jacaric acid, the expression levels of MMP-2 and MMP-9 proteins in sensitized HMC-1 cells were significantly decreased (Fig. 5B and C), and this was accompanied by an increase in the expression level of TIMP-1 protein (Fig. 5D), suggesting that jacaric acid may exert its anti-allergic response in activated HMC-1 cells via modulation of MMP-2, MMP-9 and TIMP-1 protein expression levels.

Discussion

Mast cells are the central effector cells in allergic response and they are also involved in the defense against pathogens, typically by cross-linking of surface immunoglobulin E (IgE) receptors, or through IgE-independent mechanisms

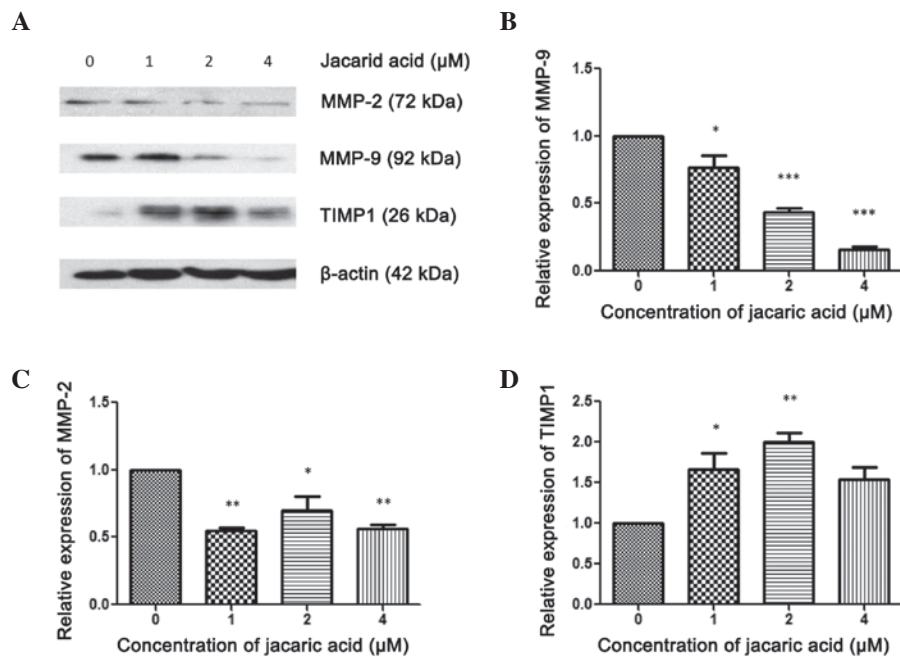


Figure 5. Jacaric acid modulates the expression levels of matrix metalloproteinase (MMP)-2, MMP-9 and tissue inhibitor of metalloproteinase-1 (TIMP-1) proteins in human mast cell line-1 (HMC-1) cells. HMC-1 cells were incubated with 1 (lane 2), 2 (lane 3) and 4 μ M jacaric acid (lane 4) at 37°C for 72 h. Cells treated with 0.02% ethanol (lane 1) acted as the control. Subsequently, treated cells were further incubated with Iono and phorbol 12-myristate 13-acetate at 37°C for 6 h. (A) Protein expression levels of MMP-2, MMP-9 and TIMP-1 were assayed by western blotting with β -actin protein as an internal control. (B-D) The relative expression levels of MMP-9, MMP-2 and TIMP-1 proteins compared to β -actin were quantified. Results represent mean \pm standard error. * P <0.05; ** P <0.01; *** P <0.001.

in non-pathogenic conditions (15,18). The HMC-1 cell line was derived from a patient with mastocytosis by Butterfield *et al* (19) in 1988, which shows numerous characteristics of immature mast cells and contains classical mast cell-associated markers, such as the c-kit receptor, heparin, histamine and tryptase, and secretes a panel of bioactive cytokines and a variety of MMP (18). In the present study, the modulatory effects and the underlying action mechanisms of jacaric acid on the human mast cells using the HMC-1 cells as the *in vitro* model were examined. Pre-treatment of HMC-1 cells with jacaric acid at lower concentrations (<5 μ M) could reduce the secretion of inflammatory mediators, including NAG and tryptase, when the HMC-1 cells were activated with Iono and PMA. The presence of allergens or stimulants elicits an inflammatory response, which leads to the increased production and release of Th2 cytokines, such as IL-4, IL-5 and IL-13, in mast cells (17). The present results showed that the release of IL-4 and IL-13 was suppressed following the pre-treatment of activated HMC-1 cells with jacaric acid. To further elucidate the anti-allergic effect of jacaric acid on HMC-1 cells, the expression levels of MMP-2, MMP-9 and TIMP-1 proteins were examined. MMPs belong to a family of Zn-dependent endopeptidases that are involved in the decomposition of extracellular matrix and basement membrane (20). Furthermore, MMP-2, MMP-9 and MMP-13 have a key role in tissue remodeling and repair through degradation of type IV collagen, which is the major component of the basement membrane (21). By contrast, the activation of MMP is inhibited by TIMP, and previous studies have suggested that MMP-2 and MMP-9 are inactivated by TIMP-1 and TIMP-2 (22,23). By western blotting, jacaric acid-treated HMC-1 cells showed an increase in the TIMP-1 protein expression level, accompanied

by a decrease in the expression levels of MMP-2 and MMP-9 proteins. To the best of our knowledge, this is the first study demonstrating the *in vitro* anti-allergic effect of CLNAs on HMC-1 cells, and this is in line with earlier findings showing that CFAs, such as CLAs, may possess anti-allergic activities. For example, Jaudszus *et al* (24) reported that 9Z, 11E-CLA could inhibit allergic sensitization and airway inflammation through peroxisome proliferator-activated receptor- γ dependent mechanism in mice, whereas others demonstrated that human healthy volunteers supplemented with 9Z, 11E-CLA resulted in a decreased plasma IgE level and reduced secretion of IL-5 by PBMCs (25,26). Despite these earlier findings, the underlying action mechanisms for the anti-allergic effects of CFAs, such as CLAs and CLNAs, remain poorly understood and further studies are required to elucidate the mechanisms, *in vitro* and *in vivo*, for the anti-allergic activity of CFAs. In addition, whether CLNA supplementation could ameliorate other types of hypersensitivity reactions, such as the *ex vivo* delayed-type hypersensitivity response as shown by CLAs (25), is also a noteworthy aspect that is currently under investigation.

Collectively, the present results suggest that jacaric acid may exhibit modulatory effects on human mast cells, as it was shown to alleviate the allergic response in HMC-1 cells activated by Iono and PMA. As jacaric acid was found to exert minimal direct cytotoxicity on normal human PBMCs, and apparently was non-toxic to mice (27), further elucidation of the immunomodulatory effects of jacaric acid, *in vitro* and *in vivo*, may provide better insights for the development of jacaric acid as a potential candidate for the treatment of certain allergic disorders with minimal toxicity and fewer side effects.

References

1. Nagao K and Yanagita T: Conjugated fatty acids in food and their health benefits. *J Biosci Bioeng* 100: 152-157, 2005.
2. Palmquist DL, Lock AL, Shingfield KJ and Bauman DE: Biosynthesis of conjugated linoleic acid in ruminants and humans. *Adv Food Nutr Res* 50: 179-217, 2005.
3. Takagi T and Itabashi Y: Occurrence of mixtures of geometrical isomers of conjugated octadecatrienoic acids in some seed oils. Analysis by open tubular gas liquid chromatography. *Lipids* 16: 546-551, 1981.
4. Dilzer A and Park Y: Implication of conjugated linoleic acid (CLA) in human health. *Crit Rev Food Sci Nutr* 52: 488-513, 2012.
5. Wang T and Lee HG: Advances in research on cis-9, trans-11 conjugated linoleic acid: A major functional conjugated linoleic acid isomer. *Crit Rev Food Sci Nutr* 55: 720-731, 2015.
6. Tanaka T, Hosokawa M, Yasui Y, Ishigamori R and Miyashita K: Cancer chemopreventive ability of conjugated linolenic acids. *Int J Mol Sci* 12: 7495-7509, 2011.
7. Hennessy AA, Ross RP, Devery R and Stanton C: The health promoting properties of the conjugated isomers of α -linolenic acid. *Lipids* 46: 105-119, 2011.
8. Saha SS and Ghosh M: Protective effect of conjugated linolenic acid isomers present in vegetable oils against arsenite-induced renal toxicity in rat model. *Nutrition* 29: 903-910, 2013.
9. Ike K, Uchida Y, Nakamura T and Imai S: Induction of interferon-gamma (IFN-gamma) and T helper 1 (Th1) immune response by bitter melon extract. *J Vet Med Sci* 67: 521-524, 2005.
10. Yamasaki M, Kitagawa T, Koyanagi N, Chujo H, Maeda H, Kohno-Murase J, Imamura J, Tachibana H and Yamada K: Dietary effect of pomegranate seed oil on immune function and lipid metabolism in mice. *Nutrition* 22: 54-59, 2006.
11. Wong CK, Tsang CM, Ip WK and Lam CW: Molecular mechanisms for the release of chemokines from human leukemic mast cell line (HMC)-1 cells activated by SCF and TNF- α : Roles of ERK, p38 MAPK, and NF- κ B. *Allergy* 61: 289-297, 2006.
12. Yanagihara Y, Kajiwaru K, Basaki Y, Ikizawa K, Akiyama K and Saito H: Induction of human IgE synthesis in B cells by a basophilic cell line, KU812. *Clin Exp Immunol* 108: 295-301, 1997.
13. Han XQ, Yue GL, Yue RQ, Dong CX, Chan CL, Ko CH, Cheung WS, Luo KW, Dai H, Wong CK, *et al*: Structure elucidation and immunomodulatory activity of a beta glucan from the fruiting bodies of *Ganoderma sinense*. *PLoS One* 9: e100380, 2014.
14. Liu WN and Leung KN: Apoptosis- and differentiation-inducing activities of jacaric acid, a conjugated linolenic acid isomer, on human eosinophilic leukemia EoL-1 cells. *Oncol Rep* 32: 1881-1888, 2014.
15. Guhl S, Babina M, Neou A, Zuberbier T and Artuc M: Mast cell lines HMC-1 and LAD2 in comparison with mature human skin mast cells - drastically reduced levels of tryptase and chymase in mast cell lines. *Exp Dermatol* 19: 845-847, 2010.
16. Stone KD, Prussin C and Metcalfe DD: IgE, mast cells, basophils, and eosinophils. *J Allergy Clin Immunol* 125 (Suppl 2): S73-S80, 2010.
17. Mori S, Pawankar R, Ozu C, Nonaka M, Yagi T and Okubo K: Expression and roles of MMP 2, MMP 9, MMP 13, TIMP 1, and TIMP 2 in allergic nasal mucosa. *Allergy Asthma Immunol Res* 4: 231-239, 2012.
18. Galinsky DST and Nechushtan H: Mast cells and cancer - no longer just basic science. *Crit Rev Oncol Hematol* 68: 115-130, 2008.
19. Butterfield JH, Weiler D, Dewald G and Gleich GJ: Establishment of an immature mast cell line from a patient with mast cell leukemia. *Leuk Res* 12: 345-355, 1988.
20. Murphy G and Docherty AJP: The matrix metalloproteinases and their inhibitors. *Am J Respir Cell Mol Biol* 7: 120-125, 1992.
21. Salib RJ and Howarth PH: Remodelling of the upper airways in allergic rhinitis: Is it a feature of the disease? *Clin Exp Allergy* 33: 1629-1633, 2003.
22. Nagase H: Activation mechanisms of matrix metalloproteinases. *Biol Chem* 378: 151-160, 1997.
23. Shimizu T, Kanai K, Asano K, Hisamitsu T and Suzuki H: Suppression of matrix metalloproteinase production in nasal fibroblasts by tranilast, an antiallergic agent, in vitro. *Mediators Inflamm* 2005: 150-159, 2005.
24. Jaudszus A, Krokowski M, Möckel P, Darcan Y, Avagyan A, Matricardi P, Jahreis G and Hamelmann E: Cis-9, trans-11-conjugated linoleic acid inhibits allergic sensitization and airway inflammation via a PPAR γ -related mechanism in mice. *J Nutr* 138: 1336-1342, 2008.
25. Song HJ, Grant I, Rotondo D, Mohede I, Sattar N, Heys SD and Wahle KWJ: Effect of CLA supplementation on immune function in young healthy volunteers. *Eur J Clin Nutr* 59: 508-517, 2005.
26. Turpeinen AM, Ylönen N, von Willebrand E, Basu S and Aro A: Immunological and metabolic effects of cis-9, trans-11-conjugated linoleic acid in subjects with birch pollen allergy. *Br J Nutr* 100: 112-119, 2008.
27. Shinohara N, Tsuduki T, Ito J, Honma T, Kijima R, Sugawara S, Arai T, Yamasaki M, Ikezaki A, Yokoyama M, *et al*: Jacaric acid, a linolenic acid isomer with a conjugated triene system, has a strong antitumor effect in vitro and in vivo. *Biochim Biophys Acta* 1821: 980-988, 2012.