Gastroprotective effects of arctigenin of *Arctium lappa* L. on a rat model of gastric ulcers

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Received April 4, 2016; Accepted July 12, 2016

**Abstract.** In the present study, the gastroprotective effects of arctigenin of *Fructus Arctii* were evaluated and the possible underlying mechanisms of action were elucidated. Arctigenin (high-performance liquid chromatography purity, >99.0%) was isolated and purified from the seeds of *Arctium lappa* L. The anti-ulcerogenic activity of arctigenin against ulcers induced by absolute ethanol and acetic acid was evaluated in a Sprague-Dawley rat model. In addition, the antioxidant activity was assessed by measuring malondialdehyde (MDA) levels in an ethanol-induced model and the anti-inflammatory effects were assessed by measuring five factors in an acetic acid-induced model. In the ethanol-induced model, arctigenin inhibited gastric lesions in a dose-dependent manner, by 53.04, 53.91 and 64.43% at doses of 0.05, 0.15 and 0.45 mg/kg, respectively. In addition, arctigenin reduced MDA (P<0.01) and increased superoxide dismutase (P<0.01) levels in serum when compared with the vehicle group. The lesion index induced by acetic acid was significantly inhibited by all doses of arctigenin (0.05, 0.15 and 0.45 mg/kg; P<0.01) in comparison to the vehicle group and in a dose-dependent manner. In addition, it was shown that the expression levels of tumor necrosis factor-α, interleukin-6 (IL-6), IL-10 and C-reactive protein were significantly decreased (P≤0.05) in the arctigenin group compared with the vehicle group. Thus, the current study indicated that arctigenin exerted anti-ulcer activity, which may be associated with its reduction in oxidative and inflammatory damage. All the results indicate that arctigenin may be used as an effective therapeutic agent to prevent gastric ulcers.

**Introduction**

Gastric ulcers are characterized by necrosis, induction of oxidative stress and secretion of inflammatory factors (1). In the Western population, the prevalence of gastric ulcers is 2.4% (2) and 6.07% in China (3). In addition, in patients with gastrointestinal symptoms, the incidence rates of gastric ulcers increases to 22.5% (4). According to the epidemiological investigation, when individuals smoke, consume alcohol or use nonsteroidal anti-inflammatory drugs, the incidence of gastric ulcer increases (5-7). Although gender differences have not been reported for gastric ulcers, the incidence rate in women has increased during the previous 10 years (8). The pathogenesis of gastric ulcers is based on a multifactorial and complex interaction between protective and aggressive factors, including mucosal integrity, secretion of gastric acid, *Helicobacter pylori*, free oxygen radicals and excess alcohol consumption (9-11). Currently, the treatment is predominantly performed by inhibition of gastric acid secretion, proton pump inhibitors and eradication of *H. pylori* with antibiotics. However, such treatments are not completely effective and produce certain adverse effects (12). Disturbing the balance between aggressive and protective factors that control cell apoptosis and proliferation results in gastric ulceration, which then activates the repairing system in the gastric mucosa. For these reasons, the search for novel therapeutic agents is relevant, and medicinal plants and natural products are included in these studies, since many extracts and isolated compounds have shown promising results in gastroprotective activities (13,14).

*Fructus Arctii*, the dried fruits of *Arctium lappa* L., are well-known detoxifying agents in traditional Chinese medicine (15). Arctigenin is a lignin constituent of *Fructus Arctii*. Arctigenin displays anti-*H. pylori* (16) and anti-inflammatory activity (17-19), inhibits T lymphocyte proliferation and the gene expression of IL-6 and interferon-γ, and exerts anti-tumor activity against pancuronium-1 cells (20,21). However, there are few studies analyzing anti-ulcer activity using arctigenin. Therefore, the present study aimed to evaluate the potential effects of arctigenin on the prevention (by the absolute ethanol model of ulcer induction) and the treatment of experimental
gastrointestinal ulcers (by the acetic acid model of ulcer induction). In addition, malondialdehyde (MDA) and superoxide dismutase (SOD) levels in serum and expression levels of cytokines, such as tumor necrosis factor-α (TNF-α), interleukin-6 (IL-6), IL-10 and C-reactive protein (CRP) in the gastric tissues were evaluated.

In the present study, the preventive effect of arctigenin on gastritis ulcers was investigated in a Sprague-Dawley (SD) rat model. The results indicate that arctigenin effectively decreases inflammatory factors in gastric ulcer rats and protects them from gastric ulcer lesions.

Materials and methods

Preparation of arctigenin. The dried seeds of *Arctium lappa* L. were bought from Cangshan (February 2012, Shandong, China). Identification of the plants was confirmed by Dr Li Shouxin (senior engineer; State Key Laboratory of Generic Manufacture Technology of Chinese Traditional Medicine, Linyi, China). A voucher specimen (no. 20110920) was deposited in the State Key Laboratory of Generic Manufacture Technology of Chinese Traditional Medicine. The Fructus Arctii was hydrolyzed by acid hydrolysis and alcohol extraction. The crude product was separated by ethyl acetate extraction, and the arctigenin with >75% purity was collected. A high purity of arctigenin was obtained by ethanol crystallization. The finished product was crystallized with anhydrous ethanol until the purity of arctigenin was >99%.

Animals. One hundred male SD rats (200±20 g; age, 8–10 weeks) were supplied by Lunan Pharmaceutical Group Co., Ltd. (Linyi, China), acclimatized to a controlled temperature (23±2°C) and maintained under a 12-h light/dark cycle. The animals were supplied with pelleted chow and water *ad libitum*. All experiments were in accordance with the experimental protocols previously approved by the Institution Animal Ethics Committee at the Pharmacological Center of Lunan Pharmaceutical Group Co., Ltd., and performed in accordance with the Declaration of Helsinki.

Ethanol-induced gastric ulcer. The model of ethanol-induced ulcers was performed as described previously with minor modifications (22). Rats were randomly divided into five groups (n=8 per group). Animals were administrated with vehicle [20% PEG400 (Shanghai Chemical Co., Shanghai, China) in saline], graded doses of arctigenin (0.05, 0.15 and 0.45 mg/kg; purity, >99%), extracted in our laboratory dissolved in saline containing 20% PEG400 or cimetidine (36 mg/kg; Shandong Fangming Pharmaceutical Group Co., Ltd., Heze, China) for 6 days. Animals were deprived of food for 24 h before experiments. One-hour post administration with 1 ml absolute ethanol, animals were anesthetized with pentobarbital (50 mg/kg; Sigma-Aldrich, St. Louis, MO, USA), the inferior vena cava was punctured and 2-ml blood samples were collected in pro-coagulation tubes to obtain the serum (by centrifugation at 1,024 x g for 10 min at 4°C). The serum was stored at -80°C. The rats were anesthetized with 3% pentobarbital (Sigma-Aldrich) the abdomen was opened, to expose the abdominal aorta, and the rats were sacrificed by bloodletting of the vena cardinalis. The stomachs were removed on ice, opened along the greater curvature, washed with saline (0.9%), and the ulcer index was evaluated according to the number and severity of lesions formed. The scoring was performed according to the following scale (23): 0, no visible ulcers; 1, petechial hemorrhage or minute pin-point ulcers; 2, striate hemorrhage and erosion <2 mm; 3, striate hemorrhage and erosion ≥2 and <3 mm; 4, striate hemorrhage and erosion ≥3 and <4 mm; 5, striate hemorrhage and erosion ≥4 and <5 mm.

The mean ulcer indices in each group were calculated and expressed as the percentage of inhibition using the following formula: Inhibition (%) = (control mean - treated mean/control mean) x 100.

Acetic acid-induced gastric ulcer. The experiment was performed as described previously (24) with minor modifications. Rats had been fasting for 12 h before the experiment and had free access to water. A longitudinal incision below the xiphoid process was made in the anesthetized rats. The anterior wall of the stomach was exposed, then 0.02 ml of 30% acetic acid was injected with a microsyringe in the subserosal layer at the junction of the fundus with the antrum, and later washed with saline. On the following day, the daily treatments were initiated (once a day for 12 days) in six groups: sham, vehicle, cimetidine (36 mg/kg), arctigenin (0.05, 0.15 and 0.45 mg/kg) and the sham group (that underwent the surgical procedure of ulcer induction without the application of acetic acid). Following the final treatment, the rats were anesthetized with 3% pentobarbital and sacrificed as described above. The stomachs were removed. The minimum and maximum diameters of the open ulcer were then measured, and the product was considered to be the ulcer index. The 100-mg gastric tissue samples obtained from the corpus region were stored at -80°C for the subsequent measurement of TNF-α, IL-6, IL-10 and CRP using ELISA.

Determination of MDA and SOD serum levels. The blood samples were centrifuged at 1,024 x g for 10 min at 4°C to obtain the serum. Serum levels of MDA and SOD were determined using a MDA Assay kit (Nanjing JianCheng Bioengineering Institute, Nanjing, China) and Total SOD Assay kit (Nanjing JianCheng Bioengineering Institute) according to the manufacturer's instructions.

Measurement of TNF-α, IL-6, IL-10 and CRP expression levels. Tissue samples were homogenized in 10 volumes of 0.9% saline and centrifuged at 1,024 X g at 4°C for 15 min. The supernatant was collected and the levels of TNF-α (Tumor Necrosis Factor-α Assay kit; Nanjing JianCheng Bioengineering Institute), IL-6 (Interleukin-6 Assay kit; Nanjing JianCheng Bioengineering Institute), IL-10 (Interleukin-10 Assay kit; Nanjing JianCheng Bioengineering Institute) and CRP (C-Reactive Protein Assay kit; Nanjing JianCheng Bioengineering Institute) were determined by ELISA. All procedures were performed according to the manufacturer's instructions.

Statistical analysis. The parameters were recorded for individuals within all of the groups. Data are presented as the mean ± standard error of the mean. Statistical analysis was
performed with a one-way analysis of variance (ANOVA) using SPSS 17.0 (SPSS, Inc., Chicago, IL, USA) and P<0.05 was considered to indicate a statistically significant difference. Continuous variables are expressed as mean±standard deviation and compared with two-tailed probability values from an overall F test from a one-way ANOVA, and pairwise comparisons with Fisher’s test of least significant difference.

Results

Ethanol-induced gastric ulcers. The gastroprotective effects of arctigenin on acetic-induced gastric damage were observed in arctigenin-treated rats (Fig. 1A). Ulcerated rats pretreated with cimetidine or arctigenin demonstrated significantly reduced (P<0.05) ulcer indices when compared with the vehicle group. The cimetidine and arctigenin at doses of 0.05, 0.15, 0.45 mg/kg significantly inhibited the appearance of lesions by 31.60, 64.43, 53.91 and 53.04%, respectively. By determination of the ulcer index, arctigenin resulted in an inverted curve, thus, the optimum gastroprotective effect was obtained at a dose of 0.45 mg/kg; Fig. 1A).

Acetic acid-induced chronic ulcer. Serosal application of acetic acid to the vehicle-treated rats resulted in extensive gastric lesions at 10 days of ulcer induction (17.56±3.56 mm²), where the ulcer erosion index was found to be significantly high when compared with the sham group rats without any lesions (P<0.001). All doses of arctigenin (0.05, 0.15 and 0.45 mg/kg) reduced the ulcer index significantly (P<0.01). However, 36 mg/kg cimetidine reduced the ulcer index significantly (P<0.05). The percentages inhibition was 65.83, 61.73, 56.09 and 42.14% for the groups treated with 0.05, 0.15 and 0.45 mg/kg and cimetidine, respectively. In the present study, arctigenin also presented an inverted curve, so that the best gastroprotective effect was obtained at a dose of 0.45 mg/kg; Fig. 1B).

Determination of the sera levels of MDA and SOD. For the group that received vehicle only, the level of MDA was 7.96±0.680 nmol/l. Pretreatment with arctigenin at doses of 0.05, 0.15, 0.45 mg/kg and cimetidine significantly decreased the levels of MDA (5.670±0.264, 4.962±0.311, 4.979±0.260 and 5.527±0.231 nmol/l, respectively) compared with the vehicle group (P<0.01; Fig. 2A). The SOD level in arctigenin-treated at 0.05 mg/kg (122.39±6.85 U/ml), 0.15 mg/kg (119.33±4.04 U/ml) and 0.45 mg/kg (145.50±6.83 U/ml) significantly increased the SOD level compared with the vehicle group (124.28±6.27 U/ml; P<0.05; Fig. 2B).

Comparison of the levels of cytokines in rat stomach tissue samples. All doses of arctigenin treatment significantly reduced the TNF-α levels (364.55±27.22, 348.43±41.45 and 355.13±60.35 ng/l; P<0.01) when compared with the vehicle group (534.17±49.91 ng/l). However, treatment of cimetidine at the doses of 36 mg/kg did not significantly decrease the TNF-α levels (Fig. 3A).

The IL-10 levels in the vehicle group increased significantly (692.21±27.36 pg/l; P<0.01) compared with the sham group (605.75±57.20 pg/l; Fig. 3B). The rats that received 0.05, 0.15 and 0.45 mg/kg arctigenin demonstrated significantly reduced IL-10 levels (612.49±40.39, 473.02±36.30 and 489.46±36.26 pg/l, respectively; P<0.001) when compared with the vehicle group. The other groups were not significantly altered compared with the vehicle group.

Treatment with arctigenin at doses of 0.05, 0.15 and 0.45 mg/kg significantly decreased the levels of CRP [2.75±0.19 µg/l (P<0.05); 1.79±0.18 µg/l (P<0.01) and 1.38±0.08 µg/l (P<0.01)] compared with the sham group. However, rats that received cimetidine showed no significant reduction in the CRP level when compared with the vehicle group (2.98±0.24 µg/l; P>0.05) (Fig. 3C).

The gastric tissue IL-6 level in the vehicle group (32.97±5.30 ng/l) demonstrated a significant increase when compared with the sham group (20.48±1.05 ng/l; P<0.05). However, cimetidine showed no significant reduction in the IL-6 level when compared with the vehicle group. In addition, 0.05, 0.15 and 0.45 mg/kg arctigenin exhibited strong inhibition effects on IL-6 (23.77±2.88, 21.20±2.94 and 21.04±2.94 ng/l) compared with vehicle group (Fig. 3D).

Discussion

To the best of our knowledge, the present study is the first to report the gastroprotective of arctigenin from Fructus Arctii, which inhibits the ulcer index induced by absolute ethanol and acetic acid (Fig. 1), and provides experimental evidence to support the traditional use of Fructus Arctii in the treatment of stomach dysfunction.
Ethanol and acetic acid are the most commonly utilized experimental models for the evaluation of anti-ulcer activity in rats (24,25). The gastroprotective effect of arctigenin in different experimental ulcer models is presented in the current study, as various mechanisms are involved in the etiology of gastric ulcers. Ethanol-induced gastric ulcers are a reliable model of acute gastric mucosal ulceration; as shown in Fig. 1A, arctigenin treatment inhibited the gastric damage induced by ethanol at all doses with similar efficacy as compared with the vehicle group.

In addition to the gastroprotective effect of arctigenin, the healing effect on the gastric mucosa in the acetic acid-induced ulcer model was investigated. This chronic ulcer model, which highly resembles human chronic ulcers in terms of pathological features and healing mechanisms, is utilized to develop novel anti-ulcer medication, which may potentially prevent ulcer relapse or enhance ulcer healing (26). Acetic acid is reported to produce ulcers by gastric obstruction leading to increased quantities of acidic gastric juice (27,28). In the current study, arctigenin caused significant reductions in gastric lesions at all doses tested, when compared with the vehicle group (Fig. 1B), indicating that arctigenin exerted curative effects. This result suggested that arctigenin may have protective effects in the treatment of chronic ulcers.

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The effects of ethanol on the gastric mucosa may be associated with the formation of reactive oxygen species (ROS), which cause an imbalance between oxidant and antioxidant cellular processes. It is known that lipid peroxidation is the result of ROS interacting with the cell membrane, subsequently producing highly reactive lipid-derived free radicals, such as MDA, to cause oxidative gastric damage (29). However, organisms possess enzymatic defenses (SOD) against the lipid peroxidation. In the current study, arctigenin lowered the elevated MDA level and increased the SOD level (Fig. 2). Thus,
it was concluded that the ability of arctigenin to reduce the MDA level and increase the SOD level may contribute to its gastroprotective effect by inhibiting oxidative gastric damage. Inflammation is generally considered to be a highly regulated defensive process characterized by the release of cytokines from the blood to the affected tissue (30). The potent pro-inflammatory cytokines (such as TNF, IL-1, interferon-γ) are involved in the occurrence and development of inflammatory responses (31,32). The acetic acid, a well-known ulcerogenic agent, stimulates inflammation via the imbalance of pro- and anti-inflammatory cytokines (33). In the current study, acetic acid-induced ulcer models were used to investigate the gastroprotective effect of arctigenin. Arctigenin effectively regulated four pro-inflammatory cytokines and attenuated acetic acid-induced ulcers. Thus, in the present study TNF-α, IL-6, IL-10 and CRP were evaluated in gastric homogenate (Fig. 3). TNF-α, which is increased in non-steroidal anti-inflammatory drug-treated rats, appears to be a key contributor on many forms of gastric mucosal damage (34,35). IL-10 is a pleiotropic cytokine with important immunoregulatory functions whose actions influence activities of many cell-types in the immune system. The levels of IL-10 are prominently increased in peptic ulcer patients (36). Numerous clinical studies reported that the levels of inflammatory factors (IL-6, CRP and TNF-α) were consistent with the pathogenic condition of gastric ulcer patients (37). It is reported that an elevated CRP level is a positive predictive value for peptic ulcers caused by active inflammation (38). These results corroborated the report of Hyam et al (39), who observed that arctigenin inhibited 2,4,6-trinitrobenzenesulfonic acid-induced IL-6 and TNF-α expression. These findings reveal that the anti-ulcerogenic effects of arctigenin may be mediated by anti-inflammatory activities.

The results of the present study indicate that arctigenin afforded remarkable ameliorative effects against gastric mucosal lesions induced by ethanol and acetic acid, thereby confirming anti-ulcer activity in SD rats. The gastroprotective effect of arctigenin may be attributed to its activities against oxidative and inflammatory damage. In conclusion, the results obtained in the present study provide support for the use of arctigenin as a gastroprotective medicine. Furthermore, the current study provides more information on the possible underlying mechanisms by which arctigenin exerts its anti-ulcer effects.

Acknowledgements

The present study was supported by the National Science and Technology Support Program (grant no. 2012CB724001) and the Shandong province Science and Technology Major Project (grant no. 2015ZDJQ05004), gratefully received from The Generic Manufacture Technology of Chinese Traditional Medicine and Lunan Pharmaceutical Group Co., Ltd.

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