

Cilostazol regulates the expressions of endothelin-1 and endothelial nitric oxide synthase via activation of the p38 MAPK signaling pathway in HUVECs

YING XUE^{1,2}, XIAOYE LI¹, ZI WANG¹ and QIANZHOU LV¹

¹Department of Clinical Pharmacy, Zhongshan Hospital, Fudan University, Shanghai 200032, P.R. China;

²Department of Pharmacy and Therapeutics, School of Pharmacy, University of Pittsburgh, Pittsburgh, PA 15213, USA

Received March 9, 2022; Accepted May 12, 2022

DOI: 10.3892/br.2022.1560

Abstract. Cilostazol is a selective inhibitor of phosphodiesterase type III that inhibits platelet aggregation. The beneficial effects of cilostazol have been attributed not only to its anti-platelet functions but also to its actions on the endothelium. Whether cilostazol regulates endothelin-1 (ET-1) and endothelial nitric oxide synthase (eNOS) through mitogen-activated protein kinase (MAPK) remains undetermined. The aim of this study was to investigate the effects of cilostazol on ET-1 and eNOS expression in HUVECs, and to assess its relationship with MAPK activity. HUVECs were cultured *in vitro*, stimulated with TNF- α , and pretreated with different concentrations of cilostazol. ET-1 and eNOS levels in the supernatant were detected by ELISA. RT-qPCR was performed to detect the mRNA expression levels of ET-1 and eNOS. The phosphorylation levels of p38/MAPK and protein expression levels of ET-1 and eNOS were assessed using western blotting. A P38 inhibitor, SB203580, was utilized to further validate the involvement of p38/MAPK in the regulation. Expression of ET-1, which was upregulated by TNF- α , was significantly suppressed by cilostazol in a dose-dependent manner, meanwhile, with as the cilostazol concentration was increased, the expression of eNOS increased as well. In addition, cilostazol also decreased phosphorylation of p38, which was upregulated by TNF- α . The observed upregulation of eNOS and ET-1 levels were completely abolished upon p38 inhibitor treatment, indicating the involvement of the p38/MAPK pathway in cilostazol-induced regulation of eNOS and ET-1 in HUVECs. The results indicated that cilostazol regulates ET-1 and eNOS production by suppressing the p38/MAPK signaling pathway

in TNF- α -stimulated HUVECs, and this may contribute to the protective effects of cilostazol on the endothelium.

Introduction

The vascular endothelium is an active monolayer of cells lining the entire circulatory system, and it is indispensable for the regulation of vascular tone and in the maintenance of vascular homeostasis (1). Endothelial dysfunction is regarded as a key early event in the development of atherosclerosis and the occurrence of atherosclerotic complications (2,3). Notably, endothelial dysfunction is an established predictor of cardiovascular outcomes (4).

The endothelium regulates the remodeling of the vessel wall by releasing a large number of vasoactive substances. Nitric oxide (NO) is an endothelium-derived relaxing factor that is produced by endothelial NO synthase (eNOS) (5). It protects the vascular system by acting as an endogenous defense against the development of atherosclerosis. The reduction in the generation and bioavailability of NO is accepted as an important factor in the process of atherosclerosis and thrombosis (6,7). Endothelin (ET)-1 is a potent vasoconstrictor produced by endothelial cells that plays a pivotal role in the etiology of atherosclerotic vascular disease (8). It induces endothelial dysfunction and contributes to atherosclerosis by stimulating platelet aggregation, cell adhesion molecule expression and the proliferation of vascular smooth muscle cells (9,10). A pathophysiological imbalance in NO and ET-1 is usually referred as endothelial dysfunction (11).

Cilostazol is a selective inhibitor of phosphodiesterase type III (PDE3) that inhibits platelet aggregation (12). It is widely used in patients after receiving percutaneous coronary intervention (PCI) to reduce the risk of thrombotic events (13,14). The beneficial effects of cilostazol have been attributed not only to its antiplatelet functions but also to its actions on the endothelium. Recent studies suggested that cilostazol could protect endothelial function by inducing NO production (15-17) and decreasing ET-1 production (18,19).

As an important member of the MAPK pathway, p38 MAPK plays an important role in inflammation and stress response. Animal studies have confirmed that p38/MAPK is closely related to the expression and secretion of ET-1 (20,21).

Correspondence to: Dr Ying Xue or Dr Qianzhou Lv, Department of Clinical Pharmacy, Zhongshan Hospital, Fudan University, 180 Fenglin Road, Shanghai 200032, P.R. China
E-mail: Xueyl4@fudan.edu.cn
E-mail: Lv.qianzhou@zs-hospital.sh.cn

Key words: cilostazol, endothelial dysfunction, endothelin-1, endothelial nitric oxide synthase, p38 MAPK

It has also been reported that p38/MAPK can regulate the expression of eNOS (22). A number of studies have shown that p38/MAPK is one of the targets of cilostazol, which may explain its endothelial protective effect (23,24). A study published by Chao *et al.* (25) suggested that cilostazol could regulate NO production via the p38/MAPK signaling pathway, however, it did not assess the effects of cilostazol on ET-1. Thus, whether cilostazol regulates NO and ET-1 via the p38/MAPK signaling pathway remains to be determined.

Taking the above evidence together, the objective of this study was to examine the effects of cilostazol on the regulation of ET-1 and eNOS, as well as its relationship with p38/MAPK. HUVECs were used in this study as they have proven invaluable in investigating endothelial damage and repair, and on the potential impact of atherosclerosis during the early stages and during atherosclerosis progression (26). Specifically, HUVECs were exposed to TNF- α and cilostazol with or without specific inhibitors of p38/MAPK, and the changes in p38/MAPK activity, and in the expression levels of ET-1 and eNOS were determined.

Materials and methods

Materials. HUVECs (cat. no. 8000) and endothelial cell medium (ECM, cat. no. #1001) were purchased from ScienCell Research Laboratories, Inc. Cilostazol (cat. no. PHR1503) and the p38 inhibitor SB203580 were obtained from Sigma-Aldrich; Merck KGaA. Antibodies were obtained from ProteinTech Group, Inc., Cell Signaling Technology, Inc., or Zhejiang Kangchen Biotech, Co., Ltd.; TRIzol[®] reagent from Invitrogen; Thermo Fisher Scientific, Inc.; and TNF- α from Sigma-Aldrich; Merck KGaA. PCR primers (Integrated DNA Technologies, Inc.); and the GoTaq[®] qPCR MasterMix kit was from Promega Corporation. The study proposal was approved by Zhongshan hospital, Fudan University.

Cell culture and treatments. HUVECs were cultured at 37°C in a humidified 5% CO₂ incubator and split in accordance with standard procedures. When they grew 60–85% confluent, HUVECs were treated with different concentrations (0, 1, 10 or 50 μ M) cilostazol (25) or TNF- α (10 μ g/l) for 24 h and then collected for further analysis.

SB203580 (10 μ M, for 2 h) (27) and TNF- α (for 24 h) (28) were added to cells prior to incubation with cilostazol. Control HUVECs were pre-treated with goat serum.

ELISA. ET-1 and eNOS levels in the cell culture supernatant were quantified by spectrophotometry using ELISA kits according to the manufacturer's protocol (cat. no. H093 and cat. no. H195, respectively; both from Nanjing Jiancheng Bioengineering Institute).

Reverse transcription-quantitative (RT-q)PCR. Total mRNA was extracted from HUVECs using TRIzol[®] reagent. Reverse transcription was performed using a Prime Script RT reagent kit (Takara Bio, Inc.) according to the manufacturer's protocols. The mRNA levels of ET-1 and eNOS were measured by qPCR using an iCycler real-time system (Bio-Rad Laboratories, Inc.). The thermocycling conditions were as follows: An initial pre-denaturation step for 10 sec at 95°C; followed by 45 cycles of denaturation at 95°C for 10 sec, annealing at 60°C for 20 sec

and extension at 70°C for 30 sec; with a final extension step at 70°C for 5 min.

The sequences of the primers used for amplification were: ET-1 forward, 5'-AAGGCAACAGACCGTGAA AAT-3' and reverse, 5'-CGACCTGGTTTGTCTTAGGTG-3'; eNOS forward, 5'-TGATGGCGAAGCGAGTGAAG-3' and reverse, 5'-ACTCATCCATACACAGGACCC-3'; and GAPDH forward, 5'-AGAAGGCTGGGGCTCATTG-3' and reverse, 5'-AGGGGCCATCCACAGTCTTC-3'.

Western blotting. HUVECs were washed three times with PBS and lysed with ice-cold RIPA buffer (Beyotime Institute of Biotechnology) supplemented with protease inhibitor cocktail (Thermo Fisher Scientific, Inc.). A BCA protein assay kit (Thermo Fisher Scientific, Inc.) was used to determine the protein concentrations in the lysates. Equivalent amounts (30 μ g) of total protein were resolved by SDS-PAGE (10% tris-glycine gels) and immunoblotted with primary antibodies at 4°C overnight (29) [anti-ET-1 (1:1,000, cat. no. 67008-1-Ig, ProteinTech Group, Inc.), anti-eNOS (dilution 1:1,000, cat. no. 27120-1AP, ProteinTech Group, Inc.), anti-p38/p-p38 (1:1,000, cat. nos. 4511 and 8690, respectively, Cell Signaling Technology, Inc.) and anti-GAPDH (1:5,000, cat. no. 5174, Cell Signaling Technology, Inc.)]. After washing three times, the horseradish peroxidase-conjugated secondary antibody (1:5,000, cat. no. KC-RB-035; Zhejiang Kangchen Biotech Co., Ltd.) was added and incubated for 1 h at room temperature.

Statistical analysis. All experiments were performed at least three times. The representative results and corresponding quantification data of one repeat is shown for each experiment. Data are presented as the mean \pm SEM and were compared using a Student's t-test or a one-way ANOVA with a post-hoc Tukey's test using GraphPad Prism version 5.01 (GraphPad software, Inc.). $P < 0.05$ was considered to indicate a statistically significant difference.

Results

Effect of cilostazol on the levels of ET-1 and eNOS in HUVECs. Compared with the untreated cells, TNF- α markedly increased the levels of ET-1, and decreased the levels of eNOS in the HUVECs culture supernatant. Cilostazol (1, 10 and 50 μ M) administration significantly decreased the levels of ET-1, and increased the levels of eNOS in HUVECs compared with the TNF- α -induced cells (Fig. 1).

Effect of cilostazol on the mRNA expression levels of ET-1 and eNOS in HUVECs. To determine whether ET-1 and eNOS expression in HUVECs was affected by cilostazol at the mRNA level, the HUVECs were incubated overnight with TNF- α and then treated with different concentrations of cilostazol (1, 10 or 50 μ M). Total RNA was isolated from HUVECs and subjected to RT-qPCR analysis. As shown in Fig. 2, HUVECs exposed to cilostazol exhibited upregulated eNOS mRNA levels and downregulated ET-1 mRNA levels in a dose-dependent manner ($P < 0.05$).

HUVECs were pre-treated with a p38 inhibitor (SB203580-10 μ M) for 2 h prior to incubation with cilostazol.

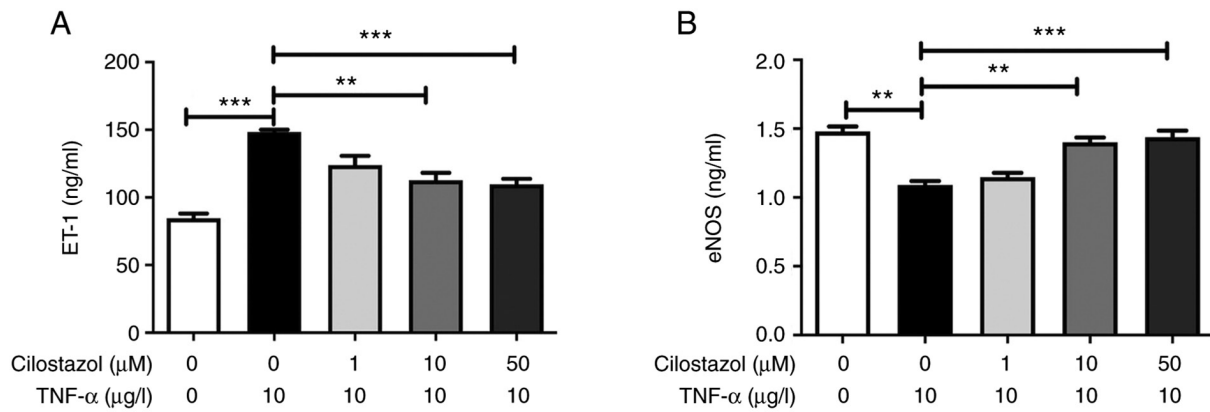


Figure 1. Effect of cilostazol on ET-1 and eNOS expression in HUVECs. Cells were pre-treated with different concentrations of cilostazol (0, 1, 10 or 50 μM) or TNF-α. (A) ET-1 and (B) eNOS levels were measured by ELISA. The results are presented as the mean ± SEM of three repeats. **P<0.01, ***P<0.001. ET-1, endothelin-1; eNOS, endothelial nitric oxide synthase.

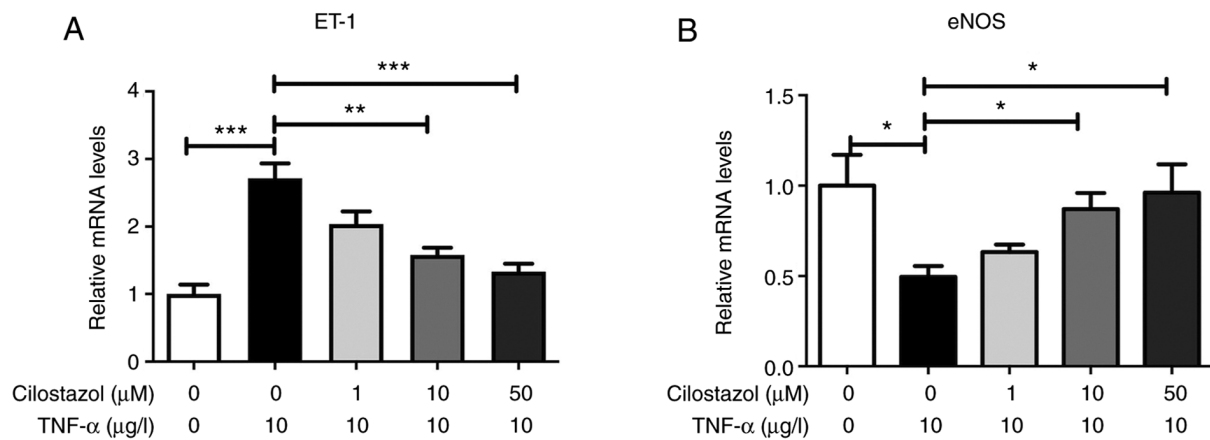


Figure 2. Effect of cilostazol on the mRNA levels of ET-1 and eNOS in HUVECs following treatment with cilostazol. (A) ET-1 mRNA levels were measured by PCR. (B) eNOS mRNA levels were measured by PCR. The results are presented as the mean ± SEM of three repeats. *P<0.05, **P<0.01, ***P<0.001. ET-1, endothelin-1; eNOS, endothelial nitric oxide synthase.

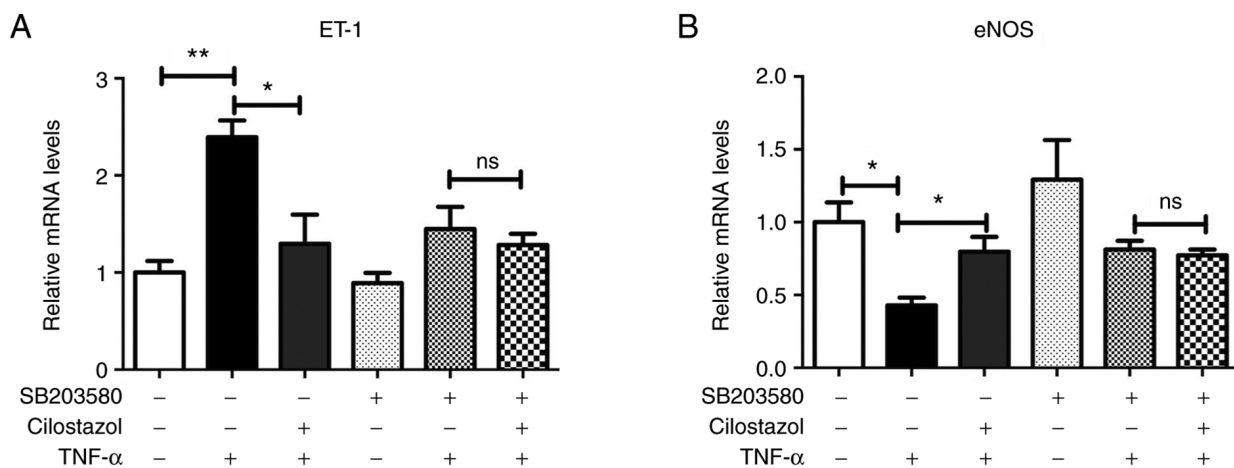


Figure 3. ET-1 and eNOS mRNA levels following treatment with cilostazol. (A) ET-1 mRNA levels following pre-incubation with/without the p38 inhibitor (SB203580). (B) mRNA levels following pre-incubation with/without p38 inhibitor (SB203580). *P<0.05, **P<0.01. ET-1, endothelin-1; eNOS, endothelial nitric oxide synthase; ns, not significant.

The increase in ET-1 mRNA expression levels were fully abolished in the presence of this p38 inhibitor indicating the involvement of this kinase in cilostazol-induced regulation

of ET-1 in HUVECs (P<0.05; Fig. 3A). Additionally, the effects of cilostazol on eNOS mRNA were also abolished by SB203580 (Fig. 3B).

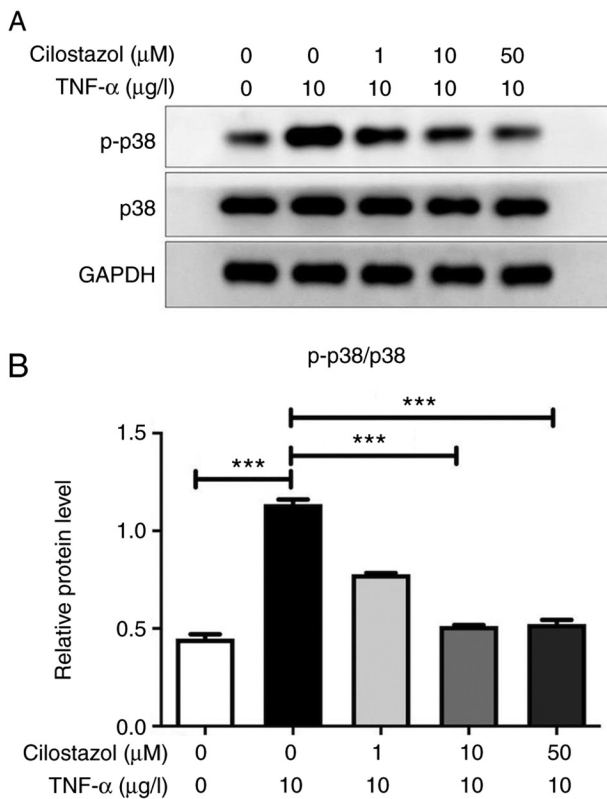


Figure 4. Effect of cilostazol administration on p-p38 levels. (A) Western blotting and the corresponding densitometry analysis for p-p38 expression in HUVECs after treatment with different concentrations of cilostazol. (B) Quantitative analysis of the western blotting results. *** $P < 0.001$.

Effect of cilostazol on p-p38 MAPK expression in HUVECs. Lysates from HUVECs treated with different cilostazol concentrations were analyzed by western blotting utilizing specific antibodies against the activated (i.e. phosphorylated) form of p38. Compared with the untreated cells, TNF- α markedly increased the p-p38 MAPK levels in HUVECs. Treatment of HUVECs with several different concentrations of cilostazol resulted in a decrease in p-p38 levels in a dose-dependent manner (Fig. 4).

Effect of cilostazol on the protein expressions levels of ET-1 and eNOS in HUVECs. To further validate the above findings, lysates from HUVECs exposed to p38 inhibitor (SB203580) and cilostazol were analyzed by western blotting. Compared with the untreated cells, TNF- α markedly increased the levels of ET-1, and decreased the levels of eNOS in HUVECs. Cilostazol administration significantly decreased the protein expression levels of ET-1, and increased the protein expression levels of eNOS in HUVECs. These effects of cilostazol on ET-1 and eNOS were also abolished by SB203580 treatment (Fig. 5).

Discussion

Endothelial dysfunction increases the vulnerability of plaque formation, triggers plaque rupture, and promotes thrombosis. It is considered a key precursor in the initiation, progression, and complication of coronary atherosclerotic heart disease (30,31). Given its role in the development of coronary artery and cerebrovascular diseases, endothelial dysfunction may be

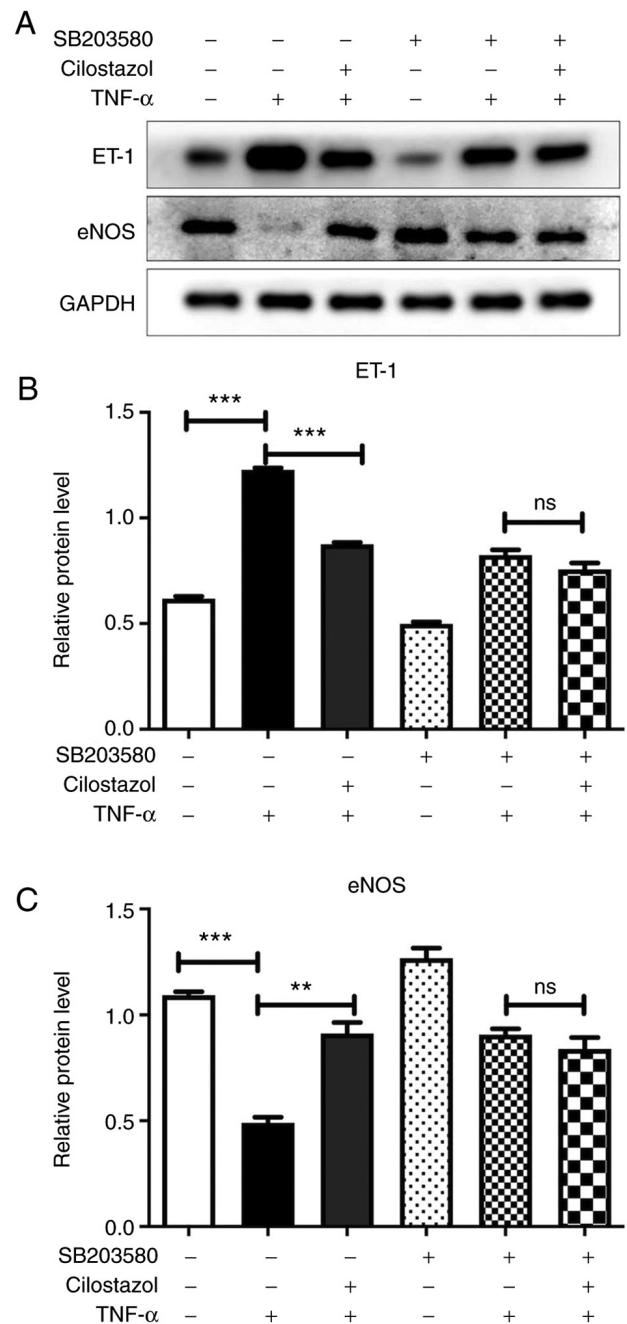


Figure 5. Effect of cilostazol on ET-1 and eNOS expression in HUVECs after pre-treatment with the p-p38 inhibitor (SB203580). (A) Representative western blots of ET-1 and eNOS. Quantitative analysis of the western blotting results of (B) ET-1 and (C) eNOS. The results are presented as the mean \pm SEM of three repeats. ** $P < 0.01$, *** $P < 0.001$. ET-1, endothelin-1; eNOS, endothelial nitric oxide synthase; ns, not significant.

an attractive target in the efforts to optimize individualized therapeutic strategies to reduce cardiovascular morbidity and mortality rates (32).

Endothelial cells can release mediators and transcription factors to regulate important functions such as the tension of blood vessels and the growth of smooth muscle cells. Among those endothelial active factors, the relaxation factor NO and the contraction factor ET-1 are two leading active substances (5,33). Endothelial dysfunction is in part characterized by enhanced ET-1 and diminished eNOS expression (34). The balance between ET-1 and eNOS expression maintains

the normal function of endothelial cells in terms of cellular integrity.

Cilostazol is a novel type of antiplatelet drug, which inhibits platelets by selectively inhibiting PDE3, and functions to expand blood vessels, inhibiting intimal hyperplasia and protecting the endothelium. Unlike other antiplatelet agents, cilostazol not only exhibits antiplatelet properties, but also appears to have beneficial effects on endothelial function (34-37). A number of studies suggested that cilostazol can induce NO production (15-17). Furthermore, it has been reported that cilostazol can decrease ET-1 production (18,19).

The inflammatory cytokine TNF- α plays a pivotal role in the disruption of macrovascular and microvascular circulation. It is suggested that TNF- α can induce the synthesis and release of ET-1 (38). It also accelerates the degradation of eNOS mRNA, decreases the activity of the gene promoter and down-regulates the expression of eNOS (39). In this study, TNF- α and HUVECs were used to study the effects of cilostazol on the expression of ET-1 and eNOS. The results showed that cilostazol reduced the expression of ET-1 and increased the expression of eNOS in a dose-dependent manner. HUVECs exposed to TNF- α exhibited increased ET-1 levels and ET-1 mRNA expression. This suggested that TNF- α -mediated increases in ET-1 expression may be achieved through regulation of ET-1 gene transcription. After incubating with cilostazol, the expression of ET-1 was significantly reduced. Similarly, cilostazol upregulated the mRNA and protein expression levels of eNOS.

The MAPKs are a family of kinases that transduce signals from the cell membrane to the nucleus in response to a wide range of stimuli, including stress and injury. The MAPK pathway regulates cell proliferation, mitosis, transformation, apoptosis and other biological activities by affecting gene transcription and regulation. p38/MAPK is an important member of the MAPK family. There is a close relationship between the p38/MAPK signaling pathway and vascular remodeling. Recent studies have shown that the p38/MAPK signaling pathway is closely related to the regulation of ET-1 and eNOS expression (20,22). It has also been suggested that p38/MAPK is one of the targets of cilostazol, which may explain the beneficial effects of cilostazol in slowing down the progression of atherosclerosis (23,24).

To explore the effects of cilostazol on the p38 signaling pathway in HUVECs, the intracellular signaling pathways involved in eNOS and ET-1 expression were examined. HUVECs were treated with the p38 inhibitor SB203580 prior to cilostazol treatment. The observed upregulation of eNOS and ET-1 mRNA levels was completely abolished upon p38 inhibitor treatment, indicating the involvement of p38/MAPK and the associated signaling cascades in cilostazol-induced regulation of eNOS and ET-1 in HUVECs. Western blotting analysis of lysates from HUVECs exposed to cilostazol further validated this finding. These results suggested that the regulation of NO and ET-1 in HUVECs by cilostazol may be mediated through the p38MAPK signaling pathway.

In summary, we found that cilostazol regulates ET-1 and eNOS production by suppressing the p38/MAPK signal pathway in TNF- α -stimulated HUVECs, and this may contribute to the protective effect of cilostazol on the

endothelium. Further experiments are required to confirm these results.

Acknowledgements

Not applicable.

Funding

This study was supported by the Natural Science Foundation of China (grant no. 81803630).

Availability of data and materials

The datasets used and/or analyzed during the present study are available from the corresponding author on reasonable request.

Authors' contributions

YX and QL designed the study. YX, ZW and XL performed the experiments. YX analyzed the data and wrote the manuscript. All authors have read and approved the final manuscript. YX, XL and ZW confirmed the authenticity of all the raw data.

Ethics approval and consent to participate

Not applicable.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

References

1. Reriani MK, Flammer AJ, Jama A, Lerman LO and Lerman A: Novel functional risk factors for the prediction of cardiovascular events in vulnerable patients following acute coronary syndrome. *Circ J* 76: 778-783, 2012.
2. el-Tamimi H, Mansour M, Wargovich TJ, Hill JA, Kerensky RA, Conti CR and Pepine CJ: Constrictor and dilator responses to intracoronary acetylcholine in adjacent segments of the same coronary artery in patients with coronary artery disease. Endothelial function revisited. *Circulation* 89: 45-51, 1994.
3. Huang PH, Leu HB, Chen JW, Wu TC, Lu TM, Yu-An Ding P and Lin SJ: Decreased heparin cofactor II activity is associated with impaired endothelial function determined by brachial ultrasonography and predicts cardiovascular events. *Int J Cardiol* 114: 152-158, 2007.
4. Martin BJ and Anderson TJ: Risk prediction in cardiovascular disease: The prognostic significance of endothelial dysfunction. *Can J Cardiol* 25 (Suppl A): 15A-20A, 2009.
5. Li H, Horke S and Förstermann U: Vascular oxidative stress, nitric oxide and atherosclerosis. *Atherosclerosis* 237: 208-219, 2014.
6. Tousoulis D, Kampoli AM, Tentolouris C, Papageorgiou N and Stefanadis C: The role of nitric oxide on endothelial function. *Curr Vasc Pharmacol* 10: 4-18, 2012.
7. Sogo N, Magid KS, Shaw CA, Webb DJ and Megson IL: Inhibition of human platelet aggregation by nitric oxide donor drugs: relative contribution of cGMP-independent mechanisms. *Biochem Biophys Res Commun* 279: 412-419, 2000.
8. Miyauchi T and Masaki T: Pathophysiology of endothelin in the cardiovascular system. *Annu Rev Physiol* 61: 391-415, 1999.

9. Lüscher TF and Barton M: Endothelins and endothelin receptor antagonists: Therapeutic considerations for a novel class of cardiovascular drugs. *Circulation* 102: 2434-2440, 2000.
10. Ross R: Atherosclerosis-an inflammatory disease. *N Engl J Med* 340: 115-126, 1999.
11. Rajendran P, Rengarajan T, Thangavel J, Nishigaki Y, Sakthisekaran D, Sethi G and Nishigaki I: The vascular endothelium and human diseases. *Int J Biol Sci* 9: 1057-1069, 2013.
12. Cone J, Wang S, Tandon N, Fong M, Sun B, Sakurai K, Yoshitake M, Kambayashi J and Liu Y: Comparison of the effects of cilostazol and milrinone on intracellular cAMP levels and cellular function in platelets and cardiac cells. *J Cardiovasc Pharmacol* 34: 497-504, 1999.
13. Guerra E, Byrne RA and Kastrati A: Pharmacological inhibition of coronary restenosis: Systemic and local approaches. *Expert Opin Pharmacother* 15: 2155-2171, 2014.
14. Jang JS, Jin HY, Seo JS, Yang TH, Kim DK, Kim DS, Kim DK, Seol SH, Kim DI, Cho KI, *et al*: A meta-analysis of randomized controlled trials appraising the efficacy and safety of cilostazol after coronary artery stent implantation. *Cardiology* 122: 133-143, 2012.
15. Hashimoto A, Miyakoda G, Hirose Y and Mori T: Activation of endothelial nitric oxide synthase by cilostazol via a cAMP/protein kinase A- and phosphatidylinositol 3-kinase/Akt-dependent mechanism. *Atherosclerosis* 189: 350-357, 2006.
16. Ito H, Hashimoto A, Matsumoto Y, Yao H and Miyakoda G: Cilostazol, a phosphodiesterase inhibitor, attenuates photothrombotic focal ischemic brain injury in hypertensive rats. *J Cereb Blood Flow Metab* 30: 343-351, 2010.
17. Bai Y, Muqier, Murakami H, Iwasa M, Sumi S, Yamada Y, Ushikoshi H, Aoyama T, Nishigaki K, Takemura G, *et al*: Cilostazol protects the heart against ischaemia reperfusion injury in a rabbit model of myocardial infarction: focus on adenosine, nitric oxide and mitochondrial ATP-sensitive potassium channels. *Clin Exp Pharmacol Physiol* 38: 658-665, 2011.
18. Pelletier S, Dubé J, Villeneuve A, Gobeil F Jr, Bernier SG, Battistini B, Guillemette G and Sirois P: Adenosine induces cyclic-AMP formation and inhibits endothelin-1 production/secretion in guinea-pig tracheal epithelial cells through A(2B) adenosine receptors. *Br J Pharmacol* 129: 243-250, 2000.
19. Shima A, Maki T, Mimura N, Yamashita H, Emoto N, Yoshifuji H and Takahashi R: A case of reversible cerebral vasoconstriction syndrome associated with anti-phospholipid antibody syndrome and systemic lupus erythematosus. *eNeurologicalSci* 24: 100351, 2021.
20. Armstead WM, Bohman LE, Riley J, Yarovo S, Higazi AA and Cines DB: tPA-S(481)A prevents impairment of cerebrovascular autoregulation by endogenous tPA after traumatic brain injury by upregulating p38 MAPK and inhibiting ET-1. *J Neurotrauma* 30: 1898-1907, 2013.
21. Jiang Y, Zeng Y, Huang X, Qin Y, Luo W, Xiang S, Sooranna SR and Pinhu L: Nur77 attenuates endothelin-1 expression via downregulation of NF- κ B and p38 MAPK in A549 cells and in an ARDS rat model. *Am J Physiol Lung Cell Mol Physiol* 311: L1023-L1035, 2016.
22. Solone X, Wells B and Chrestensen C: MAP kinases mediate regulation of eNOS through phosphorylation of different sites. *Biochem Mol Biol* 33(S1): 478.10, 2019.
23. Lee KM, Lee HJ, Kim MK, Kim HS, Jung GS, Hur SH, Kim HT, Cho WH, Kim JG, Kim BW, *et al*: Cilostazol inhibits high glucose- and angiotensin II-induced type 1 plasminogen activator inhibitor expression in artery wall and neointimal region after vascular injury. *Atherosclerosis* 207: 391-398, 2009.
24. Lim JH, Woo JS and Shin YW: Cilostazol protects endothelial cells against lipopolysaccharide-induced apoptosis through ERK1/2- and P38 MAPK-dependent pathways. *Korean J Intern Med* 24: 113-122, 2009.
25. Chao TH, Tseng SY, Li YH, Liu PY, Cho CL, Shi GY, Wu HL and Chen JH: A novel vasculo-angiogenic effect of cilostazol mediated by cross-talk between multiple signalling pathways including the ERK/p38 MAPK signalling transduction cascade. *Clin Sci (Lond)* 123: 147-159, 2012.
26. Medina-Leyte DJ, Domínguez-Pérez M, Mercado I, Villarreal-Molina MT and Jacobo-Albavera L: Use of human umbilical vein endothelial cells (HUVEC) as a model to study cardiovascular disease: A review. *Appl Sci* 10, 938, 2020.
27. Xiong T, Zhang Z, Zheng R, Huang J and Guo L: N-acetyl cysteine inhibits lipopolysaccharide-induced apoptosis of human umbilical vein endothelial cells via the p38MAPK signaling pathway. *Mol Med Rep* 20: 2945-2953, 2019.
28. Cho HY, Park CM, Kim MJ, Chinzorig R, Cho CW and Song YS: Comparative effect of genistein and daidzein on the expression of MCP-1, eNOS, and cell adhesion molecules in TNF- α -stimulated HUVECs. *Nutr Res Pract* 5: 381-388, 2011.
29. Kong LJ, Liu XQ, Xue Y, Gao W and Lv QZ: Muramyl dipeptide induces reactive oxygen species generation through the NOD2/COX-2/NOX4 signaling pathway in human umbilical vein endothelial cells. *J Cardiovasc Pharmacol* 71: 352-358, 2018.
30. Halcox JP, Schenke WH, Zalos G, Mincemoyer R, Prasad A, Waclawiw MA, Nour KR and Quyyumi AA: Prognostic value of coronary vascular endothelial dysfunction. *Circulation* 106: 653-658, 2002.
31. Widlansky ME, Gokce N, Keaney JF Jr and Vita JA: The clinical implications of endothelial dysfunction. *J Am Coll Cardiol* 42: 1149-1160, 2003.
32. Bonetti PO, Lerman LO and Lerman A: Endothelial dysfunction: A marker of atherosclerotic risk. *Arterioscler Thromb Vasc Biol* 23: 168-175, 2003.
33. Chua BH, Chua CC, Diglio CA and Siu BB: Regulation of endothelin-1 mRNA by angiotensin II in rat heart endothelial cells. *Biochim Biophys Acta* 1178: 201-206, 1993.
34. Madden JA: Role of the vascular endothelium and plaque in acute ischemic stroke. *Neurology* 79 (Suppl 1): S58-S62, 2012.
35. Goto S: Cilostazol: Potential mechanism of action for antithrombotic effects accompanied by a low rate of bleeding. *Atheroscler (Suppl 6)*: 3-11, 2005.
36. Kim KY, Shin HK, Choi JM and Hong KW: Inhibition of lipopolysaccharide-induced apoptosis by cilostazol in human umbilical vein endothelial cells. *J Pharmacol Exp Ther* 300: 709-715, 2002.
37. Kawanabe Y, Takahashi M, Jin X, Abdul-Majeed S, Nauli AM, Sari Y and Nauli SM: Cilostazol prevents endothelin-induced smooth muscle constriction and proliferation. *PLoS One* 7: e44476, 2012.
38. Hohlfeld T, Klemm P, Thiernemann C, Warner TD, Schrör K and Vane JR: The contribution of tumour necrosis factor- α and endothelin-1 to the increase of coronary resistance in hearts from rats treated with endotoxin. *Br J Pharmacol* 116: 3309-3315, 1995.
39. Zhang H, Park Y, Wu J, Chen Xp, Lee S, Yang J, Dellsperger KC and Zhang C: Role of TNF- α in vascular dysfunction. *Clin Sci (Lond)* 116: 219-230, 2009.