

Role of hyaluronic acid in periodontal therapy (Review)

ASHOK BHATI¹, HYTHAM FAGEEH¹, WAEEL IBRAHEEM¹, HAMMAM FAGEEH¹,
HARNEET CHOPRA² and SUMAN PANDA¹

¹Department of Preventive Dental Sciences, College of Dentistry, Jazan University, Jazan 45142, Saudi Arabia;

²Department of Adult Restorative Dentistry, Oman Dental College, Muscat 116, Oman

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Abstract. Hyaluronic acid (HA) is essential for the function of extracellular matrices in both hard and soft periodontal components. HA plays an important role in the mechanisms underlying inflammation and wound healing. HA is located in periodontal tissues in differing amounts, including non-mineralized tissues, such as gingiva and periodontal ligament, and lower levels located in mineralized tissues, such as cementum and alveolar bone. According to preliminary findings, HA exhibits potential in the regulation of periodontal tissue regeneration and in the treatment of periodontal disease. HA promotes symptomatic relief in both marginal gingiva and deeper periodontal tissues. The present review aimed to examine the role of HA in periodontal therapy, and investigate the current literature supporting its use in periodontal regeneration.

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1. Introduction

Hyaluronic acid (HA), also referred to as hyaluronan, is one of the most predominant glycosaminoglycans in the extracellular matrix (1). In 1934, Karl Meyer and John Palmer extracted HA from the vitreous fluid of the eye of a cow (2). 'HA', as it originated from hyalos (meaning glass in Greek), possessed two

sugar molecules, one of them being uronic acid. Moreover, this linear polysaccharide exhibits numerous repeating disaccharide units (3). Notably, a healthy adult (weight, 70 kg) possesses ~15 g HA in the body (4). The majority of cells in the human body synthesize HA; thus, it may play a role in a variety of key biological processes. This evidence demonstrates the potential of HA as a therapeutic option (5).

At present, HA is used for the treatment of chronic inflammatory diseases. Notably, periodontal disease is an inflammatory disease involving the periodontium. The mineralized periodontal tissues of alveolar bone and cementum contain very little HA; however, it is necessitated by the extracellular matrix of the gingiva and the periodontal ligament (4). Wound healing is accelerated following treatment with HA, due to the subsequent impact on HA receptors, which play a role in cellular migration, angiogenesis and inflammation. HA relieves symptoms, both in the marginal gingiva and deeper periodontal tissues (6). This wound-healing characteristic of HA has also been used in different periodontal treatments, involving non-surgical and surgical therapy, and soft and hard tissue regeneration. Moreover, HA plays a crucial role in cell signaling and hemostasis, and manages the cell-matrix and cell-cell exchanges. HA also plays role in the inflow and outflow of nutrients and waste products (7). Notably, HA may be used in periodontal research due to the extensive scope of applications, which are summarized in the present review. No systemic search strategy was performed for the review. An online search was performed for studies, without a specific time frame, using key words (hyaluronic acid, periodontics, periodontal therapy, periodontal regeneration and wound healing) in combination.

2. Properties of HA

HA plays a variety of roles in the body, leveraging physico-chemical and biological properties. These biological functions vary from basic structural roles in the extracellular matrix to developmental regulation via effects on cell behaviour through tissue macro and microenvironment control. These functions of HA also exert direct receptor-mediated effects on gene expression (8,9). HA exhibits hygroscopic and viscoelastic properties. Hydrogen and adjacent carboxyl and N-acetyl group bonding occurs after HA is absorbed by an aqueous solution, allowing HA to maintain conformational rigidity through water retention (10). Through maintaining

Correspondence to: Dr Ashok Bhati, Department of Preventive Dental Sciences, College of Dentistry, Jazan University, Shawajrah Campus, Almareefah Road, Jazan 45142, Saudi Arabia
E-mail: gums_ashh@yahoo.com

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gaps and preserving surfaces, HA modifies the surrounding cellular and extracellular micro and macro environments. HA exhibits bacteriostatic activity, and this was demonstrated to be the highest when high concentrations of medium and lower molecular weight HA were used (11). HA is biocompatible and non-immunogenic in nature. Esterification and cross-linking are two modifications of HA that provide a gel-like structure and rigidity for cell seeding (12). In addition, HA exhibits anti-inflammatory properties. It performs scavenging actions, through the clearing of prostaglandins, metalloproteinases and other bioactive molecules (12). HA also exhibits anti-edematous properties associated with osmotic action (13), and antioxidant properties. HA scavenges reactive oxygen species; thus, stabilizing the granulation tissue matrix (14).

3. Synthesis of HA

HA is a negatively charged glycosaminoglycan that differs from other glycosaminoglycans. HA synthesis occurs in the cellular plasma membrane in mammals, whereas glycosaminoglycan synthesis often occurs in the Golgi apparatus. Moreover, HA synthesis occurs via three hyaluronan synthase isoenzymes (HAS1, 2 and 3) (7). HA exhibits a high molecular weight of 10^3 - 10^4 kDa, a length of 2-25 μ m, and it does not contain any sulphate groups (8). The synovial fluid, epidermis, umbilical cord and other tissues exhibit the highest concentrations of HA, while the blood serum exhibits the lowest concentration (15). A membrane-bound protein present in plasma membranes produces HA through the transportation of activated monosaccharides to glycosaminoglycan chains, and the release of uridine diphosphate, secreted directly into the extracellular space. Lymphatic drainage into the circulatory system or local metabolism causes HA turnover in tissues. Depending on its removal, HA exhibits a tissue half-life ranging from 12 h to 2-3 days (16).

4. Role of HA in wound healing

HA plays a role in numerous physiological and biological processes, serving as a structural component of cartilage and other tissues. To produce proteoglycans, HA interacts with proteins rich in numerous forms of glycosaminoglycans. It increases inflammatory cell and extracellular matrix infiltration, assisting inflammation. Thus, HA exhibits the potential to impact cellular behaviour via influencing the environment surrounding cells (16). HA is engaged in numerous cell functions which increase tissue healing, such as cell proliferation, locomotion and recognition. This makes HA increasingly susceptible to colonization by tissue repair cells (17). In its highly purified form, HA has been used in medicine for a number of years, due to its physiochemical characteristics and non-immunogenicity. As HA retains water in large amounts, it affects and improves tissue regeneration; thus, preventing the production of scabs and scars (18,19). It has been proposed that HA stimulates angiogenesis, leading to increased levels of wound healing in the bone matrix. At a low molecular weight, HA is angiogenic, whereas at a high molecular weight, HA is anti-angiogenic (20). High molecular weight HA enhances osteo-induction or bone production during wound healing (21). Results of previous studies demonstrated that exogenous

HA exerted satisfying wound healing benefits (22-24). In cosmetic dermatology, HA is also used as a dermal filler (25). As it forms an integral part of cell migration, organogenesis and development, HA exhibits potential in tissue engineering (26). The esterification and crosslinking of HA are two modifications that provide the gel-like structure and rigidity for cell seeding. These biopolymers are biodegradable which help fibroblasts, chondrocytes and mesenchymal stem cells to proliferate (27). HA has been employed as a chemotherapeutic agent in the treatment of gingivitis. Moreover, osseointegration of dental implants indicates the involvement of HA (28). As HA exhibits bone induction properties, it may exhibit potential as a biomaterial scaffold in guided bone regeneration and tissue engineering (29). In 2022, Ibraheem *et al* (30) demonstrated improved wound healing in extraction socket wounds following treatment with HA. Moreover, HA exhibited dose-dependent bacteriostatic effects on a range of microorganisms in the planktonic phase (31).

Hyaluronan generates cell responses via interaction with cell receptors. Notably, there are numerous different cell receptors involved in HA signaling. The most common receptor involved in HA signaling is CD44 (32). CD44 signaling plays a crucial role in wound healing, as CD44 in fibroblasts is required for emigration into the affected region (33). Notably, receptor for hyaluronan-mediated motility (RHAMM), also known as CD168, plays a key role in signaling. RHAMM-hyaluronan interactions are vital for tissue repair and inflammation (34). In addition, hyaluronan receptor for endocytosis is involved in hyaluronan endocytosis, and lymphatic vessel endothelial hyaluronan receptor 1 plays a role in the regulation of tissue hydration and other biochemical properties, via the absorption of HA in the lymphatic system (35,36).

Toll-like receptors are involved in innate immune response, tissue metabolism and tissue haemostasis (37). Toll-like receptors prompt the synthesis of defensins, which possess antibacterial properties and employ regenerative stimuli for cells (38). The molecular weight of HA is an important aspect in HA signaling. Notably, HA induces different signaling pathways at different molecular weights (39).

5. Role of HA and overview of studies in periodontal therapy

HA is located in the periodontium in differing amounts. There are higher levels in the gingiva and periodontal ligament, compared with the cementum and alveolar bone. Moreover, high amounts of HA in the circulatory blood serum represent a serum overload factor of gingival crevicular fluid (40).

Scaling and root planing (SRP) exerts a positive effect on periodontal parameters, as it decreases the pathogens present in the periodontal pocket and alters the microflora to be less pathogenic (41). Results of previous studies investigating SRP combined with non-mechanical therapy demonstrated that the use of local or systemic antibacterial medicines further improved clinical parameters (42,43). However, systemic antibiotics should not be utilized to treat all types of periodontitis. Limiting the use of systemic antibiotics is vital as the development of resistance and unwarranted drug interactions may occur. In patients with gingivitis and chronic periodontitis, HA is utilized as an adjuvant therapy following SRP. Treatment

Table I. Overview of studies using HA in periodontal therapy.

Study	Sample size	HA Form	Intervention	Inference	Periodontal Therapy
Nguyen <i>et al</i> , 2021 (57)	28 chronic periodontitis patients	0.2% HA gel	HA + SRP vs. SRP	HA showed a greater reduction in BOP and PPD	Non-surgical periodontal therapy
Al-Shammari <i>et al</i> , 2018 (49)	24 chronic periodontitis patients	0.8% HA gel	HA + SRP vs. SRP	HA showed a greater reduction in BOP and PPD	Non-surgical periodontal therapy
Rajan <i>et al</i> , 2014 (58)	33 chronic periodontitis patients	HA gel	HA + SRP vs. SRP	HA showed a greater reduction in BOP and PPD	Non-surgical periodontal therapy
Pilloni <i>et al</i> , 2019 (59)	30 patients single Miller's Class I recession	HA gel	HA + CAF vs. CAF	80% complete root coverage for HA and 33.3% complete root coverage for control sites	Recession coverage
Kumar <i>et al</i> , 2014 (60)	10 patients with Miller's Class I recession	HA gel	HA + CAF vs. CAF	HA group clinically more stable	Recession coverage
Pitale <i>et al</i> , 2021 (61)	25 patients with Class I and II papillary recession	Injection HA gel	-	Significant difference in black triangle height and width	Papilla regeneration
Mansouri <i>et al</i> , 2013 (62)	21 interdental papilla deficiencies	HA gel	-	43% of samples showed 50% or higher improvement	Papilla regeneration
Babgi <i>et al</i> , 2020 (63)	15 single-rooted teeth	HA gel	SRP or EDTA or CHX gel	SRP highest improvement followed by the HA and CHX groups	Root conditioning
Mamajiwala <i>et al</i> , 2021 (64)	20 chronic periodontitis patients with 40 infrabony defects	HA gel	HA gel + OFD vs. OFD + Placebo	HA showed significantly greater CAL gain and bone defect fill	Regenerative therapy
Bhowmik and Rao, 2021 (65)	32 graftable defects	HA gel	HA + NHA bone graft (H-NHA) vs. NHA alone	H-NHA group showed a greater reduction in PPD and defect depth	Regenerative therapy
Briguglio <i>et al</i> , 2013 (66)	40 subjects with a two-wall infrabony defect	HA gel	HA + OFD vs. OFD	Test group showed significantly greater CAL gain and PPD	Regenerative therapy
Sehdev <i>et al</i> , 2016 (67)	24 infrabony defects	Esterified HA in the form of fibers	HA + bioresorbable membrane vs. bioresorbable membrane alone	HA showed significantly higher CAL of 2.20 mm	Regenerative therapy
Sánchez-Fernández <i>et al</i> , 2021 (68)	100 dental implants	HA gel	HA vs. excipient-based gel vs. no gel	PPD was significantly lower in the HA group	Therapy for peri-implantitis cases
Dogan <i>et al</i> , 2017 (69)	13 systemically healthy patients requiring bilateral two-stage maxillary sinus augmentation (residual crest height ≤4 mm)	HA matrix	Hyaluronic matrix and collagenated heterologous bone graft vs. collagenated heterologous bone graft	Higher percentage of new bone formed in the HA group after 4 months	Sinus lift augmentation

HA, hyaluronic acid; SRP, scaling and root planing; BOP, bleeding on probing; PPD, probing pocket depth; CAF, coronally advanced flap; EDTA, ethylenediaminetetraacetic acid; CHX, chlorhexidine; OFD, open flap debridement; NHA, nanohydroxyapatite; CAL, clinical attachment level.

with HA results in a reduction in prostaglandins, metalloproteinases and bioactive materials. This results in the impediment of tissue destruction; thus, promoting healing (44).

Notably, Sahayata *et al* (45) used 0.2% HA gel (Gengigel®) topically in patients with gingivitis. Results of this previous study demonstrated that HA treatment led to decreased gingival bleeding, reduced gingival fluid flow and improved gingival health (43,44). Moreover, Pilloni *et al* (46) investigated the effects of HA on periodontal parameters in patients with periodontitis. Results of this previous study demonstrated improved outcomes in the HA group compared with the non-HA group, indicated by significant improvements in bleeding on probing (46). In addition, Pistorius *et al* (47) also demonstrated improvements in bleeding following treatment with HA. Eick *et al* (48) evaluated the effects of HA with SRP and SRP alone in patients with chronic periodontitis. Results of this previous study demonstrated a significantly higher reduction in probing pocket depth (PPD) in the SRP + HA group, compared with the use of SRP alone (48).

Improvements have been observed in healing, periodontal indices and clinical attachment level (CAL) following treatment with HA. In a split-mouth study, Al-Shammari *et al* (49) evaluated the effects of SRP + HA and SRP alone in patients with chronic periodontitis. Improvements in gingival indices, CAL and PPD were observed following 6 and 12 weeks in the test group (49). In addition, Madkour *et al* (50) assessed the effects of HA in patients with chronic periodontitis under medication. Comparable with the results obtained by Al-Shammari *et al* (49), the levels of gingival indices, PPD and CAL were improved in both groups, but these levels were most improved in the SRP + HA group (50). Eliezer *et al* (51) also conducted a systematic review, which demonstrated that HA offers beneficial effects in pocket depth reduction, CAL gain and reduction in bleeding on probing during both surgical and non-surgical periodontal therapy.

HA improved CAL and boosted keratinization. Treatment with injectable HA gel of various doses led to improvements in papillary regeneration, as HA promotes neovascularization (52). Çankaya and Tamam compared the interdental papillary fill after HA was injected into the maxillary and mandibular jaws. The injection was delivered until the color of the gingiva changed to white. After 3, 12 and 24 months, the interdental area demonstrated 54.21, 73.22 and 79.35% coverage remaining in the maxilla, and 57.24, 71.40 and 78.71% coverage remaining in the mandible, respectively (52). Hyaluronan promotes fibroblast attachment to the cementum, while also acting as an antibacterial agent. Moreover, sodium hyaluronate enhances chemical signaling between cells. As a result, sodium hyaluronate is often used in the guided tissue regeneration membrane (GTR) (53). Notably, Vanden Bogaerde (54) examined the clinical effectiveness of esterified HA fibers in the treatment of 18 periodontal defects. The mean PPD was decreased by 5.8 mm after a follow-up of 12 months, and the CAL increase was 2.8 mm (54). In a histological examination in experimental animals, novel alveolar bone growth was observed in bone lesions (55). In 2022, a systemic review by Rodríguez-Aranda *et al* (56) also demonstrated favourable results with HA in periodontal regeneration. This systemic review revealed improvements in terms of CAL, PPD, BOP, and radiographic parameters, when

HA was used alone or in combination with bone graft or other biomaterials (56).

Collectively, the results of previous studies have demonstrated the effective use of HA in periodontal therapy. An overview of studies using HA as a periodontal therapeutic intervention with different sample sizes and HA forms are presented in Table I (49,57-69).

6. Conclusion

HA is a vital biomaterial used in periodontal therapy. Treatment with HA leads to clinical improvement in patients with gingivitis, periodontitis, implants and periodontal defects. Treatment with HA accelerates wound healing, resulting in improved postoperative results and high levels of patient comfort. However, further research into the therapeutic effects of HA in periodontal disease is required. This will further determine the specific uses, the optimal administration of HA for the postoperative treatment of periodontal conditions and the potential for complete periodontal tissue regeneration.

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AB, HyF, HaF and WI conceived the review. AB performed the literature search. AB, HyF, HaF and WI wrote the manuscript. HC, SP and AB reviewed and edited the manuscript. All authors have read and approved the final manuscript. Data authentication is not applicable.

Ethics approval and consent to participate

Not applicable.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

References

1. Amorim S, Reis CA, Reis RL and Pires RA: Extracellular matrix mimics using hyaluronan-based biomaterials. *Trends Biotechnol* 39: 90-104, 2021.
2. Meyer K and Palmer JW: The polysaccharide of the vitreous humor. *J Biol Chem* 107: 629-634, 1934.

3. Bansal J, Kedige SD and Anand S: Hyaluronic acid: A promising mediator for periodontal regeneration. *Indian J Dent Res* 21: 575-578, 2010.
4. Aydinlyurt HS, Akbal D, Altindal D, Bozoglan A, Ertugrul AS and Demir H: Evaluation of biochemical and clinical effects of hyaluronic acid on non-surgical periodontal treatment: A randomized controlled trial. *Ir J Med Sci* 189: 1485-1494, 2020.
5. Abatangelo G, Vindigni V, Avruscio G, Pandis L and Brun P: Hyaluronic acid: Redefining its role. *Cells* 9: 1743, 2020.
6. Casale M, Moffa A, Vella P, Sabatino L, Capuano F, Salvinelli B, Lopez MA, Carinci F and Salvinelli F: Hyaluronic acid: Perspectives in dentistry. A systematic review *Int J Immunopathol Pharmacol* 29: 572-582, 2016.
7. Vigetti D, Karousou E, Viola M, Deleonibus S, De Luca G and Passi A: Hyaluronan: Biosynthesis and signaling. *Biochim Biophys Acta* 1840: 2452-2459, 2014.
8. Toole BP: Hyaluronan is not just a goo! *J Clin Invest* 106: 335-336, 2000.
9. Stern R, Asari AA and Sugahara KN: Hyaluronan fragments: An information rich system. *Eur J Cell Biol* 85: 699-715, 2006.
10. Sutherland IW: Novel and established applications of microbial polysaccharides. *Trends Biotechnol* 16: 41-46, 1998.
11. Pirnazar P, Wolinsky L, Nachmani S, Haake S, Piloni A and Bernard GW: Bacteriostatic effects of hyaluronic acid. *J Periodontol* 70: 370-374, 1999.
12. Laurent TC, Laurent UB and Fraser JR: Functions of hyaluronan. *Ann Rheum Dis* 54: 429-432, 1995.
13. Jentsch H, Pomowski R, Kundt G and Göcke R: Treatment of gingivitis with hyaluronan. *J Clin Periodontol* 30: 159-164, 2003.
14. Waddington RJ, Moseley R and Embery G: Reactive oxygen species: A potential role in the pathogenesis of periodontal diseases. *Oral Dis* 6: 138-151, 2000.
15. Laurent TC and Fraser JRE: The properties and turnover of hyaluronan. *Ciba Found Symp* 124: 9-29, 1986.
16. Fraser JR, Laurent TC and Laurent UB: Hyaluronan: Its nature, distribution, functions and turnover. *J Intern Med* 242: 27-33, 1997.
17. Samuel SK, Hurta RA, Spearman MA, Wright JA, Turley EA and Greenley AH: TGF-beta 1 stimulation of cell locomotion utilizes the hyaluronan receptor RHAMM and hyaluronan. *J Cell Biol* 123: 749-758, 1993.
18. Nakamura M, Hikida M, Nakano T, Ito S, Hamano T and Kinoshita S: Characterization of water retentive properties of hyaluronan. *Cornea* 12: 433-436, 1993.
19. Adzick NS and Longaker MT: Scarless wound healing in the fetus: The role of extracapsular matrix. *Prog Clin Biol Res* 365: 177-192, 1991.
20. West DC and Kumar S: Hyaluronan and angiogenesis. *Ciba Found Symp* 143: 187-207, 1989.
21. Sakaki T and Watanabe C: Stimulation of osteoinduction in bone wound healing by high-molecular hyaluronic acid. *Bone* 16: 9-15, 1995.
22. Abatangelo G, Martelli M and Vecchia P: Healing of hyaluronic acid-enriched wounds: Histological observations. *J Surg Res* 35: 410-416, 1983.
23. King SR, Hickerson WL, Proctor KG and Newsome AM: Beneficial actions of exogenous hyaluronic acid on wound healing. *Surgery* 109: 76-84, 1991.
24. Nakamura M, Hikida M, Lyngstaadas SP, Paine ML and Snead ML: Principles and applications of cell delivery systems for periodontal regeneration. *Periodontol* 2000 41: 123-135, 2006.
25. Klingner MM, Rahemtulla F, Prince CW, Lucas LC and Lemonas JE: Proteoglycans at the bone-implant interface. *Crit Rev Oral Med* 9: 449-463, 1988.
26. Hunt DR, Jovanovic SA, Wikesjö UM, Wozney JM and Bernard GW: Hyaluronan supports recombinant human bone morphogenetic protein-2 induced bone reconstruction of advanced alveolar ridge defects in dogs. A pilot study. *J Periodontol* 72: 651-658, 2001.
27. Ibraheem W, Jedaiba WH, Alnami AM, Hussain Baiti LA, Ali Manqari SM, Bhati A, Almarghlani A and Assaggaf M: Efficacy of hyaluronic acid gel and spray in healing of extraction wound: A randomized controlled study. *Eur Rev Med Pharmacol Sci* 26: 3444-3449, 2022.
28. Carlson GA, Dragoo JL, Samimi B, Bruckner DA, Bernard GW, Hedrick M and Benhaim P: Bacteriostatic properties of biomaterials against common orthopaedic pathogens. *Biochem Biophys Res Commun* 321: 472-478, 2004.
29. Arfullo A, Stamenkovic I, Melnick M, Underhill CB and Seed B: CD44 is the principal cell receptor for hyaluronate. *Cell* 61: 1303-1313, 1990.
30. Clark RA, Lin F, Greiling D, An J and Couchman JR: Fibroblast invasive migration into fibronectin/fibrin gels requires a previously uncharacterized dermatan sulfate-CD44 proteoglycan. *J Invest Dermatol* 122: 266-277, 2004.
31. Zaman A, Cui Z, Foley JP, Zhao H, Grimm PC, Delisser HM and Savani RC: Expression and role of the hyaluronan receptor RHAMM in inflammation after bleomycin injury. *Am J Respir Cell Mol Biol* 33: 447-454, 2005.
32. Zhao B, Weigel JA, Saxena A and Weigel PH: Molecular cloning and functional expression of the rat 175-kDa hyaluronan receptor for endocytosis. *Mol Biol Cell* 13: 2853-2868, 2002.
33. Prevost R, Banerji S, Ferguson DJ, Clasper S and Jackson DG: Mouse LYVE1 is an endocytic receptor for hyaluronan in lymphatic endothelium. *J Biol Chem* 276: 19420-19430, 2001.
34. Rakoff-Nahoum S, Paglino J, Eslami-Varzaneh F, Edberg S and Medzhitov R: Recognition of commensal microflora by toll like receptors is required for intestinal hemostasis. *Cell* 118: 229-234, 2004.
35. Gariboldi S, Plazzo M, Zanobbio L, Selleri S, Sommariva M, Sfondrini L, Cavacchini S, Balsari A and Rucio C: Low molecular weight hyaluronic acid increases the self defense of skin epithelium by induction of beta defensin 2 via TLR2 and TLR 4. *J Immunol* 181: 2103-2110, 2008.
36. Itano N: Simple primary structure, complex turnover regulation and multiple roles of hyaluronan. *J Biochem* 144: 131-137, 2008.
37. Embery G, Waddington RJ, Hall RC and Last KS: Connective tissue elements as diagnostic aids in periodontology. *Periodontol* 2000 24: 193-214, 2000.
38. Cugini MA, Haffajee AD, Smith C, Kent RL Jr and Socransky SS: The effect of scaling and root planing on the clinical and microbiological parameter of periodontal diseases: 12-month results. *J Clin Periodontol* 27: 30-36, 2000.
39. Bonito AJ, Lux L and Lohr KN: Impact of local adjuncts to scaling and root planing in periodontal disease therapy: A systematic review. *J Periodontol* 76: 1227-1236, 2005.
40. Johannsen A, Tellefsen M, Wikesjö U and Johannsen G: Local delivery of hyaluronan as an adjunct to scaling and root planing in the treatment of chronic periodontitis. *J Periodontol* 80: 1493-1497, 2009.
41. Laurent TC, Laurent UB and Fraser JR: The structure and function of hyaluronan: An overview. *Immunol Cell Biol* 74: A1-A7, 1996.
42. Sahayata VN, Bhavsar NV and Brahmabhatt NA: An evaluation of 0.2% hyaluronic acid gel (Gengigel®) in the treatment of gingivitis: A clinical & microbiological study. *Oral Health Dent Manag* 13: 779-785, 2014.
43. Piloni A, Annibali S, Dominici F, Di Paolo C, Papa M, Cassini MA and Polimeni A: Evaluation of the efficacy of an hyaluronic acid-based biogel on periodontal clinical parameters. A randomized-controlled clinical pilot study. *Ann Stomatol (Roma)* 2: 3-9, 2011.
44. Pistorius A, Martin M, Willershausen B and Rockmann P: The clinical application of hyaluronic acid in gingivitis therapy. *Quintessence Int* 36: 531-538, 2005.
45. Eick S, Renatus A, Heinicke M, Pfister W, Stratul SI and Jentsch H: Hyaluronic Acid as an adjunct after scaling and root planing: A prospective randomized clinical trial. *J Periodontol* 84: 941-949, 2013.
46. Al-Shammari NM, Shafshak SM and Ali MS: Effect of 0.8% Hyaluronic acid in conventional treatment of moderate to severe chronic periodontitis. *J Contemp Dent Pract* 19: 527-534, 2018.
47. Madkour GG, EL Refaie I and Mostafa B: Adjunctive use of hyaluronic acid with scaling and root planing in treatment of chronic periodontitis patients with diabetes mellitus type 2: A randomized controlled trial. *Egypt Dent J* 64: 4057-4065, 2018.
48. Eliezer M, Imber JC, Sculean A, Pandis N and Teich S: Hyaluronic acid as adjunctive to non-surgical and surgical periodontal therapy: A systematic review and meta-analysis. *Clin Oral Invest* 23: 3423-3435, 2019.

52. Turgut Çankaya Z and Tamam E: An examination of the 2-year results obtained from hyaluronic acid filler injection for interdental papilla losses. *Quintessence Int* 51: 274-284, 2020.
53. Aveic S, Craveiro RB, Wolf M and Fischer H: Current trends in in vitro modeling to mimic cellular crosstalk in periodontal tissue. *Adv Healthc Mater* 10: e2001269, 2021.
54. Vanden Bogaerde L: Treatment of infrabony periodontal defects with esterified hyaluronic acid: Clinical report of 19 consecutive lesions. *Int J Periodontics Restorative Dent* 29: 315-323, 2009.
55. Sukumar S and Ivo Dřížhal I: Hyaluronic acid and periodontitis. *Acta Medica (Hradec Kralove)* 50: 225-228, 2007.
56. Rodríguez-Aranda M, Iborra-Badia I, Alpiste-Illueca F and Lopez-Roldan A: Hyaluronic acid for periodontal tissue regeneration in intrabony defects. A systematic review. *Dentistry Review* 2: 100057, 2022.
57. Nguyen TT, Ho HT, Huynh NC, Dien VHA and Vo TL: Hyaluronic acid 0.2% application enhanced periodontitis treatment in non-surgical phase. *J Stoma* 74: 76-83, 2021.
58. Rajan P, Baramappa R, Rao NM, Pavaluri AK, P I and Rahaman SM: Hyaluronic Acid as an adjunct to scaling and root planing in chronic periodontitis. A randomized clinical trial. *J Clin Diagn Res* 8: ZC11-ZC14, 2014.
59. Pilloni A, Schmidlin PR, Sahrman P, Sculean A and Rojas MA: Effectiveness of adjunctive hyaluronic acid application in coronally advanced flap in Miller class I single gingival recession sites: A randomized controlled clinical trial. *Clin Oral Invest* 23: 1133-1141, 2019.
60. Kumar R, Srinivas M, Pai J, Suragimath G, Prasad K and Polepalle T: Efficacy of hyaluronic acid (hyaluronan) in root coverage procedures as an adjunct to coronally advanced flap in Millers Class I recession: A clinical study. *J Indian Soc Periodontol* 18: 746-750, 2014.
61. Pitale U, Pal PC, Thakare G, Verma M, Dhakad S and Pandey R: Minimally invasive therapy for reconstruction of lost interdental papilla by using injectable hyaluronic acid filler. *J Indian Soc Periodontol* 25: 22-28, 2021.
62. Sadat Mansouri S, Ghasemi M, Salmani Z and Shams N: Clinical application of hyaluronic acid gel for reconstruction of interdental papilla at the esthetic zone. *J Iran Dent Asso* 25: 208-213, 2013.
63. Babgi W, Alhajaji M, Al-Mehmadi L, Elbaqli R, Khayat N, Aldahlawi S and Youssef AR: Effect of root conditioning agents hyaluronic acid, EDTA and chlorhexidine on the attachment of human gingival fibroblasts to healthy root surface. *Saudi Dent J* 33: 342-347, 2021.
64. Mamajiwala AS, Sethi KS, Raut CP, Karde PA and Mamajiwala BS: Clinical and radiographic evaluation of 0.8% hyaluronic acid as an adjunct to open flap debridement in the treatment of periodontal intrabony defects: Randomized controlled clinical trial. *Clin Oral Invest* 25: 5257-5271, 2021.
65. Bhowmik E and Rao DPC: Clinicoradiographic evaluation of hyaluronan-nano hydroxyapatite composite graft in the management of periodontal infrabony defects. *J Indian Soc Periodontol* 25: 220-227, 2021.
66. Briguglio F, Briguglio E, Briguglio R, Cafiero C and Isola G: Treatment of infrabony periodontal defects using a resorbable biopolymer of hyaluronic acid: A randomized clinical trial. *Quintessence Int* 44: 231-240, 2013.
67. Sehdev B, Bhongade ML and Ganji KK: Evaluation of effectiveness of hyaluronic acid in combination with bioresorbable membrane (poly lactic acid-poly glycolic acid) for the treatment of infrabony defects in humans: A clinical and radiographic study. *J Indian Soc Periodontol* 20: 50-56, 2016.
68. Sánchez-Fernández E, Magán-Fernández A, O'Valle F, Bravo M and Mesa F: Hyaluronic acid reduces inflammation and crevicular fluid IL-1 β concentrations in peri-implantitis: A randomized controlled clinical trial. *J Periodontal Implant Sci* 51: 63-74, 2021.
69. Dogan E, Dursun E, Tosun E, Bilgic E, Akman AC, Orhan K, Celik HH, Korkusuz P and Caglayan F: Evaluation of hyaluronic matrix efficacy in sinus augmentation: A randomized-controlled histomorphometric and micro-computed tomography analysis. *Int J Oral Maxillofac Surg* 46: 931-937, 2017.



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