

Reduced expression of TRF1 is associated with tumor progression and poor prognosis in oral squamous cell carcinoma

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Abstract. The functions of telomeric repeat-binding factor 1 (TRF1) and 2 (TRF2) in oral carcinogenesis are largely unexplored. This study examined the relationship between the expression of TRF1 and TRF2 and clinicopathological variables and survival in oral cavity squamous cell carcinoma (OCSCC). Western blotting and immunohistochemistry were used to evaluate the protein expression of TRF1 and TRF2 in paired OCSCC patient specimens. Expression of TRF1 and TRF2 was assessed by immunohistochemistry in 256 OCSCC patients who underwent tumor resection without previous radiotherapy. The results were analyzed using Fisher's exact test. Protein expression of TRF1 and TRF2 was significantly lower in the OCSCC than in the adjacent non-tumor tissue. Reduced TRF1 and TRF2 levels in 256 patients, as revealed by immunohistochemistry, were significantly associated with aggressive clinicopathological features, such as advanced tumor stage ($p<0.001$) and advanced tumor node metastasis stage ($p<0.001$). According to Kaplan-Meier analysis, reduced TRF1 expression was significantly correlated with an unfavorable cumulative 5-year overall survival rate ($p<0.001$). In conclusion, decreased expression of TRF1 was significantly associated with tumor progression and poor prognosis in OCSCC patients.

Introduction

Oral cavity squamous cell carcinoma (OCSCC) accounts for at least 90% of all oral malignancies. It is a multifactorial

condition with etiological links to a wide variety of external causes of cancer, including alcohol, tobacco and betel nut use, and certain viral infections. The high and increasing prevalence of OCSCC in Taiwan has been attributed to the popularity of betel nut chewing. It was estimated that, in 2006, more than 4,000 people in Taiwan were diagnosed with oral cancer. This represents 5.49% of all newly diagnosed malignancies. Despite advances in technology and the implementation of multidisciplinary treatment programs, only modest improvements in survival rates have been achieved, and these are primarily due to earlier diagnosis, rather than improved therapeutic interventions (1). Moreover, the rate of recurrence of advanced tumors remains relatively high. Salvage outcomes are unsatisfactory, although they depend on the stage of the recurrent tumors (2). Investigation of OCSCC progression from a genetic perspective has identified distinct patterns and timings of genetic alterations (3). The most important prognostic factors in OCSCC are those that form part of the grading system, including tumor stage and lymph node status (4-6). The identification of new prognostic factors linked to OCSCC initiation and progression may aid in the development of new diagnostic tools and treatment strategies.

Among the various molecular factors implicated in carcinogenesis, telomere dysfunction has emerged as an early event associated with genetic instability. Telomeres stabilize the ends of chromosomes, protect them from end-to-end fusion and mediate chromosome pairing during cell division (7-10). Recently, telomere-associated proteins, such as telomeric repeat-binding factor 1 (TRF1) and 2 (TRF2), have been identified as putative modulators of telomerase activity and have been suggested to play key roles in the maintenance of the telomere function (8,9,11,12). Several reports have indicated that the altered expression of TRF1 and TRF2 proteins is associated with tumor progression in various human carcinomas, including lung, stomach, adrenal and pancreatic cancer; the altered expression has also been identified in malignant hematopoietic cells and colorectal pre-neoplastic lesions (13-20). However, the relationship between TRF1 and TRF2 and OCSCC remains unclear. The aim of the present study was to examine TRF1 and TRF2 expression in OCSCC and to determine its relationship with clinicopathological variables and survival.

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Materials and methods

Patients and tumor samples. The study population included 256 OCSCC patients who underwent primary surgical resection without previous radiotherapy and/or chemotherapy between October 1996 and August 2005. Clinicopathological information for each subject, including gender, age, tumor (T) stage, nodal (N) status, tumor node metastasis (TNM) stage and overall survival, was obtained retrospectively from clinical records and pathological reports. TNM status was classified according to the 1997 American Joint Committee on Cancer (AJCC) system. The study was approved by the Medical Ethics and Human Clinical Trial Committee at Chang Gung Memorial Hospital, Taipei, Taiwan. The patient group comprised 17 women and 239 men, with an average age of 50.9 years (range, 26-87 years). Thirty-nine patients were diagnosed with T1 tumors, 55 with T2, 64 with T3 and 98 with T4. A total of 153 patients had an N status of N0, 38 had N1, 48 had N2b, 13 had N2c and 4 had N3. Thirty-four patients had stage I tumors, 38 stage II, 61 stage III and 123 stage IV.

Immunoblot analysis. For tissue protein extraction, frozen samples (adjacent non-tumor and tumor tissues) were homogenized in RIPA lysis buffer (50 mM Tris-HCl, pH 7.5, 150 mM NaCl, 1% NP-40, 0.5% Na-deoxycholate and 0.1% SDS), and the protein concentrations were quantified using a Bio-Rad Protein assay (Bio-Rad, Hercules, CA, USA). Immunoblotting was performed according to standard procedures. Anti-TRF1 and -TRF2 polyclonal antibodies and the anti-GAPDH monoclonal antibody (Santa Cruz Biotechnology Santa Cruz, CA, USA) were used. The bound primary antibody was detected by incubation with HRP-conjugated secondary antibody (Bio/Can Scientific, Mississauga, ON, Canada). Blots were developed using the Western Lighting reagent, and protein bands were visualized using X-ray film.

Immunohistochemical analysis. OCSCC and adjacent non-cancerous tissue samples were identified by a pathologist based on diagnosis and microscopic morphology. Tumor tissues were fixed in 10% buffered formalin, embedded in paraffin and decalcified in 10% EDTA. Formalin-fixed paraffin-embedded tissue was sectioned to a thickness of 4 μ m, and the sections were deparaffinized in xylene and rehydrated in a graded series of ethanol (100, 90, 80 and 70%). The sections were washed in phosphate-buffered saline (PBS) and treated with 3% H₂O₂ for 30 min to block the endogenous peroxidase activity. Antigens were retrieved by microwaving the sections in 10 mM citrate buffer, pH 6.0. The sections were then incubated with anti-TRF1 and -TRF2 antibodies (diluted 1:100) for 1 h, washed in PBS and incubated for 30 min with a horse-radish peroxidase-Fab polymer conjugate (PicTure™-Plus kit; Zymed, South San Francisco, CA, USA). After the sections were washed in PBS, the immunoreactive bands were visualized by incubation with 3,3'-diaminobenzidine for 5 min. As a negative control, the primary antibody was omitted. Two pathologists blinded to the subjects' clinical information independently evaluated the reactivity level of the immunostained tissues in 15-20 high-power fields. Criteria were developed for quantitating the immunoreactivity of the TRF1 and TRF2

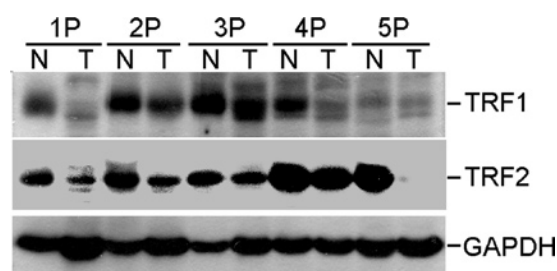


Figure 1. Expression of TRF1 and TRF2 was down-regulated in OCSCC. Western blot analysis of the protein expression of TRF1 and TRF2 in paired specimens from 5 oral cancer patients. The total protein extracted from cancerous (T) and adjacent non-cancerous tissues (N) was probed using polyclonal antibodies against TRF1 and TRF2. GAPDH was used as a loading control.

staining in the adjacent non-tumor and tumor sections using a score range of 0 to +3. A percentage value of 0 indicated 0-25% of the area stained; +1, 25-50%; +2, 50-75% and +3, >75% stained. Similarly, the staining intensity was graded as +0, +1, +2 or +3. High expression of TRF1 and TRF2 was defined as $\geq +2$ in both scoring methods. Low expression of TRF1 and TRF2 was defined as $\leq +1$ in both scoring methods.

Statistical analysis. Fisher's exact test was used to evaluate the correlation between TRF1 and TRF2 expression and various clinicopathological variables, including gender, age, N status, T stage and TNM stage. A p-value <0.05 was considered to indicate statistical significance. TRF1 and TRF2 expression and the clinicopathological variables were used in a Kaplan-Meier analysis of survival, and statistical significance ($p < 0.05$) was assessed by the log-rank test. To determine the effects of specific prognostic factors on survival, a multivariate analysis was performed using Cox's regression model.

Results

Down-regulation of TRF1 and TRF2 expression in OCSCC tissues. To investigate the potential roles of TRF1 and TRF2 in the pathogenesis of OCSCC, their expression was assessed in representative and paired tumor and adjacent non-cancerous tissue samples by Western blot analysis using anti-human TRF1 and TRF2 polyclonal antibodies. The protein expression levels of TRF1 and TRF2 were lower in the tumor samples compared to those in the paired non-cancerous tissues (Fig. 1).

TRF1 and TRF2 expression in the tumor and adjacent non-cancerous tissues from the 256 OCSCC patients was also examined immunohistochemically. Representative results of TRF1 and TRF2 immunostaining are presented in Fig. 2A. Staining was stronger in the adjacent non-cancerous tissues (Fig. 2A-a and -f) than the tumor tissues (Fig. 2A-b and -c, TRF1; Fig. 2A-g and -h, TRF2). Moreover, TRF1 and TRF2 expression levels in the tumor samples were negatively correlated with T stage (Fig. 2A-b and -c, TRF1; Fig. 2A-g and -h, TRF2) and N stage (Fig. 2A-d and -e, TRF1; Fig. 2A-i and -j, TRF2). Notably, TRF1 and TRF2 were focally expressed in the nuclei of both tumor and non-cancerous cells (Fig. 2A-b and -c, TRF1; Fig. 2A-g and -h, TRF2).

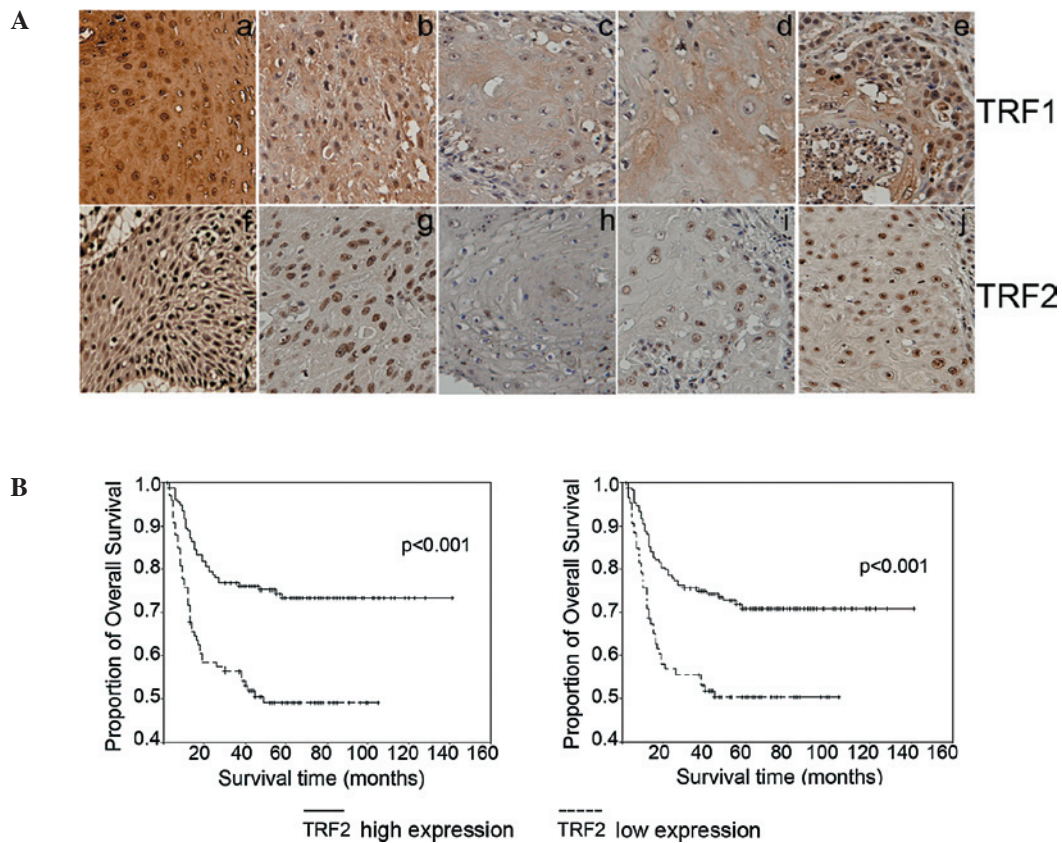


Figure 2. Immunohistochemical staining of TRF1 and TRF2 and overall survival in OCSCC patients. (A) Immunohistochemical staining intensities of TRF1 and TRF2 in tumor and non-cancerous tissues. Non-cancerous tissues displayed extremely strong TRF1 and TRF2 staining in the nucleus (a and f). Staining intensity for TRF1 and TRF2 was lower in the tumor tissues (b,c and g,h). Significant nuclear expression of TRF1 and TRF2 was observed in early (b and g) and late (c and h) stage tumors. TRF1 and TRF2 expression was present in patients with (d and i) and without (e and j) lymph node metastases. Original magnification, x100. (B) The survival time of patients with low (-/+) expression of TRF1 and TRF2 was significantly shorter ($p<0.001$, log-rank test) than that of patients displaying higher (++/+++) expression levels (n=256).

To investigate whether the expression of TRF1 and TRF2 is associated with various prognostic factors, including age, gender and TNM pathologic classification, we classified the patients into two groups based on the immunohistochemical analysis: low (-/+) and high (++/+++) TRF1 or TRF2 expression. Low TRF1 expression was correlated with advanced T stage ($p<0.001$) and advanced TNM stage ($p<0.001$). Low TRF2 expression was correlated with advanced T stage ($p<0.001$) and advanced TNM stage ($p<0.001$), as well as positive lymph node metastasis ($p=0.022$). Neither TRF1 nor TRF2 expression was correlated with age or gender (Table I). These findings suggest that TRF1 and TRF2 expression levels may be linked to tumor progression in OCSCC.

TRF1 and TRF2 expression and OCSCC patient survival. In view of the finding that TRF1 and TRF2 expression levels were associated with T stage, we investigated whether TRF1 and TRF2 expression was correlated with patient prognosis. As shown in Fig. 2B, Kaplan-Meier overall survival analysis revealed that the prognosis of patients with low (-/+) tumor expression of TRF1 and TRF2 was significantly poorer than that of patients displaying higher (++/+++) expression ($p<0.001$). Univariate analysis revealed that advanced T stage ($p<0.001$), positive N stage ($p<0.001$), advanced TNM stage ($p<0.001$), low TRF1 expression and low TRF2 expression

each predicted a significantly worse prognosis for OCSCC patients (Table II). The prognosis was not associated with age or gender. Cox regression analysis revealed that T stage (95% CI, 1.585-5.037; RR=2.826; $p<0.001$), N status (95% CI, 1.966-4.681; RR=3.034; $p<0.001$) and TRF1 expression (95% CI, 0.391-0.924; RR=0.601; $p=0.02$) were independent prognostic factors for survival. These results clearly indicate that the clinical prognosis for OCSCC patients is affected by the tumor expression of TRF1 and TRF2, and suggest that TRF1 and TRF2 may be good prognostic indicators in OCSCC.

Discussion

To our knowledge, this is the first investigation of TRF1 and TRF2 expression in primary OCSCC specimens from a large cohort of patients. Our results indicate that the reduced expression of TRF1 and TRF2 is associated with increased tumor aggressiveness and poor prognosis in OCSCC patients. Low expression of TRF1 and TRF2 and advanced T stage and N stage were correlated with a poor prognosis. As it is difficult to determine a prognosis for these patients, TRF1 and TRF2 staining of oral cancer cells may be helpful in selecting an appropriate therapeutic strategy following surgery. Our findings suggest that TRF1 is a good prognostic indicator in OCSCC and a candidate molecular target for oral cancer therapy.

Table I. Correlation between the clinicopathological features and expression of TRF1 and TRF2 in the oral squamous cell carcinoma cases.

Variables	No. of patients	TRF1 expression		P-value	TRF2 expression		P-value
		Low	High		Low	High	
Gender				0.452			0.434
Male	239	95	144		83	156	
Female	17	5	12		4	13	
Age (years)				0.272			0.333
<60	202	75	127		72	130	
≥60	54	25	29		15	39	
Tumor stage				<0.001 ^a			<0.001 ^a
T1 and T2	94	15	79		11	83	
T3 and T4	162	85	77		76	86	
Nodal stage				0.090			0.022 ^a
Negative	153	53	100		43	110	
Positive	103	47	56		44	59	
TNM stage				<0.001 ^a			<0.001 ^a
I and II	66	5	61		1	65	
III and IV	190	95	95		86	104	

^aStatistically significant

Table II. Cumulative 5-year overall survival rate according to clinicopathological features.

Variables	No. of patients	Cumulative 5-year overall survival rate (%)	P-value
Gender			0.1360
Male	239	62.5	
Female	17	81.9	
TRF1 expression			<0.0010 ^a
Low	100	73.3	
High	156	49.1	
TRF2 expression			0.0003 ^a
Low	87	70.1	
High	169	50.3	
Age (years)			0.1300
<60	202	66.2	
≥60	54	55.1	
Tumor stage			<0.0010 ^a
T1 and T2	94	84.0	
T3 and T4	162	56.5	
Nodal stage			<0.0010 ^a
Negative	153	77.0	
Positive	103	44.7	
TNM stage			<0.0010 ^a
I and II	66	92.4	
III and IV	190	53.9	

^aStatistically significant.

Semi-quantitative RT-PCR analysis of *TRF1* and *TRF2* mRNA expression in total RNA from tumor samples and matched adjacent non-cancerous tissues from 5 OCSCC patients demonstrated that *TRF1* and *TRF2* mRNA levels did not differ significantly between the tumor and adjacent non-cancerous tissues (data not shown). This observation suggests that altered TRF1 and TRF2 protein expression during the development of OCSCC may be realized post-transcriptionally.

Telomere-binding proteins have attracted increasing interest due to their essential roles in regulating the length of telomeric DNA tracts and protecting against chromosomal end-to-end fusion (21). In cancer, telomeres become dysfunctional due to the loss or alteration of telomere-binding proteins involved in telomere maintenance or to DNA damage caused by oxidative stress (22). The telomere-binding proteins TRF1 and TRF2 are crucial for the protection and maintenance of telomeres in mammalian cells (8,23). TRF1 and TRF2 contain a Myb-like helix-turn-helix domain in the C-terminus of the protein, and a conserved central domain that is responsible for the formation of homodimers (24). Previous studies have indicated that TRF1 and TRF2 are down-regulated in malignant tissues (13,14,25-28). To our knowledge, the present study is the first to report not only that TRF1 and TRF2 are down-regulated in OCSCC, but also that their expression levels are correlated with the clinical characteristics of tumors. The correlation of TRF1 and TRF2 expression with clinical T stage may be explained at the cellular level by the roles of TRF1 and TRF2 in regulating the growth of cancer cells, while the correlation with N stage may reflect the participation of TRF1 and TRF2 in the control of metastasis. In contrast to our results, other studies have revealed that TRF1 and TRF2 are up-regulated

in aggressive adenocarcinoma (29-32). This apparent disparity may be the result of differences in the tumors examined and in their microenvironments.

In the present study, decreased expression of TRF1 and TRF2 was detected in OCSCC patients based on Western blotting and immunohistochemistry. TRF1 and TRF2 were strongly expressed in the nucleus of adjacent non-cancerous tissues, and weakly expressed in human OCSCC specimens. Additionally, expression of TRF1 and TRF2 was correlated with 5-year overall survival and clinical prognosis. Notably, TRF1 expression was an independent prognostic indicator for OCSCC in this cohort. These results indicate that TRF1 and TRF2 may be critical regulators of disease progression in OCSCC, making them potential therapeutic targets. Future studies of the physiological targets of TRF1 and TRF2 and their potential roles in the pathogenesis of OCSCC may facilitate the development of novel therapeutic strategies.

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