

# Sulfotransferase 1A1 Arg<sup>213</sup>His polymorphism and prostate cancer risk

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Abstract. Sulfotransferase 1A1 (SULT1A1) is a member of the sulfotransferase family that plays an important role in the biotransformation of numerous carcinogenic and mutagenic compounds through sulfation. A transition, G to A at position 638, in the SULT1A1 gene, results in the Arg<sup>213</sup>His change. This single nucleotide polymorphism reduces the activity and thermostability of the SULT1A1 enzyme. In the present study, the relationship between the SULT1A1 Arg<sup>213</sup>His polymorphism and prostate cancer was investigated using PCR-RFLP. No significant difference in genotype and allele distribution was noted between the prostate cancer and control populations (P=0.072; P=0.099, respectively). The risk of prostate cancer in individuals carrying the SULT1A1\*2 allele (His<sup>213</sup> allele) was determined by combining the SULT1A1\*1/SULT1A1\*2 (Arg/His<sup>213</sup>) and SULT1A1\*2/SULT1A1\*2 (His/His<sup>213</sup>) genotypes. No association was observed between SULT1A1 Arg213His polymorphism and prostate cancer incidence (P=0.24; OR, 1.36; 95% CI, 0.84-2.25). However, the His<sup>213</sup> allele was found to increase the risk of prostate cancer by 1.36-fold. In smoker and non-smoker populations, no significant relationship was determined between the prostate cancer and control population (P=0.45; P=0.34, respectively).

#### Introduction

Human sulfotransferases (SULTs) catalyze the conjugation of sulfate groups to a variety of endogen and exogenous substrates, including many drugs, neurotransmitters, thyroid and steroid hormones and pro-carcinogenic agents (1,2). SULTs are genetically polymorphic and are expressed in a wide variety of tissues, such as the liver, lung, brain, kidney, and platelets (3). To date, 13 human cytosolic SULT isoforms have been identified and grouped as four major families: SULT1, SULT2, SULT4 and SULT6 (4). The SULT1A1 gene mapped to chromosome 16p12.1-p11.2 encodes four different allozymes: SULT1A1<sup>\*</sup>1 (wild-type), SULT1A1<sup>\*</sup>2, SULT1A1<sup>\*</sup>3 and SULT1A1<sup>\*</sup>4. The SULT1A1 enzyme catalyzes the sulfation of certain carcinogenic and mutagenic compounds including heterocyclic and aromatic amines, and polycyclic aromatic hydrocarbons (2). A genetic polymorphism in exon 7 of the SULT1A1 gene at the nucleotide of 638 (codon 213), results in a substitution of histidine by arginine (Arg<sup>213</sup>His). SULT1A1<sup>\*</sup>2 (His<sup>213</sup> allele) is associated with less enzymatic activity and thermal stability compared with the wild-type allele (Arg<sup>213</sup> allele) in platelets (5,6).

Prostate cancer, a serious health problem in the Western world and Turkey, has shown an increasing incidence over the last decade (7). Some reports suggest that the risk of prostate cancer development is influenced by both genetic and environmental factors, such as diet, hormone levels, drinking habit, ethnicity and genetic background (8). It has been suggested by researchers that the SULT1A1 Arg<sup>213</sup>His polymorphism may affect an individual's capacity in the metabolism of numerous endogenous and exogenous compounds consequently resulting in the susceptibility of an individual to cancer (2). Studies have demonstrated the relationship between genetic polymorphisms of SULT1A1 Arg213His and several cancer types including prostate cancer (9-16). On the other hand, the findings of these studies remain controversial. This study investigated, for the first time, the relationship between the SULT1A1 Arg<sup>213</sup>His polymorphism and prostate cancer susceptibility in a Turkish population.

## Materials and methods

Study population. The study population consisted of a total of 255 Turkish men (104 cases and 151 controls). The prostate cancer patients were treated at the Urology Department, Cumhuriyet University Hospital (Central Anatolia) during the year 2004. The patients were newly diagnosed and histologically confirmed to have prostate cancer and were previously untreated (by radiotherapy or chemotherapy). The prostate cancer patients had elevated serum levels of prostate-specific antigen (PSA). The controls were selected randomly from healthy individuals without a history of cancer and having serum levels of PSA <4 ng/ml. Members of the study populations were informed in regards to the aim of this study. During

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the study period, critical information, such as age and smoking habit were collected from the members using a standardized questionnaire. This study was approved by the Ethics Committee of Cumhuriyet University.

SULTIA1 genotyping. Genomic DNA of the study populations was extracted from blood leukocytes using the standard phenolchloroform method (17). SULTIA1 Arg<sup>213</sup>His genotypes were determined using polymerase chain reaction (PCR)-based restriction fragment length polymorphism (RFLP) assay. The PCR reaction was carried out in a total volume of  $25 \ \mu$ l containing ~100 ng genomic DNA, 200 mM deoxynucleotide triphosphates (dATP, dCTP, dGTP and dTTP), 0.2 m of each SULT1A1 primer (forward, 5'-GGG TCT CTA GGA GAG GTG GC-3'; reverse, 5'-GCT GTG GTC CAT GAA CTC CT-3'), 1X reaction buffer [75 mM Tris-HCI pH 8.8 at 25°C, 20 mM (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, 0.01% Tween-20, MBI Fermentas], 1.5 mM MgCl<sub>2</sub>, and 2 units Taq polymerase (MBI Fermentas) in a thermal cycler (Techne, UK). PCR conditions consisted of 94° for, 5 min, followed by 35 cycles of 30 sec at 94°C, 30 sec at 62°C, 30 sec at 72°C, with a final extension step at 72°C for 5 min. Amplification products (333 bp) were observed in a 2% agarose gel. The PCR product (10  $\mu$ l) was digested with 10 units of HaeII restriction enzyme (New England Biolabs, Beverly, MA) overnight. The fragments were separated in a 2.5% agarose gel. The SULT1A1\*1/SULT1A1\*1 (Arg/Arg<sup>213</sup>) genotype yielded two distinct digestion products (168 and 165 bp), the SULT1A1\*1/SULT1A1\*2 (Arg/His<sup>213</sup>) genotype yielded three distinct digestion products (333, 168 and 165 bp), and the SULT1A1\*2/SULT1A1\*2 (His/His<sup>213</sup>) genotype yielded no digestion products (333 bp) (Fig. 1).

Statistical Package for the Social Sciences (SPSS) release 10.0.1 software was used to perform the statistical analyses. Hardy-Weinberg equilibrium, genotype frequencies and allele frequencies were tested by the Pearson's  $\chi^2$  test. The statistical significance of the differences in SULT1A1 Arg<sup>213</sup>His genotypes among the cases and controls was determined by the  $\chi^2$  test. Probability values <0.05 were regarded as statistically significant. Odds ratios and 95% confidence intervals (CIs) for prostate cancer were calculated by using a multivariate logistic regression analysis adjusting several confounding variables such as age and smoking status.

## Results

Demographic characteristics of the cases and controls are summarized in Table I. Mean ages of the cases and controls were  $65.2\pm13.2$  years (range, 42-88) and  $61\pm11.4$  years (range, 41-82), respectively. No significant relationship was found between the cases and controls in terms of smoking status (P=0.83). Mean PSA levels were  $3.4\pm0.5$  and  $30.6\pm11.2$  ng/ml in the controls and patients, respectively.

SULT1A1 Arg<sup>213</sup>His allele and genotype frequencies are indicated in Table II. The genotype and allele frequencies were found to be in Hardy-Weinberg equilibrium. In the cases, the frequencies of the homozygous wild-type genotypes (Arg/Arg<sup>213</sup>), the heterozygous genotype (Arg/His<sup>213</sup>) and the homozygous variant genotype (His/His<sup>213</sup>) were 52.8, 36.6 and 10.6%, respectively; in the controls, these frequencies were 60.3, 35.8 and 3.9%, respectively.

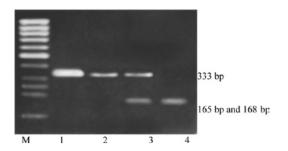


Figure 1. SULT1A1 genotypes determined by PCR-RFLP analysis. Lanes 1 and 2, SULT1A1\*2/SULT1A1\*2 genotype; lane 3, SULT1A1\*1/SULT1A1\*2 genotype; lane 4, SULT1A1\*1/SULT1A1\*1 genotype; and M, molecular weight marker (50 bp DNA ladder, Fermentas).

Table I. Characteristics of the prostate cancer patients and controls.

	Controls		Patients	
Sample size (n)				
Males	151		104	
Age (year)				
Range	41-82		42-88	
Mean ± SD	61±11.4		65.2±13.2	
Smoking history, n (%)				
Smokers	79 (52.3)		53 (50.9)	
Non-smokers	72 (47.7)		51 (49.1)	
$\chi^2$		0.045		
P-value <sup>a</sup>		0.830		
PSA (ng/ml)	3.4±0.5		30.6±11.2	

<sup>a</sup>P-values were calculated using the  $\chi^2$  test.

The risk of prostate cancer in individuals carrying the  $His^{213}$  allele was determined by combining Arg/His<sup>213</sup> and  $His/His^{213}$  genotypes. No statistically significant difference was found between the cases and controls in comparison of the genotype combination (P=0.24; OR, 1.36; 95% CI, 0.84-2.25). Concerning the smoker and non-smoker populations, no significant relationship was evident between the case and control groups regarding genotypic combinations (P=0.45; OR, 1.32 and P=0.34; OR, 1.39, respectively) (Table III).

## Discussion

The incidence of prostate cancer displays large ethnic variations worldwide. While the lowest incidence rate of prostate cancer is observed for Chinese men, African-American men have the highest rate of incidence (7). It is believed that advanced age, an intact androgen metabolism, ethnicity and genetic background are risk factors for prostate cancer development (8). The majority of studies suggest that genetic polymorphisms in xenobiotic metabolizing enzymes may play an important role in the susceptibility of individuals to cancer (18). In our previous study, we found that the GSTM1



Table II. Genotype and allele frequencies for SULTA1 locus in cases and controls.

Genotype	Controls (n=151)	Cancer patients (n=104)	P-value	$\chi^2$
Allele frequency, n (%)			0.072	3.23
Arg allele	236 (78.1)	148 (71.1)		
His allele	66 (21.9)	60 (38.9)		
Genotype frequency, n (%)			0.099	4.62
Arg/Arg	91 (60.3)	55 (52.8)		
Arg/His	54 (35.8)	38 (36.6)		
His/His	6 (3.9)	11 (10.6)		

Table III. Risk estimates for SULTA1 genotypes categorized according to total cases and smoking status.

Variable	Genotype combinations	Controls n (%)	Cancer patients n (%)	$\chi^2$	P-value	<sup>a</sup> OR (95% CI)
Total	Arg/Arg	91 (60.2)	55 (52.8)	1.37	0.24	1.36 (0.84-2.25)
	Arg/His + His/His	60 (39.8)	49 (47.2)			
Smoking status						
Smokers	Arg/Arg	44 (29.1)	26 (25)	0.56	0.45	1.32 (0.68-2.65)
	Arg/His + His/His	35 (23.17)	27 (25.9)			
Non-smokers	Arg/Arg	47 (31.12)	29 (27.8)	0.89	0.34	1.39 (0.62-2.91)
	Arg/His + His/His	25 (16.55)	22 (21.1)			

null genotype may play an important role as a risk factor for prostate cancer development in the Turkish population (14).

Allelic frequencies of the His<sup>213</sup> allele differ ranging from 5 to 32% among ethnic populations (5,19,20). The frequency of the His<sup>213</sup> allele was reported to be 18.5 and 22% in a study of a Turkish population (21,22). In the present study, the frequency of this allele in the control population was determined to be 21.9% which was higher than the frequency in Chinese, Taiwanese and Koreans while lower than the frequency reported for Caucasian and Nigerian populations (5,19,20). Distributions of the SULT1A1 Arg<sup>213</sup>His genotypes and alleles were also not significantly different between the cases and controls in the present study.

In many studies, a significant relationship has been demonstrated between the His<sup>213</sup> allele and various cancer types, including gastric, lung, colorectal, and breast (9,10,13,21). In addition, a statistical significant association was noted between the His<sup>213</sup> allele and primary brain tumor and lung cancer incidence in our previous studies (21,22). However, in the present study, no significant relationship was determined between the His<sup>213</sup> allele and prostate cancer although His<sup>213</sup> allele frequencies were higher in the patients than in the controls. Another study which is in agreement with ours was reported by Steiner *et al* (15). In contrast, Nowell *et al* (11) found a positive association between the Arg<sup>213</sup> allele (rapid sulfation allele) and prostate cancer risk. These controversial results may be due to the dual role (bioactivation and detoxification) of SULT1A1 in the metabolism of various carcinogens (23). Although SULT1A1 is considered as a phase II enzyme, it has been demonstrated that this enzyme acts to bioactivate various pro-carcinogens and pro-mutagens such as dietary carcinogen 2-amino-1-methyl-6-phenyl-imidazo (4,5-b) pyridine which induces prostate tumors in rats (24). Chou et al (25) and Ozawa et al (26) found that SULT1A1 catalyzes the sulfation of N-hydroxy derivates of arylamines and heterocyclic amines, to form more reactive DNA adduct-forming compounds. In this context, differing environmental parameters may also influence the function of a given allele. Thus, a different genetic background and different carcinogen exposure may play an important role in the different risk estimates associated with polymorphisms. In light of this knowledge, we believe that the influence of the SULT1A1 polymorphism on cancer development risk may differ according to the type of exposed carcinogen.

In the present study, there was no statistically significant association between the SULT1A1 Arg<sup>213</sup>His polymorphism and smoking status of the study population which is in accordance with previously published studies (21,22).

In summary, our results suggest that the SULT1A1 Arg<sup>213</sup>His polymorphism does not play a role in prostate cancer susceptibility in the Turkish population. However, since this study is the first report carried out in a Turkish population, it may contribute to the understanding of the relationship between the SULT1A1 polymorphism and prostate cancer. In order to elucidate the role of genetic polymorphisms in carcinogen metabolizing enzymes in prostate cancer development more accurately, environmental exposure to specific carcinogens must be investigated in larger studies.

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#### References

- 1. Falany CN: Enzymology of human cytosolic sulfotransferases. FASEB J 11: 206-216, 1997.
- Glatt H, Engelke CE, Pabel U, Teubner W, Jones AL, Coughtrie MW, Andrae U, Falany CN and Meinl W: Sulfotransferases: genetics and role in toxicology. Toxicol Lett 112: 341-348, 2000.
- 3. Richard K, Hume R, Kaptein E, Stanley EL, Visser TJ and Coughtrie MW: Sulfation of thyroid hormone and dopamine during human development: ontogeny of phenol sulfotransferases and arylsulfatase in liver, lung, and brain. J Clin Endocrinol Metab 86: 2734-2742, 2001.
- 4. Blanchard RL, Freimuth RR, Buck J, Weinshilboum RM and Coughtrie MW: A proposed nomenclature system for the cytosolic sulfotransferase (SULT) superfamily. Pharmacogenetics 14: 199-211, 2004.
- Raftogianis RB, Wood TC, Otterness DM, Van Loon JA and Weinshilboum RM: Phenol sulfotransferase pharmacogenetics in human: association of common SULT1A1 alleles with TS PST phenotype. Biochem Biophys Res Commun 239: 298-304, 1997.
- Nowell S, Ambrosone CB, Ozawa S, MacLeod SL, Mrackova G, Williams S, Plaxco J, Kadlubar FF and Lang NP: Relationship of phenol sulfotransferase activity (SULT1A1) genotype to sulfotransferase phenotype in platelet cytosol. Pharmacogenetics 10: 789-797, 2000.
- National Cancer Institute: Annual Report to the Nation Finds Declines in Cancer Incidence and Death Rates; Special Feature Reveals Wide Variations in Lung Cancer Trends across States. www.cancer.gov/newscenter/pressreleases/2008/ reportnation2008release.
- 8. Dearnaley DP: Cancer of the prostate. BMJ 308: 780-784, 1994. Erratum in: BMJ 308: 975, 1994.
- 9. Bamber DE, Fryer AA, Strange RC, Elder JB, Deakin M, Rajagopal R, Fawole A, Gilissen RA, Campbell FC and Coughtrie MW: Phenol sulfotransferase SUL1A1\*1 genotype is associated with reduced risk of colorectal cancer. Pharmacogenetics 11: 679-685, 2001.
- Boccia S, Persiani R, La Torre G, Rausei S, Arzani D, Gianfagna F, Romano-Spica V, D'Ugo D and Ricciardi G: Sulfotransferase 1A1 polymorphism and gastric cancer risk: a pilot case-control study. Cancer Lett 229: 235-243, 2005.
- 11. Nowell S, Ratnasinghe DL, Ambrosone CB, Williams S, Teague-Ross T, Trimble L, Runnels G, Carrol A, Green B, Stone A, Johnson D, Greene G, Kadlubar FF and Lang NP: Association of SULT1A1 phenotype and genotype with prostate cancer risk in African-Americans and Caucasians. Cancer Epidemiol Biomarkers Prev 13: 270-276, 2004.

- 12. Peng CT, Chen JC, Yeh KT, Wang YF, Hou MF, Lee TP, Shih MC, Chang JY and Chang JG: The relationship among the polymorphisms of SULT1A1, 1A2 and different types of cancers in Taiwanese. Int J Mol Med 11: 85-89, 2003.
- Seth P, Lunetta KL, Bell DW, Gray H, Nasser SM, Rhei E, Kaelin CM, Iglehart DJ, Marks JR, Garber JE, Haber DA and Polyak K: Phenol sulfotransferases: hormonal regulation, polymorphism, and age of onset of breast cancer. Cancer Res 60: 6859-6863, 2000.
- Silig Y, Pinarbasi H, Güneş S, Ayan S, Bagci H and Çetinkaya Ö: Polymorphisms of CYP1A1, GSTM1, GSTT1, and prostate cancer risk in Turkish population. Cancer Invest 24: 41-45, 2006.
- Steiner M, Bastian M, Schulz WA, Pulte T, Franke KH, Röhring A, Wolff JM, Seiter H and Schuff-Werner P: Phenol sulphotransferase SULT1A1 polymorphism in prostate cancer: lack of association. Arch Toxicol 74: 222-225, 2000.
- 16. Zheng W, Xie D, Cerhan JR, Sellers TA, Wen W and Folsom AR: Sulfotransferase 1A1 polymorphism, endogenous estrogen exposure, well-done meat-intake, and breast cancer risk. Cancer Epidemiol Biomarkers Prev 10: 89-94, 2001.
- Sambrook J, Fritsch E and Maniatis T (eds): Molecular Cloning A Laboratory Manual. 2nd edition, Cold Spring Harbor Laboratory Press, Plainview, 1989.
- Wormhoudt LW, Commandeur JN and Vermeulen NP: Genetic polymorphisms of human N-acetyltransferase, cytochrome P450, glutathione-S-transferase, and epoxide hydrolase enzymes: relevance to xenobiotic metabolism and toxicity. Crit Rev Toxicol 29: 59-124, 1999.
- Coughtrie MW, Gilissen RA, Shek B, Strange RC, Fryer AA, Jones PW and Bamber DE: Phenol sulphotransferase SULT1A1 polymorphism: molecular diagnosis and allele frequencies in Caucasian and African populations. Biochem J 337: 45-49, 1999.
- Arslan S, Siliğ Y and Pinarbaşı H: An investigation of the relationship between SULT1A1 Arg<sup>213</sup>His polymorphism and lung cancer susceptibility in a Turkish population. Cell Biochem Funct 27: 1-5, 2009.
- 21. Kim KA, Lee SY, Park PW, Ha JM and Park JY: Genetic polymorphisms and linkage disequilibrium of sulfotransferase SULT1A1 and SULT1A2 in a Korean population: comparison of other ethnic groups. Eur J Clin Pharmacol 61: 743-747, 2005.
- 22. Bardakci F, Arslan S, Bardakci S, Binatli AO and Budak M: Sulfotransferase 1A1 (SULT1A1) polymorphism and susceptibility to primary brain tumors. J Cancer Res Clin Oncol 134: 109-114, 2008.
- Banoglu E: Current status of the cytosolic sulfotransferases in the metabolic activation of promutagens and procarcinogens. Curr Drug Metab 1: 1-30, 2000.
- Shirai T, Tamano S, Sano M, Masui T, Hasegawa R and Ito N: Carcinogenicity of 2-amino-1-methyl-6-phenylimidazo[4,5-b] pyridine (PhIP) in rats: dose-response studies. Princess Takamatsu Symp 23: 232-239, 1995.
  Chou HC, Lang NP and Kadlubar FF: Metabolic activation of
- Chou HC, Lang NP and Kadlubar FF: Metabolic activation of the N-hydroxy derivative of the carcinogen 4-aminobiphenyl by human tissue sulfotransferases. Carcinogenesis 16: 413-417, 1995.
- 26. Ozawa S, Chou HC, Kadlubar FF, Nagata K, Yamazoe Y and Kato R: Activation of 2-hydroxyamino-1-methyl-6-phenylimidazo[4,5-b] pyridine by cDNA-expressed human and rat arylsulfotransferases. Jpn J Cancer Res 85: 1220-1228, 1994.