

Bathing effects of east saline groundwater concentrates on allergic (atopic) dermatitis-like skin lesions induced by 2,4-dinitrochlorobenzene in hairless mice

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Abstract. In the present study, it was evaluated whether east saline groundwater concentration solution (ESGWc) exerted a favorable inhibitory effect on 2,4-dinitrochlorobenzene (DNCB)-induced allergic/atopic-like dermatitis (AD). AD was induced and boosted by sensitization with DNCB via topical application on the dorsal back skins. Mice with DNCB-induced AD were bathed in 100-, 200- and 400-fold diluted ESGWc. After 6 weeks bathing, changes to body weight, clinical skin severity scores, scratching behavior, serum total immunoglobulin (Ig)E levels, submandibular lymph node and spleen weights, splenic cytokine levels, skin cytokine mRNA expressions, antioxidant defense systems and superoxide anion productions were recorded to determine the effects of bathing on the histopathology of dorsal back skin tissues. All DNCB-induced mice demonstrated that the induction of AD through IgE-mediated hypersensitivities, oxidative stresses, activation of MMP and apoptosis of keratinocytes resulted in no significant differences in body weight between the different groups at each time point following initial sensitization.

However, markers of DNCB-induced AD were significantly inhibited ($P<0.05$) in a concentration-dependent manner following bathing in all concentrations of ESGWc. The results obtained in the present study suggest that bathing in ESGWc may have favorable protective effects against DNCB-induced AD due to favorable systemic and local immunomodulatory effects, active cytoprotective anti-apoptotic effects, inhibitory effects of matrix metalloproteinase activity, and anti-inflammatory and antioxidative effects.

Introduction

Allergic/atopic dermatitis (AD) is characterized by various symptoms, including itching, erythema, eczema skin lesions, chronic relapse and pruritus (1). Patients with AD, may experience itchy skin, inducing scratching behavior and increasing the induction of pro-inflammatory cytokines (1,2). This, in turn, triggers immune cells and activates an inflammatory cycle of allergic dermatitis that is accompanied by erythema, keratosis and scaling of the skin (2). Due to an imbalanced immune response to various allergens (3), there are a number of symptoms, such as erythema, keratosis and scaling, that occur during AD. Another characteristic of the allergic immune system is the ability to produce a greater number of immunoglobulin E (IgE) antibodies and type 2 helper T cells (Th2), which are pivotal for IgE synthesis (4,5). Th2 cells predominantly induce the production of interleukin (IL) families and these cytokines are associated with the specific function of immune cells in AD (5). IgE is used as an indicator of AD severity and abnormalities of the skin barrier, a key feature of AD (6). During allergic inflammation, IgE and IgE-mediated mast cell and eosinophil activation contribute to AD (4,5). In addition, IgE is able to sensitize mast cells in the skin when cell-bound IgE is cross-linked by allergens, culminating in the production of inflammatory mediators, such as interleukins (IL)-4, -5 and -13 and tumor necrosis factor (TNF)- α (5). The cytokines IL-4 and IL-13 released by mast cells contribute to

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the Th2 response (5). TNF- α produced by macrophages also serves an important role in the acute phase of AD (7) and it has been demonstrated that oxidative stress is involved in the pathogenesis of AD (3,8).

To relieve the symptoms of AD, adrenocorticosteroids and anti-histamine agents have been utilized due to their ameliorating effects. However, they evoke severe side effects (9), therefore alternative therapies and natural products have been investigated for the treatment of AD (3,6). Hydrotherapy is regarded as a complementary therapy for AD in the therapeutic field (10). A number of studies have investigated the association between different water baths and their beneficial effects on dermatological disorders (11,12). Different types of water have demonstrated unique and distinct characteristics, although each water type differs in composition (13). Hydrotherapy may regulate lymphocyte proliferation and cytokine production (13) due to the antioxidant effects of the mineral composition of different types of water (14,15). Various seawaters exhibit protective effects against various types of dermatitis (16,17). Notably, the present study is a follow-up to a previous study by the present authors, in which it was demonstrated that bathing in four different types of seawater collected around Korea induces favorable systemic and local immunomodulatory effects, active cytoprotective anti-apoptotic effects, inhibitory effects of matrix metalloproteinase (MMP) activity, anti-inflammatory and antioxidative effects against 2,4-dinitrochlorobenzene (DNCB)-induced AD (18). The four types of water included: West surface seawater (WSSW) collected around Wepo-ri, Ganghwa-do, Korea; west saline groundwater (WSGW) collected at Yonggungoncheon, Seokmo-do, Korea; east surface seawater (ESSW) collected around Nagok-ri, Uljin, Korea; and east saline groundwater (ESGW) collected around Hoojeong-ri, Uljin, Korea (18). The greatest protective effects were demonstrated by ESGW, followed by WSGW, ESSW and WSSW, respectively (18).

As ESGW demonstrated the most protective effect among the four different types of seawater collected around Korea (18), the objective of the present study was to evaluate the potential concentration-dependent inhibitory effect of three different dilutions (100-, 200- and 400-fold dilutions) of ESGW concentration solution (50X; ESGWc) on DNCB-induced AD-like skin lesions in hairless mice following 6 weeks of bathing (once daily for 42 days for 20 min/day). Bathing is regarded as a method of developing a novel alternative therapy for AD patients (10); in the present study, it was used to determine the inhibitory effect of ESGWc against DNCB-induced AD-like skin lesions. Results were compared with those of 1% dexamethasone (DEXA) topically applied to the mice (once a day for 42 days) as a control treatment (18). Among animal models, DNCB-stimulated mice exhibit diverse symptoms similar to those of AD, including skin infiltration of inflammatory cells and increases in mast cell numbers and serum IgE levels (19). Therefore, a hairless murine model of DNCB-induced AD was selected to identify the bathing effects of various types of seawaters.

Materials and methods

Animal model. A total of 110 6-week SKH-1 female hairless mice were obtained from Orient Bio, Ltd., (Seungnam, Korea; body weight ranged in 20~23 g upon receipt) and 6 groups

(each, n=8) were selected 5 weeks following DNCB sensitization based on body weight (intact control: 27.51 \pm 1.07 g; DNCB treated mice: 27.37 \pm 0.87 g), clinical skin severity scores (intact control: 1.00 \pm 0.76 scores; DNCB treated mice: 12.45 \pm 1.01 scores) and scratching behavior (intact control: 11.88 \pm 5.46 frequencies/30 min; DNCB treated mice: 478.93 \pm 94.64 frequencies/30 min). Animals were acclimatized for 10 days and placed in polycarbonate cages (4 mice/cage) in a temperature (20-25°C) and humidity (50-55%) controlled room. Mice were housed under a 12 h light/dark cycle and standard rodent chow (Samyang Corp., Seoul, Korea) and water were available *ad libitum*. All laboratory animals were treated according to the national regulations of the usage and welfare of laboratory animals, and approved by the Institutional Animal Care and Use Committee of Daegu Haany University (Gyeongsan, Korea; approval no. DHU2015-068). The mice were subdivided into six groups, comprising three control groups (intact, DNCB treatment for negative control and DEXA treatment for reference control) and three treatment groups (ESGWc 100-, 200- and 400-fold treatment).

Preparation of seawaters and DEXA. Colorless clear solutions of ESGWc were prepared by AriBio Inc. (Seoul, Korea). Briefly, ESGW was collected around Hoojeong-ri (Gyeongsang, Korea) and a 50-fold concentrated solution was created using a rotary evaporator (N-1200B; Tokyo Rikakikai Co., Ltd., Tokyo, Japan). In addition, appropriate amounts of Ca-lactate and NaCl were separately prepared to supplement for the reduction in minerals during the concentration process. Salinity and mineral compositions of ESGWc compared with ESGW and general underground water are listed in Table I. White powders of water-soluble DEXA (Sigma-Aldrich; Merck KGaA, Darmstadt, Germany) were obtained and used in the present study as a potent reference agent. All test materials used in the present experiment were stored at 4°C in a refrigerator to protect from light and humidity until usage.

Induction of AD. AD-like dermatitis was induced in mice by sensitization with 10 mg/ml DNCB (Sigma-Aldrich; Merck KGaA; dissolved in a 3:1 mixture of acetone and olive oil) once a day for 1 week and boosted by 5 mg/ml DNCB three times a week for 28 days, according to established previous methods (3,18). DNCB solutions were topically applied on the dorsal back skins of mice in a volume of 200 μ l/mouse. In control mice, vehicle (3:1 mixtures of acetone and olive oil) was topically applied, instead of DNCB solution.

Bathing and topical application of DEXA. ESGWc 100-fold dilutions (2-fold dilutions of original ESGW) were selected as the highest concentrations based on a previous study investigating the effects of ESGW on DNCB hairless mice, in which favorable anti-AD effects were demonstrated at this concentration (18). In addition, 200- and 400-fold dilutions were selected as the middle and lowest concentrations, respectively, using common ratio 2. ESGWc were 100-, 200- and 400-fold diluted with distilled water, using 10, 5 and 2.5 ml ESGWc in 990, 995 and 997.5 ml of distilled water, respectively. Appropriate amounts of Ca-lactate and NaCl were added to supplement for the reduction in minerals during the concentration process with a Ca:Mg balance, salinity and conductance adjustment of

Table I. Salinity and mineral compositions of different seawaters.

Seawater	Salinity, %	Conductance, ms/cm	Balance, Ca:Mg	Mineral compositions, mg/l			
				Ca	Mg	K	Na
GUGW	-	-	0.67:1	81.00	121.00	12.00	569.00
ESGW	24.20	38.02	1.80:1	1,720.00	950.00	100.00	6,000.00
ESGWc	-	-	1.19:1	53,707.00	45,080.00	2,800.00	15,000.00

GUGW, general underground water; ESGW, east saline groundwater; ESGWc, east saline groundwater concentration solution (50X).

2.11 and 7.40, 1.06 and 3.70, 0.53 and 1.85 g in 100-, 200- and 400-fold dilutions of ESGWc, respectively. All mice in the ESGWc groups were allowed to swim freely in the ESGWc in mouse polycarbonate cages (200x260x130 mm; DJ-101; Daejong Instrument Industry Co., Seoul, Korea), which contained ~1,900 ml of warm water at ~37°C and bottom plates 4 cm deep, for 20 min/day.

DEXA treatment was performed as follows: Water-soluble DEXA was dissolved in distilled water to form a 1% solution and topically applied to the dorsal back skins (200 µl/mouse) once daily for 6 weeks, 5 weeks after initial DNCB sensitization. Intact and DNCB control mice bathed in distilled water instead of test waters in the present experiment to induce the same swimming stresses. Test waters and 1% DEXA solutions were warmed to ~37°C at least 30 min prior to bathing to avoid cooling irritations. A total bathing period of 6 weeks was selected for the present study, based on previous findings of the bathing effects of mineral-rich water (20) and the results of a previous study into the bathing effects of various seawaters collected around Korea (18).

Changes in body weight. Changes in body weight were measured once a week, from 1 day prior to initial DNCB sensitization to the end of the 6-week bathing period in the three different concentrations of ESGWc, distilled water or topical application of 1% DEXA, using an automatic electronic balance (Precisa Gravimetrics AG, Dietikon, Switzerland). To reduce individual differences, the total body weight gains during the 11-week whole experimental period and the total body weight gains during the 6-week bathing or topical application of 1% DEXA period were calculated.

Evaluation of clinical skin severity scores. Five signs of skin lesions, including pruritus/itching, erythema/hemorrhage, edema, excoriation/erosion and scaling/dryness were graded as follows: 0, (no symptoms); 1, (mild); 2, (moderate); and 3, (severe). Total scores (maximum=15) were regarded as the clinical skin severity score, in accordance with previous reports (3,18), with some modifications. Scoring was conducted once a week for 6 weeks, starting 5 weeks after DNCB sensitization and ending 24 h after the final bathing (11 weeks after DNCB sensitization). An additional scoring was conducted 3 days after initial bathing (38 days after initial DNCB sensitization).

Evaluation of scratching behavior. Each mouse from each group was placed individually in a routine polycarbonate

mouse cage and their behavior was monitored for 30 min once a week, from 5 weeks after initial DNCB sensitization to 24 h after final bathing (11 weeks after first DNCB sensitization) including 3 days after initial bathing (38 days after first DNCB sensitization), respectively. Scratching of the rostral back and biting of the caudal back were observed; scratching movements by the hind paw were defined as a scratching bout that ended when the mouse either licked its hind paw or placed its hind paw back on the floor. A series of one or more biting movements was counted as one episode that ended when the mouse returned to the straight-forward position (3).

Serum total IgE level measurement. At 6 weeks after the initial bathing, ~1 ml venous blood was collected from the vena cava under anesthesia with 2 to 3% isoflurane (Hana Pharm Co., Ltd., Hwasung, Korea) and maintained with 1 to 1.5% isoflurane in a mixture of 70% N₂O and 28.5% O₂, and serum was separated by centrifugation at 21,000 x g for 10 min at 4°C using a clotting-activated serum tube (BD Biosciences, San Jose, CA, USA). Total IgE levels in sera were determined by sandwich ELISA using the mouse IgE ELISA kit (cat. no. 555248, BD Biosciences), according to previous methods (3,6,18). Briefly, plates were coated with capture antibody in ELISA coating buffer and incubated overnight at 4°C. Subsequently, plates were washed with phosphate-buffered saline (PBS)-Tween-20 (0.05%) and blocked with 10% fetal bovine serum (FBS; Gibco; Thermo Fisher Scientific, Inc., Waltham, MA, USA) in PBS for 1 h at 20°C. Serial dilutions of standard antigen or sample in dilution buffer (10% FBS in PBS) were added to the plates and plates were incubated for 2 h at 20°C. Following three washes with PBS-Tween-20 (0.05%), biotin-conjugated anti-mouse IgE and streptavidin-horseradish peroxidase conjugate were added to the plates, according to the ELISA kit protocols, and plates were incubated for 1 h at 20°C. Finally, tetramethylbenzidine substrate solution was added to the plates and following 15-min incubation in the dark, a 2 N H₂SO₄ solution was added to stop the reaction. Optical densities were measured at 450 nm using an automated ELISA plate reader (Tecan Group, Ltd., Männedorf, Switzerland).

Lymphatic organ weight measurements. At 24 h after the end of the forty second bath, the mice were sacrificed by exsanguination through vena cava under anesthesia with 3% isoflurane and dissected. The spleen and left submandibular lymph node (LN) in each mouse were collected following

elimination of the surrounding connective tissues, muscles and debris. Individual weights of lymphatic organs were measured in grams to record absolute wet-weight. To reduce the individual body weight differences, the relative weight (% of body weight) of lymphatic organs was calculated using body weight at sacrifice and absolute organ weight according to previously established methods (18).

Splenic cytokine content measurements. Splenic concentrations of TNF- α , IL-1 β and IL-10 were measured by ELISA using commercially available kits, including a mouse TNF- α ELISA kit (cat. no. 558534, BD Biosciences), a mouse IL-1 β ELISA kit (cat. no. MBS824757) and a mouse IL-10 ELISA kit (cat. no. MBS8244590; both MyBioSource, Inc., San Diego, CA, USA), respectively, as previously described (18). Tissue samples (10–15 mg) were homogenized in a tissue grinder containing 1 ml lysis buffer (PBS containing 2 mM phenylmethylsulfonyl fluoride and 1 mg/ml aprotinin, leupeptin, and pepstatin A, all obtained from Sigma-Aldrich; Merck KGaA), as described by Clark *et al.* (21). Analysis was performed with 100 μ l standard (diluted in lysis buffer) or 10, 50 or 100 μ l tissue homogenate. Each sample was run in duplicate and a portion of the sample was analyzed for protein. Data were expressed as pg/mg of protein. For each assay, a standard curve was generated and based on replicates of the measured absorbance, demonstrated an average coefficient of variance of <10%.

Reverse transcription-quantitative polymerase chain reaction (RT-qPCR). Total RNA was extracted from skin tissue samples using TRIzol[®] reagent (Invitrogen; Thermo Fisher Scientific, Inc.), according to a previously described method (18). cDNA was synthesized from 2 μ g total RNA in a final reaction volume of 20 μ l. RNA was reverse transcribed using a High-Capacity cDNA Reverse Transcription kit (cat. no. 43-749-67; Applied Biosystems; Thermo Fisher Scientific, Inc.), according to the manufacturer's instructions. Briefly, a mixture of 2 μ l RT buffer, 0.8 μ l dNTP mix, 2 μ l RT primers, 1 μ l MultiScribe[™] reverse transcriptase, 1 μ l RNase inhibitor and 3.2 μ l nuclease-free distilled water was prepared and mixed with 10 μ l RNA sample. Reverse transcription was performed as follows: 25°C for 10 min, 37°C for 120 min, 85°C for 5 min. RT-qPCR analysis was carried out with a CFX96[™] Real-Time System (Bio-Rad Laboratories, Inc., Hercules, CA, USA) using iTaq[™] SYBR-Green (Bio-Rad Laboratories, Inc.). Nuclear free water, forward primer (10 μ M), reverse primer (10 μ M) and SYBR Green were mixed with cDNA in a 10 μ l reaction volume. The PCR cycling conditions included an initial pre-denaturation of 95°C for 1 min, denaturation for 15 sec, annealing of 55–65°C for 20 sec, and extension of 72°C for 30 sec. A total of 50 cycles were performed. All reactions were done in triplicate. The expression of GAPDH mRNA was used as a control for tissue integrity in all samples. The sequences of the PCR oligonucleotide primers for TNF- α , IL-4, IL-5, IL-13 and GAPDH are listed in Table II. For quantitative analysis, the intact control skin tissue was used as the control, and the relative expression of TNF- α , IL-4, IL-5 and IL-13 was calculated using the $2^{-\Delta\Delta C_q}$ method (22).

Glutathione (GSH) assay. Cutaneous GSH levels were determined using a fluorescence assay, as previously described (23).

Firstly, skin (1:3, w/w dilution) was homogenized in 100 mM NaH₂PO₄ (pH 8.0; Sigma-Aldrich; Merck KGaA) containing 5 mM EDTA (buffer 1). Homogenates were then treated with 30% trichloroacetic acid (Sigma-Aldrich; Merck KGaA) and centrifuged twice, at 1,940 \times g for 6 min at 4°C and at 485 \times g for 10 min at 4°C and the fluorescence of the resulting supernatant was measured using a fluorescence spectrophotometer (RF-5301PC; Shimadzu Corp., Kyoto, Japan). Briefly, 100 μ l supernatant was mixed with 1 ml buffer 1 and 100 μ l o-phthalaldehyde (1 mg/ml in methanol; Sigma-Aldrich; Merck KGaA). Fluorescence was determined after 15 min (k_{exc} =350 nm; k_{em} =420 nm). The standard curve was prepared with various concentrations of GSH (0.0–75.0 μ M). Protein levels in the skin homogenates were measured using the method described in a study by Lowry *et al.* (24). Results were presented as μ M GSH/mg of protein.

Lipid peroxidation. Firstly, the protein content of homogenate (10 mg/ml in 1.15% KCl) was measured using the method described by Lowry *et al.* (24). The thiobarbituric acid reactive substances (TBARS) measurement was used to evaluate lipid peroxidation, as previously described (25). For this assay, 10% trichloroacetic acid (Sigma-Aldrich; Merck KGaA) was added to the homogenate to precipitate proteins. This mixture was then centrifuged for 3 min at 1,000 \times g at 4°C. The protein-free sample was extracted and 0.67% thiobarbituric acid (Sigma-Aldrich; Merck KGaA) was added. The mixture was kept in a water bath at 100°C for 15 min. Levels of malondialdehyde (MDA), an intermediate product of lipoperoxidation, were determined by difference between absorbances at 535 and 572 nm on a microplate spectrophotometer reader (Tecan Group, Ltd., Männedorf, Switzerland) and the results were reported as nM/mg of protein (18).

Superoxide anion production. The quantification of superoxide anion production in skin tissue homogenates (10 mg/ml in 1.15% KCl; tissue was obtained after sacrifice) was performed using the nitroblue tetrazolium (NBT) assay, as previously described (26). Briefly, 50 μ l homogenate was incubated with 100 μ l NBT (1 mg/ml; Sigma-Aldrich; Merck KGaA) in 96-well plates at 37°C for 1 h. Subsequently, the supernatant was carefully removed and the reduced formazan solubilized by adding 120 μ l 2 M KOH and 140 μ l dimethyl sulfoxide. NBT reduction was measured at 600 nm using a microplate spectrophotometer reader (Tecan Group, Ltd.). Protein content was used for data normalization.

Histopathology. Samples from dorsal back skins, spleen and left submandibular LN were separated and fixed in 10% neutral buffered formalin at room temperature for 24 h, embedded in paraffin, sectioned into 3–4- μ m sections and stained with hematoxylin and eosin (H&E) for general histopathology, Masson's trichrome (MT) for collagen fiber or toluidine blue for mast cells, according to previously described methods (18). Following this, the histopathological profiles of each sample were observed under a light microscope (Eclipse 80i; Nikon Corporation, Tokyo, Japan). Mean epithelial thicknesses of the epidermis (μ m), mean numbers of inflammatory and mast cells infiltrated in the dermis (cells/mm² of dermis), total splenic thicknesses (mm/central regions), mean numbers of white pulp

Table II. Oligonucleotides for reverse transcription-quantitative polymerase chain reaction.

Target	Direction	Sequence (5'-3')	Gene bank number
Tumor necrosis factor- α	Forward	CCTGTAGCCCACGTCGTAGC	NM_013693.2
	Reverse	5TTGACCTCAGCGCTGAGTTG	
Interleukin-4	Forward	GAATGTACCAGGAGCCATATC	NM_021283.2
	Reverse	CTCAGTACTACGAGTAATCCA	
Interleukin-5	Forward	TCACCGAGCTCTGTTGACAA	NM_010558.1
	Reverse	CCACACTTCTCTTTTGGCG	
Interleukin-13	Forward	GACCCAGAGGATATTGCATG	NM_008355.3
	Reverse	CCAGCAAAGTCTGATGTGAG	
Glyceraldehyde 3-phosphate dehydrogenase	Forward	CATCTTCCAGGAGCGAGACC	NM_008084.2
	Reverse	AAGCCATGCCAATGTTGTCT	

(white pulps/mm² of splenic parenchyma) and red pulp cells ($\times 10^3$ cells/mm² of splenic parenchyma), total submandibular LN thicknesses (mm/central regions), cortex lymphoid follicle numbers (follicles/mm² of cortex) and mean thicknesses (μ m/LN) of submandibular LN were calculated for general histomorphometrical analysis using a computer-assisted image analysis program (iSolution FL v. 9.1; IMT i-solution Inc., Irvine, CA, USA) under H&E stain and collagen fiber occupied regions in the dermis (%/mm² of dermis) under an MT stain, according to previously established methods (18,27). The histopathologist was blinded to the groups when this analysis was performed.

Immunohistochemistry. Following deparaffinization of prepared skin using xylene, submandibular LN or spleen histological paraffin sections, citrate buffer antigen (epitope) retrieval pretreatment was conducted, as previously described (18). Briefly, a water bath with a staining dish containing 10 mM citrate buffer (pH 6.0) was pre-heated until the temperature reached 95-100°C. Slides were immersed in the staining dish and the lid was placed loosely on the staining dish. Slides were incubated in the water bath for 20 min and then the water bath was turned off. Subsequently, the staining dish was placed at room temperature for 20°C to allow the slides to cool. Following epitope retrieval, sections were immunostained using avidin-biotin complex (ABC) methods for caspase-3, poly ADP ribose polymerase (PARP), nitrotyrosine (NT), 4-hydroxynonenal (4-HNE), MMP-9, interferon (IFN)- γ , inducible nitric oxide synthase (iNOS), IL-1 β , IL-2 and TNF- α , following protocols recorded in previous studies (18,27). Briefly, endogenous peroxidase activity was blocked by incubation in methanol and 0.3% H₂O₂ at room temperature for 30 min, and non-specific binding of immunoglobulin was blocked with 1% normal horse serum blocking solution (1:100; Vector Laboratories, Inc., Burlingame, CA, USA) at room temperature for 1 h in a humidity chamber. Primary antiserum (listed in Table III) was treated overnight at 4°C in a humidity chamber and then incubated with biotinylated universal secondary antibody (Vectastain Elite ABC kit; 1:50; Vector Laboratories, Inc.) and ABC reagents (Vectastain Elite ABC kit; 1:50; Vector Laboratories, Inc.) for 1 h at room temperature in a humidity chamber. Finally, a DAB peroxidase substrate kit (cat.

no. SK-4100; Vector Laboratories, Inc.) was applied for 3 min at room temperature. All sections were rinsed in 0.01 M PBS three times between steps. The cells or fibers occupied by >30% of immunoreactivities, the density of each antiserum (for caspase-3, PARP, NT, 4-HNE, MMP-9, IFN- γ , iNOS, IL-1 β , IL-2 and TNF- α) as compared with intact dermal keratinocytes or dermal fibers, were regarded as positive, and the mean numbers of caspase-3, PARP, NT and 4-HNE immunoreactive cells in the epidermis (cells/100 epithelial cells), and mean IFN- γ , iNOS, IL-1 β , IL-2 and TNF- α immunolabeled cell numbers in the dermis (cells/mm² of dermis), spleen (cells/mm² of spleen) and submandibular LN (cells/mm² of LN) were also counted using an automated image analysis process (iSolution FL v. 9.1; IMT i-Solution Inc., Irvine, CA, USA), as previously described (18,27) with some modifications. In addition, the occupied percentages by MMP-9 immunoreactive fibers were also calculated in the dermis (%/mm² of dermis) as MMP-9 immunoreactivities in the present experiment. The histopathologist was blinded to the group distribution when performing the analysis.

Statistical analyses. All data were expressed as the mean \pm standard deviation of eight hairless mice. Multiple comparison tests for different dose groups were conducted. Variance homogeneity was examined using Levene's tests. If the Levene's test indicated no significant deviations from variance homogeneity, the obtained data were analyzed by one way analysis of variance, followed by least-significant differences multi-comparison test to determine which pairs of group comparison were significantly different. In the case of significant deviations from variance homogeneity being observed in the Levene's test, a non-parametric comparison test, Kruskal-Wallis H test, was conducted. When a significant difference was observed with the Kruskal-Wallis H test, the Mann-Whitney U test was conducted to determine the specific pairs of group comparisons that were significantly different. Statistical analyses were conducted using SPSS v. 14.0 for Windows (SPSS, Inc., Chicago, IL, USA). In addition, the percentage changes between intact vehicle and DNCB control were calculated to observe the severities of AD-like lesions induced by DNCB in the present study, and the percentage changes as compared with DNCB control and hairless mice bathing in seawaters or receiving

Table III. Primary antiserum and detection kits used in this study.

Product	Catalog number	Supplier	Dilution
Primary antisera			
Anti-cleaved caspase-3 (Asp175) polyclonal antibody	9661	Cell Signaling Technology Inc., Danvers, MA, USA	1:400
Anti-cleaved poly(ADP-ribose) polymerase (Asp214) specific antibody	9545	Cell Signaling Technology Inc.	1:100
Anti-4-Hydroxynonenal polyclonal antibody	Ab46545	Abcam, Cambridge, UK	1:100
Anti-Nitrotyrosine polyclonal antibody	06-284	EMD Millipore, Billerica, MA, USA	1:200
Anti- Matrix metalloproteinase-9 mouse antibody	Ab38898	Abcam	1:100
Anti-interferon γ antibody	AF781	R&D Systems, Inc., Minneapolis, MN, USA	1:100
Anti-IL-2 antibody	BAF1815	R&D Systems, Inc.	1:100
Anti-tumor necrosis factor- α antibody	sc-52746	Santa Cruz Biotechnology, Inc., Dallas, TX, USA	1:200
Anti-nitric oxide synthase2 (N-20) polyclonal antibody	sc-651	Santa Cruz Biotechnology, Inc.	1:100
Anti-IL-1 β (H-153) polyclonal antibody	sc-7884	Santa Cruz Biotechnology, Inc.	1:100
Detection kits			
Vectastain Elite ABC Kit	PK-6200	Vector Laboratories, Inc., Burlingame, CA, USA	1:50
Peroxidase substrate kit	SK-4100	Vector Laboratories, Inc.	1:50

All antisera were diluted with 0.01 M phosphate buffered saline. IL, interleukin.

topical application of 1% DEXA were also calculated to aid the understanding of the efficacy of seawater treatment, as described previously (28).

Results

Changes in body weight. No significant differences in body weight gains were observed among any of the groups during the total 11-week experimental period. During the 6-week bathing period, DNCB control mice demonstrated similar body weights to the intact vehicle control mice, apart from significant decreases in body weight identified in the DNCB-sensitized (DEXA-treated and ESGWc 100-, 200-, or 400-fold) mice on days 6 and 7 days after initial DNCB sensitization compared with the intact vehicle control mice ($P < 0.05$). Topical application of 1% DEXA and bathing with all three concentrations of ESGWc did not significantly alter body weight compared with mice in the DNCB control group throughout the entire experimental period of the present experiment (Fig. 1).

Clinical skin severity score changes. DNCB control mice exhibited significant increases in clinical skin severity scores from 24 h before initial bathing up until the end of the experimental period compared with intact vehicle control mice ($P < 0.05$; Table IV). However, significant decreases in clinical

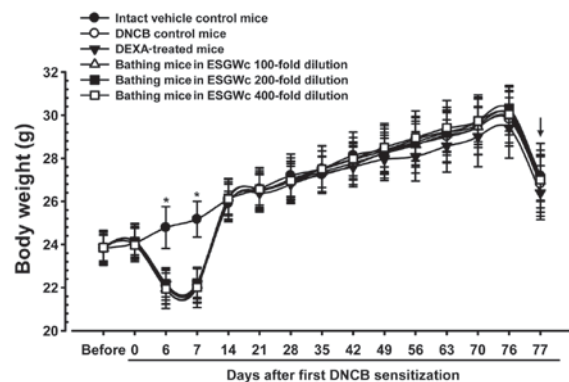


Figure 1. Body weight changes during 6 weeks bathing in ESGWc or topical application of 1% DEXA in mice with DNCB-induced AD. On day 7, DNCB boosting was initiated and bathing in ESGWc or application of DEXA began on day 35. The arrow indicated that all animals were fasted overnight on day 77 prior to sacrifice. Values are expressed as the mean \pm standard deviation ($n=8$). * $P < 0.05$ vs. all treatment groups. AD, allergic/atopic-like dermatitis; DNCB, 2,4-dinitrochlorobenzene; DEXA, dexamethasone; ESGWc, east saline groundwater concentration solution (50X); Before, 1 day before initial DNCB sensitization.

skin severity scores were observed in mice receiving topical application of 1% DEXA, from 1 week after initial topical application up until the end of the experiment, as compared with DNCB control mice ($P < 0.05$). Significant decreases in

Table IV. Changes to clinical skin severity scores during 6 weeks of bathing in ESGWc or topical application of DEXA in mice with DNCB-induced AD.

Days following initial DNCB sensitization	Controls			Bathing in ESGWc dilutions		
	Intact	DNCB	DEXA	100-fold	200-fold	400-fold
35	1.00±0.76	12.38±1.06 ^a	12.50±0.93 ^a	12.38±0.92 ^a	12.50±1.20 ^a	12.50±1.25 ^a
38	1.00±0.76	12.38±1.06 ^a	12.13±0.83 ^a	12.13±1.25 ^a	12.13±0.83 ^a	12.13±1.36 ^a
42	1.13±0.99	11.75±1.04 ^a	9.88±0.99 ^{a,b}	10.88±0.99 ^a	11.00±1.07 ^a	11.13±1.13 ^a
49	0.88±0.64	11.25±1.28 ^a	7.38±1.06 ^{a,b}	8.88±1.13 ^{a,b}	9.63±1.19 ^{a,b}	9.88±0.99 ^{a,b}
56	0.88±0.64	10.38±0.92 ^a	6.00±0.76 ^{a,b}	7.00±1.20 ^{a,b}	8.25±1.28 ^{a,b}	9.00±1.07 ^{a,b}
63	1.25±0.71	9.75±1.28 ^a	4.25±1.04 ^{a,b}	5.38±0.92 ^{a,b}	7.13±1.55 ^{a,b}	8.00±0.93 ^{a,b}
70	1.38±0.74	9.50±1.31 ^a	3.38±1.19 ^{a,b}	4.88±0.64 ^{a,b}	6.38±0.92 ^{a,b}	7.25±0.71 ^{a,b}
77	1.38±0.74	8.88±1.46 ^a	2.88±1.13 ^{a,b}	3.88±0.83 ^{a,b}	5.13±0.99 ^{a,b}	6.63±0.74 ^{a,b}

Values are expressed as the mean ± standard deviation (n=8). Maximum scores were 15. ^aP<0.05 vs. Intact control; ^bP<0.05 vs. DNCB control. AD, allergic/atopic-like dermatitis; DNCB, 2,4-dinitrochlorobenzene; DEXA, dexamethasone; ESGWc, east saline ground water concentration solution (50X).

Table V. Changes in scratching behaviors during 6 weeks of bathing in ESGWc or topical application of DEXA in mice with DNCB-induced AD.

Days after initial DNCB sensitization	Controls			Bathing in ESGWc dilutions		
	Intact	DNCB	DEXA	100-fold	200-fold	400-fold
35	11.88±5.46	477.63±101.18 ^a	478.88±77.58 ^a	478.75±113.11 ^a	476.13±105.78 ^a	483.25±98.13 ^a
38	10.88±4.05	474.25±72.20 ^a	452.63±68.02 ^a	459.38±105.89 ^a	442.63±87.99 ^a	453.63±79.81 ^a
42	12.63±3.70	467.50±73.14 ^a	366.50±50.54 ^{a,b}	407.25±88.80 ^a	419.88±79.74 ^a	420.13±68.88 ^a
49	13.38±4.00	442.38±73.82 ^a	281.50±34.79 ^{a,b}	341.25±72.07 ^{a,b}	364.50±54.56 ^{a,b}	373.38±37.08 ^{a,b}
56	14.50±3.70	424.75±72.51 ^a	204.50±20.35 ^{a,b}	255.38±53.83 ^{a,b}	320.75±60.23 ^{a,b}	339.38±33.87 ^{a,b}
63	14.88±4.16	405.88±73.21 ^a	188.38±16.65 ^{a,b}	204.13±29.86 ^{a,b}	241.75±51.25 ^{a,b}	280.63±44.71 ^{a,b}
70	15.63±3.93	384.38±54.37 ^a	146.13±32.39 ^{a,b}	174.88±33.47 ^{a,b}	195.13±20.70 ^{a,b}	252.38±47.88 ^{a,b}
77	18.25±4.77	380.25±49.69 ^a	76.25±23.66 ^{a,b}	138.88±42.58 ^{a,b}	169.88±23.04 ^{a,b}	218.25±46.37 ^{a,b}

Values are expressed as the mean ± standard deviation scratching behavior frequencies/30 min (n=8). ^aP<0.05 vs. Intact control; ^bP<0.05 vs. DNCB control. AD, allergic/atopic-like dermatitis; DNCB, 2,4-dinitrochlorobenzene; DEXA, dexamethasone; ESGWc, east saline ground water concentration solution (50X).

clinical skin severity scores were detected from 2 weeks after initial bathing in ESGWc 100-, 200- and 400-fold dilutions, as compared with DNCB control mice throughout the experimental period (P<0.05; Table IV).

Changes in scratching behavior. DNCB control mice exhibited significant increases in scratching behavior from 24 h before initial bathing up until the end of the experimental period, as compared with intact vehicle control mice (P<0.05; Table V). However, these increases in scratching behavior were significantly inhibited by topical application of 1% DEXA from 1 week after initial topical application until the end of the experimental period, compared with DNCB control mice (P<0.05). In addition, significant decreases in scratching behaviors were detected from 2 weeks after initial bathing in all three concentrations of ESGW in a concentration-dependent

manner, compared with DNCB control mice throughout the experimental period (P<0.05; Table V).

Effects on total serum IgE levels. Significant increases in total serum IgE levels were detected in DNCB control mice compared with intact vehicle control mice (P<0.05; Fig. 2). However, significant decreases of total serum IgE levels were detected by topical treatment of 1% DEXA and also by bathing in ESGWc at 100-, 200- and 400-fold dilutions, in a concentration-dependent manner, compared with DNCB control mice (P<0.05; Fig. 2).

Changes in submandibular LN and spleen weights. Significant increases of submandibular LN and spleen absolute and relative weights were detected in DNCB control mice compared with intact vehicle control mice (P<0.05; Table VI). However,

Table VI. Changes in lymphatic organ weights after 6 weeks of bathing in ESGWc or topical application of DEXA in mice with DNCB-induced AD.

Group	Absolute weight, g		Relative weight, % of body weight	
	Spleen	Submandibular LN	Spleen	Submandibular LN
Controls				
Intact	0.093±0.012	0.008±0.002	0.341±0.051	0.028±0.009
DNCB	0.242±0.023 ^a	0.020±0.002 ^a	0.905±0.099 ^a	0.074±0.008 ^a
Reference				
DEXA	0.125±0.017 ^{a,b}	0.011±0.003 ^{a,b}	0.474±0.081 ^{a,b}	0.040±0.013 ^{a,b}
ESGWc dilutions				
100-fold	0.162±0.013 ^{a,b}	0.013±0.002 ^{a,b}	0.604±0.057 ^{a,b}	0.049±0.009 ^{a,b}
200-fold	0.181±0.017 ^{a,b}	0.015±0.002 ^{a,b}	0.668±0.052 ^{a,b}	0.055±0.009 ^{a,b}
400-fold	0.198±0.014 ^{a,b}	0.016±0.001 ^{a,b}	0.739±0.090 ^{a,b}	0.060±0.006 ^{a,b}

Values are expressed as the mean ± standard deviation (n=8). ^aP<0.05 vs. Intact control; ^bP<0.05 vs. DNCB control. AD, allergic/atopic-like dermatitis; DNCB, 2,4-dinitrochlorobenzene; DEXA, dexamethasone; ESGWc, east saline ground water concentration solution (50X); LN, lymph node.

Table VII. Changes in splenic cytokine contents after 6 weeks of bathing in ESGWc or topical application of DEXA in mice with DNCB-induced AD.

Group	Splenic cytokine contents, pg/mg protein		
	Tumor necrosis factor- α	Interleukin-1 β	Interleukin-10
Controls			
Intact	173.33±31.33	53.97±16.10	263.06±128.36
DNCB	548.44±120.26 ^a	207.87±44.12 ^a	718.54±141.35 ^a
Reference			
DEXA	202.07±19.19 ^{a,b}	97.47±20.38 ^{a,b}	324.84±83.72 ^b
ESGWc dilutions			
100-fold	331.46±39.19 ^{a,b}	127.57±16.39 ^{a,b}	435.17±103.85 ^{a,b}
200-fold	369.29±51.93 ^{a,b}	136.52±19.78 ^{a,b}	493.76±96.21 ^{a,b}
400-fold	416.41±20.43 ^{a,b}	153.86±17.10 ^{a,b}	535.12±101.32 ^{a,b}

Values are expressed as the mean ± standard deviation (n=8). ^aP<0.05 vs. Intact control; ^bP<0.05 vs. DNCB control. AD, allergic/atopic-like dermatitis; DNCB, 2,4-dinitrochlorobenzene; DEXA, dexamethasone; ESGWc, east saline ground water concentration solution (50X).

significant decreases in both absolute and relative submandibular LN and spleen weights were detected following topical application of 1% DEXA, and also following bathing in all three ESGWc dilutions, concentration-dependently, compared with DNCB control mice (P<0.05; Table VI).

Effects on splenic cytokine contents. Significant increases in splenic tissue TNF- α , IL-1 β and IL-10 contents were detected in DNCB control mice compared with intact control vehicle mice (P<0.05). However, significant decreases in splenic tissue TNF- α , IL-1 β and IL-10 contents were detected following topical treatment of 1% DEXA and also concentration-dependent decreases were observed following bathing in ESGWc 100-, 200- and 400-fold dilutions, compared with DNCB control mice (P<0.05; Table VII).

Changes to skin tissue cytokine mRNA expression levels (RT-qPCR analysis). Significant increases in the expression of TNF- α , IL-4, IL-5 and IL-13 mRNA in the dorsal back skin tissue were detected in DNCB control mice compared with intact vehicle control mice according to RT-qPCR analysis (P<0.05; Table VIII). However, significant decreases in expression of TNF- α , IL-4, IL-5 and IL-13 mRNA were observed in skin tissues following topical treatment with 1% DEXA. Concentration-dependent decreases were also observed following bathing in the three dilutions of ESGWc, compared with DNCB control mice (P<0.05; Table VIII).

Effects on the skin tissue antioxidant defense systems. Significant decreases in dorsal back skin tissue GSH content and significant increases in skin lipid peroxidation and

Table VIII. Changes in skin mRNA expressions after 6 weeks of bathing in ESGWc or topical application of DEXA in mice with DNCB-induced AD.

Groups	Skin mRNA expressions (relative expressions/GAPDH mRNA)			
	TNF- α	IL-4	IL-5	IL-13
Controls				
Intact	0.98 \pm 0.08	1.03 \pm 0.13	1.01 \pm 0.09	0.99 \pm 0.07
DNCB	6.47 \pm 0.97 ^a	5.27 \pm 1.08 ^a	5.92 \pm 1.19 ^a	3.30 \pm 0.64 ^a
Reference				
DEXA	2.97 \pm 0.71 ^{a,b}	2.03 \pm 0.33 ^{a,b}	2.45 \pm 0.72 ^{a,b}	1.56 \pm 0.32 ^{a,b}
ESGWc dilutions				
100-fold	3.15 \pm 0.81 ^{a,b}	2.66 \pm 0.36 ^{a,b}	3.31 \pm 0.52 ^{a,b}	1.80 \pm 0.14 ^{a,b}
200-fold	3.93 \pm 0.80 ^{a,b}	3.30 \pm 0.56 ^{a,b}	3.79 \pm 0.62 ^{a,b}	1.95 \pm 0.13 ^{a,b}
400-fold	4.71 \pm 1.16 ^{a,b}	3.70 \pm 0.83 ^{a,b}	4.17 \pm 0.51 ^{a,b}	2.36 \pm 0.46 ^{a,b}

Values are expressed as the mean \pm standard deviation. (n=8). ^aP<0.05 vs. Intact control; ^bP<0.05 vs. DNCB control. AD, allergic/atopic-like dermatitis; DNCB, 2,4-dinitrochlorobenzene; DEXA, dexamethasone; ESGWc, east saline ground water concentration solution (50X); TNF, tumor necrosis factor; IL, interleukin.

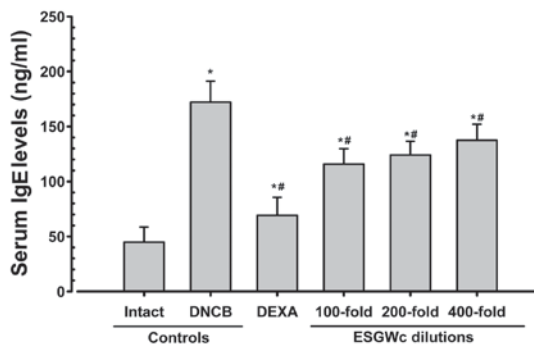


Figure 2. Changes in serum IgE levels following 6 weeks bathing in ESGWc or topical application of DEXA in mice with DNCB-induced AD. Values are expressed as the mean \pm standard deviation (n=8). ^{*}P<0.05 vs. Intact control; ^{**}P<0.05 vs. DNCB control. AD, allergic/atopic-like dermatitis; DNCB, 2,4-dinitrochlorobenzene; DEXA, dexamethasone; ESGWc, east saline groundwater concentration solution (50X); IgE, immunoglobulin E.

superoxide anion production were observed in DNCB control mice compared with intact vehicle control mice (P<0.05; Table IX). However, compared with DNCB control mice, there were significant increases in GSH content and significant decreases in lipid peroxidation and superoxide anion production in the skin tissues of mice bathing in ESGWc 100-, 200- and 400-fold dilutions, in a concentration-dependent manner (P<0.05). Topical application of 1% DEXA did not significantly influence skin tissue antioxidant defense systems compared with DNCB control mice (Table IX).

Histopathological changes on the dorsal back skin tissues. A significant increase in mean epithelial thickness due to hyperplasia/hypertrophy of epidermal keratinocytes was detected in the dorsal back skin tissues of DNCB control mice, with significant increases in the numbers of mast and inflammatory cells infiltrating into dermis, compared with mice in the intact vehicle control group (P<0.05; Table X; Fig. 3).

Abnormal collagen depositions, elevations of caspase-3, PARP, NT and 4-HNE immunoreactive epidermal cells, dermal MMP-9 immunoreactivities, infiltrations of dermal IFN- γ , iNOS, IL-1 β , IL-2 and TNF- α immunolabeled cells were also observed in the DNCB control group compared with the intact vehicle control group (Figs. 3-5). These results were confirmed by histomorphometrical analysis, which demonstrated significant increases in mean epithelial thickness, numbers of dermal infiltrated mast and inflammatory cells, percentages of collagen fiber-occupied dermal regions, caspase-3, PARP, NT and 4-HNE immunoreactive epidermal cells, dermal IFN- γ , iNOS, IL-1 β , IL-2 and TNF- α immunolabeled cells in DNCB control mice compared with intact vehicle control mice (all P<0.05; Table XI). However, these histopathological hypersensitivity-related AD signs were significantly inhibited by bathing in the ESGWc 100-, 200- and 400-fold dilutions, in a concentration-dependent manner, compared with DNCB control mice (P<0.05; Tables X and XI). Topical application of 1% DEXA also significantly reduced the increases of mean epithelial thickness, numbers of dermal infiltrated mast and inflammatory cells, caspase-3, PARP, NT and 4-HNE immunoreactive epidermal cells, dermal MMP-9 immunoreactivities, dermal IFN- γ , iNOS, IL-1 β , IL-2 and TNF- α immunolabeled cells induced by DNCB treatment (P<0.05; Tables X and XI); however, 1% DEXA did not significantly influence the percentage of collagen fiber-occupied dermal regions, as compared with DNCB control mice (Table X).

Histopathological changes in splenic tissues. Marked hypertrophic changes due to hyperplasia of red pulp lymphoid cells were detected in the splenic tissues of DNCB control mice, with marked increases of the numbers of IFN- γ , iNOS, IL-1 β , IL-2 and TNF- α immunolabeled cells (Figs. 6 and 7). These results were confirmed by histomorphometrical analysis, which demonstrated significant increases in total splenic thicknesses, numbers of red pulp lymphoid cells,

Table IX. Changes in skin antioxidant defense systems after 6 weeks of continuous bathing in ESGWc or topical application of DEXA in mice with DNCB-induced AD.

Groups	Skin antioxidant defense systems		
	Glutathione, $\mu\text{M}/\text{mg}$ of protein	Lipid peroxidation-Malondialdehyde, nM/mg of protein	Superoxide anion production, NBT reduction/OD at 600 nm
Controls			
Intact	1.56 \pm 0.32	0.44 \pm 0.16	0.42 \pm 0.19
DNCB	0.37 \pm 0.15 ^a	2.87 \pm 0.69 ^a	1.80 \pm 0.19 ^a
Reference			
DEXA	0.34 \pm 0.07 ^a	3.00 \pm 0.95 ^a	1.85 \pm 0.18 ^a
Bathing in ESGWc dilutions			
100-fold	0.97 \pm 0.17 ^{a,b}	1.26 \pm 0.27 ^{a,b}	0.91 \pm 0.14 ^{a,b}
200-fold	0.82 \pm 0.11 ^{a,b}	1.49 \pm 0.26 ^{a,b}	1.12 \pm 0.22 ^{a,b}
400-fold	0.68 \pm 0.11 ^{a,b}	1.83 \pm 0.20 ^{a,b}	1.26 \pm 0.20 ^{a,b}

Values are expressed as the mean \pm standard deviation (n=8). ^aP<0.05 vs. Intact control; ^bP<0.05 vs. DNCB control. AD, allergic/atopic-like dermatitis; DNCB, 2,4-dinitrochlorobenzene; DEXA, dexamethasone; ESGWc, east saline ground water concentration solution (50X); TNF, tumor necrosis factor; OD, optical density; NBT, nitroblue tetrazolium.

Table X. General skin tissue histomorphometry after 6 weeks of bathing in ESGWc or topical application of DEXA in mice with DNCB-induced AD.

Index	Controls			Bathing in ESGWc dilutions		
	Intact	DNCB	DEXA	100-fold	200-fold	400-fold
Epithelial Th, μm	34.69 \pm 8.73	117.28 \pm 19.66 ^a	37.07 \pm 7.07 ^{a,b}	48.49 \pm 11.00 ^{a,b}	54.71 \pm 12.06 ^{a,b}	70.53 \pm 12.16 ^{a,b}
Collagen fOP, %/mm ² of dermis	41.48 \pm 3.36	77.28 \pm 10.51 ^a	77.58 \pm 10.65 ^a	50.24 \pm 10.01 ^b	55.78 \pm 11.16 ^{a,b}	61.86 \pm 10.16 ^{a,b}
Infiltrated cells, cells/mm ² of dermis						
IF	27.13 \pm 14.81	409.00 \pm 76.80 ^a	74.75 \pm 27.06 ^{a,b}	132.75 \pm 33.48 ^{a,b}	211.50 \pm 44.26 ^{a,b}	283.25 \pm 34.05 ^{a,b}
Mast	134.34 \pm 40.05	842.75 \pm 109.18 ^a	216.75 \pm 44.52 ^b	381.88 \pm 136.05 ^{a,b}	483.75 \pm 77.75 ^{a,b}	570.25 \pm 111.70 ^{a,b}

Values are expressed as the mean \pm standard deviation (n=8). ^aP<0.05 vs. Intact control; ^bP<0.05 vs. DNCB control. AD, allergic/atopic-like dermatitis; DNCB, 2,4-dinitrochlorobenzene; DEXA, dexamethasone; ESGWc, east saline ground water concentration solution (50X); Th, thickness; fOP, fiber occupied region percentages; IF, inflammatory.

numbers of IFN- γ , iNOS, IL-1 β , IL-2 and TNF- α immunolabeled cells in DNCB control mice compared with intact vehicle control mice (P<0.05; Table XII). However, these hypersensitivity-related splenic hypertrophic signs were significantly inhibited following topical treatment with 1% DEXA, and also by bathing in the three dilutions of ESGWc, concentration-dependently, as compared with DNCB control mice (P<0.05). No significant changes in the number of white pulp cells were demonstrated following treatment with DNCB compared with the intact vehicle control mice, nor by topical application of 1% DEXA or bathing in ESGWc 100-, 200- and 400-fold dilutions, as compared with DNCB control mice (Table XII).

Histopathological changes in submandibular LN tissues. Marked hypertrophic changes due to hyperplasia of cortex lymphoid cells were detected in submandibular LN tissues in DNCB control mice, with marked increases of the numbers of IFN- γ , iNOS, IL-1 β , IL-2 and TNF- α immunolabeled cells (Figs. 8 and 9). These observations were confirmed by histomorphometrical analysis, which demonstrated significant increases in total submandibular LN thicknesses, numbers of cortex lymphoid follicles, cortex thicknesses, numbers of IFN- γ , iNOS, IL-1 β , IL-2 and TNF- α immunolabeled cells in submandibular LN tissues of DNCB control mice compared with intact vehicle control mice (P<0.05; Table XIII). However, these submandibular LN hypersensitivity-related hypertrophic signs were significantly inhibited by topical treatment of

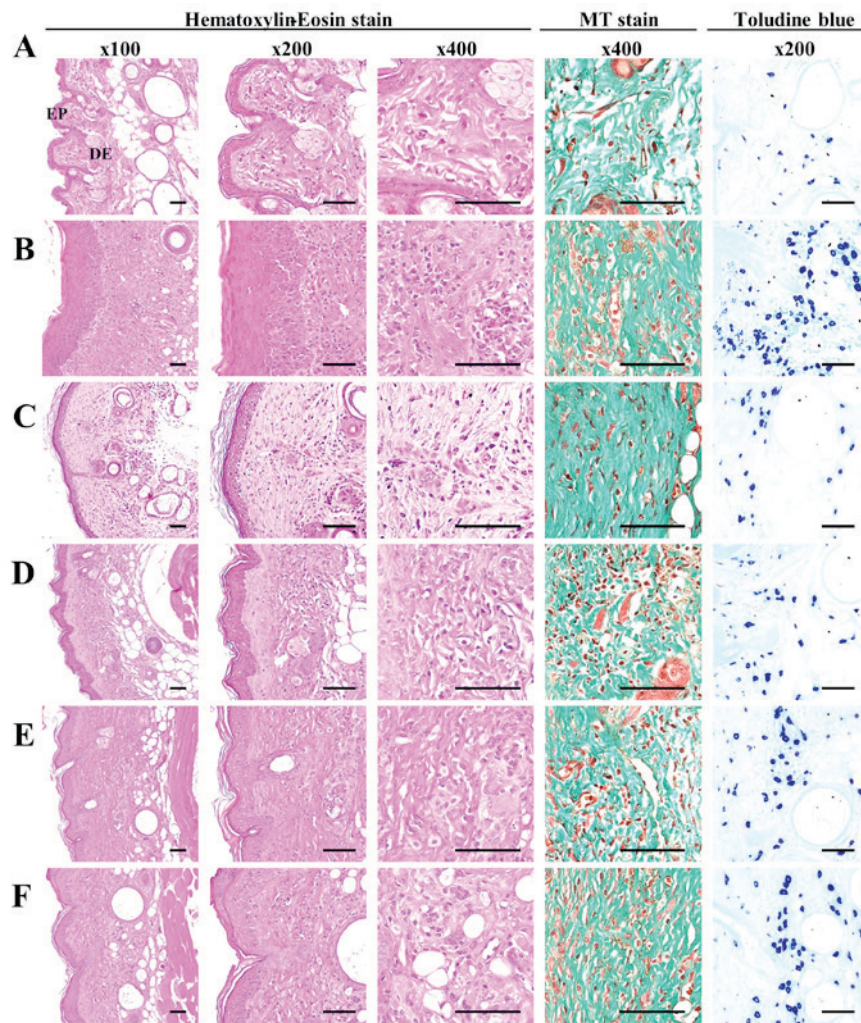


Figure 3. Representative histological images of dorsal back skin tissues taken from unexposed intact mice or mice with DNCB-induced AD following bathing in distilled water or ESGWc, or topical application of DEXA. Histological images from (A) intact vehicle control mice bathing in distilled water, (B) DNCB control mice bathing in distilled water, (C) mice with DNCB-induced AD treated with topical application of 1% DEXA and mice with DNCB-induced AD following bathing in ESGWc at (D) 100-, (E) 200- and (F) 400-fold dilution. Scale bar, 40 μ m. AD, allergic/atopic-like dermatitis; DNCB, 2,4-dinitrochlorobenzene; DEXA, dexamethasone; ESGWc, east saline groundwater concentration solution (50X); EP, epidermis; DE, dermis; MT, Masson's trichrome.

1% DEXA, and also concentration-dependently by bathing in all three dilutions of ESGWc, compared with DNCB control mice ($P < 0.05$; Table XIII).

Discussion

In the present study, clinical skin severity and scratching behaviors were significantly reduced, and serum total IgE levels were decreased in mice with DNCB-induced AD following bathing in all ESGWc dilutions, suggesting that bathing in ESGWc inhibited DNCB-induced dermatitis symptoms. Clinical skin severity scores and scratching behaviors were used as an index to select the mice that had developed AD at the end of DNCB sensitization and boosting. Major characteristics of skin lesions of AD are considered to be infiltration with various inflammatory cells, including mast cells, basophils, eosinophils and T cells (29), and symptoms, including itching, erythema, eczema skin lesions, chronic relapse and pruritus (1). DNCB-treated skin exhibits inflammation and dermis sclerosis with these common symptoms (30). Thus, clinical skin severity has been used as a meaningful indicator for predicting the progression

of AD with scratching behaviors (3,18). In the current study, clinical skin severity was graded based on five signs, which were pruritus/itching, erythema/hemorrhage, edema, excoriation/erosion and scaling/dryness. Mast cell-mediated inflammatory responses cause hypersensitivity and allergic reactions. Upon the cross-linking of their surface IgE by antigens, IgE-dependent mast cells may be stimulated to degranulate and release histamine, leukotrienes, proteases, prostaglandins and cytokines (6,18). Mast cell activation results in the release of numerous inflammatory mediators through the cross-linking of IgE receptor complexes at the high-affinity IgE receptor I. Of these mediators, histamine is considered as a marker of mast cell degranulation in immediate allergic reactions and is a violent inducer of itching (6,18). Histamine is a characteristic major mediator in mast cell storage granules and directly triggers type I allergic responses (6,18). Marked increases of infiltrated mast cells have been observed in various types of atopic dermatitis (6,18).

Furthermore, the present study demonstrated that bathing in ESGWc dilutions significantly suppressed DNCB-induced systemic and local hypersensitivities in

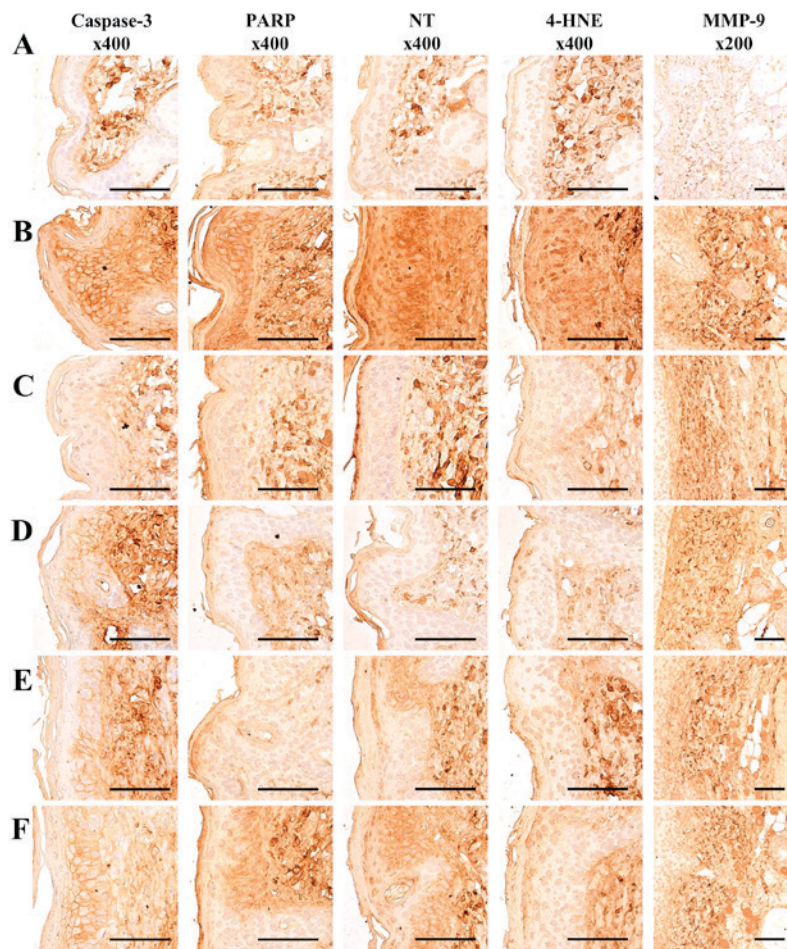


Figure 4. Representative immunohistochemical images of caspase-3, PARP, NT, 4-HNE and MMP-9 in the dorsal back skin tissues taken from unexposed intact mice or mice with DNCB-induced AD following bathing in distilled water or ESGWc, or topical application of DEXA. Immunohistochemical images from (A) intact vehicle control mice bathing in distilled water, (B) DNCB control mice bathing in distilled water, (C) mice with DNCB-induced AD treated with topical application of 1% DEXA and mice with DNCB-induced AD following bathing in ESGWc at (D) 100-, (E) 200- and (F) 400-fold dilution. Scale bar, 40 μ m. Stain, avidin-biotin complex immunostain. AD, allergic/atopic-like dermatitis; DNCB, 2,4-dinitrochlorobenzene; DEXA, dexamethasone; ESGWc, east saline groundwater concentration solution (50X); PARP, cleaved poly(ADP-ribose) polymerase; NT, nitrotyrosine; 4-HNE, 4-Hydroxynonenal; MMP-9, matrix metalloproteinase-9.

concentration-dependent patterns. TNF- α , IL-1 β and IL-10 levels in spleen tissue were markedly reduced, concomitantly TNF- α , IL-4, IL-5 and IL-13 mRNA expression in skin tissue were decreased following bathing in ESGWc dilutions. In addition, bathing in ESGWc dilutions inhibited the IFN- γ , iNOS, IL-1 β , IL-2 and TNF- α immunoreactive cells in the local skin, submandibular LN and spleen tissues. It has previously been reported that hydrotherapy, including with ESGW, was able to modulate lymphocyte proliferation and cytokine production (13,18). AD is a form of delayed-type hypersensitivity reaction induced in susceptible individuals by a variety of low molecular weight chemicals (31). The pathogenesis of AD is recognized as a complex process mediated by genetic, environmental and immunological factors (32). IL-4, IL-5 and IL-13 produced by Th2 cells may be responsible for the onset and development of AD (32). Although the etiology and pathology of AD remain unclear, research suggests that typical symptoms of AD include increased levels of Th2-mediated cytokines and a deficiency in Th1-mediated cytokines (33). Th2 cells are primarily involved in the acute phase of AD, while Th1 cells are activated and attribute to pathogenesis in the chronic phase (34). Increased IgE levels

are an important indicator of AD and the activation of IL-4 contributes to this elevation. IL-4 triggers IgE production in B cells and IgE released from B cells binds to mast cells. When sufficient antigen has bound to the IgE, the mast cells degranulate and release various biological mediators, including histamine and heparin, which is mediated by IgE in AD (35). AD is dependent on the release of the cytokines IL-4, IL-5, and IL-13 by Th2 cells that are produced from precursors (36). Many patients with AD have experienced an increase in eosinophils and IgE due to elevated IL-4, IL-5 and IL-13 levels (36). TNF- α , a 17-kDa protein that is classified as one of the major products of activated macrophages (37), is a presentative pro-inflammatory cytokine (38). Previous research has demonstrated that TNF- α is markedly increased in DNCB-induced dermatitis (39).

TNF- α -mediated inflammation is stimulated by the release of eicosanoids and cytokines, including IL-1 and TNF- α (38). IL-1 may stimulate neutrophils and macrophages by increasing the production and release of reactive oxygen species (ROS) and nitric oxide, which has been implicated in local tissue damage (40). IL-2 is generally produced by T cells during an immune response and is involved in the growth, proliferation

Table XI. Immunohistochemical skin tissue histomorphometry after 6 weeks of bathing in ESGWc or topical application of DEXA in mice with DNCB-induced AD.

Antiserum	Controls			Bathing in ESGWc dilutions		
	Intact	DNCB	DEXA	100-fold	200-fold	400-fold
Caspase-3 ^a	4.75±3.20	83.50±11.35 ^d	35.38±13.36 ^{d,e}	44.25±10.01 ^{d,e}	55.13±12.89 ^{d,e}	65.13±10.27 ^{d,e}
PARP ^a	6.00±2.27	84.63±10.10 ^d	23.00±4.72 ^{d,e}	35.63±10.95 ^{d,e}	42.25±12.89 ^{d,e}	59.38±13.75 ^{d,e}
NT ^a	17.13±3.52	85.00±10.98 ^d	18.00±5.07 ^c	25.13±10.92 ^c	51.63±11.98 ^{d,e}	61.88±18.65 ^{d,e}
4-HNE ^a	5.75±2.49	81.83±11.10 ^d	24.63±4.75 ^{d,e}	31.00±12.49 ^{d,e}	40.75±15.57 ^{d,e}	54.63±10.46 ^{d,e}
MMP-9 ^b	9.24±3.15	59.49±11.07 ^d	42.12±5.25 ^{d,e}	27.15±6.50 ^{d,e}	38.79±5.56 ^{a,e}	45.97±5.54 ^{d,e}
IFN- γ ^c	8.15±2.14	153.93±12.21 ^d	41.29±11.76 ^{d,e}	66.54±16.83 ^{d,e}	84.70±21.85 ^{d,e}	120.74±16.68 ^{d,e}
iNOS ^c	7.15±1.47	240.40±34.21 ^d	78.05±20.34 ^{d,e}	93.23±15.87 ^{d,e}	165.69±23.82 ^{d,e}	189.61±22.66 ^{d,e}
IL-1 β ^c	5.41±1.54	199.48±17.25 ^d	49.44±12.07 ^{d,e}	85.69±16.49 ^{d,e}	126.81±25.93 ^{d,e}	160.10±20.52 ^{d,e}
IL-2 ^c	4.13±1.15	87.08±21.38 ^d	19.71±5.55 ^{d,e}	37.24±9.30 ^{d,e}	51.58±11.99 ^{d,e}	60.74±11.01 ^{d,e}
TNF- α ^c	5.64±1.50	68.81±14.21 ^d	18.21±3.25 ^{d,e}	23.74±5.63 ^{d,e}	35.66±10.87 ^{d,e}	42.26±10.10 ^{d,e}

^aNumber of immunoreactive cells/100 epithelial cells; ^b% immunoreactivity/mm² of dermis; ^cx10 immunoreactive cells/mm² of dermis; ^dP<0.05 vs. Intact control; ^eP<0.05 vs. DNCB control. Values are expressed as the mean ± standard deviation (n=8). AD, allergic/atopic-like dermatitis; DNCB, 2,4-dinitrochlorobenzene; DEXA, dexamethasone; ESGWc, east saline ground water concentration solution (50X); PARP, cleaved poly(ADP-ribose) polymerase; NT, nitrotyrosine; 4-HNE, 4-hydroxynonenal; MMP, matrix metalloprotease; IFN, interferon; IL, interleukin; iNOS, inducible nitric oxide synthase; TNF, tumor necrosis factor.

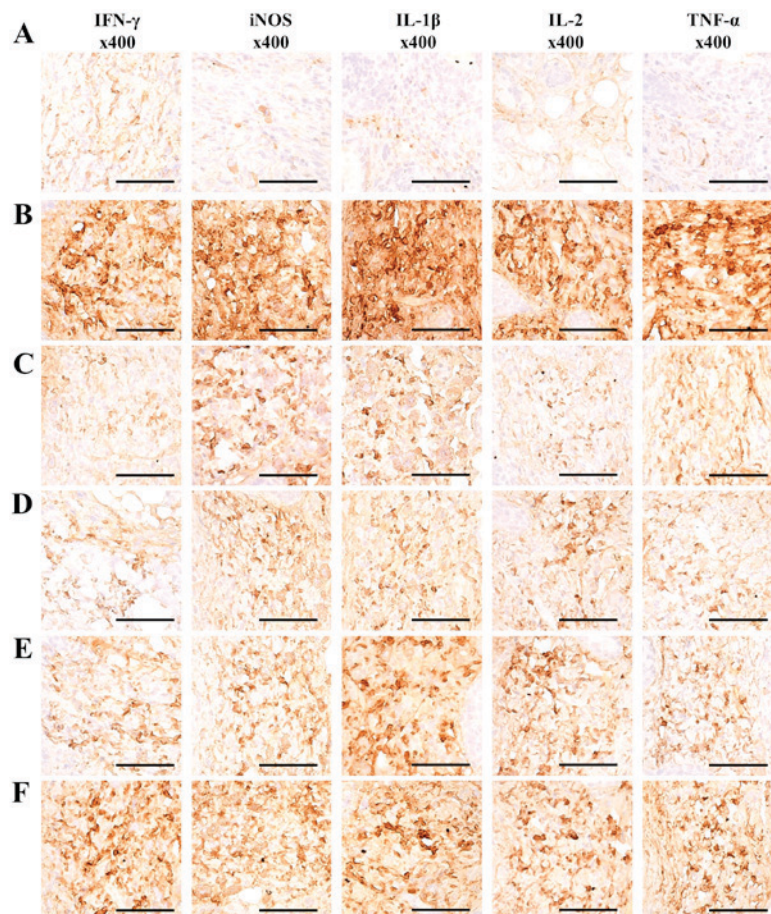


Figure 5. Representative immunohistochemical images of dermal IFN- γ , iNOS, IL-1 β , IL-2 and TNF- α in the dorsal back skin tissues taken from unexposed intact mice or mice with DNCB-induced AD following bathing in distilled water or ESGWc, or receiving topical application of DEXA. Immunohistochemical images from (A) intact vehicle control mice bathing in distilled water, (B) DNCB control mice bathing in distilled water, (C) mice with DNCB-induced AD treated with topical application of 1% DEXA and mice with DNCB-induced AD following bathing in ESGWc at (D) 100-, (E) 200- and (F) 400-fold dilution. Scale bar, 40 μ m. Stain, avidin-biotin complex immunostain. AD, allergic/atopic-like dermatitis; DNCB, 2,4-dinitrochlorobenzene; DEXA, dexamethasone; ESGWc, east saline groundwater concentration solution (50X); IFN, interferon; IL, interleukin; iNOS, inducible nitric oxide synthase; TNF, tumor necrosis factor.

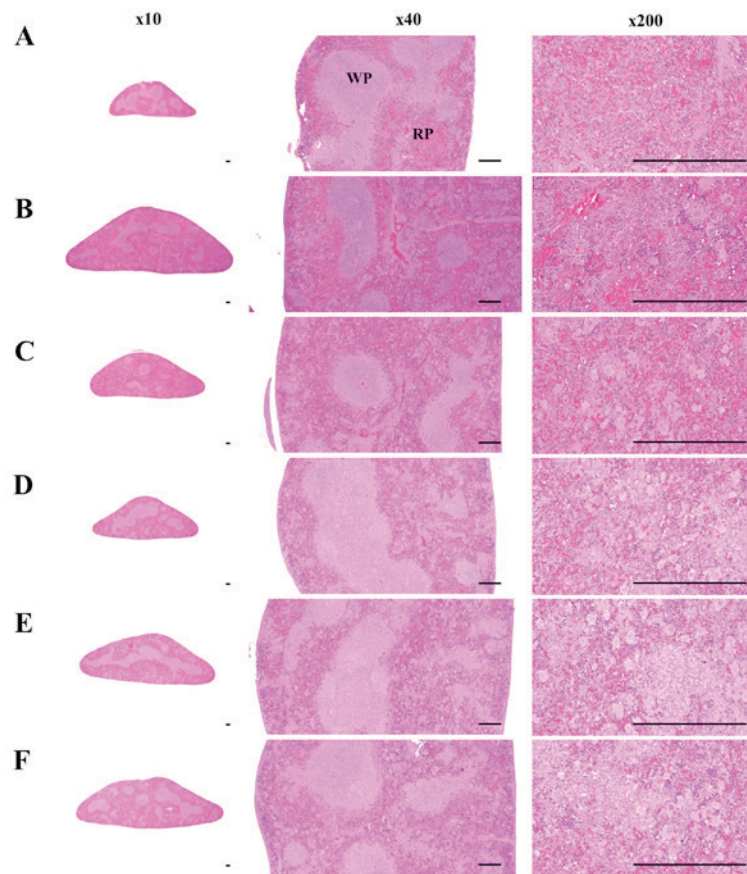


Figure 6. Representative histological images of splenic tissues taken from unexposed intact mice or mice with DNCB-induced AD following bathing in distilled water or ESGWc, or receiving topical application of DEXA. Histological images from (A) intact vehicle control mice bathing in distilled water, (B) DNCB control mice bathing in distilled water, (C) mice with DNCB-induced AD treated with topical application of 1% DEXA and mice with DNCB-induced AD following bathing in ESGWc at (D) 100-, (E) 200- and (F) 400-fold dilution. Scale bar, 200 μ m. Stain, hematoxylin and eosin. AD, allergic/atopic-like dermatitis; DNCB, 2,4-dinitrochlorobenzene; DEXA, dexamethasone; ESGWc, east saline groundwater concentration solution (50X); WP, white pulp; RP, red pulp.

and differentiation of T cells into effector T cells (41). In addition, IL-2 serves an important role in the induction of pruritus in atopic dermatitis (42). IFN- γ is produced by CD8+ T cells, Th1 cells and natural killer (NK) cells in response to IL-2. IFN- γ stimulates NK cell and macrophage activities and also exerts various effects on B and T cells (43). Increases in IFN- γ activities have also been detected in DNCB-induced dermatitis (44). iNOS is involved in the production of nitric oxide from L-arginine. High-output production of iNOS occurs under oxidative conditions. Excess levels of NO may increase the likelihood of interaction with superoxide, leading to peroxynitrite formation and cell toxicity (45). Pro-inflammatory agents, including endotoxin, IL-1 β , TNF- α and IFN- γ , may stimulate an increase in iNOS activity (46) and iNOS is also involved in the pathogenesis of atopic dermatitis (18). Therefore, the downregulation of iNOS, IL-2, TNF- α and IFN- γ expression may have protective effects in various allergic diseases. In addition, AD-induced systemic hypersensitivities induce marked proliferation of central and peripheral lymphocytes, particularly T cells (18,31).

As a result, in the present study, depletion of endogenous antioxidant GSH and increases of superoxide anion were significantly inhibited by bathing in ESGWc dilution, which acted as an antioxidant. In addition, epidermal NT and 4-HNE immunoreactive cells were significantly inhibited by bathing

in ESGWc dilutions. These findings suggest that bathing in ESGWc dilutions reduces DNCB-induced dermatitis symptoms through potent antioxidative effects. It has been reported that hydrotherapy has potent antioxidant effects on various forms of dermatitis (15,18). Further supporting the antioxidant effects of bathing in ESGWc, previous research has demonstrated that bathing in ESGW inhibited lipid peroxidation, as determined by the MDA concentration of TBARS assay (18). Many toxic substances induce lipid peroxidation damage to the surrounding tissues (47) and AD also causes an imbalance between ROS production and endogenous antioxidant capacity, resulting in depletion of endogenous antioxidants, such as GSH, an epidermal marker that is sensitive to oxidative stress (48). GSH, an important endogenous antioxidant, protects against tissue damage by maintaining ROS at low levels and at certain cellular concentrations (49). The highly polarizable sulfhydryl group of GSH facilitates the scavenging of radicals directly by hydrogen transfer, making it an optimum nucleophile for reactions with electrophilic chemicals (23). Acting as a cofactor for GSH peroxidase and GSH reductase, it may remove hydrogen peroxide and lipid hydroperoxides (50).

The α , β -unsaturated hydroxyalkenal 4-HNE, is generated as a result of lipid peroxidation and is a biomarker for oxidative stress (51). It is also regarded as a potential causal agent of various diseases, including chronic inflammation,

Table XII. Splenic tissue histomorphometry after 6 weeks of bathing in ESGWc or topical application of DEXA in mice with DNCB-induced AD.

Index	Controls			Bathing in ESGWc dilutions cells ^c		
	Intact	DNCB	DEXA	100-fold	200-fold	400-fold
Total Th, mm/ central regions	1.46±0.29	3.47±0.39 ^d	1.73±0.20 ^e	2.01±0.30 ^{d,e}	2.24±0.28 ^{d,e}	2.39±0.38 ^{d,e}
White pulp ^a	12.13±1.55	13.50±2.56	12.63±1.41	12.00±1.77	13.38±1.69	13.63±2.26
Red pulp ^b	2.69±0.56	34.30±10.11 ^d	4.98±1.33 ^{d,e}	9.65±2.99 ^{d,e}	16.15±2.82 ^{d,e}	22.29±5.10 ^{d,e}
Immunoreactive cells ^c						
TNF- α	15.38±2.00	471.13±88.58 ^d	55.38±13.14 ^{d,e}	127.00±24.17 ^{d,e}	277.25±60.30 ^{d,e}	334.38±65.07 ^{d,e}
IFN- γ	61.38±14.05	704.50±155.85 ^d	144.13±25.39 ^{d,e}	301.88±40.13 ^{d,e}	380.00±54.65 ^{d,e}	513.13±68.77 ^{d,e}
iNOS	64.00±15.09	829.38±85.72 ^d	147.88±27.88 ^{d,e}	356.25±100.05 ^{d,e}	556.63±113.03 ^{d,e}	640.50±115.92 ^{d,e}
IL-1 β	143.38±25.16	893.88±170.86 ^d	144.38±42.08 ^e	368.88±107.94 ^{d,e}	471.13±140.56 ^{d,e}	626.13±159.79 ^{d,e}
IL-2	75.13±14.82	878.00±134.90 ^d	150.63±25.40 ^{d,e}	412.63±60.59 ^{d,e}	538.00±141.18 ^{d,e}	651.38±150.39 ^{d,e}

^aNumber of cells/mm² of splenic parenchyma; ^bx10³ cells/mm² of splenic parenchyma; ^cnumber of immunoreactive cells/mm² of splenic parenchyma; ^dP<0.05 vs. Intact control; ^eP<0.05 vs. DNCB control. Values are expressed as the mean \pm standard deviation (n=8). AD, allergic/atopic-like dermatitis; DNCB, 2,4-dinitrochlorobenzene; DEXA, dexamethasone; ESGWc, east saline ground water concentration solution (50X); Th, thickness; IFN, interferon; IL, interleukin; iNOS, inducible nitric oxide synthase; TNF, tumor necrosis factor.

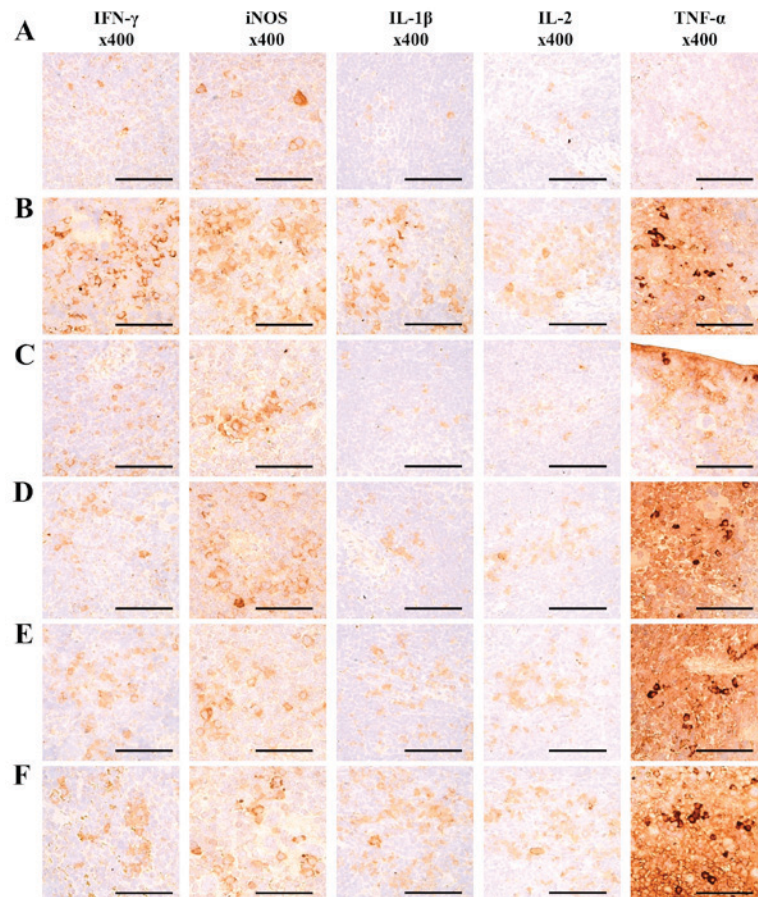


Figure 7. Representative immunohistochemical images of IFN- γ , iNOS, IL-1 β , IL-2 and TNF- α in the splenic tissues taken from unexposed intact mice or mice with DNCB-induced AD following bathing in distilled water or ESGWc, or receiving topical application of DEXA. Immunohistochemical images from (A) intact vehicle control mice bathing in distilled water, (B) DNCB control mice bathing in distilled water, (C) mice with DNCB-induced AD treated with topical application of 1% DEXA and mice with DNCB-induced AD after bathing in ESGWc at (D) 100-, (E) 200- and (F) 400-fold dilution. Scale bar, 40 μ m. Stain, avidin-biotin complex immunostain. AD, allergic/atopic-like dermatitis; DNCB, 2,4-dinitrochlorobenzene; DEXA, dexamethasone; ESGWc, east saline groundwater concentration solution (50X); IFN, interferon; IL, interleukin; iNOS, inducible nitric oxide synthase; TNF, tumor necrosis factor.

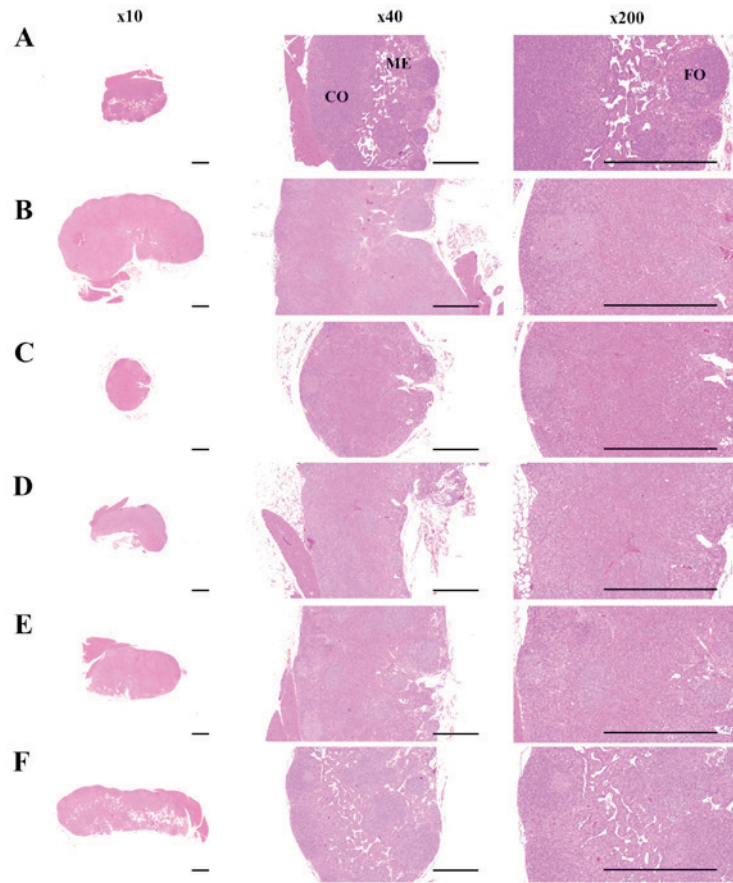


Figure 8. Representative histological images of submandibular LN tissues taken from unexposed intact mice or mice with DNCB-induced AD following bathing in distilled water or ESGWc, or receiving topical application of DEXA. Histological images from (A) intact vehicle control mice bathing in distilled water, (B) DNCB control mice bathing in distilled water, (C) mice with DNCB-induced AD treated with topical application of 1% DEXA and mice with DNCB-induced AD following bathing in ESGWc at (D) 100-, (E) 200- and (F) 400-fold dilution. Scale bar=400 μ m. Stain, hematoxylin and eosin. AD, allergic/atopic-like dermatitis; DNCB, 2,4-dinitrochlorobenzene; DEXA, dexamethasone; ESGWc, east saline groundwater concentration solution (50X); LN, lymph node; CO, cortex; FO, lymphoid follicle; ME, medulla.

neurodegenerative disease, adult respiratory distress syndrome, atherogenesis, diabetes and various types of cancer (51). NT is a product of tyrosine nitration mediated by reactive nitrogen species, including peroxynitrite anion and nitrogen dioxide. It is accepted as a marker of iNOS-dependent, reactive nitrogen species-induced nitrative stress in many pathological conditions (52,53).

To examine the cytoprotective anti-apoptotic effects of bathing in ESGWc dilutions in the present study, the activities of caspase-3 and PARP were examined in mice with DNCB-induced AD. Bathing in ESGWc dilutions exerted potent keratinocyte protection activity against DNCB-induced epidermal apoptosis by modulating the activities of caspase-3 and PARP. It has been demonstrated that the cleavage of PARP is accompanied by a decrease in enzymatic repair function and consequently leads to the progression of apoptosis (54). Caspase-3 contributes to the cleavage of key nuclear targets in the apoptotic pathway. It contains the inhibitor of caspase-activated deoxynuclease, which causes nuclear fragmentation and PARP cleavage, consequently leading to a decrease in DNA repair (55). Thus, caspase-3 is considered to be an important executioner of apoptosis, as it is involved in the proteolytic cleavage of pivotal proteins, such as PARP (56). PARP is a recognized

target molecule of caspase-3, which induces the activation of the DNA repair system by caspase-mediated cleavage (57). However, previous studies have reported that overexpression of PARP proteins induced necrosis as intracellular energy levels were decreased through the over-consumption of its substrate, nicotinamide adenine dinucleotide (58-60). Although the association of PARP with apoptosis and necrosis remains unclear, the suppression of PARP is regarded as a useful strategy for protecting against various forms of skin damage (61) induced by apoptotic changes in DNCB-induced allergic dermatitis (18,30).

As MMPs are able to degrade various extracellular matrix components and are known to serve important roles in tissue remodeling during developmental morphogenesis, angiogenesis, tissue repair, arthritis, skin aging and tumor invasion (62), the present study examined whether regulation of MMP activity by bathing in ESGWc was involved in DNCB-induced skin sclerosis. The results indicated that bathing in ESGWc (up to 400-fold dilution) exerted strong inhibitory activity against DNCB-induced skin sclerosis by modulating the activities of MMP-9. MMPs, a family of structurally related endopeptidases (63) are categorized into collagenases, gelatinases, stromelysins, membrane-type MMPs and other MMPs, according to their structures and substrate specificities (64).

Table XIII. Submandibular LN tissue histomorphometry after 6 weeks of bathing in ESGWc or topical application of DEXA in mice with DNCB-induced AD.

Index	Controls			Bathing in ESGWc dilutions		
	Intact	DNCB	DEXA	100-fold	200-fold	400-fold
Total Th, mm/ central regions	0.73±0.13	1.89±0.23 ^d	1.02±0.17 ^{d,e}	1.13±0.19 ^{d,e}	1.21±0.15 ^{d,e}	1.39±0.13 ^{d,e}
Follicles ^a	12.50±1.93	35.00±5.40 ^d	16.00±2.45 ^{d,e}	17.88±2.75 ^{d,e}	19.63±2.97 ^{d,e}	26.25±6.30 ^{d,e}
Cortex Th ^b	364.96±112.92	987.27±146.47 ^d	431.98±86.57 ^e	566.65±143.70 ^{d,e}	759.54±109.37 ^{d,e}	804.96±92.38 ^{d,e}
Immunoreactive cells ^c						
TNF- α	7.23±1.26	139.63±34.18 ^d	23.23±8.57 ^{d,e}	34.21±9.08 ^{d,e}	46.71±12.43 ^{d,e}	75.45±21.96 ^{d,e}
IFN- γ	9.31±2.37	217.28±23.00 ^d	21.20±7.38 ^{d,e}	74.94±21.86 ^{d,e}	119.58±26.28 ^{d,e}	177.78±22.73 ^{d,e}
iNOS	9.39±3.04	167.09±31.30 ^d	21.43±8.37 ^{d,e}	42.69±15.02 ^{d,e}	55.13±12.98 ^{d,e}	63.11±12.43 ^{d,e}
IL-1 β	14.11±3.65	135.53±21.48 ^d	49.26±13.39 ^{d,e}	54.50±15.18 ^{d,e}	72.50±10.25 ^{d,e}	99.44±15.76 ^{d,e}
IL-2	3.24±1.35	89.48±12.43 ^d	19.19±7.30 ^{d,e}	26.16±11.27 ^{d,e}	39.79±11.69 ^{d,e}	48.91±18.70 ^{d,e}

^aNumbers/mm² of cortex; ^b μ m/LN; ^cx10 cells/mm² of LN parenchyma; ^dP<0.05 vs. Intact control; ^eP<0.05 vs. DNCB control. Values are expressed as the mean \pm standard deviation (n=8). LN, lymph node; AD, allergic/atopic-like dermatitis; DNCB, 2,4-dinitrochlorobenzene; DEXA, dexamethasone; ESGWc, east saline ground water concentration solution (50X); Th, thickness; IFN, interferon; IL, interleukin; iNOS, inducible nitric oxide synthase; TNF, tumor necrosis factor.

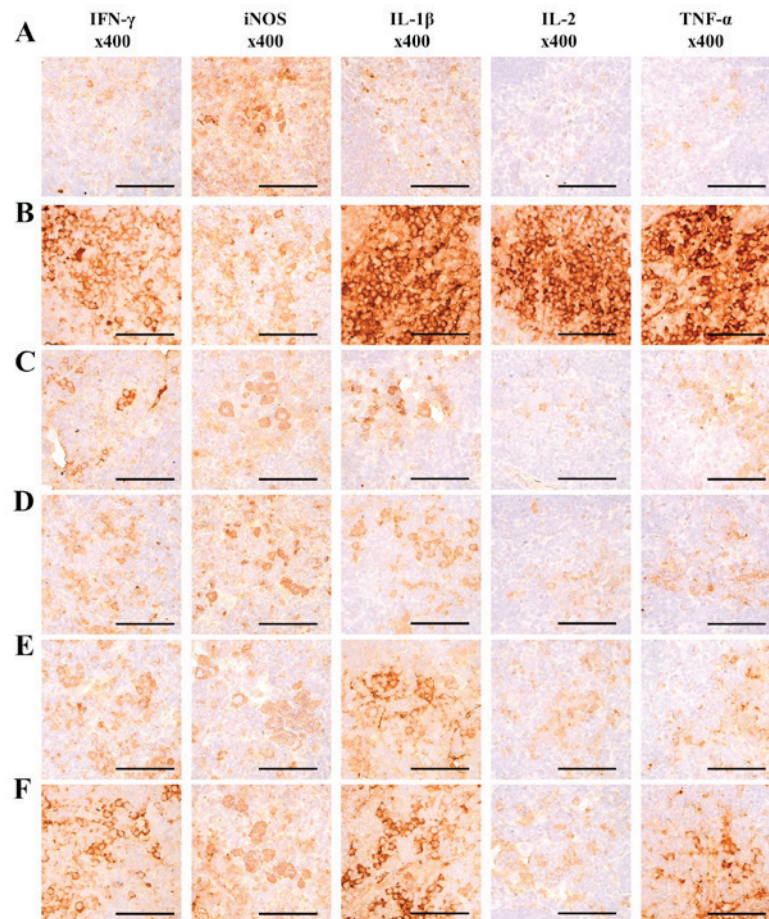


Figure 9. Representative immunohistochemical images of IFN- γ , iNOS, IL-1 β , IL-2 and TNF- α in the submandibular LN tissues taken from unexposed intact mice or mice with DNCB-induced AD following bathing in distilled water or ESGWc, or receiving topical application of DEXA. Immunohistochemical images from (A) intact vehicle control mice bathing in distilled water, (B) DNCB control mice bathing in distilled water, (C) mice with DNCB-induced AD treated with topical application of 1% DEXA and mice with DNCB-induced AD after bathing in ESGWc at (D) 100-, (E) 200- and (F) 400-fold dilution. Scale bar=40 μ m. Stain, avidin-biotin complex immunostain. AD, allergic/atopic-like dermatitis; DNCB, 2,4-dinitrochlorobenzene; DEXA, dexamethasone; ESGWc, east saline groundwater concentration solution (50X); IFN, interferon; IL, interleukin; iNOS, inducible nitric oxide synthase; TNF, tumor necrosis factor.

Levels of MMPs are usually low in unstimulated skin tissues and levels are upregulated following exposure to various extracellular stimuli, including ultraviolet light, growth factors, cytokines and tumor promoters (64,65). Previous studies have indicated that there was a significant increase in a number of MMPs, particularly MMP-8 and MMP-9, in AD patients (66,67). Such research suggests that reducing MMP activity may be a protective approach to attenuate AD-related skin sclerosis (18,68).

The results obtained in the present study suggest that bathing in all three concentrations (100-, 200- and 400-fold dilutions) of ESGWc have favorable protective effects against DNCB-induced AD. It was demonstrated that ESGWc has favorable systemic and local immunomodulatory effects, active cytoprotective anti-apoptotic effects, inhibitory effects of MMP activity, anti-inflammatory and antioxidative effects, in a concentration-dependent manner. In conclusion, bathing in ESGWc, may provide an alternative therapy for patients with AD in the future, at least under the conditions of this experiment. Topical application of 1% DEXA also significantly inhibited the DNCB-induced AD; however, it did not influence dermal collagen depositions and skin tissue antioxidant defense systems in the present experiment.

Acknowledgements

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