

Value of serum *Mycoplasma pneumoniae* immunoglobulin in the diagnosis of mycoplasma-related pneumonia in newborns

YAN DONG, WEI LV and ZHEN LIN

Department of Paediatrics, Shanghai Ninth People's Hospital, Shanghai Jiao Tong University School of Medicine, Shanghai 201900, P.R. China

Received March 24, 2017; Accepted June 2, 2017

DOI: 10.3892/etm.2017.4654

Abstract. *Mycoplasma pneumoniae* (*M. pneumoniae*) is an important pathogen of neonatal acquired pneumonia in newborns. Rapid and accurate diagnosis of *M. pneumoniae* infection is critical because timely antibiotic therapy can reduce drug overuse and prevent the development of bacterial resistance. Anti-*M. pneumoniae* immunoglobulin M (IgM) is an indicator of early infection that can persist for several months. Studies have shown that anti-*M. pneumoniae* IgA in adults is a reliable indicator of early *M. pneumoniae* infection. The aim of this study was to assess the association between *M. pneumoniae* IgA, IgM and IgG in mycoplasma-associated pneumonia. We recruited 80 newborns with pneumonia with potency of serum *M. pneumoniae* IgM positive or two sera anti-*M. pneumoniae* IgG increased by 4-fold. The potency of serum *M. pneumoniae* IgA, IgM and IgG were detected. The initial positive rates of IgM and IgA in *M. pneumoniae* were 63.6 and 33.8%, respectively, after infection. The positive rate of IgM and IgA in *M. pneumoniae* increased to 97.5 and 56.3%, respectively, at one week after infection. Compared with anti-*M. pneumoniae* IgA, anti-*M. pneumoniae* IgM has higher sensitivity in the diagnosis of neonatal mycoplasma-associated pneumonia. Detection of two sera can more effectively improve the diagnostic accuracy.

Introduction

Mycoplasma pneumoniae (*M. pneumoniae*) is one of the most common pathogens causing pneumonia, especially neonatal pneumonia. The typical clinical manifestation is cough without runny nose, but the symptoms vary widely from asymptomatic respiratory infections to severe pulmonary infections (1). *M. pneumoniae* is sensitive to macrolide or

tetracycline antibiotics, but because of the lack of appropriate diagnostic methods, these antibiotics are not used in a timely manner. *M. pneumoniae* is also sensitive to fluoroquinolone antibiotics, but fluoroquinolones cannot be used in neonates because of cytotoxic side effects (2). In recent years, the extensive use of macrolide antibiotics has led to the increase of macrolide-resistant *M. pneumoniae* around the world (3). The proportion of macrolide-resistant *M. pneumoniae* in Asia is 10-20% (4,5), whereas in some parts of China it is as high as 90%. Therefore, there is an urgent need for a rapid and accurate diagnosis method of *M. pneumoniae* infection to choose appropriate antibiotics to treat *M. pneumoniae*-associated pneumonia, thereby reducing the abuse of antibiotics and drug-resistant strains.

Anti-*M. pneumoniae* immunoglobulin M (IgM) is used to detect acute infection. However, the levels of IgM antibody in blood were too low to be detected in some patients in the early stage of acute infection and in reinfection (6). In addition, the IgM antibody can only be detected several months after infection in the blood of some patients (7). So, the clinical diagnosis of *M. pneumoniae* infection is very difficult. In the last few decades, only a small number of studies have reported the use of *M. pneumoniae* IgA antibodies to diagnose *M. pneumoniae* infection. Reports indicate that detection of anti *M. pneumoniae* IgA antibodies in adults is more sensitive than IgM in the diagnosis of acute mycoplasma-associated pneumonia (8). However, the sensitivity of *M. pneumoniae* IgA was low for diagnosing of mycoplasma-associated pneumonia in neonates (9).

The incidence of *M. pneumoniae*-associated pneumonia is high in neonates, but no studies have reported the clinical value of anti-*M. pneumoniae* Ig in the diagnosis of *M. pneumoniae*-associated pneumonia in neonates. The purpose of this study was to assess the clinical significance and efficacy of anti-*M. pneumoniae* Ig in the diagnosis of *M. pneumoniae* in neonates.

Patients and methods

Study subjects. We recruited 80 newborns in Shanghai Ninth People's Hospital, Shanghai JiaoTong University School of Medicine from May 2013 to June 2016. The cohort included 31 boys and 49 girls. Mean age: 16.6±5.3 months, age ranged from 8-27 days. All newborns had cough and fever. Bronchial

Correspondence to: Dr Zhen Lin, Department of Paediatrics, Shanghai Ninth People's Hospital, Shanghai Jiao Tong University School of Medicine, 280 Mohe Road, Baoshan, Shanghai 201900, P.R. China
E-mail: jsrpsj@163.com

Key words: *Mycoplasma pneumoniae* IgA, pneumonia, newborn, *Mycoplasma pneumoniae* IgM

Table I. Clinical data and blood test results of neonates with mycoplasma-associated pneumonia.

Clinical data	Mean \pm standard error (range)	P-value
Age (days)	16.6 \pm 5.3 (8-27)	0.859
Sex (Male/Female)	31/49	
Pre-hospital fever duration (days)	4.3 \pm 2.7 (0-10)	0.014
Total fever duration (days)	5.7 \pm 3.4 (0-15)	0.426
Days of hospitalization	7.3 \pm 5.0 (2-14)	0.210
Red blood cells (million/ μ l)	4.6 \pm 0.4 (3.4-5.7)	0.526
Hemoglobin (g/dl)	12.6 \pm 1.0 (10.1-14.4)	0.855
Platelets (1,000/ μ l)	271.2 \pm 76.4 (114.0-454.0)	0.004
White blood cells (1,000/ μ l)	9.2 \pm 3.9 (3.8-20.7)	0.158
Lymphocytes (%)	26.3 \pm 11.3 (5.0-65.0)	0.465
Eosinophils (%)	1.9 \pm 2.8 (0.0-8.6)	0.100
Basophils (%)	0.3 \pm 0.4 (0.0-3.0)	0.216
Monocytes (%)	7.0 \pm 3.0 (1.8-14.0)	0.089
AST (μ l)	34.6 \pm 11.3 (22.0-65.0)	0.634
ALT (μ l)	20.2 \pm 13.2 (9.0-66.0)	0.603
CRP (mg/l)	43.5 \pm 42.6 (1.4-215.4)	0.328

The P-value represents the correlation between the patient's clinical data and the initial value of *M. pneumoniae* IgM. The duration of fever before admission and the platelet number were closely correlated with the *M. pneumoniae* IgM potency. *M. pneumoniae*, *Mycoplasma pneumoniae*; IgM, *M. pneumoniae* immunoglobulin M.

pneumonia or lobar pneumonia was identified in all newborns by chest X-ray. Body temperature was measured with an infrared tympanic thermometer and temperature above 38.0°C was considered as fever. The patients had continued fever ($\geq 38^\circ\text{C}$) for 4.3 \pm 2.7 days before admission (information provided by the relatives of the patients). All patients with *M. pneumoniae* infection showed serum antibody positive or increased antibody potency at least two weeks after the infection: 26 were serum positive, in 72 IgM potency increased 2-fold, and in 31 IgG potency increased 4-fold. The patients involved in this study had no other disease that could alter the clinical disease process. The clinical data of the patients included age, fever duration, length of hospitalization, laboratory examination, liver zymogram and CRP test results (Table I). This study was approved by the Ethics Committee of Shanghai Ninth People's Hospital. Signed written informed consents were obtained from the patients and/or guardians before the study.

Methods. All children received macrolide antibiotics and the clinical symptoms improved after treatment. The levels of anti-*M. pneumoniae* IgA, IgM, and IgG in serum were measured at different time points at early stages, during the development of pneumonia, and after pneumonia. The levels of anti-*M. pneumoniae* IgM and IgG in serum were measured by ELISA (Ben-Bio, San Diego, CA, USA). The levels of anti-*M. pneumoniae* IgA in serum were measured with the CHORUS kit according to the manufacturer's instructions (Diesse Diagnostica Senese, Siena, Italy). The positive cutoff values for anti-*M. pneumoniae* IgA, IgM and IgG were 18 AU/ml (the upper and lower limits were 10 and 100 AU/ml, respectively), 950 and 320 AU/ml, respectively.

Continuous variables are expressed as mean \pm standard error of the mean (mean \pm SEM). The initial positive rate of IgA and IgM in *M. pneumoniae* was analyzed by chi-square test. *M. pneumoniae* IgM potency, clinical features, blood laboratory test results, and the correlation with CPR were analyzed using Pearson's correlation analysis and multivariate logistic regression analysis. All experimental data were analyzed with SPSS 13.0. $P < 0.05$ was considered to be statistically significant.

Results

***M. pneumoniae* IgM potency and pre-hospital fever duration.** To investigate the correlation between *M. pneumoniae* IgM potency and pre-hospital fever duration, we divided the patients into three groups according to the duration of fever before admission: i) 0-3 days, ii) 4-6 days, and iii) 7-10 days. Most children (40%; 32/80) had < 3 days of fever before admission; 27 patients (33.8%) had 4-6 days of pre-hospital fever; and 21 patients (26.3%) had 7-10 days of pre-hospital fever. The results showed that the potency of *M. pneumoniae* IgM, but not IgA or IgG, was positively correlated with pre-hospital fever duration ($r = 0.377$, $P = 0.002$) (Fig. 1).

***M. pneumoniae* patients show a higher positive rate for IgM.** The positive rates for anti-*M. pneumoniae* IgA, IgM, and IgG were 33.8 (27/80), 63.8 (51/80) and 32.5% (26/80), respectively, in the 80 newborns before admission. IgA and IgM showed relatively higher positive rate. In addition, the positive rates of *M. pneumoniae* IgM were higher than those of *M. pneumoniae* IgA in all groups (Fig. 2), indicating that anti-*M. pneumoniae* IgM had a higher positive rate than

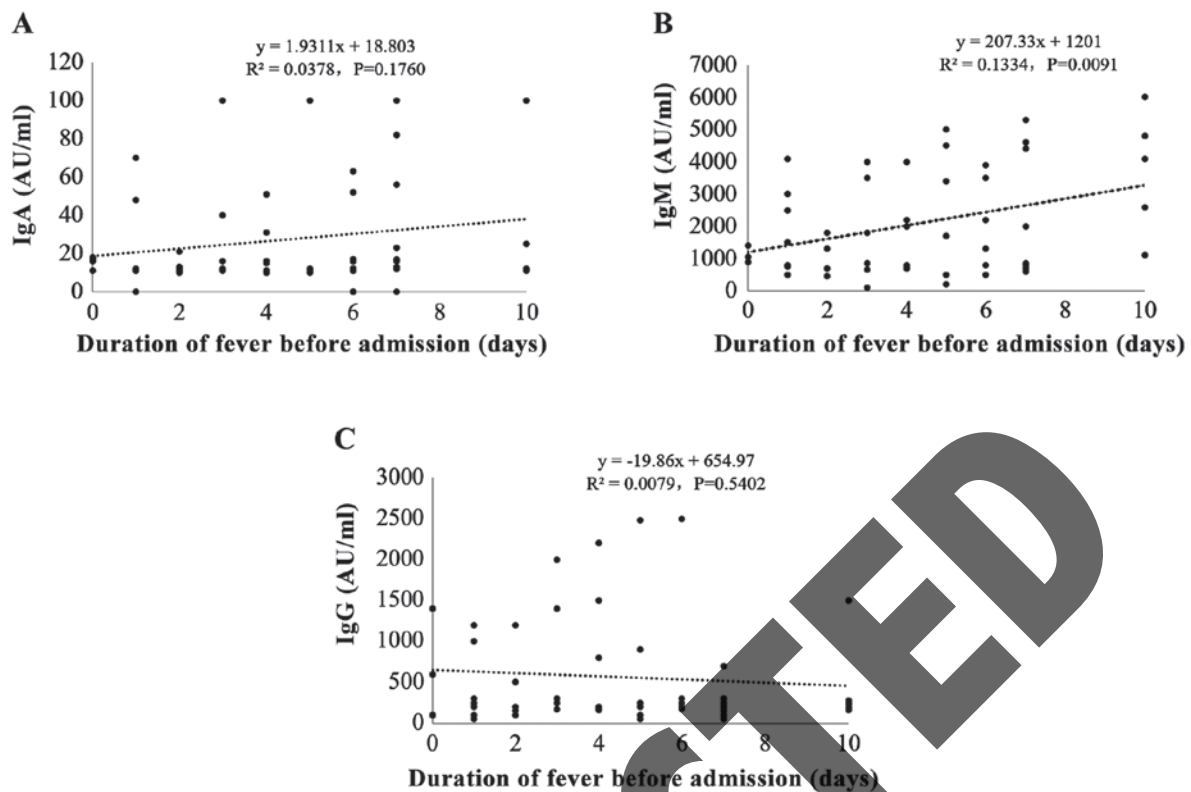


Figure 1. Correlation between initial values of *M. pneumoniae* IgA, IgM and IgG and the duration of fever before admission. *M. pneumoniae* IgM potency and fever duration before admission were positively correlated. The correlations between *M. pneumoniae* IgA and IgG potency and fever duration before admission were not significant. *M. pneumoniae*, *Mycoplasma pneumoniae*; IgM, *M. pneumoniae* immunoglobulin M.

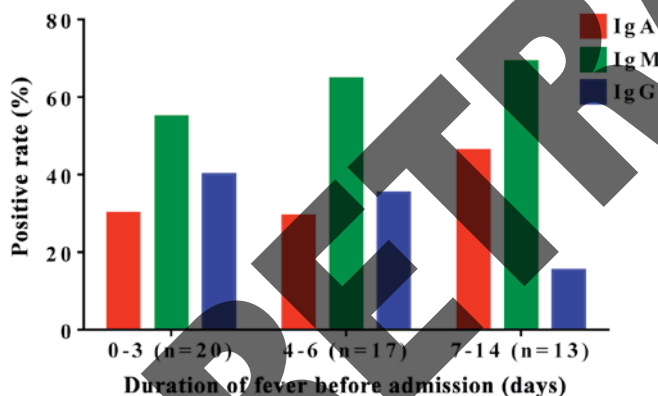


Figure 2. The positive rates of *M. pneumoniae* IgA, IgM and IgG. The positive rate of *M. pneumoniae* IgA, IgM and IgG in patients with different duration of fever before admission are presented. Patients were divided into 3 groups (0-3, 4-6 and 7-10 days) according to the duration of fever before admission. The results showed that the positive rates of IgM were higher than those of IgA in all groups. *M. pneumoniae*, *Mycoplasma pneumoniae*; IgM, *M. pneumoniae* immunoglobulin M.

that of anti-*M. pneumoniae* IgA in the diagnosis of neonatal mycoplasma-associated pneumonia.

Two sera test of the patients with clinical symptoms and *M. pneumoniae* IgM negative. To compare the cumulative positive rates of *M. pneumoniae* IgA, IgM, and IgG, we compared the values within 14 days after admission. We collected blood samples at different time points before and after admission. The samples were divided into 5 groups according to the

time of sampling: i) the day of admission, ii) 2-4 days after admission, iii) 5-7 days after admission, iv) 8-10 days after admission, and v) 11-14 days after admission. The initial positive rate of *M. pneumoniae* IgM was 63.8% (51/80) (Fig. 3A). The cumulative positive rates of *M. pneumoniae* IgM in groups 2 and 3 were 85.0 and 97.5%, respectively (Fig. 3A). Serum *M. pneumoniae* IgM for 26 cases (32.5%) turned positive one week after admission. These results suggest that it is necessary to collect double serum for patients with clinical symptoms, but initially negative for anti-*M. pneumoniae* IgM. In addition, the results also showed that the cumulative positive rate of serum *M. pneumoniae* IgM was higher than that of IgA.

Of the 80 children, 26 (32.5%) cases had serum *M. pneumoniae* IgM negative at admission and turned positive two weeks after admission. Of these 26 patients, 4 (15.4%) had initial serum *M. pneumoniae* IgA positive. However, the positive rate of serum *M. pneumoniae* IgM was higher than that of IgA in all patients 2-4 days after admission (61.5 vs. 46.2%) (Fig. 3B), indicating the importance of two sera tests in the clinical diagnosis.

Discussion

The course of mycoplasma infection is often relatively long. In adults, *M. pneumoniae* can still exist one week after medical treatment (8). Our results showed that most newborns with mycoplasma-associated pneumonia were admitted to the hospital within one week (4.3 ± 2.7 days) after *M. pneumoniae* infection. The possible explanation is that fever is the main

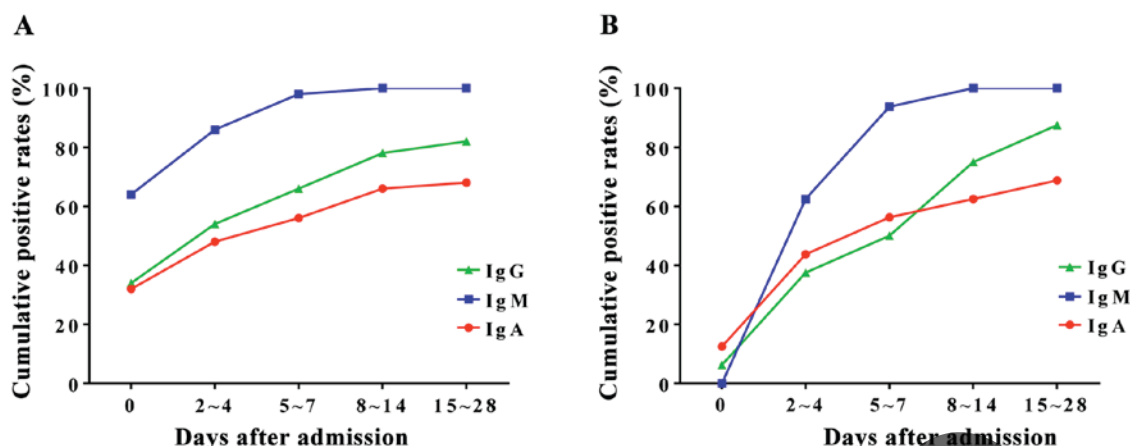


Figure 3. The cumulative positive rates of *M. pneumoniae* IgA, IgM and IgG. (A) The cumulative positive rate of *M. pneumoniae* IgA, IgM and IgG in 80 children at 5 time points after admission. (B) The positive rate of *M. pneumoniae* IgA, IgM and IgG in 26 patients. The 26 patients had initial *M. pneumoniae* IgM negative but turned positive at one week after admission. The cumulative positive rate of *M. pneumoniae* IgM was higher than that of IgA at 2 weeks after admission. *M. pneumoniae*, *Mycoplasma pneumoniae*; IgM, *M. pneumoniae* immunoglobulin M.

clinical symptoms in children, but persistent cough is the typical symptom in adults. Compared with adults, newborns with infection can be admitted earlier to hospital. Our results showed that many children (21/80, 26.3%) were negative for *M. pneumoniae*-specific antibodies at admission. To avoid false negatives, two serum samples were used to test *M. pneumoniae* IgA, IgM and IgG after admission.

Previous studies reported that serum anti-*M. pneumoniae* IgA is a good indicator for the detection of *M. pneumoniae* infection in adults (10-12). Detection of serum *M. pneumoniae* IgA in adults is more sensitive for the diagnosis of *M. pneumoniae* infection than detection of IgM (8). However, this conclusion is inconsistent with the results reported by Yamazaki *et al* (9) who found that serum *M. pneumoniae* IgA was a poor indicator of *M. pneumoniae* infection. Here, we examined the efficacy of *M. pneumoniae* IgA in the diagnosis of neonatal mycoplasma-associated pneumonia. The positive rates of serum *M. pneumoniae* IgM and IgA were 63.8 and 33.8%, respectively, on the day of admission. These rates were positively correlated with the duration of fever before admission. We also found that the positive rate of serum *M. pneumoniae* IgM was higher than that of IgA in all the groups classified by pre-hospitalization fever duration, suggesting that detection of serum *M. pneumoniae* IgA is less sensitive than IgM in the diagnosis of neonatal mycoplasma-associated pneumonia. This may be explained by the immature immune system of newborns.

In our study, the positive rate of *M. pneumoniae* IgM was 63.8% in patients with an average duration of 4.3 ± 2.7 days before admission. This result is consistent with a previous report (13). That is, the positive rate of *M. pneumoniae* IgM was 62.2% in the first week after mycoplasma infection and ranged from 70.9 to 81.8% in the second week after infection (14,15). The sensitivity of serological testing is limited by the specimen, the standard diagnostic method and the method of detection. This may be used to explain the high sensitivity of *M. pneumoniae* IgM in patients with longer hospitalization. We found that the positive rate of *M. pneumoniae* IgA was positively correlated with the pre-hospitalization fever duration, although this

correlation was lower than the correlation between IgM and pre-hospitalization fever duration.

The 4-fold increase of *M. pneumoniae* IgG in the acute phase and the reversion of the disease is considered the gold standard for diagnosis of *M. pneumoniae* respiratory tract infection (16). Medjo *et al* (14) reported that 90% of the patients with 4-fold increase of *M. pneumoniae* IgG antibody potency in two sera also showed throat swab *M. pneumoniae* positive in PCR detection. While Ma *et al* (15) reported that only 2.4% of the patients with a four-fold increase of *M. pneumoniae* IgG antibody potency in two sera also showed throat swab *M. pneumoniae* positive. However, 38.8% of patients in this study had a 4-fold increase in *M. pneumoniae* IgG antibody potency in two sera. Thus, anti-*M. pneumoniae* IgG cannot provide a timely diagnosis of *M. pneumoniae* infection. Given the complications of obtaining two sera from newborns, *M. pneumoniae* IgG is not the best indicator to diagnose *M. pneumoniae* infection.

In conclusion, detection of *M. pneumoniae* IgM has higher sensitivity in the diagnosis of neonatal mycoplasma-associated pneumonia than that of the detection of *M. pneumoniae* IgA. Two sera detection can more effectively improve the diagnostic accuracy.

References

1. Ali NJ, Sillis M, Andrews BE, Jenkins PF and Harrison BD: The clinical spectrum and diagnosis of *Mycoplasma pneumoniae* infection. *Q J Med* 58: 241-251, 1986.
2. Bradley JS and Jackson MA; Committee on Infectious Diseases; American Academy of Pediatrics: The use of systemic and topical fluoroquinolones. *Pediatrics* 128: e1034-e1045, 2011.
3. Pereyre S, Goret J and Bébér C: *Mycoplasma pneumoniae*: Current knowledge on macrolide resistance and treatment. *Front Microbiol* 7: 974, 2016.
4. Wu PS, Chang LY, Lin HC, Chi H, Hsieh YC, Huang YC, Liu CC, Huang YC and Huang LM: Epidemiology and clinical manifestations of children with macrolide-resistant *Mycoplasma pneumoniae* pneumonia in Taiwan. *Pediatr Pulmonol* 48: 904-911, 2013.
5. Wu HM, Wong KS, Huang YC, Lai SH, Tsao KC, Lin YJ and Lin TY: Macrolide-resistant *Mycoplasma pneumoniae* in children in Taiwan. *J Infect Chemother* 19: 782-786, 2013.
6. Sillis M: The limitations of IgM assays in the serological diagnosis of *Mycoplasma pneumoniae* infections. *J Med Microbiol* 33: 253-258, 1990.

7. Thacker WL and Talkington DF: Analysis of complement fixation and commercial enzyme immunoassays for detection of antibodies to *Mycoplasma pneumoniae* in human serum. Clin Diagn Lab Immunol 7: 778-780, 2000.
8. Granström M, Holme T, Sjögren AM, Ortqvist A and Kalin M: The role of IgA determination by ELISA in the early serodiagnosis of *Mycoplasma pneumoniae* infection, in relation to IgG and mu-capture IgM methods. J Med Microbiol 40: 288-292, 1994.
9. Yamazaki T, Narita M, Sasaki N, Kenri T, Arakawa Y and Sasaki T: Comparison of PCR for sputum samples obtained by induced cough and serological tests for diagnosis of *Mycoplasma pneumoniae* infection in children. Clin Vaccine Immunol 13: 708-710, 2006.
10. Lieberman D, Lieberman D, Ben-Yaakov M, Shmarkov O, Gelfer Y, Varshavsky R, Ohana B, Lazarovich Z and Boldur I: Serological evidence of *Mycoplasma pneumoniae* infection in acute exacerbation of COPD. Diagn Microbiol Infect Dis 44: 1-6, 2002.
11. Lieberman D, Lieberman D, Korsonsky I, Ben-Yaakov M, Lazarovich Z, Friedman MG, Dvoskin B, Leinonen M, Ohana B and Boldur I: A comparative study of the etiology of adult upper and lower respiratory tract infections in the community. Diagn Microbiol Infect Dis 42: 21-28, 2002.
12. Watkins-Riedel T, Stanek G and Daxboeck F: Comparison of SeroMP IgA with four other commercial assays for serodiagnosis of *Mycoplasma pneumoniae* pneumonia. Diagn Microbiol Infect Dis 40: 21-25, 2001.
13. Chang HY, Chang LY, Shao PL, Lee PI, Chen JM, Lee CY, Lu CY and Huang LM: Comparison of real-time polymerase chain reaction and serological tests for the confirmation of *Mycoplasma pneumoniae* infection in children with clinical diagnosis of atypical pneumonia. J Microbiol Immunol Infect 47: 137-144, 2014.
14. Medjo B, Atanaskovic-Markovic M, Radic S, Nikolic D, Lukac M and Djukic S: *Mycoplasma pneumoniae* as a causative agent of community-acquired pneumonia in children: Clinical features and laboratory diagnosis. Ital J Pediatr 40: 104, 2014.
15. Ma YJ, Wang SM, Cho YH, Shen CF, Liu CC, Chi H, Huang YC, Huang LM, Huang YC, Lin HC, *et al*: Taiwan Pediatric Infectious Disease Alliance: Clinical and epidemiological characteristics in children with community-acquired mycoplasma pneumonia in Taiwan: A nationwide surveillance. J Microbiol Immunol Infect 48: 632-638, 2015.
16. Gavranich JB and Chang AB: Antibiotics for community acquired lower respiratory tract infections (LRTI) secondary to *Mycoplasma pneumoniae* in children. Cochrane Database Syst Rev (3): CD004875, 2005.

RETRACTED