

# Extraction of flavonoids from *Chrysanthemum morifolium* and antitumor activity *in vitro*

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**Abstract.** The aim of the present study was to optimize flavonoid extraction from *Chrysanthemum morifolium* and to study the antitumor effects of flavonoids on human gastric cancer MKN45 cells *in vitro*. A single factor experiment was designed and the extraction process was optimized using an orthogonal test. MKN45 cells were treated with different concentrations of flavonoid from *Chrysanthemum morifolium* for 24 and 48 h and the inhibitory effect on the MKN45 cells was evaluated using an MTT assay. Following staining with Annexin V-fluorescein isothiocyanate/propidium iodide, flow cytometry was performed. The optimized flavonoid extraction conditions were as follows: Duration of ultrasonic treatment: 35 min; ethanol concentration: 75%; extraction temperature: 80°C and liquid-to-solid ratio 25: 1. Under the above conditions, the extraction rate of flavonoids was 5.24%. When compared with a blank control group, flavonoids extracted from *Chrysanthemum morifolium* inhibited the proliferation of MKN45 cells in a dose- and time-dependent manner. Furthermore, in cell groups treated with low, moderate and high concentrations of flavonoid, it was observed that the proportion of apoptotic cells increased in a dose-dependent manner. The extraction process optimized by the orthogonal test achieved a high yield and satisfactory extraction efficiency. Additionally, the experiment demonstrated that flavonoids from *Chrysanthemum morifolium* inhibited the growth of MKN45 cells and induced their apoptosis. Thus, flavonoids from *Chrysanthemum morifolium* exerted antitumor effects on MKN45 cells, which may be exploited as a potential antitumor therapeutic for gastric cancer.

## Introduction

*Chrysanthemum morifolium* is a species of perennial plant from the Asteraceae family. Also known as mums, *Chrysanthemum morifolium* is among the four most famous chrysanthemum species in China. According to traditional Chinese medicine principles, *Chrysanthemum morifolium* has been indicated to be 'cool' in nature and 'acid' in taste, and exhibits affinity for the lung and liver systems (1). Furthermore, according to traditional Chinese medicine, chrysanthemum is able to improve eyesight and prevent fatigue (2). Chrysanthemum may be used to treat wind-heat type common cold, headache and dizziness, red and painful eyes and toxin-induced swelling. The main ingredients of *Chrysanthemum morifolium* are volatile oils (3), flavonoids (4), chlorogenic acid (5), polysaccharides (6), phenols (7) and trace elements (8). Flavonoid compounds, triterpenoids and volatile oils are the main active components (9). Flavonoids from *Chrysanthemum morifolium* may significantly improve the activity of antioxidases in the tissues of rats with lead poisoning, relieve lipid peroxidation and antagonize oxidative injury of the brain, liver and kidney (10). In addition, flavonoids from *Chrysanthemum morifolium* downregulated high glucose-induced B-cell lymphoma 2 (Bcl-2)-associated X protein expression and upregulated Bcl-2 in vascular endothelial cells (VECs), while inducing the apoptosis of VECs (11). These benefits may be exploited for the treatment of diabetic angiopathy. Thus, *Chrysanthemum morifolium* may be used as a clinical medicine and for health purposes due to its medicinal value.

Extraction and separation of the active components from medicinal herbs are key to understanding the composition and biological activities of the herbs (12,13). Even for the same medicinal herb, the yield of active components may differ under different extraction conditions. The orthogonal test design is an effective tool for determining the optimal combination of extraction conditions and their optimum levels (14). To understand the effect of 4 independent factors each with 3 factor level values (3<sup>4</sup>), 81 runs are typically required, excluding the repeats for each combination (15). With an L<sub>9</sub>(3<sup>4</sup>) orthogonal array, 9 runs are required (16). The present study used an orthogonal test design to evaluate flavonoid extraction from *Chrysanthemum morifolium* via ultrasonic treatment.

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**Key words:** *Chrysanthemum morifolium*, flavonoids, orthogonal test, flow cytometry, cell apoptosis

The optimal extraction conditions were determined to achieve the optimal yield and extraction efficiency.

In the last 10 years, the incidence of cancer has increased due to the accelerating pace of urban life, environmental pollution and food safety issues, and greater proportions of young adults are now being diagnosed with cancer (17). *Chrysanthemum morifolium* is considered to have a number of pharmacological activities, including the ability to remove free radicals (18) and anti-inflammatory (19), immunoregulatory (20) and antitumor effects (21). For instance, flavonoids extracted from *Chrysanthemum morifolium* exhibited significant cytotoxicity in human colorectal cancer cells (22). The present study evaluated the flavonoids extracted from *Chrysanthemum morifolium*, and an antitumor effect against human gastric cancer MKN45 cells was determined by an MTT assay. In addition, the apoptosis of MKN45 cells was evaluated by flow cytometry. The purpose of this was to confirm the clinical effects of *Chrysanthemum morifolium* on cancer cells.

## Materials and methods

**Materials.** *Chrysanthemum morifolium* was purchased from Tongxiang Datong Industrial & Trading Co., Ltd. (Tongxiang, China); rutin reference standard (batch number 42045) was purchased from the National Institutes for Food and Drug Control (Beijing, China); human gastric cancer MKN45 cells were provided by the Microbiology and Immunology Teaching and Research Section at the School of Basic Medical Sciences of Zhejiang Chinese Medical University (Hangzhou, China).

**Reagents.** Reagents purchased for the present study were 0.25% trypsin-0.02% EDTA (Jinuo Biomedical Technology Co., Ltd., Hangzhou, China), fetal bovine serum (FBS), Roswell Park Memorial Institute (RPMI) 1640 medium (both from Hangzhou Sijiqing Biological Engineering Co., Ltd., Hangzhou, China), an MTT assay kit (Amresco LLC, Solon, OH, USA), an Annexin V-fluorescein isothiocyanate/propidium iodide (FITC/PI) apoptosis detection kit (Nanjing KeyGen Biotech Co., Ltd., Nanjing, China). All other reagents were manufactured domestically and analytically pure.

**Equipment.** A TU-1950 UV-Vis spectrophotometer (Beijing Purkinje General Instrument Co., Ltd., Beijing, China), M5800H-C ultrasonic cleaner (Branson Ultrasonics Co., Ltd., Shanghai, China), Varioskan Flash microplate reader (Thermo Fisher Scientific, Inc., Waltham, MA, USA), IX70 inverted phase-contrast microscope (Olympus Corporation, Tokyo, Japan), FACSCalibur flow cytometer (BD Biosciences, Franklin Lakes, NJ, USA) were used.

**Grouping.** Four cell groups were used: Group A (blank control), which was treated with RPMI-1640 medium alone; group B (low-dose treatment), treated with RPMI-1640 medium containing 5 mg/ml flavonoids; group C (moderate-dose treatment), treated with RPMI-1640 medium containing 10 mg/ml flavonoids; and group D (high-dose treatment), treated with RPMI-1640 medium containing 20 mg/ml flavonoids.

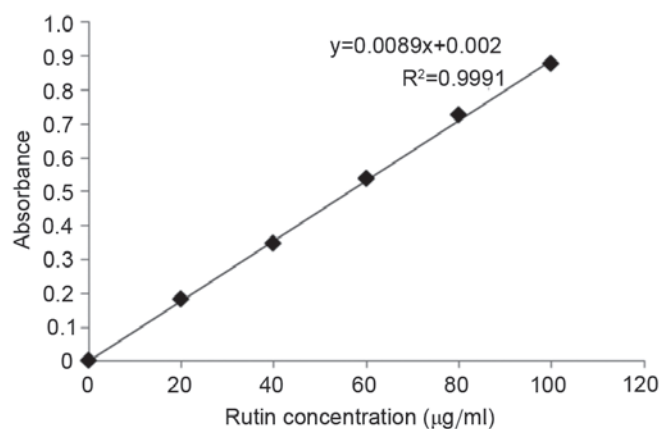


Figure 1. Standard curve regression equation.

## Optimization of the extraction process

**Extraction procedures.** *Chrysanthemum morifolium* weighing 2.0 g was cut into pieces and placed into a 100-ml round-bottom flask. Ultrasonic extraction was performed at different temperatures with the addition of ethanol using a M5800H-C ultrasonic cleaner (Branson Ultrasonics Co., Ltd., Shanghai, China) and at a frequency of 80 kHz. After extraction, a 100 mesh screen (pore size, 0.150 mm) was used to obtain the filtrate, which was centrifuged at 12,000 g at 4°C for 30 min to remove the impurities. Ethanol was removed by rotary evaporation. The supernatant was then collected and condensed at a reduced pressure (-0.08-0.1MPa, 50-65°C for 1 h) to obtain the crude extract of flavonoids.

**Plotting of standard curve.** A rutin reference standard of 0.5 mg was used (rutin was dried at 120°C to 5.0 mg) and placed into a 25.0 ml volumetric flask. Ethanol (60%) was added to a constant volume (25.0 ml) to obtain a 0.2 mg/ml standard solution of rutin. Volumes of the standard rutin solution (0.0, 1.0, 2.0, 3.0, 4.0 and 5.0 ml) were placed into 10.0 ml volumetric flasks. A total of 0.3 ml of 5% sodium nitrite (Sinopharm Chemical Reagent Co., Ltd., Shanghai, China) solution was then added, mixed and allowed to stand at room temperature for 6 min, after which 0.3 ml of 10% aluminum nitrite (Sinopharm Chemical Reagent Co., Ltd.) solution was added, mixed and allowed to stand at room temperature for 6 min. Following the addition of 4.0 ml sodium hydroxide (1.0 mol/l), 60% ethanol was added to a constant volume (10 ml). After standing at room temperature for 10 min, absorbance was measured using a TU-1950 UV-Vis spectrophotometer at 510 nm and a standard curve was plotted with concentration on the x axis and absorbance on the y axis (Fig. 1). A regression equation was fitted as follows:  $y=0.0089x+0.002$  and  $R^2=0.9991$ .

**Determination of flavonoid content.** Crude extract of flavonoids (1 ml) was placed into a tube and subjected to the same treatment protocol of the rutin reference standard. The absorbance was measured and flavonoid concentration was calculated from the standard curve. The extraction efficiency (%) was also calculated as follows: extraction efficiency (%) = extracted flavonoid content / total mass of *Chrysanthemum morifolium* powder x 100 (16).

Table I.  $L_9(3^4)$  orthogonal array.

Level	Factor			
	Duration, min	Ethanol concentration, %	Temperature, °C	Liquid-to-solid ratio
1	30	65	70	20:1
2	35	70	75	25:1
3	40	75	80	30:1

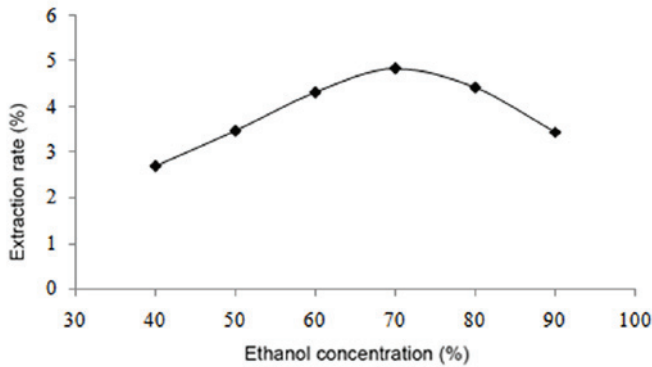


Figure 2. Effect of ethanol concentration of flavonoid extraction.

**Single factor experiment.** The initial extraction parameters were as follows: 60% ethanol, 10:1 liquid-to-solid ratio, 80°C extraction temperature and 30 min duration of ultrasonic treatment. The above conditions were optimized in order to maximize the yield. The following 6 levels were assessed for each factor: 40, 50, 60, 70, 80 and 90% ethanol concentration; 10:1, 15:1, 20:1, 25:1, 30:1 and 35:1 liquid-to-solid ratio; 50, 60, 70, 80, 90 and 100°C temperature; and 10, 20, 30, 40, 50 and 60 min duration. The effect of each factor on the yield was assessed to determine the optimum level for each factor.

**Orthogonal test design.** Based on the above single factor experiment, 4 factors (duration, ethanol concentration, temperature and liquid-to-solid ratio) were selected to form an  $L_9(3^4)$  orthogonal array (Table I).

**Cell Morphology changes.** Human gastric cancer MKN45 cells were seeded at a density of  $1 \times 10^3$  cells/well into 96-well plates and allowed to adhere for 24 h at 37°C in an atmosphere containing 5%  $CO_2$ . Flavonoid solutions, at concentrations of 0, 5, 10 and 20 mg/ml were added to the wells, and the cells were cultured for 24 h at 37°C. Morphological changes were observed using IX70 inverted phase-contrast microscope (magnification, x200) in a blinded manner.

**Detection of antitumor effects of flavonoids.** Human gastric cancer MKN45 cells were placed in RPMI-1640 medium supplemented with 10% FBS and cultured at 37°C in a 5%  $CO_2$  incubator. The culture medium was discarded when MKN45 cells reached 80% confluence, and the cells were washed once/twice with PBS and digested with 2.0 ml 0.25% trypsin-0.02% EDTA to prepare a cell suspension. With cell

density adjusted to  $5-6 \times 10^4$  cells/ml, the cells were inoculated into 96-well plates (100  $\mu$ l/well), and the culture medium was replaced by fresh medium after 24 h 37°C. Subsequently, 0, 5, 10 and 20 mg/ml of crude flavonoids were added to the blank control group (group A) and the three treatment groups (groups B, C and D). Each treatment was replicated three times at 37°C. MTT solution (5 mg/ml) was added 24 or 48 h later (at 37°C) and the culture medium was discarded 4 h later. Dimethyl sulfoxide was then added to dissolve the precipitate and the cells were oscillated for 10 min. The absorbance was measured at 570 nm using Varioskan Flash microplate reader, and the cell inhibition rate was calculated as follows: Cell inhibition rate (%) =  $(1 - OD_{570 \text{ nm}} \text{ value of the treatment group} / OD_{570 \text{ nm}} \text{ value of the blank control group}) \times 100$  (23).

**Flow cytometry of cell apoptosis.** Log-phase MKN45 cells were harvested with cell density adjusted to  $1 \times 10^5$  cells/ml. The cells were inoculated into 6-well plates at 1 ml/well containing RPMI-1640 medium supplemented with 10% FBS. The cells were treated with flavonoids of different concentrations (0.0, 5.0, 10.0 and 20.0 mg/ml) after cell culture for 24 h at 37°C. The cells were then digested with EDTA-free 0.25% trypsin (Jinuo Biomedical Technology Co., Ltd., Hangzhou, China) and centrifuged at  $100 \times g$  for 5 min at 25°C. The supernatant was discarded and the cells were washed twice with pre-cooled PBS. A total of 500  $\mu$ l binding buffer (eBioscience; Thermo Fisher Scientific, Inc.) was added to  $1 \times 10^5$  cells to resuspend them, followed by staining with 5.0  $\mu$ l Annexin V-FITC and 5.0  $\mu$ l PI at room temperature for 15 min in the dark. A total of 400  $\mu$ l binding buffer was then added to cells. As a negative control, a group of cells was not stained with Annexin V-FITC or PI. Cell apoptosis was detected using a FACSCalibur flow cytometer (BD Biosciences, Franklin Lakes, NJ, USA) within 1 h.

**Apoptosis assay using Annexin V-FITC/PI.** Normal, necrotic and apoptotic cells were discriminated by staining with Annexin V-FITC/PI. A dual parameter dot plot was prepared based on the intensity of fluorescence emitted by FITC and PI. Cells were divided into 4 zones: B1, with mechanically damaged cells (Annexin V-FITC-/PI+); B2, with late apoptotic or necrotic cells (Annexin V-FITC+/PI+); B3, with living cells (Annexin V-FITC-/PI-); and B4, with early apoptotic cells (Annexin V-FITC+/PI-). The proportions of the different cells were compared between the treatment groups.

**Statistical analysis.** Statistical analyses were performed using SPSS 19.0 software (IBM Corp., Armonk, NY, USA). The data were presented as the mean  $\pm$  standard deviation of three independent experiments. Statistical differences between the means were analyzed using one-way analysis of variance with Turkey post hoc test.  $P < 0.05$  was considered to indicate statistical significance.

**Results**

**Single factor experiment.** The results demonstrated that the yield of flavonoids increased up to an ethanol concentration of

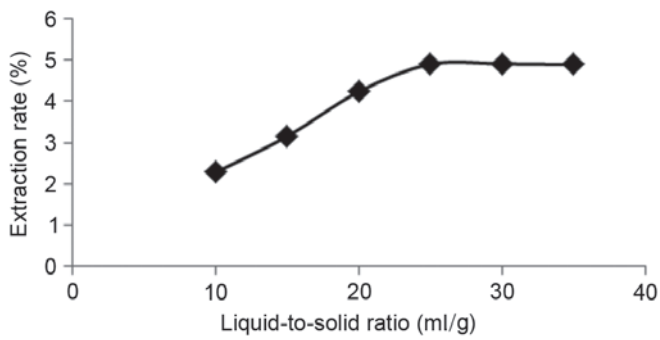


Figure 3. Effect of the liquid-to-solid ratio on flavonoid extraction.

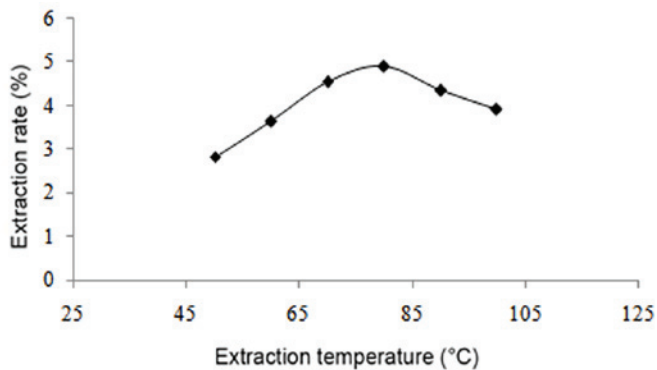


Figure 4. Effect of temperature on flavonoid extraction.

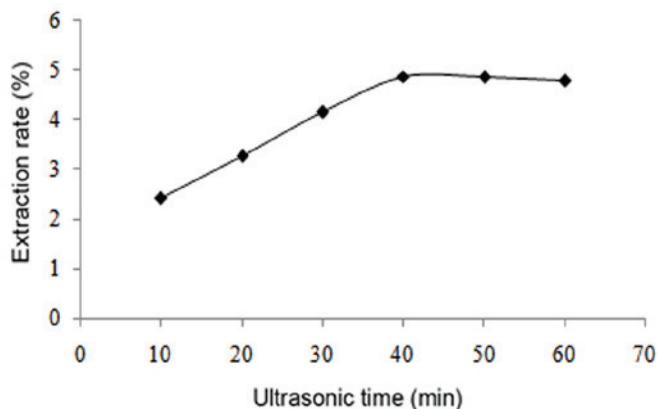


Figure 5. Effect of duration of ultrasonic treatment on flavonoid extraction.

70%, after which the yield of flavonoids decreased (Fig. 2). As the liquid-to-solid ratio increased from 10:1 to 25:1, the yield of flavonoids also increased; however, when the liquid-to-solid ratio was above 25:1, the yield remained mostly unchanged (Fig. 3). With regard to the influence of temperature, the yield increased with a temperature increase from 50 to 80°C, though began to decline when the temperature was >80°C (Fig. 4). Furthermore, the yield of flavonoids increased with a longer duration of ultrasonic treatment, reaching a maximum at 40 min, after which the yield slightly declined (Fig. 5). These results indicated that the optimal extraction conditions were 70% ethanol concentration, 25:1 liquid-to-solid ratio, 80°C temperature and 40 min duration of ultrasonic treatment.

Table II. Orthogonal test design.

Run no.	Factor				Yield, %
	A	B	C	D	
1	1	1	1	1	3.46
2	1	2	2	2	3.96
3	1	3	3	3	4.48
4	2	1	2	3	4.81
5	2	2	3	1	5.03
6	2	3	1	2	4.82
7	3	1	3	2	4.32
8	3	2	1	3	3.64
9	3	3	2	1	3.86
K					
1	3.97	4.20	3.97	4.12	
2	4.89	4.21	4.21	4.37	
3	3.94	4.39	4.61	4.31	
R	0.95	0.19	0.64	0.25	

Data are presented as the mean. 1, 2 and 3 represent the factor levels 1, 2 and 3, respectively (30, 35 and 40 min duration; 65, 70 and 75% ethanol concentration, 70, 75 and 80°C temperature and 20:1, 25:1 and 30:1 liquid-to-solid ratio'. A, duration of ultrasonic treatment; B, ethanol concentration; C, temperature; D, liquid-to-solid ratio; K, the mean of the corresponding levels of flavonoids yield at each factor; R, the difference between the maximum and minimum values of K under each factor.

*Results of orthogonal test.* The duration of ultrasonic treatment had the greatest impact on flavonoids yield (R=0.95), followed by temperature (R=0.64), liquid-to-solid ratio (R=0.25) and ethanol concentration (R=0.19) (Table II). The optimal combination of flavonoid extraction conditions was determined to be 35 min duration of ultrasonic treatment, 75% ethanol concentration, 80°C temperature and 25:1 liquid-to-solid ratio (A<sub>2</sub>B<sub>3</sub>C<sub>3</sub>D<sub>2</sub>, with the letter denoting the factor and number denoting the level). The optimal combination was not included in the orthogonal table, however, the results of a different analysis (the 'extreme difference analysis') indicated what the optimum conditions were, and subsequent experiments demonstrated that the extraction yield was highest under those conditions. In the sixth orthogonal experiment, only the extraction temperature did not meet the optimum combination, but this experiment produced the higher extraction yield (4.82%), which suggested these experimental conditions (Duration of ultrasonic treatment: 35 min; ethanol concentration: 75%; extraction temperature: 80°C and liquid-to-solid ratio 25:1, A<sub>2</sub>B<sub>3</sub>C<sub>3</sub>D<sub>2</sub>) may be optimal. In order to redefine the optimum conditions, the extraction experiments were performed under this condition (A<sub>2</sub>B<sub>3</sub>C<sub>3</sub>D<sub>2</sub>), and the extraction yield of flavonoids reached 5.24%. So this deduced condition was rationally confirmed to be the best combination of different parameters. However, one-way analysis of variance indicated that only the duration of ultrasonic treatment had a significant impact on flavonoid extraction yield (P<0.05; Table III).



Table III. One-way analysis of variance of the orthogonal test data.

Factor	Sum of squares	Degree of freedom	Mean square	F-value	P-value
A	1.743	2	0.872	16.922	a
B	0.067	2	0.034	0.650	-
C	0.621	2	0.311	6.029	-
D	0.103	2	0.052	1.000	-
Error	0.100	2	0.050	-	-

$F_{0.01}(2, 2)=99.00$ ,  $F_{0.05}(2, 2)=19.00$ ; a:  $P<0.05 [F>F_{0.05}(2, 2)]$ .  $F_{0.01}(2, 2)=99.00$ ,  $F_{0.05}(2, 2)=19.00$  are the value obtained by referring to the critical value table of F. P-value of less than 0.05 by the test was considered a statistically significant difference. A, duration of ultrasonic treatment; B, ethanol concentration; C, temperature and D, liquid-to-solid ratio.

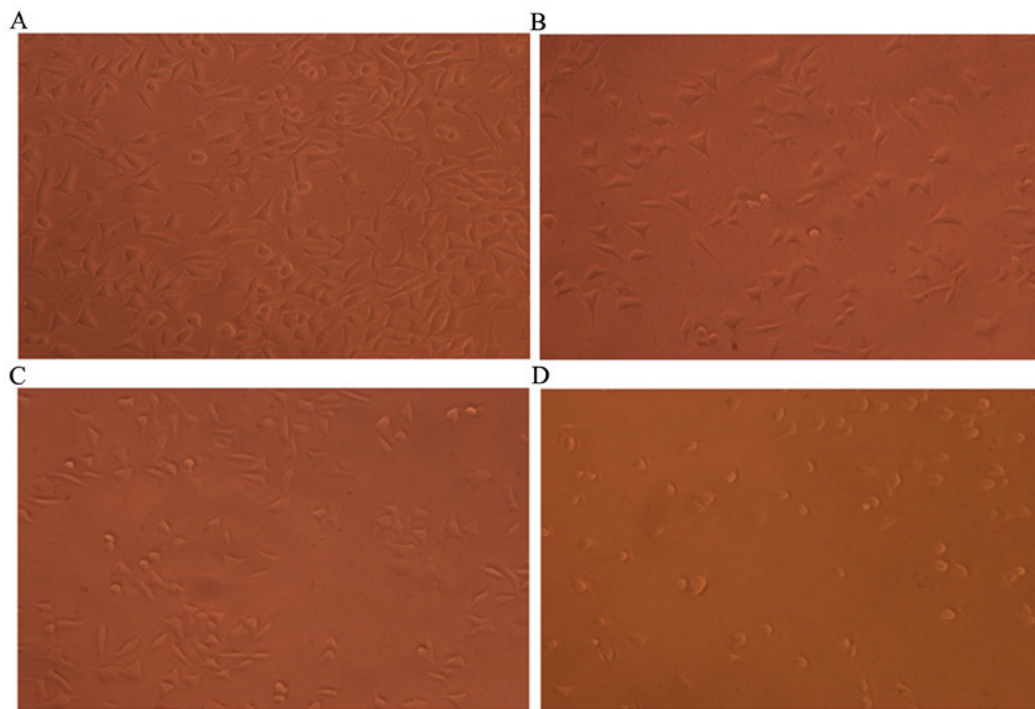


Figure 6. Cell growth in the different treatment groups. (A) Blank; (B) low-dose (5 mg/ml); (C) moderate-dose (10 mg/ml); and (D) high-dose (20 mg/ml) groups. Morphology of MKN45 cells treated with flavonoid (0.0, 5.0, 10.0 and 20.0 mg/ml) for 24 h was observed using a IX70 inverted phase-contrast microscope (magnification, x200).

**Effects of flavonoid concentration on cell morphology.** Morphological observation of MKN45 cells treated with flavonoid (0.0, 5.0, 10.0 and 20.0 mg/ml) for 24 h was performed using IX70 inverted phase-contrast microscopy. A total of 24 h after cells treated with flavonoid, the cells in different concentrations indicated different degrees of adherence. In the blank control group, cells grew normally, exhibiting tight adherence, a regular pattern and distinct cell morphology, and a large number of cells was observed (Fig. 6A). In the low-dose group, a small number of cells lost the original morphology and exhibited a widened intercellular space and irregular arrangement, and a small number of apoptotic cells were present (24) (Fig. 6B). Morphological characteristics of apoptosis were observed in cells: Cell shrinkage, cell size reduction, and membrane blebbing. Compared with the low-dose group, an increased

number of cells in the moderate-dose group lost the original morphology, and more suspended cells and less adherent cells were observed. The number of apoptotic cells was also increased (Fig. 6C). In the high-dose group, the majority of cells lost the original morphology. The cells were round and shrunken, and the large majority were apoptotic (Fig. 6D). These results suggested that flavonoids extracted from *Chrysanthemum morifolium* exerted antitumor effects in a dose-dependent manner.

**Proliferation of MKN45 cells following treatment with different concentrations of flavonoids.** Following drug treatment for 24 and 48 h, cell growth was significantly inhibited in groups B ( $P<0.05$ ), C and D (both  $P<0.01$ ) when compared with the blank control group (group A). Inhibition of cell growth was also significantly greater in groups C and D when

Table IV. Effect of flavonoids on the proliferation activity of MKN45 cells.

Group	24 h		48 h	
	Absorbance	Inhibition rate, %	Absorbance	Inhibition rate, %
A	0.696±0.021	-	0.686±0.012	-
B	0.656±0.046 <sup>a</sup>	5.7	0.650±0.007 <sup>a</sup>	6.6
C	0.548±0.018 <sup>b,c</sup>	21.3	0.506±0.046 <sup>b,c</sup>	27.3
D	0.343±0.020 <sup>b,c</sup>	50.7	0.249±0.017 <sup>b,c</sup>	64.2

Data are presented as the mean ± standard deviation, n=3. <sup>a</sup>P<0.05 and <sup>b</sup>P<0.01 vs. group A; <sup>c</sup>P<0.01 vs. group B. A, blank control; B, low-dose; C, moderate-dose; D, high-dose.

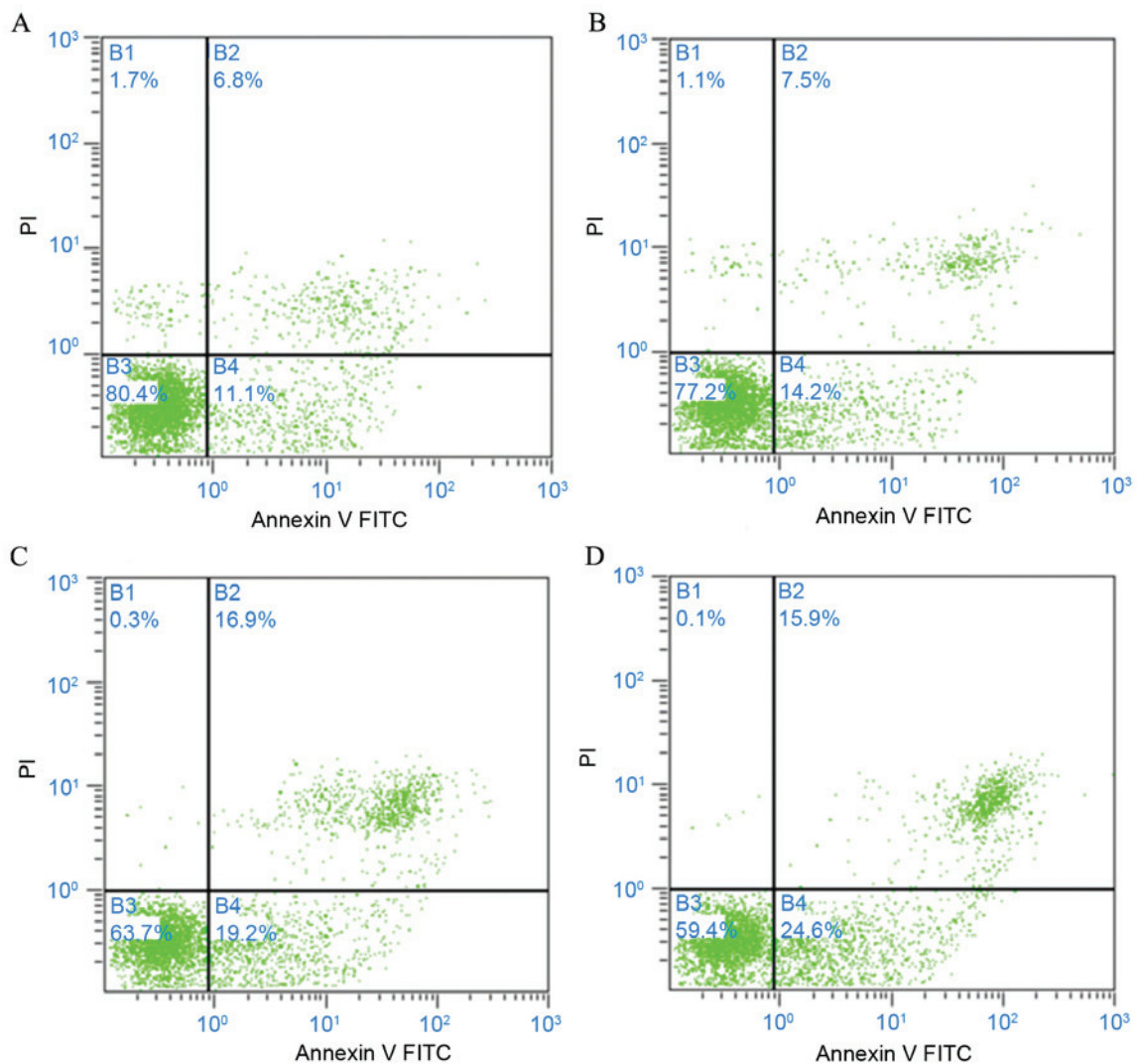


Figure 7. Detection of cell apoptosis by Annexin V-FITC/PI staining of a single experiment. (A) 0; (B) 5; (C) 10; and (D) 20 mg/ml flavonoid treatment groups. B1, mechanically damaged cells (Annexin V-FITC/PI<sup>+</sup>); B2, late apoptotic or necrotic cells (Annexin V-FITC<sup>+</sup>/PI<sup>+</sup>); B3, living cells (Annexin V-FITC/PI<sup>-</sup>); and B4, with early apoptotic cells (Annexin V-FITC<sup>+</sup>/PI<sup>-</sup>). FITC, fluorescein isothiocyanate; PI, propidium iodide.

compared with group B ( $P<0.01$ ). Furthermore, the rate of inhibition markedly increased with time at each concentration (Table IV). These results indicated that flavonoids promoted the apoptosis of MKN45 cells in a dose- and time-dependent manner.

*Pro-apoptotic effects of flavonoids on MKN45 cells.* As observed from dual-parameter dot plots, MKN45 cells of the blank control group were primarily distributed in the B3 zone, indicating that few cells had undergone apoptosis when compared with the other treatment groups (Fig. 7). Analysis

Table V. Pro-apoptotic effect of flavonoids on MKN45 cells.

Group	Apoptotic rate (%)
A	11.73±0.60
B	13.70±0.46 <sup>a</sup>
C	19.43±0.49 <sup>b,c</sup>
D	24.23±0.40 <sup>b,c</sup>

Apoptotic rate is based upon the percentage of cells stained as Annexin V-FITC<sup>+</sup>/PI<sup>+</sup> (necrotic). Data are presented as the mean ± standard deviation, n=3. <sup>a</sup>P<0.05 and <sup>b</sup>P<0.01 vs. group A; <sup>c</sup>P<0.01 vs. group B. A, blank control; B, low-dose; C, moderate-dose; D, high-dose.

of the flow cytometry data indicated that pretreatment with flavonoids induced the apoptosis of MKN45 cells, as demonstrated by a significantly increased rate of apoptosis in groups B (P<0.05), C and D (both P<0.01) when compared with the control group. Furthermore, the rate of apoptosis increased as flavonoid concentration increased, with a significantly greater apoptotic rate observed in groups C and D when compared with group B (P<0.01; Table V).

## Discussion

*Chrysanthemum morifolium* is mainly produced in Tongxiang city in Zhejiang province, and is among the first genuine and edible medicinal materials to be approved by the Ministry of Health (Beijing, China) (25). As one of ‘the eight genuine traditional Chinese Medicines of Zhejiang’ (26), *Chrysanthemum morifolium* may be eaten as food and used as a medicine. *Chrysanthemum morifolium* was recently included in the Chinese Pharmacopoeia (2010 version) (2), and contains flavonoids and triterpenoids as the main active ingredients (27).

Ultrasonic extraction is a novel method used to enhance production yield and to avoid thermal damage (28). It has been used in previous studies to extract natural products (29) and improve solvent extraction, mainly due to the mechanical effects of cavitation bubble collapse, causing better solvent penetration into plant materials (30,31). In the single factor experiment of the present study, the yield of flavonoids reached the maximum 4.87% by using ultrasonication for 40 min. Compared with the traditional extraction method, ultrasonication was simple and time-saving. The result is consistent with previous studies (32,33). Therefore, ultrasonication exhibits strong potential as a method for enhancing production yield during extraction from traditional Chinese medicine. The present findings indicated the effect of ultrasonic extraction on the yield of flavonoids from *Chrysanthemum morifolium*. However, to obtain more active ingredients from *Chrysanthemum morifolium*, further studies such as the addition of ultrasonic frequency are required.

Mathematical modeling is an effective statistical model for investigating the influences of different factors on extraction, as well as indicating the optimal conditions of extraction (34). Among the current modeling methodologies, orthogonal

experimental design is an efficient way for addressing multi-factorial experiments and screening optimum levels by using the orthogonal design table and statistical analysis (14). Factors that typically influence the extraction of flavonoids from *Chrysanthemum morifolium* include the duration of ultrasonic treatment, ethanol concentration, temperature and liquid-to-solid ratio (32,35). Using a single factor experiment and orthogonal test design, the extraction process was optimized in the present study, and the following was determined as the optimal combination of factors: Duration of ultrasonic treatment, 35 min; ethanol concentration, 75%; temperature, 80°C; and liquid-to-solid ratio, 25:1. Under the above conditions, the yield of flavonoids reached a maximum of 5.24%. The duration of ultrasonic treatment exerted the greatest impact on flavonoid yield, followed by temperature, liquid-to-solid ratio and ethanol concentration. Orthogonal experimental design was also used to optimize extraction process in previous reports (32,36,37). Thus, these findings suggest that the optimum conditions for extracting flavonoids may be determined by using orthogonal test design.

The pharmacological activities of *Chrysanthemum morifolium* include reductive effects on blood lipid, blood sugar (38) and blood pressure, and anti-oxidative (39), antitumor (40), neuroprotective (41), hepatoprotective (42) and cardioprotective effects (43). Through an *in vitro* antitumor activity test, the present study observed that flavonoids extracted from *Chrysanthemum morifolium* inhibited the proliferation of MKN45 cells in a dose- and time-dependent manner. Similarly, it had been reported that the flavonoids from *Chrysanthemum morifolium* indicated significant cytotoxicities against human breast cancer cells, liver cancer cells (44) and colon cancer cells (22) by using the MTT assay. Furthermore, Annexin V-FITC/PI staining and flow cytometry were used to discriminate apoptotic and necrotic cells, and results indicated that flavonoids may induce tumor cell apoptosis in a dose-dependent manner. To conclude, *Chrysanthemum morifolium* can be exploited for its medicinal properties against cancer.

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## References

1. Editor Committee of Jiangsu New Medical College Encyclopedia of Traditional Chinese Medicine. Shanghai Science and Technology Press, Shanghai, pp2008-2011, 1995.
2. Chinese Pharmacopoeia Commission, 2010: Pharmacopoeia of the people's Republic of China Version 2010. Chinese Medicine Science Technology Press, Beijing, pp292, 2010.
3. Sun QL, Hua S, Ye JH, Zheng XQ and Liang YR: Flavonoids and volatiles in *Chrysanthemum morifolium* Ramat flower from Tongxiang County in China. Afr J Biotechnol 9: 3817-3821, 2010.
4. Sugawara T and Igarashi K: Identification of major flavonoids in petals of edible *Chrysanthemum* flowers and their suppressive effect on carbon tetrachloride-induced liver injury in mice. Food Sci Technol Res 15: 499-506, 2009.



5. Wang L, Wang T, Guo QS, Huang Y and Xu HK: Comparative study on four major active compounds of *Sanvitalia procumbens* and *Chrysanthemum morifolium* cv 'Hangju' and 'Gongju'. *Zhongguo Zhong Yao Za Zhi* 38: 3442-3445, 2013 (In Chinese).
6. Zheng C, Dong Q, Chen H, Cong Q and Ding K: Structural characterization of a polysaccharide from *Chrysanthemum morifolium* flowers and its antioxidant activity. *Carbohydr Polym* 130: 113-121, 2015.
7. Lin LZ and Harnly JM: Identification of the phenolic components of chrysanthemum flower (*Chrysanthemum morifolium* Ramat). *Food Chem* 120: 319-326, 2010.
8. Niu YF, Han CM, Shao Y and Tao YD: Comparative study on eight trace elements in twelve flower medicines. *Guang Pu Xue Yu Guang Pu Fen Xi* 29: 1997-2000, 2009 (In Chinese).
9. Jin JZ, Wen M and Shen TC: Research progress in chemical components in *Chrysanthemum Morifolium*. *Sci Technol Food Industry* 35: 386-394, 2014.
10. Xia DZ, Lv GY, Yu XF, Wang HM and Yang Q: Antagonism of total flavonoids from *Chrysanthemum morifolium* against lead induced oxidative injury in mice. *Zhongguo Zhong Yao Za Zhi* 33: 2803-2808, 2008 (In Chinese).
11. Liu YH, Zhou DY, Shou CM and Mou X: Effect of flavonoids from *Chrysanthemum morifolium* on Bax and Bcl-2 mRNA expressions induced by high glucose in vascular endothelial cells. *China J Trad Chinese Med Pharm* 30: 2504-2507, 2015 (In Chinese).
12. Tu PF, Shi SP and Jiang Y: Material basis for efficacy of traditional Chinese medicine research ideas and methods overview. *Chinese Herb Med* 43: 209-215, 2012 (In Chinese).
13. Wang SJ, Wu ZF, Yang M, Wang YQ, Wang F, Ke G and Huang JM: Research status and translational application of new extraction techniques of traditional Chinese medicine. *Zhongguo Zhong Yao Za Zhi* 39: 1360-1367, 2014 (In Chinese).
14. Ji LJ, Si YF, Liu HF, Song XL, Zhu W and Zhu AP: Application of orthogonal experimental design in synthesis of mesoporous bioactive glass. *Micropor Mesopor Mat* 184: 122-126, 2014.
15. Gao X, Li Q, Cheng W, Han G and Xuan L: Optimization of high temperature and pressurized steam modified wood fibers for high-density polyethylene matrix composites using the orthogonal design method. *Materials (Basel)* 9: E847, 2016.
16. Wang HJ, Pan MC, Chang CK, Chang SW and Hsieh CW: Optimization of ultrasonic-assisted extraction of cordycepin from *Cordyceps militaris* using orthogonal experimental design. *Molecules* 19: 20808-20820, 2014.
17. Torre LA, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J and Jemal A: Global cancer statistics, 2012. *CA Cancer J Clin* 65: 87-108, 2015.
18. Pongjit K, Ninsontia C, Chaotham C and Chanvorachote P: Protective effect of Glycine max and *Chrysanthemum indicum* extracts against cisplatin-induced renal epithelial cell death. *Hum Exp Toxicol* 30: 1931-1944, 2011.
19. Wu XN, Yu CH and Bao QN: Anti-inflammatory effect and mechanism study of total flavonoids in *Chrysanthemum morifolium* Ramat. *Chinese J Clin Pharmacol Ther* 14: 1000-1004, 2009 (In Chinese).
20. Cheng W, Li J, You T and Hu C: Anti-inflammatory and immunomodulatory activities of the extracts from the inflorescence of *Chrysanthemum indicum* Linné. *J Ethnopharmacol* 101: 334-337, 2005.
21. Yu HN, Sun C, Shan WG and Shen SR: Semi-bionic extraction of active components against stomach cancer from chrysanthemum. *J Zhejiang Univ Technol* 40: 630-633, 2012 (In Chinese).
22. Xie YY, Yuan D, Yang JY, Wang LH and Wu CF: Cytotoxic activity of flavonoids from the flowers of *Chrysanthemum morifolium* on human colon cancer Colon205 cells. *J Asian Nat Prod Res* 11: 771-778, 2009.
23. Wang H, Zhang X, Li Y, Chen R, Ouyang S, Sun P, Pan L, Ren H and Yang B: Antitumor activity of a polysaccharide from longan seed on lung cancer cell line A549 in vitro and in vivo. *Tumour Biol* 35: 7259-7266, 2014.
24. Alenzi FQ: Links between apoptosis, proliferation and the cell cycle. *Br J Biomed Sci* 61: 99-102, 2004.
25. Wu XJ, Yu CH and Bao QN: Anti-inflammatory effect and mechanism study of total flavonoids in *Chrysanthemum morifolium* Ramat. *Chinese J Clin Pharmacol Ther* 14: 1000-1003, 2009 (In Chinese).
26. Yu SK, Zhang Y and Wu XQ: Nutritional components and biological activities of *Chrysanthemum morifolium*. *Food Nutri China* 2: 51-52, 2002 (In Chinese).
27. Wang S, Hao LJ, Zhu JJ, Zhang QW, Wang ZM, Zhang X and Song XM: Study on the effects of sulfur fumigation on chemical constituents and antioxidant activity of *Chrysanthemum morifolium* cv. Hang-ju. *Phytomedicine* 21: 773-779, 2014.
28. Liao JQ, Qu BD, Liu D and Zheng NQ: New method to enhance the extraction yield of rutin from *Sophora japonica* using a novel ultrasonic extraction system by determining optimum ultrasonic frequency. *Ultrason Sonochem* 27: 110-116, 2015.
29. Li X, Wang Z, Wang L, Walid E and Zhang H: Ultrasonic-assisted extraction of polysaccharides from *Hohenbuehelia serotina* by Response surface methodology. *Int J Biol Macromol* 51: 523-530, 2012.
30. Paniwnyk L, Cai H, Albu S, Mason TJ and Cole R: The enhancement and scale up of the extraction of anti-oxidants from *Rosmarinus officinalis* using ultrasound. *Ultrason Sonochem* 16: 287-292, 2009.
31. Hsieh CW, Cheng JY, Wang TH, Wang HJ and Ho WJ: Hypoglycaemic effects of *Ajuga* extract in vitro and in vivo. *J Funct Food* 6: 224-230, 2014.
32. Yin H, Hu YZ, Yang XJ, Sheng J and Wang Z: Study on ultrasonic extraction technics of flavonoids in *Chrysanthemum morifolium*. *Zhongguo Zhong Yao Za Zhi* 29: 424-425, 2004 (In Chinese).
33. Zhang GH, Qiao JW and Zhang RS: Optimization of ultrasonic extraction of flavonoids from *Chrysanthemum morifolium* by response surface methodology. *Asia-Pacific Trad Med* 5: 20-22, 2009.
34. Tao Y, Wu D, Zhang QA and Sun DW: Ultrasound-assisted extraction of phenolics from wine lees: Modeling, optimization and stability of extracts during storage. *Ultrason Sonochem* 21: 706-715, 2014.
35. Yang J, Jiang HD and Xu JH: Study on extraction technics and variety trends of flavonoids in *Chrysanthemum morifolium*. *Zhong Cao Yao* 33: 988-990, 2002 (In Chinese).
36. Yin H, Hu YZ, Yang XJ, Tian YL and Wang Z: Study on extraction technology for extract and flavonoids in *Chrysanthemum morifolium* by orthogonal design. *Zhongguo Zhong Yao Za Zhi* 29: 737-739, 2004 (In Chinese).
37. Xu XQ, Chen HH and Li XM: Optimization of ultrasound-assisted extraction of total flavonoids from *Chrysanthemum indicum* L. *Asia-Pacific Trad Med* 7: 21-22, 2011.
38. Yamamoto J, Tadaishi M, Yamane T, Oishi Y, Shimizu M and Kobayashi-Hattori K: Hot water extracts of edible *Chrysanthemum morifolium* Ramat. exert antidiabetic effects in obese diabetic KK-Ay mice. *Biosci Biotechnol Biochem* 79: 1147-1154, 2015.
39. Kuang CL, Lv D, Huang X, Gao W, Shen GH and Zhang ZQ: A study on the anti-oxidative activity of *Chrysanthemum morifolium* extract using different solvents. *Sci Technol Food Industry* 36: 83-87, 2015.
40. Hu WX, Mao WF, He C, Zhang L, Huang XF, Du WD and Fang BL: *Chrysanthemum morifolium* Ramat sensitizes TNF-related apoptosis inducing ligand-induced apoptosis in human colon cancer cell line DLD-1. *Chin J Pathophysiol* 22: 742-747, 2006.
41. Kim IS, Koppula S, Park PJ, Kim EH, Kim CG, Choi WS, Lee KH and Choi DK: *Chrysanthemum morifolium* Ramat (CM) extract protects human neuroblastoma SH-SY5Y cells against MPP+ induced cytotoxicity. *J Ethnopharmacol* 126: 447-454, 2009.
42. Kang WY, Huang H, Lian TT and Xu QT: A study on hepatoprotective effect of *Chrysanthemum morifolium*. *Natural Product Res Develop* 24: 1634-1636, 2012.
43. Liu YH, Zhou DY, Shou CM and Mou X: The influence of flavonoids from *Chrysanthemum morifolium* on the apoptosis of HUVEC. *Zhejiang J Integrated Traditional Chinese Western Med* 24: 571-572, 2014.
44. Peng YF, Zou WT and Xu CL: Evaluation of flavonoids from flos chrysanthemum ion cell proliferation in human cancer cells. *Chin Pharm J* 45: 1454-1459, 2010 (In Chinese).

