

Glucose affects cell viability, migration, angiogenesis and cellular adhesion of human retinal capillary endothelial cells via SPARC

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Abstract. The expression of secreted protein acidic and rich in cysteine (SPARC) has been recently identified to be associated with the pathology of diabetic retinopathy. Therefore, the present study aimed to evaluate the regulatory role of SPARC in human retinal capillary endothelial cells (HRCECs), following exposure to a high glucose environment *in vitro*. The cell viability, migration, angiogenesis, permeability and SPARC expression levels of HRCECs were measured following treatment with different concentrations of glucose (25, 50 or 100 mM). Lentiviral vectors (LV185-pL_shRNA_mKate2-SPARC-543; target sequence, GGATGAGGACAACAACCTTCT) that inhibit the expression of SPARC were constructed, and HRCECs were evaluated when infected by viruses carrying the lentiviral vectors. Cell viability was examined using the Cell Counting Kit-8 assay. The expression of SPARC in HRCECs increased as the concentration of glucose in the culture medium increased. Relatively high concentrations of glucose significantly inhibited cell proliferation ($P<0.05$), migration ($P<0.05$), angiogenesis ($P<0.01$), and the expression of ZO, occludin, claudin and JAM1 in tight junctions ($P<0.01$), gap junctions (Cx37 and Cx43; $P<0.01$) and adherens junctions (VE-cadherin, CTNNA1 and CTNNB1; $P<0.05$). However, when SPARC was downregulated by lentiviral vectors, the inhibitions induced by high concentrations of glucose were partially reversed. To conclude, the inhibitory effects on cell viability, migration, angiogenesis and cellular adhesion of HRCECs induced by high concentrations of glucose were reversed once the expression of SPARC was inhibited. These

findings suggest that SPARC may serve an important role in pathogenesis of diabetic retinopathy.

Introduction

Diabetic retinopathy is recognized as the most important manifestation of diabetic microvascular disease and the leading cause of vision loss, particularly among adults aged 20-74 years (1). Recently, the disease has become the fifth most common cause of severe visual impairment worldwide (2,3). For patients with diabetic retinopathy, neovascularization is an important event involving various secretory proteins, including multiple growth factors, and angiogenesis has been recognized as the therapeutic target for diabetic retinopathy (4,5).

Secreted protein acidic and rich in cysteine (SPARC), also known as osteonectin or BM-40, is a widely expressed profibrotic protein with pleiotropic roles that has been studied under a variety of conditions (6,7). **Previous results have demonstrated** that SPARC may be a key component in the pathologies associated with obesity and diabetes (8). Furthermore, elevated plasma SPARC levels have been indicated to be associated with insulin resistance, dyslipidemia and inflammation in gestational diabetes mellitus (9). A previous study suggested that elevated plasma levels of SPARC were exhibited in patients with newly diagnosed type 2 diabetes mellitus (10). SPARC gene expression has also been revealed to be increased in diabetes-associated mesenteric vascular hypertrophy (11). However, the role of SPARC in glucose-mediated damage of endothelial cells remains unclear.

To study the role of SPARC in diabetic retinopathy, an *ex vivo* model of human retinal capillary endothelial cells (HRCECs) in the presence of high concentrations of glucose was established. The expression of SPARC in HRCECs was altered and cell viability, migration, angiogenesis and cellular adhesion were evaluated.

Materials and methods

Construction of lentiviral vectors. Viral vectors were constructed using 293T cells (cat. no. CRL-11268; American Type Culture Collection, Manassas, VA, USA). The 293A cells (cat. no. R70507; Invitrogen; Thermo Fisher Scientific, Inc.,

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Waltham, MA, USA) were used for titration of lentiviral vectors by reverse transcription-quantitative-polymerase chain reaction (RT-qPCR). The 293T and 293A cell lines were cultured at 37°C in Opti-MEM (cat. no. 31985; Gibco; Thermo Fisher Scientific, Inc.) supplemented with 10% fetal bovine serum (cat. no. SH30084.03, Hyclone Laboratories, Inc., Logan, UT, USA). Lentiviruses were produced using a second-generation packaging system as described previously (12). Briefly, 6×10^6 293T cells were plated in a 15-cm dish and transfected with 9 μ g Packaging Mix (cat. no. K4975-00, Invitrogen; Thermo Fisher Scientific, Inc.), 3 μ g transfer plasmid and 36 μ l Lipofectamine 2000 (cat. no. 11668019, Invitrogen; Thermo Fisher Scientific, Inc.), which were combined in 5 ml Opti-MEM. The transfection mix was removed once cells were cultured for 6 h at 37°C, and fresh culture medium was added. Virus-containing medium was collected at 48 h post-transfection. Subsequently, the collected medium was filtered through a cellulose acetate membrane (pore size, 0.45 μ m).

The 293A cell lines were used to detect titration of lentiviral vectors. The 293A cells were plated into each well of a 24-well plate and transfected using various quantities of lentivirus (1, 0.1 and 0.01 μ l). Genomic DNA was extracted and the provirus titer was calculated using RT-qPCR at 72 h post-transfection. β -actin was used as the housekeeping gene. The target sequence for SPARC knockdown was as follows: 5'-GGATGAGGACAACAACCTTCT-3'.

Expression of SPARC in HRCECs. HRCECs (BeNa Culture Collection, Shanghai, China; http://www.bncc.org.cn/pro/p92/p_339792.html; primary cells, passage 4-5) were cultured in Dulbecco's modified Eagle's medium without glucose (cat. no. 11966025, Gibco; Thermo Fisher Scientific, Inc.) supplemented with 10% fetal bovine serum in a cell incubator with 5% CO₂ at 37°C. The 3rd to 5th passaged cells were used for all experiments as described previously (13,14). Different concentrations of glucose (25, 50 or 100 mM) were added in culture medium, respectively. RT-qPCR was performed to evaluate SPARC expression in HRCECs treated with different concentrations of glucose. HRCECs were then infected with viruses carrying LV185-pL_shRNA_mKate2-SPARC-543. HRCECs infected with empty vectors (negative control) or HRCECs that were not infected (blank control) served as controls. The expression of SPARC was assessed 48 h following this.

Cell viability assay. To determine the effects of SPARC on cell viability following treatment with different concentrations of glucose, HRCECs (1×10^4) were plated in a 96-well plate and infected with LV185-pL_shRNA_mKate2-SPARC-543 or empty vectors. Subsequently, 10 μ l of Cell Counting Kit-8 (CCK-8) reagent (cat. no. CK04-05, Dojindo Molecular Technologies, Inc., Kumamoto, Japan) was added into each well for each of the three groups, respectively. The absorbance at 450 nm was measured to evaluate cell viability at 24, 48 and 72 h, respectively. Experiments were performed a total of three times.

Transwell migration assay. Cell migration was assessed by Transwell assay. Transwell plate was pretreated using Dulbecco's modified Eagle's medium (DMEM) without

glucose (cat. no. 11966025; Thermo Fisher Scientific, Inc.) for 30 min. HRCECs transfected with viruses carrying LV185-pL_shRNA_mKate2-SPARC-543, or negative control vectors were resuspended at a cell density of 1×10^5 cells/ml in DMEM without glucose. Cell suspension (200 μ l) was added to the upper chamber and 800 μ l culture medium with different concentrations of glucose (25, 50 or 100 mM) was added to the lower chamber. Cells were allowed to migrate through the membrane for 24 h. Following incubation, cells on top of the filter were scraped off using a cotton swab. The membrane was subsequently fixed in 4% paraformaldehyde for 10 min at room temperature, and stained with hematoxylin and eosin for 10 min at room temperature. The inserts were mounted on a slide, and the number of cells migrated to the bottom of filter was determined by counting 10 high-power fields with a bright field microscope (magnification, x200).

Measurement of angiogenesis. HRCECs were trypsinized and resuspended in extracellular matrix (ECM) culture media with 10% fetal bovine serum (Hyclone). Cells from the experimental groups were plated at a density of 1.5×10^4 cells/well in triplicate in 96-well plates, respectively. Plates were incubated for 24 h at 37°C prior to photomicroscopy. Images from each well were taken at a magnification of x4 using an Olympus microscope (cat. no. CKX31; Olympus Corporation, Tokyo, Japan) every 2 h. Image-Pro Plus software (version 6.0.0.260; Media Cybernetics, Inc., Rockville, MD, USA) was utilized to calculate the length of blood vessels formed (15,16). Statistical analysis was subsequently performed.

RT-qPCR. HRCECs were cultured in medium containing 25, 50 or 100 mM glucose, respectively. Following 24 h, total RNA was extracted from HRCECs. The expression levels of the following genes were detected using RT-qPCR: Genes associated with tight junctions, including zonula occludens (ZO)-1, occludin, claudin and junctional adhesion molecule 1 (JAM1); genes associated with gap junctions, including connexin (Cx)37, Cx40 and Cx43; genes associated with adherens junctions, including vascular endothelial (VE)-cadherin, catenin alpha 1 (CTNNA1), catenin beta 1 (CTNNB1) and catenin delta 1 (CTNND1). Total RNA was extracted using TRIzol (cat. no. 15596018; Invitrogen; Thermo Fisher Scientific, Inc.) according to the manufacturer's instructions. A cDNA synthesis kit (SuperScript First-Strand Synthesis System; cat. no. 11904-018; Invitrogen; Thermo Fisher Scientific, Inc.) was used for RT. Each reaction contained 0.5 μ l of random primers (0.2 μ g/ μ l) and 1 μ l of SuperScript III reverse transcriptase (200 U/ μ l). The specific primers used are listed in Table I. qPCR was performed by utilizing the SYBR® Premix Ex Taq (cat. no. RR420L; Takara Bio, Inc., Otsu, Japan). The following PCR conditions were used: Pre-denaturing at 95°C for 2 min; denaturing at 95°C for 10 sec; and annealing and polymerization at 60°C for 30 sec and 70°C for 45 sec. A total of 40 PCR cycles were performed. PCR was performed using a CFX96 Touch Real-Time PCR Detection System. Gene expression was determined as the ratio of relative optical density of target gene to β -actin. The 2^{- $\Delta\Delta$ C_q} method was utilized to measure PCR results (17).

Table I. Primers used in reverse transcription-quantitative polymerase chain reaction.

Genes	Primers	Sequences (5' to 3')
SPARC	SPARC-F	AGGAAACCGAAGAGGAGG
	SPARC-R	GCAAAGAAGTGGCAGGAA
ZO-1	ZO-1-F	TATTCACGCAGTTACGAGCAAG
	ZO-1-R	AAGGTATCAGCGGAGGGACA
Occludin	Occludin-F	AAGCCAAACCTCTGTGAGCA
	Occludin-R	GGAGTGTTCAGCCCAGTTGA
Claudin	Claudin-F	TTGGGCTTCATTCTCGCCTT
	Claudin-R	GAGGATGCCAACCACCATCA
JAM1	JAM1-F	AATCTGCACTCAACTGCCCA
	JAM1-R	CATTTTCAGCCGCTCTAGCCT
Cx37	Cx37-F	ACCATGCCCCACCTACAATG
	Cx37-R	CAGCCAGACTTCTCAGGACC
Cx40	Cx40-F	GCATTGTGCTTCTCGGGTTC
	Cx40-R	CTATCACACACCTCTCGGC
Cx43	Cx43-F	TTCAAGGGCGTTAAGGATCG
	Cx43-R	TAACCAGCTTGACCCAGGA
VE-cadherin	VE-cadherin-F	AAGCAGAGCTGGGTCCTTG
	VE-cadherin-R	GAAGCTCTTGGCTCTGGAGG
CTNNA1	CTNNA1-F	TAACAGCATCGGGGTCTGGTTG
	CTNNA1-R	GGCTGGTGTAAATCAGCACT
CTNNB1	CTNNB1-F	GTCTGAGGACAAGCCACAAGA
	CTNNB1-R	GCCACCCATCTCATGTTCCA
CTNND1	CTNND1-F	AGCTGGCTGCCTTAGGAATG
	CTNND1-R	ATCTTTGCCCTTCTTGGCCC

SPARC, secreted protein acidic and rich in cysteine; ZO, zonula occludens; JAM1, junctional adhesion molecule 1; Cx, connexin; VE, vascular endothelial; CTNNA1, catenin alpha 1; CTNNB1, catenin beta 1; CTNND1, catenin delta 1; F, forward; R, reverse.

Western blot analysis. Protein expression levels of SPARC in HRCECs were detected using western blot analysis. HRCECs were lysed in lysis buffer (cat. no. P0013, Beyotime Institute of Biotechnology, Haimen, China) at 4°C with inhibitors of phosphatase and protease (cat. no. P1045, Beyotime Institute of Biotechnology). The lysis mixture was centrifuged at 4°C for 10 min at 10,000 x g, and the supernatant containing cellular proteins was utilized in the following experiments. The protein concentration was measured using a BCA kit. Proteins were separated using SDS-PAGE (10% gels, 40 µg per lane; 120 V). The separated proteins were then transferred to polyvinylidene fluoride membranes (100 V for 120 min; cat. no. FFP24, Beyotime Institute of Biotechnology). The membranes were blocked with 5% non-fat milk at room temperature for 1 h and subsequently incubated with anti-SPARC (cat. no. ab225716; Abcam; 1:1,000) and anti-β-actin (cat. no. ab8227; Abcam; 1:1,000) primary antibodies against SPARC at 4°C overnight. Membranes were washed with Tris-buffered saline containing Tween 20 and incubated with horseradish peroxidase-conjugated goat anti-rabbit secondary antibody (cat. no. ab6721; Abcam; 1:2,000) at room temperature for 1 h. Membranes were incubated in enhanced chemiluminescence solution (cat. no. P0018A, Beyotime Institute of Biotechnology). Images were captured on film (cat. no. FF057, Beyotime Institute of

Biotechnology) in a dark room. Experiments were repeated for 3 times. Blot images were quantified in greyscale using ImageJ (version 1.5.2; National Institute of Health, Bethesda, MD, USA).

Statistical analyses. Data were analysed using GraphPad Prism 5.0 software (GraphPad Software Inc., La Jolla, CA, USA). Data were presented as mean ± standard deviation. Differences among three groups were measured by analysis of variance followed by the Bonferroni's post-hoc test for multiple comparisons. Differences between two groups were measured using a Student's t-test. P<0.05 was considered to indicate a statistically significant difference.

Results

Treatment with glucose increases SPARC expression levels in HRCECs. The mRNA expression levels of SPARC in HRCECs was detected by RT-qPCR at 24, 48 and 72 h, respectively. Compared with the 25 mM glucose group, the mRNA expression levels of SPARC in the 100 mM glucose group were significantly increased at 24, 48 and 72 h (P<0.05 and P<0.01; Fig. 1A). In addition, protein expression levels of SPARC in HRCECs were detected by western blot analysis at 72 h. Compared with the

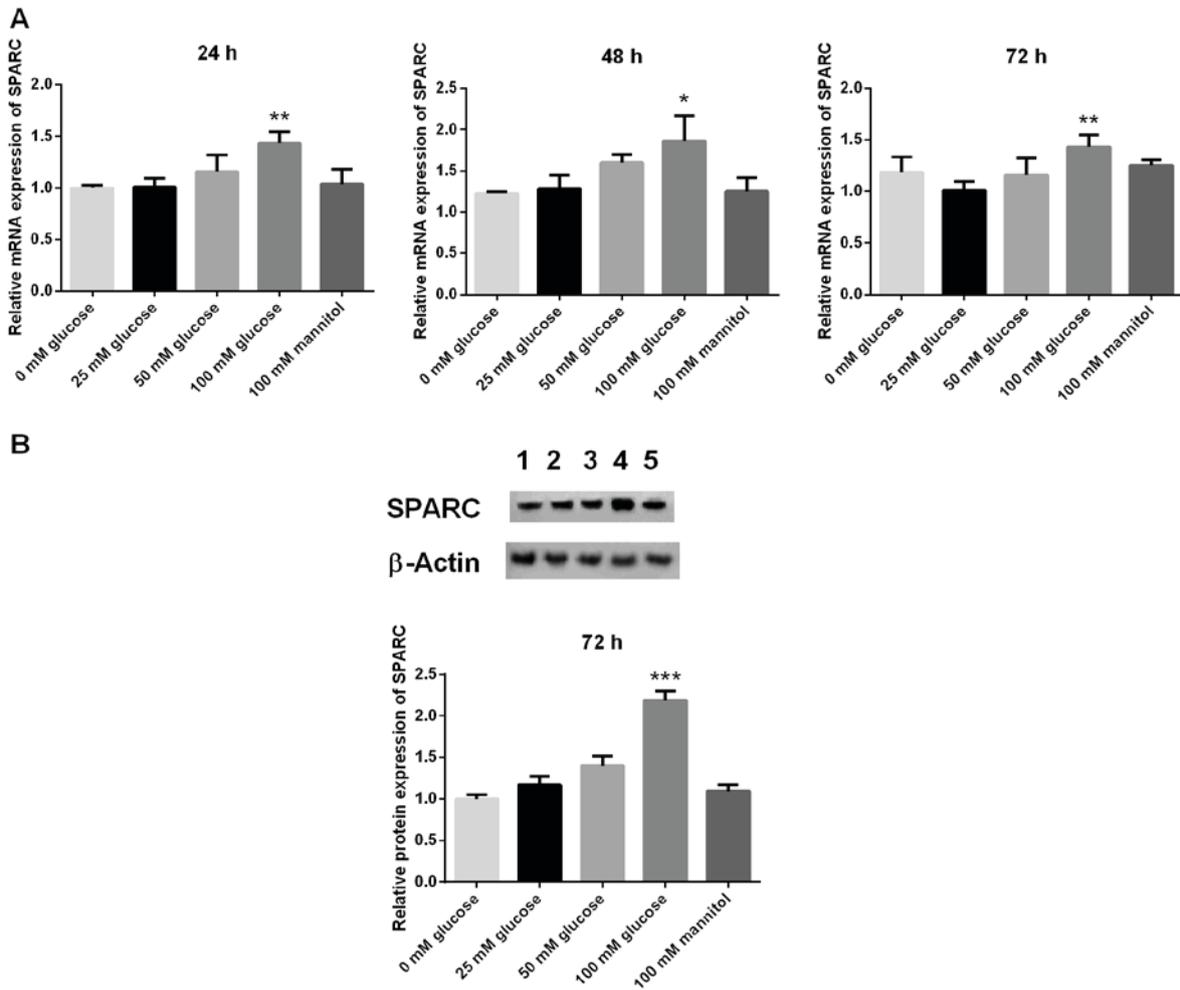


Figure 1. Expression of SPARC in HRCECs treated with 100 mM glucose is significantly increased. (A) mRNA and (B) protein expression levels of SPARC. The mRNA expression levels of SPARC in HRCECs was detected by reverse transcription-quantitative polymerase chain reaction at 24, 48, and 72 h, respectively. Western blot analysis was used to detect the protein expression levels of SPARC. Data were presented as the mean \pm standard deviation (n=3). *P<0.05, **P<0.01 and ***P<0.001 vs. 25 mM glucose group. SPARC, secreted protein acidic and rich in cysteine; HRCECs, human retinal capillary endothelial cells; 1, blank; 2, 25 mM glucose; 3, 50 mM glucose; 4, 100 mM glucose; 5, 100 mM mannitol.

25 mM glucose group, the protein expression levels of SPARC in the 100 mM glucose group were significantly increased at 72 h (P<0.001; Fig. 1B). HRCECs without treatment (blank group) and 100 mM mannitol served as controls.

SPARC interference increases the viability of HRCECs treated with glucose. The success of SPARC interference was confirmed by RT-qPCR. The expression of SPARC in the SPARC interference group was significantly decreased compared with blank or negative control groups (P<0.001; Fig. 2). The interference efficiency was 89.34% (Fig. 2). Cell viability was detected using the CCK-8 assay. The SPARC interference group exhibited significantly increased cell viability when compared with the blank or negative control groups at 24, 48 and 72 h (all P<0.05; Fig. 3).

SPARC interference enhances the migration of HRCECs treated with glucose. The migration ability of HRCECs was detected using the Transwell assay. The SPARC interference group exhibited significantly increased cell migration following treatment with 25, 50 and 100 mM glucose respectively, compared with the negative control group (P<0.05;

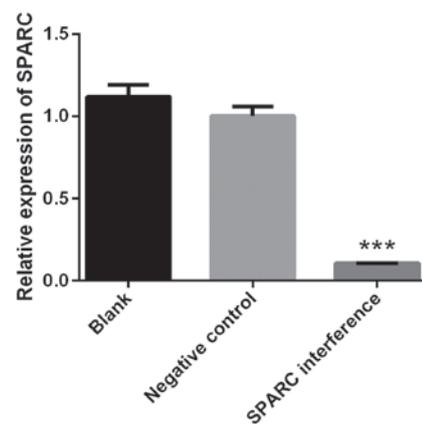


Figure 2. Confirmation of SPARC interference. The success of SPARC interference was confirmed by reverse transcription-quantitative polymerase chain reaction. Data were presented as the mean \pm standard deviation (n=3). ***P<0.001 vs. blank or negative control group. SPARC, secreted protein acidic and rich in cysteine.

Fig. 4). Results indicated that the migration ability was higher when the concentration of glucose was lower.

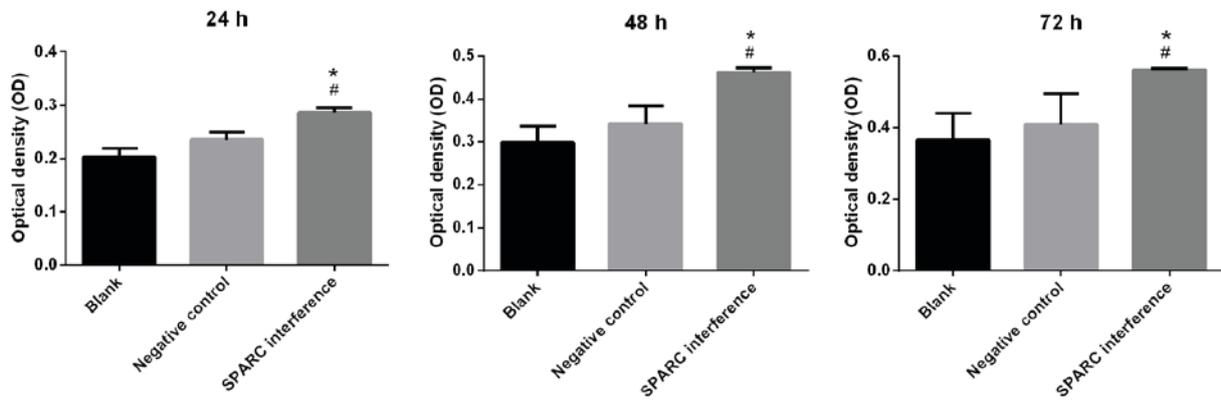


Figure 3. SPARC interference group increases the viability of human retinal capillary endothelial cells. Cell viability was detected using the Cell Counting Kit-8 assay. Data were presented as the mean \pm standard deviation (n=3). *P<0.05 vs. blank group; #P<0.05 vs. the negative control group. SPARC, secreted protein acidic and rich in cysteine.

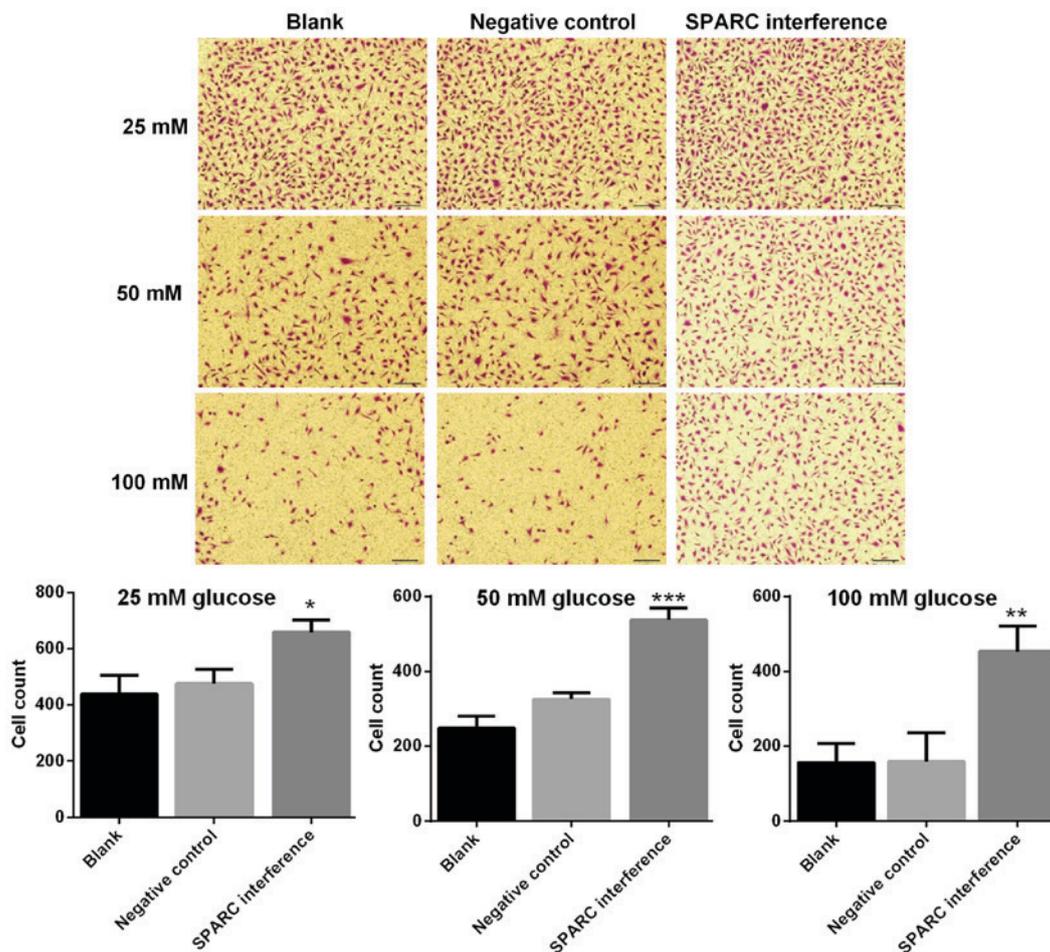


Figure 4. SPARC interference increases the migration ability of HRCECs treated with glucose. HRCECs were incubated with glucose for 24 h. Cell migration ability of HRCECs was detected using the Transwell assay (magnification, x100). Data were presented as mean \pm standard deviation (n=3). *P<0.05, **P<0.01, and ***P<0.001 vs. negative control group. SPARC, secreted protein acidic and rich in cysteine; HRCECs, human retinal capillary endothelial cells.

SPARC interference increases the length of blood vessels treated with glucose. Image-Pro Plus software was utilized to calculate the length of blood vessels formed with HRCECs. Compared with the negative control group, the SPARC interference group exhibited significantly increased vascular length following treatment with 25, 50 and 100 mM glucose (P<0.01 and P<0.001; Fig. 5).

SPARC interference increases the expression of genes associated with tight junctions, gap junctions and adherens junctions. The expression levels of genes associated with tight junctions (ZO-1, occludin, claudin and JAM1), gap junctions (Cx37, Cx40 and Cx43), and adherens junctions (VE-cadherin, CTNNA1, CTNNB1 and CTNND1) were detected using RT-qPCR. The gene expression levels of SPARC in the

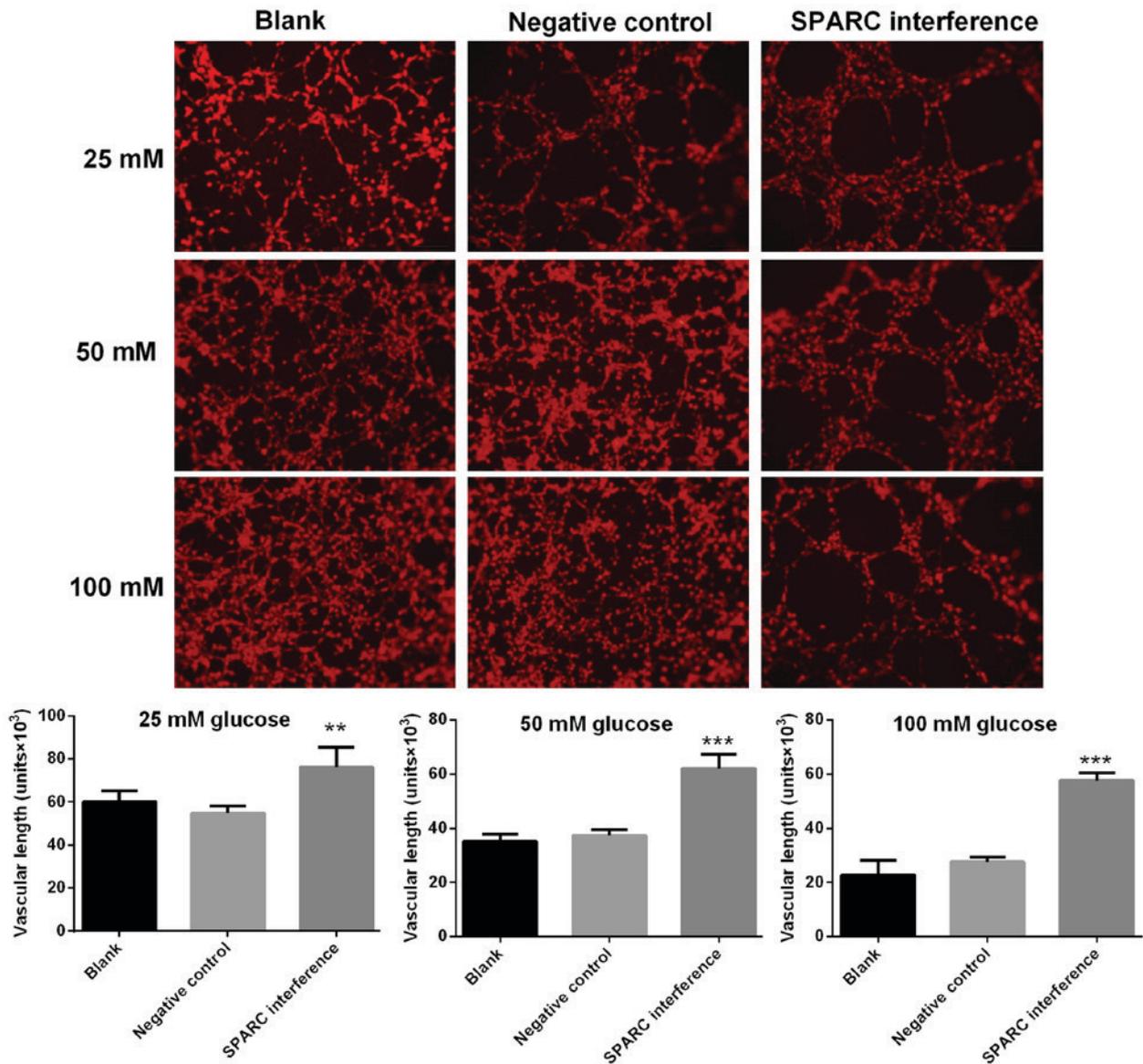


Figure 5. SPARC interference increases the vascular length of HRCECs treated with glucose. HRCECs were incubated with glucose for 24 h. Image-Pro Plus software was utilized to calculate the length of blood vessels formed. Data were presented as mean ± standard deviation (n=3). **P<0.01 and ***P<0.001 vs. negative control group. SPARC, secreted protein acidic and rich in cysteine; HRCECs, human retinal capillary endothelial cells.

SPARC interference group that received treatment with 25, 50 and 100 mM glucose were significantly decreased compared with the negative control group (P<0.01 and P<0.001; Fig. 6). Furthermore, following treatment with 25, 50 and 100 mM glucose, the SPARC interference group demonstrated significantly increased expression levels of genes associated with tight junctions (ZO, occludin, claudin and JAM1; P<0.01 and P<0.001; Fig. 7). In addition, following treatment with glucose, the SPARC interference group exhibited increased expression levels of genes associated with gap junctions when compared with the negative control group. These differences were indicated to be significant with regard to Cx37 in the presence of 25, 50 and 100 mM glucose and Cx43 in the presence of 25, 50 and 100 mM (P<0.01 and P<0.001; Fig. 8). Furthermore, following treatment with glucose, the SPARC interference group displayed significantly increased expression levels of genes associated with adherens junctions compared with the negative control group (VE-cadherin and CTNNA1, significant

in the presence of 50 and 100 mM glucose; CTNND1 and CTNND1, significant in the presence of 25, 50 and 100 mM glucose; P<0.05, P<0.01 and P<0.001; Fig. 9).

Discussion

The present study demonstrated that the cell viability, migration, cellular adhesion, and angiogenesis were significantly inhibited by high concentrations of glucose, and the inhibition could be reversed once the expression of SPARC was inhibited.

SPARC is a widely expressed profibrotic protein with pleiotropic roles, which have been studied in a variety of conditions (18,19). Previous evidence has demonstrated that SPARC was associated with human obesity and diabetes (11). In the present study, HRCECs were cultured *in vitro* and a stably transfected SPARC interference cell line was established. Notably, culture medium may contain glucose (20). However, the same volume and type of culture medium was added into

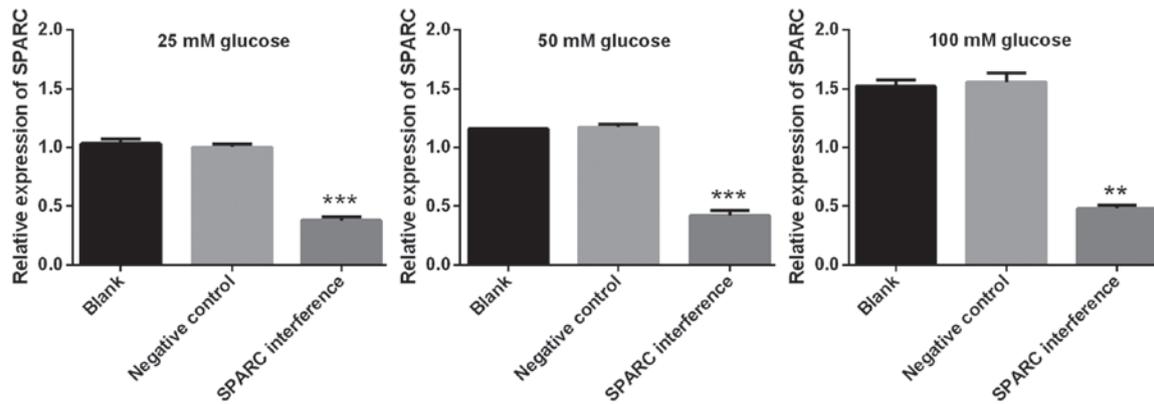


Figure 6. Expression of SPARC in HRCECs treated with glucose. HRCECs were incubated with glucose for 24 h. Data were presented as mean \pm standard deviation (n=3). **P<0.01 and ***P<0.001 vs. negative control group. SPARC, secreted protein acidic and rich in cysteine; HRCECs, human retinal capillary endothelial cells.

blank, negative control and SPARC groups. Therefore, the presence of any glucose in culture medium did not alter the comparisons among three experimental groups.

SPARC is a modulator of cell-surface interaction (21). Tissue physiology can be modulated by SPARC by altering cell proliferation and migration, and interactions between cell and ECM (22). A previous study revealed that adipocytes were recognized as the major source for producing circulating SPARC, and insulin resistance in adipose tissue may be caused by increased SPARC expression (23). Notably, the role of SPARC in diabetic retinopathy has been rarely studied. However, the role of SPARC in diabetes has been extensively studied. Xu *et al* (9) and Wu *et al* (10) have demonstrated that SPARC is an independent negative indicator of diabetes, which correlates significantly with inflammation, dyslipidemia and insulin resistance. In the present study, the expression of SPARC was significantly increased when HRCECs were stimulated with high concentrations of glucose. Watanabe *et al* (24) demonstrated that the expression of SPARC was high in proliferative diabetic retinopathy using oligo-cap signal sequence trap strategy. These findings indicate that SPARC may serve a potential regulatory role in patients with diabetic retinopathy, which is a major complication among patients with diabetes (25).

The present study revealed that high concentrations of glucose inhibited cell viability and migration, which are essential for the turnover and regeneration of capillary endothelial cells (26). HRCECs may be vulnerable to damage induced by pathological processes of diabetic retinopathy, including oxidative stress, inflammation, ischemia-reperfusion and hypoxia (27). Furthermore, impaired endothelial cells, along with other vascular dysfunctions, can act as an important implication for early diabetic retinopathy (28). The present study suggested that downregulation of SPARC increased cell viability and migration, which may have protective effects on capillary endothelial cells in diabetic retinopathy.

The present data revealed that SPARC was highly expressed, and angiogenesis was inhibited, when HRCECs were cultured with high concentrations of glucose. Furthermore, when the expression of SPARC was inhibited, angiogenesis was recovered. Previous data demonstrated that SPARC was cell-type specific in angiogenesis (29).

Until now, to the best of our knowledge no direct evidence has revealed that SPARC is associated with angiogenesis in retinopathy. In gastric cancer, Zhang *et al* (30) demonstrated that the suppression of angiogenesis by SPARC may be mediated by downregulating the expression of vascular endothelial growth factor (VEGF) and matrix metalloproteinases (MMP-7). Notably, overexpression of SPARC has also been demonstrated to be involved in radiation-reduced tumor angiogenesis by downregulating VEGF-A via microRNA-410 (31). The pathway that is associated with angiogenesis in retinopathy and is regulated by SPARC remains unclear. However, the present data suggested that the cell viability, migration and permeability was significantly associated with SPARC expression. Further studies are required to fully explore the possible molecular mechanisms of SPARC in angiogenesis in patients with diabetic retinopathy.

The present findings also indicated that high concentrations of glucose significantly inhibited the expression of genes associated with cell tight junctions, gap junctions and adherens junctions. However, when SPARC was downregulated, the expression levels of those genes increased. Previous research has demonstrated that the collapse of cell junctions caused hyperpermeability of retinal capillary endothelial cells and vascular leakage (32). In patients with diabetic retinopathy, such hyperpermeability caused leakage of macromolecules, proteins and electrolytes into tissue space with sera, thereby increasing interstitial osmolarity and exacerbating vascular leakage. This in turn formed severe diabetic macular edema (DME) (33). Nowadays, DME is known as one of most severe complications and leading cause of vision loss in diabetic retinopathy (34). SPARC, as a key regulator in cell permeability, may be promising in treatment of DME in future clinical practice.

In conclusion, the present data demonstrated that cell viability, migration, cellular adhesion and angiogenesis of retinal capillary endothelial cells exposed to high concentrations of glucose may be mediated, at least in part, by changes in the expression of SPARC. Alterations of SPARC expression may contribute to the development and pathogenesis of diabetic retinopathy. Although further studies are required to fully clarify the role of SPARC in neovascularization and

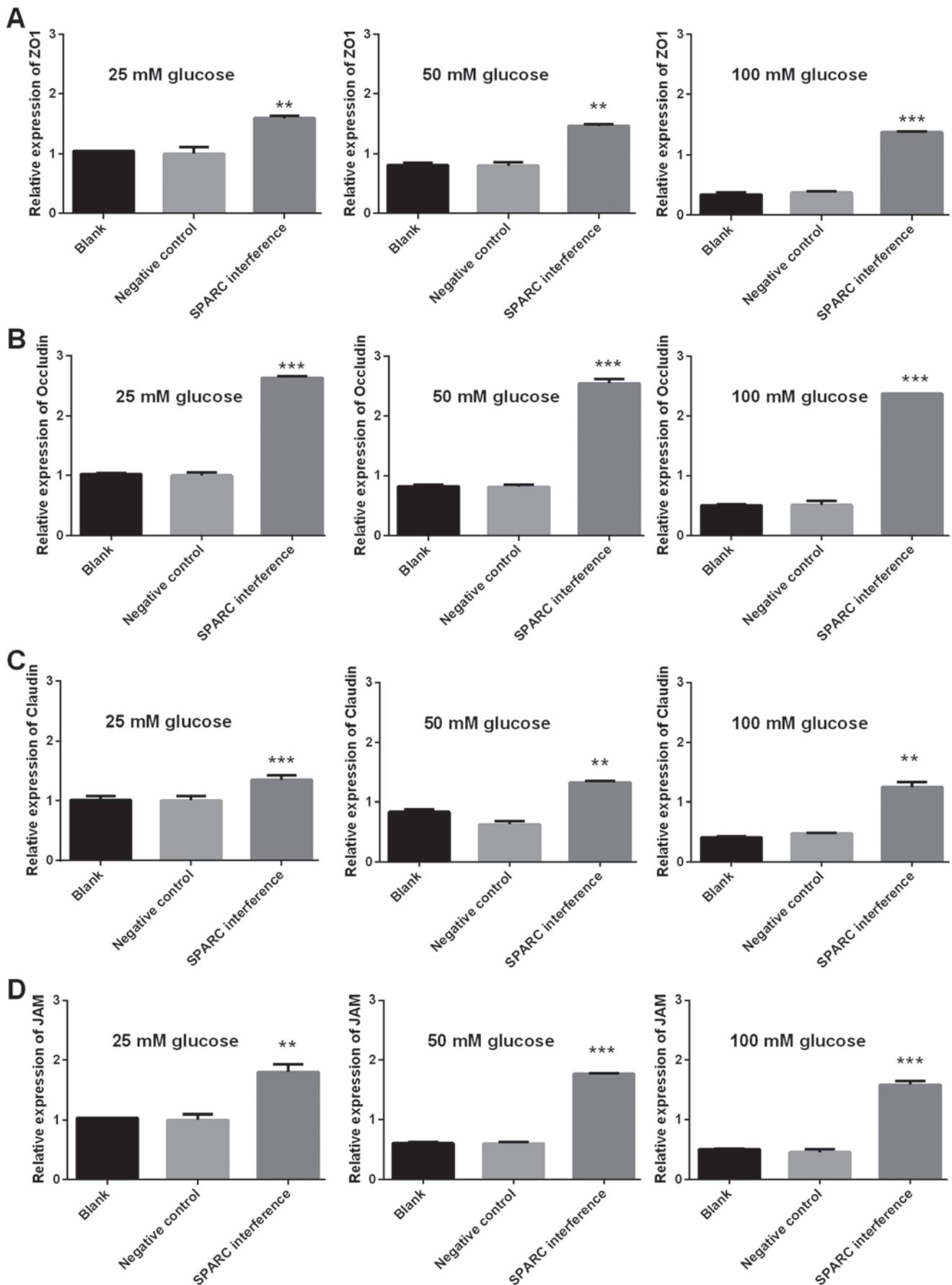


Figure 7. SPARC interference increases the expression of tight junction-associated expression of genes in HRCECs treated with glucose. (A) Expression of ZO1, (B) occluding, (C) claudin and (D) JAM1. HRCECs were incubated with glucose for 24 h. Gene expression was detected using reverse transcription-quantitative polymerase chain reaction. Data were presented as mean \pm standard deviation (n=3). **P<0.01 and ***P<0.001 vs. negative control group. SPARC, secreted protein acidic and rich in cysteine; HRCECs, human retinal capillary endothelial cells; JAM, junctional adhesion molecule; ZO-1, zonula occludens.

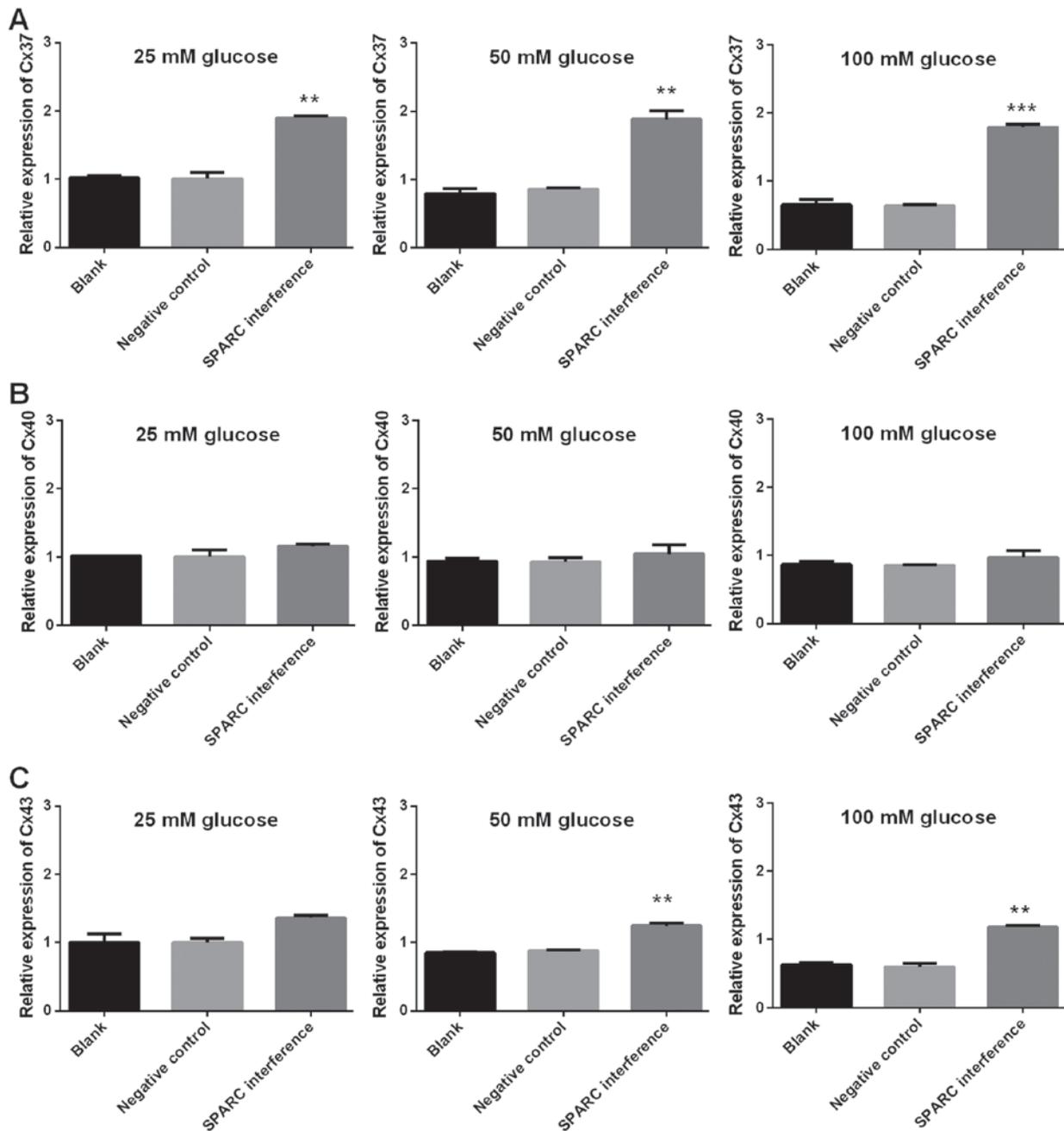


Figure 8. SPARC interference increases the expression of gap junction-associated genes associated in HRCECs treated with glucose. (A) Expression of Cx37, (B) Cx40 and (C) Cx43. HRCECs were incubated with glucose for 24 h. Gene expression was examined using reverse transcription-quantitative polymerase chain reaction. Data were presented as mean \pm standard deviation (n=3). **P<0.01 and ***P<0.001 vs. negative control group. SPARC, secreted protein acidic and rich in cysteine; HRCECs, human retinal capillary endothelial cells; Cx, connexin.

fibrosis, the present analysis on the role of SPARC expression may provide insights on future therapeutic approaches for diabetic retinopathy.

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

XX designed and performed the experiments. YF and MT interpreted the data, drafted the manuscript and provided experimental support. XQX and KL collected and analyzed the data. All authors read and approved the final manuscript.

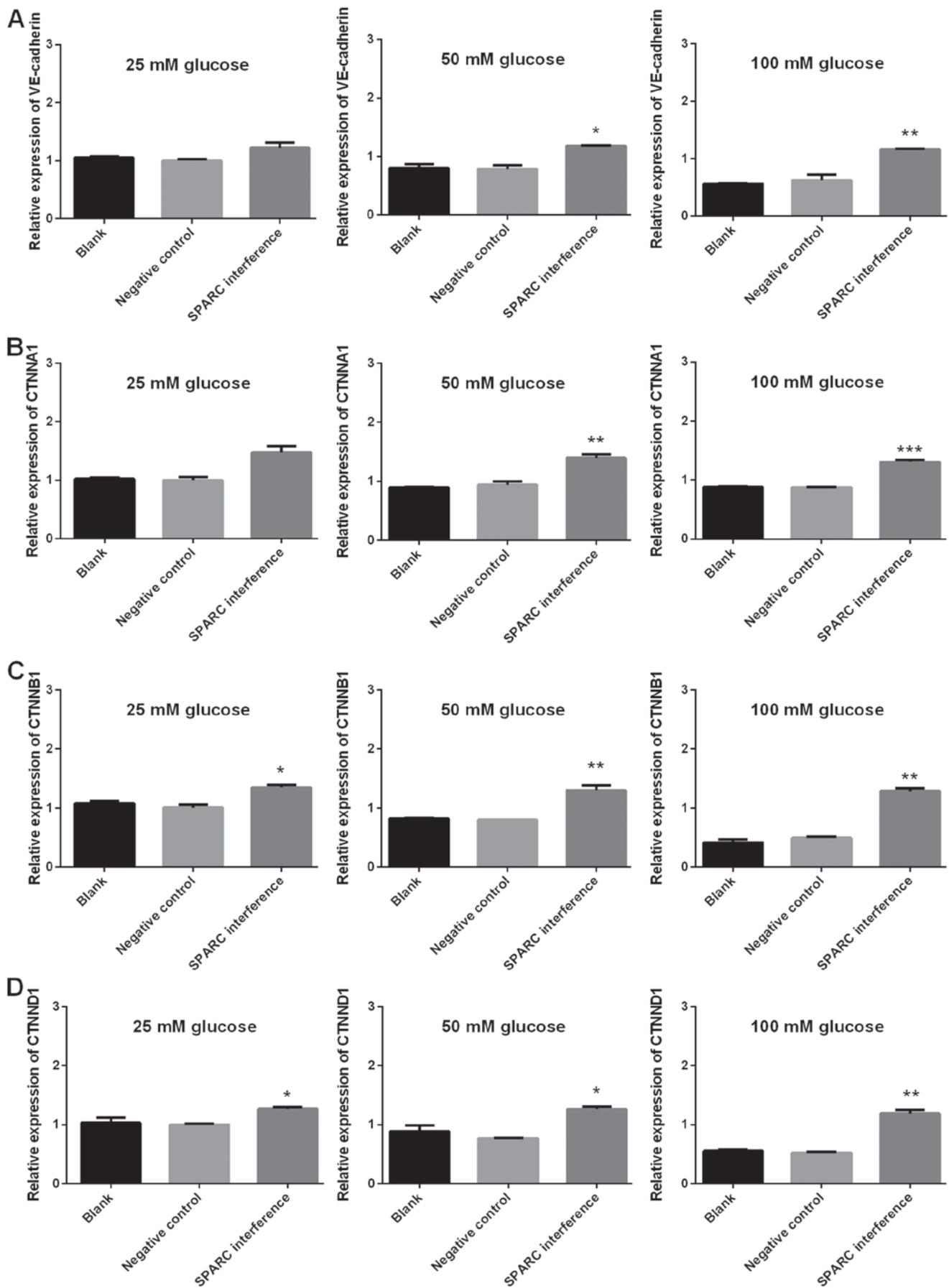


Figure 9. SPARC interference increases the expression of genes associated with adherens junctions in HRCECs treated with glucose. (A) Expression of VE-cadherin, (B) CTNNA1, (C) CTNNB1 and (D) CTNND1. HRCECs were incubated with glucose for 24 h. Data were presented as mean \pm standard deviation ($n=3$). * $P<0.05$, ** $P<0.01$ and *** $P<0.001$ vs. negative control group. SPARC, secreted protein acidic and rich in cysteine; HRCECs, human retinal capillary endothelial cells; VE, vascular endothelial; CTNNA1, catenin alpha 1; CTNNB1, catenin beta 1 CTNND1, catenin delta 1.

Ethics approval and consent to participate

The present study was approved by the Ethics Committee of Shanghai General Hospital, (Shanghai, China; Registration no. 2015 K061).

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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