Abstract. Phosphodiesterase (PDE)3-5 are expressed in cardiac tissue and play critical roles in the pathogenesis of heart failure and atrial fibrillation. PDE inhibitors are widely used in the clinic, but their effects on the electrical activity of the heart are not well understood. The aim of the present study was to examine the effects of various PDE inhibitors on spontaneous cardiac activity and compare those effects between sinoatrial nodes (SANs) and pulmonary veins (PVs). Conventional microelectrodes were used to record action potentials in isolated rabbit SAN and PV tissue preparations, before and after administration of different concentrations (0.1, 1 and 10 µM) of milrinone (PDE3 inhibitor), rolipram (PDE4 inhibitor) and sildenafil (PDE5 inhibitor), with or without the application of isoproterenol (cAMP and PKA activator), KT5823 (PKG inhibitor) or H89 (PKA inhibitor). Milrinone (1 and 10 µM) increased the spontaneous activity in PVs by 10.6±4.9 and 16.7±4.6%, respectively. In addition, milrinone (1 and 10 µM) induced the occurrence of triggered activity (0/8 vs. 5/8; P<0.005) in PVs. Rolipram increased PV spontaneous activity by 7.5±1.3‑9.5±4.0%, although this was not significant, and did not alter SAN spontaneous activity. Sildenafil reduced spontaneous activity in PVs to a greater extent than that seen in SANs. Both KT5823 and H89 suppressed milrinone‑increased PV spontaneous activity. In the presence of isoproterenol, milrinone did not alter isoproterenol‑induced PV arrhythmogenesis, suggesting that the effects of PDE3 are mediated by the protein kinase G and protein kinase A signaling pathways. In conclusion, inhibitors of different PDE subtypes exert diverse electrophysiological effects on PV and SAN activities.

Introduction

Atrial fibrillation (AF) is the most common sustained cardiac arrhythmia that causes cardiac dysfunction and strokes (I). Heart failure (HF) is characterized by chamber dilatation, which may promote the occurrence of AF through mechanoelectrical feedback and electrical or structural remodeling (2-6). Different 3',5'-cyclic nucleotide phosphodiesterase (PDE) isoforms exert distinctive biological functions, therefore, pharmacological inhibition of these PDEs might offer novel therapeutic strategies through their abilities to modulate cardiovascular diseases, including HF (7,8). In particular, PDE3-5 were reported to be expressed in cardiomyocytes and might play a role in both HF and AF (7,8). The PDE3 inhibitor, milrinone, exerts positive inotropic, vasodilating and minimal chronotropic effects, which are expected to improve HF (9). However, milrinone has potentially fatal adverse effects, including ventricular arrhythmias, which limit its use (10-14). PDE4 is expressed by human atrial myocytes, and its D isofrom (PDE4D) has been linked to stroke risk in a number of genome-wide association

Various subtypes of phosphodiesterase inhibitors differentially regulate pulmonary vein and sinoatrial node electrical activities

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differentially regulate pulmonary vein and sinoatrial node electrical activities

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Abstract. Phosphodiesterase (PDE)3-5 are expressed in cardiac tissue and play critical roles in the pathogenesis of heart failure and atrial fibrillation. PDE inhibitors are widely used in the clinic, but their effects on the electrical activity of the heart are not well understood. The aim of the present study was to examine the effects of various PDE inhibitors on spontaneous cardiac activity and compare those effects between sinoatrial nodes (SANs) and pulmonary veins (PVs). Conventional microelectrodes were used to record action potentials in isolated rabbit SAN and PV tissue preparations, before and after administration of different concentrations (0.1, 1 and 10 µM) of milrinone (PDE3 inhibitor), rolipram (PDE4 inhibitor) and sildenafil (PDE5 inhibitor), with or without the application of isoproterenol (cAMP and PKA activator), KT5823 (PKG inhibitor) or H89 (PKA inhibitor). Milrinone (1 and 10 µM) increased the spontaneous activity in PVs by 10.6±4.9 and 16.7±5.3% and in SANs by 9.3±4.3 and 20.7±4.6%, respectively. In addition, milrinone (1 and 10 µM) induced the occurrence of triggered activity (0/8 vs. 5/8; P<0.005) in PVs. Rolipram increased PV spontaneous activity by 7.5±1.3‑9.5±4.0%, although this was not significant, and did not alter SAN spontaneous activity. Sildenafil reduced spontaneous activity in PVs to a greater extent than that seen in SANs. Both KT5823 and H89 suppressed milrinone‑increased PV spontaneous activity. In the presence of isoproterenol, milrinone did not alter isoproterenol‑induced PV arrhythmogenesis, suggesting that the effects of PDE3 are mediated by the protein kinase G and protein kinase A signaling pathways. In conclusion, inhibitors of different PDE subtypes exert diverse electrophysiological effects on PV and SAN activities.

Introduction

Atrial fibrillation (AF) is the most common sustained cardiac arrhythmia that causes cardiac dysfunction and strokes (I). Heart failure (HF) is characterized by chamber dilatation, which may promote the occurrence of AF through mechanoelectrical feedback and electrical or structural remodeling (2-6). Different 3',5'-cyclic nucleotide phosphodiesterase (PDE) isoforms exert distinctive biological functions, therefore, pharmacological inhibition of these PDEs might offer novel therapeutic strategies through their abilities to modulate cardiovascular diseases, including HF (7,8). In particular, PDE3-5 were reported to be expressed in cardiomyocytes and might play a role in both HF and AF (7,8). The PDE3 inhibitor, milrinone, exerts positive inotropic, vasodilating and minimal chronotropic effects, which are expected to improve HF (9). However, milrinone has potentially fatal adverse effects, including ventricular arrhythmias, which limit its use (10-14). PDE4 is expressed by human atrial myocytes, and its D isofrom (PDE4D) has been linked to stroke risk in a number of genome-wide association
Rabbit PV and atrial tissue preparations. The present study was approved of the Institutional Animal Care and Use Committee (approval no. IACUC-19-124) of the National Defense Medical Center, Taipei, Taiwan and conformed to the institutional Guide for the Care and Use of Laboratory Animals and the ‘Guide for the Care and Use of Laboratory Animals’ published by the United States National Institutes of Health (8 ed. Washington DC, 2011). Male New Zealand white rabbits (n=47; weight, 2.0-3.0 kg; age, 6-8 months) were used in the present study. All of the rabbits had access to food and water ad libitum, and were maintained in a temperature and humidity-controlled environment (20-22°C; 50-70% humidity) with a 12 h light/dark cycle, and were raised in stainless steel cages. After rabbits were euthanized by avertin (5% in NaOH) from a precision vaporizer as an overdose of Zoletil 50 (10 mg/kg) and xylazine (5 mg/kg), hearts were rapidly removed using intramuscular injections of a mixture of Zoletil 50-70% humidity) with a 12 h light/dark cycle, and were saturated with a gas mixture of 97% O₂/3% CO₂. The temperature was maintained at 37°C and the preparations were allowed to equilibrate for 1 h before the electrophysiological assessment.

Electrophysiological and pharmacological studies. Transmembrane action potentials (APs) of the PVs were recorded by machine-pulled glass capillary microelectrodes filled with 3 mol/l KCl, which were connected to a Duo 773 electrometer (World Precision Instruments, Ltd.) under a tension of 1.47 mN (150 mg). Electrical and mechanical events (contractile force and diastolic tension) were simultaneously displayed on a 4072 oscilloscope (Gould) and a TA11 recorder (Gould). Using a data acquisition system, signals were recorded with direct coupling and a filter with a 10-kHz low-pass cut-off frequency. Signals were recorded digitally with a 16-bit accuracy, at a rate of 125 kHz. Electrical stimulation was provided using a Grass S88 stimulator through a SI1USB stimulus isolation unit (Grass Instruments Co.). Different concentrations of a PDE3 inhibitor (milrinone; 0.1, 1 and 10 µM; Sigma-Aldrich; Merck KGaA), PDE4 inhibitor (rolipram; 0.1, 1 and 10 µM; Sigma-Aldrich; Merck KGaA) or PDE5 inhibitor (sildenafil; 0.1, 1 and 10 µM; Sigma-Aldrich; Merck KGaA) were sequentially superfused to test for pharmacological responses. For each concentration, PV and SAN preparations were treated for at least 30 min. The electrical activity in isolated rabbit PVs was recorded before and after the application of 10 µM milrinone with or without isoproterenol (1 µM; Sigma-Aldrich; Merck KGaA), KT5823 (1 µM; Tocris Bioscience) [a potent selective inhibitor of cyclic guanosine monophosphate (cGMP)-dependent protein kinase G (PKG)] or H89 (a protein kinase A [PKA] inhibitor; 10 µM; Sigma-Aldrich; Merck KGaA) (29).

Spontaneous activity was defined as the constant occurrence of spontaneous APs in the absence of any electrical stimuli. Early afterdepolarizations (EADs) were defined as the interruption of the smooth contour of phase 2 or 3 of the APs. Delayed afterdepolarizations (DADs) were defined as the presence of a spontaneous hump-shaped depolarization of the impulse after full repolarization had occurred. Burst firing was defined as the occurrence of an accelerated spontaneous potential (faster than the basal rate) with sudden onset and termination.

Statistical analysis. All continuous variables are expressed as the mean ± SEM. Repeated-measures ANOVA followed by Duncan’s post hoc test was used to compare the difference before and after drug administration. Electrophysiological and mechanical characteristics were compared between different groups by a Wilcoxon rank-sum test or an unpaired t-test, depending on the outcome of the normality test. Nominal
Figure 1. Effects of the PDE3 inhibitor (PDE3-I), milrinone, on the electrical activity of isolated PVs. (A) Representative data of the beating rate, diastolic tension and contractility of PVs after the application of different concentrations (0.1, 1 and 10 µM) of the PDE3-I (n=8). (B) Representative data of the triggered activity of delayed afterdepolarizations (arrow), early afterdepolarizations (arrowheads) and burst firings (asterisk), induced by PDE3-I. PDE3, phosphodiesterase 3; PVs, pulmonary veins.
Variables were compared by a χ² analysis with Fisher's exact test. P<0.05 was considered to indicate a statistically significant difference. Statistical analysis was performed using SigmaPlot software (version 12.0; Systat Software, Inc.).

Results

Effects of the PDE3 inhibitor on the electrical activities of isolated PVs and SANs. The PDE3 inhibitor (milrinone; 1 and 10 µM) significantly increased PV spontaneous activity by 10.6±4.9% (P=0.018) and 16.7±5.3% (P<0.001) and contractility by 31.7±9.8% (P=0.019) and 58.8±19% (P<0.001) compared with the baseline, respectively, but had no significant effect on the diastolic tension of PVs (Fig. 1A). Milrinone also significantly increased PV diastolic tension by 8.9±3.7% (P=0.002) at 10 µM. Moreover, milrinone (≥1 µM) induced triggered activity including EAD, or DAD or burst firing in PVs (0/8 vs. 5/8; P<0.05; Fig. 1B). Similarly, the PDE3 inhibitor increased SAN activity by 9.3±4.3% and 20.7±4.6% at 1 and 10 µM compared with the baseline, respectively (Fig. 2). However, milrinone did not induce burst firing in SANs.

Application of KT5823 (1 µM), a selective PKG inhibitor, abolished milrinone-accelerated PV electrical activity (Fig. 3A). In addition, H89 (10 µM), a PKA inhibitor, also suppressed milrinone-accelerated PV electrical activity (Fig. 3B). Isoproterenol increased PV spontaneous activity, diastolic tension (Fig. 3B) and the occurrence of burst firing compared with the baseline (4/5 vs. 0/5; P<0.05; Fig. 3C). However, in the presence of isoproterenol, milrinone (10 µM) did not significantly alter the PV spontaneous activity or diastolic tension (P>0.05) compared with the baseline. Taken together, PDE3 inhibition may regulate PV electrical activity through adrenergic activity and activation of the PKG and PKA signaling pathways.

Discussion

HF is a common risk factor for AF (2–4). Milrinone potentiates the effect of cyclic adenosine monophosphate (cAMP) and enhances the relaxation of the left ventricle by increasing Ca²⁺-ATPase activity in the cardiac sarcoplasmic reticulum, by increasing calcium ion uptake (7). Although the PDE3 inhibitor, milrinone, improves HF through positive inotropic effects, milrinone also increases the risk of AF in patients with HF (13). In the present study, milrinone increased PV and SAN spontaneous activities to a similar extent. The accelerating effect of milrinone on PVs was abolished by the PKG inhibitor (KT5823; 1 µM) and the PKA inhibitor (H89; 10 µM), suggesting that the electrophysiological effects of
milrinone could involve the PKG and PKA signaling pathways. Additionally, the effect of milrinone on PV arrhythmogenesis in the presence of isoproterenol indicated that milrinone and isoproterenol might regulate PV electrical activity in a similar manner, via the activation of cAMP.

Previous studies reported that milrinone might relax the pulmonary arterial and venous vascular beds in guinea pig and human lung tissues by activating the ATP-sensitive potassium channel (30,31). However, the present study suggested that milrinone increased diastolic tension in PVs. The discrepancy may have been caused by the differences between distal, with simple venous structures, and proximal PVs, since the atrial-PV junction used in the present study contained the myocardial sleeve surrounding the PV vascular components. Milrinone may increase PV diastolic tension via positive inotropic effects on cardiomyocytes, as shown by increased PV contractility.

Figure 3. Effects of the protein kinase G inhibitor, KT5823, and the protein kinase A inhibitor, H89, on PDE3 inhibitor (PDE3-I)-modulated electrical activity of isolated PVs. (A) Representative data of the beating rate and diastolic tension of PVs after the application of 10 µM PDE3-I, milrinone, and 1 µM KT5823. (B) Representative data of the beating rate and diastolic tension of PVs after the application of 10 µM PDE3-I and 10 µM H89. (C) Examples of burst firing (asterisks) in PVs after the administration of 1 µM isoproterenol and 10 µM PDE3-I. PDE3, phosphodiesterase 3; PV, pulmonary vein; Iso, isoproterenol.
Figure 4. Effects of the PDE4 inhibitor (PDE4-I), rolipram, on the electrical activities of isolated PVs and SANs. (A) Representative data of the beating rate, diastolic tension and contractility of PVs after the application of different concentrations (0.1, 1 and 10 µM) of the PDE4-I (n=6). (B) Representative data of the beating rate of SANs after the application of different concentrations (0.1, 1 and 10 µM) of the PDE4-I (n=6). PDE4, phosphodiesterase 4; PV, pulmonary vein; SAN, sinoatrial node.
Figure 5. Effects of the PDE5 inhibitor (PDE5-I), sildenafil, on the electrical activity of isolated PVs. (A) Representative data of the beating rate, diastolic tension and contractility of PVs after the application of different concentrations (0.1, 1 and 10 µM) of the PDE5-I (n=7). (B) Examples of burst firing (asterisks) in PVs after the application of 1 µM isoproterenol and 10 µM PDE5-I (1 µM; n=9). PDE5, phosphodiesterase 5; PV, pulmonary vein; Iso, isoproterenol.
PDE4 inhibition increased intracellular cAMP levels and the I_{Ca,L} in atrial myocytes, as well as increasing the frequency of spontaneous Ca^{2+} release (18), which may predispose individuals to AF. However, in the present study, the PDE4 inhibitor, rolipram, had no significant effect on PV and SAN spontaneous activities. The results from the present study were consistent with that of a previous study, which suggested that PDE4 inhibition did not significantly alter SAN electrical activity (32). Therefore, clinical observations of an increased risk of AF in patients receiving PDE4 inhibition therapy might be explained by AF substrate modifications, rather than the enhancement of the arrhythmogenesis that AF triggers (33).

PDE5 expression is strongly increased in HF (34-36). Previous studies reported that inhibition of PDE5 ameliorates cardiac dysfunction and sildenafil, as a cGMP-specific PDE5 blocker (7), abolished isoproterenol-induced increases in PV activity. This result was in line with a previous report, which indicated that sildenafil can attenuate β-receptor agonist-induced cAMP generation and accelerate the beating rate of PVs (38). Similar to the present study, PDE5 inhibition was previously reported to have a negative chronotropic effect on mice SANs (26). Accordingly, sildenafil may potentially protect individuals from AF by reducing PV arrhythmogenesis.

Acute infusion of the kinase inhibitors, including KT5823 and H89, was performed in the present study to investigate the effect of milrinone on PVs. The suppression of the PDE3 accelerating effect on PVs following acute infusion of the kinase inhibitors suggested a role for the cGMP and PKA signaling pathways in PDE3 inhibitor mediated PV arrhythmogenesis. However, it is not clear whether the PDE inhibitors may have further unknown effects or whether the kinase inhibitors exert their effects at the cellular level. Since only the acute effects of the kinase inhibitors were evaluated in the present study, the protein expression of components of the cGMP and PKA signaling pathways in PVs at such a short exposure time is highly unlikely to be affected or to be shown by western blot analysis. Therefore, this requires further investigation.

In conclusion, different subtypes of PDE inhibitors regulate PV and SAN electrical activities in distinct manners and may contribute to susceptibility to atrial arrhythmogenesis.

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Availability of data and materials
The datasets used and/or analyzed during the present study are available from the corresponding author on reasonable request.
Authors' contributions

YKL and CCC contributed to the experimental design, performed the in vitro experiments, analyzed the experimental results and wrote the paper. YKL and JHII gave various contributions in the statistical analysis, and interpretation of the results and discussion. YAC contributed to the in vitro experiments and provided technical assistance in the study. SAC and YJC contributed to the experimental design, analysis of the results and final revision of the paper for publication. YYL and YCC conceived and designed the study, and reviewed the paper prior to submission.

Ethics approval and consent to participate

The present study was approved of the Institutional Animal Care and Use Committee by the local review board (approval no. IACUC-19-124) of the National Defense Medical Center, Taipei, Taiwan and conformed to the institutional Guide for the Care and Use of Laboratory Animals published by the United States National Institutes of Health (8 ed. Washington DC, 2011).

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

References


