Single-nucleotide polymorphisms and haplotypes in the interleukin-33 gene are associated with a risk of allergic rhinitis in the Chinese population

HE RAN1*, HUA XIAO2*, XING ZHOU3, LIJUN GUO2 and SHUANG LU2

1Department of Otolaryngology Head and Neck Surgery, The Second Hospital of Jingzhou, Jingzhou, Hubei 434000; 2Hubei College of Chinese Medicine, Jingzhou, Hubei 434020; 3Department of Otolaryngology Head and Neck Surgery, Shishou People's Hospital, Jingzhou, Hubei 434400, P.R. China

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Abstract. Allergic rhinitis (AR) is a common upper airway disease attributed to a variety of risk factors, such as environmental exposures and genetic susceptibility. The commonly observed comorbidity of asthma and AR in the clinic suggests the presence of shared genetic risk factors and biological mechanisms between these diseases. Interleukin (IL)-33 has been indicated to be an important factor driving asthma susceptibility and pathogenesis using both genome-wide association studies and functional studies in model animals. Although previous studies have reported the putative association of this gene with AR, evidence for the association of genetic variations of IL-33 with the disease is still missing. To examine whether variations in the IL-33 gene confer a genetic risk of AR, a total of 769 patients with AR and 769 age- and sex-matched healthy controls were recruited among Han Chinese residents in the Hubei province, and 14 single-nucleotide polymorphisms (SNPs) spanning the IL-33 gene were examined for their association with the risk of AR. The results indicated that five SNPs, which were in a moderate linkage disequilibrium and were located in the 5'-flanking region of IL-33, exhibited significant associations with the risk of AR, and these associations were additionally supported by genotypic and haplotypic analyses. Notably, three of the five IL-33 SNPs have been previously reported to exhibit genome-wide associations with asthma, and their alleles were also revealed to confer an increased risk of AR in the present study. In summary, the results of the current study suggested that certain variations in the IL-33 gene represent a potential risk for AR, and indicated a shared genetic basis between AR and asthma.

Introduction

Allergic rhinitis (AR) is a common upper airway disease, which is characterized by sneezing, rhinorrhea and obstruction of the nasal passages (1). A previous study revealed that AR was mediated by immunoglobulin (IgE)-induced upper airway inflammation, which is commonly triggered by exposure to allergens (1). AR has been indicated to coexist with other allergic and/or airway conditions, such as asthma, rhinosinusitis, nasal polyps and lower airway infection, among others (2-4). AR is highly prevalent with an incidence rate of 10-20% worldwide (1,5), which has rapidly increased along with industrialization (6). Therefore, AR is considered a major and emerging public health concern, especially in developed countries (6,7). The prevalence of AR in urban China has substantially increased in the past few years (from 2005 to 2011) (8). An epidemiological study has indicated that the overall prevalence of AR in adults in major cities across mainland China has increased from 11.1% in 2005 to 17.6% in 2011, and patients with AR have been revealed to be more susceptible to allergic complications compared with the general population (8). Therefore, subjects diagnosed with AR have been indicated to experience a reduction in their quality of life, although not always severe (9). However, AR cannot be completely treated, which poses a substantial socioeconomic burden (1).

Previous studies have revealed that environmental stimuli serve pivotal roles in the allergic airway responses that are essential for the initiation and exacerbation of AR (10,11). Nevertheless, a potent genetic component has been also associated with this disease (12). The heritability of AR has been estimated to be >65% (13,14). A number of studies have suggested that sequence polymorphisms in multiple genes and regions are associated with an increased risk of AR (15-22), although another study has reported the poor reproducibility of single-nucleotide polymorphism (SNP) associations with this disease (23). Nevertheless, certain genetic variations that have been associated with AR support the hypothesized
pathophysiology of this disease (24). Therefore, genetic and physiological investigations cross-validate the putative mechanisms of AR, providing essential insights into its pathogenesis, treatment and prevention (25).

A previous study has suggested that type 2 T helper (Th2) cell-mediated inflammatory responses against environmental stimuli, including pollutants and allergens, contributed in AR pathophysiology (26). According to the classical model for allergic airway diseases, including AR and allergens, which are presented by antigen-presenting cells, result in the activation and expansion of Th2 cells. These Th2 cells have been indicated to subsequently produce Th2 cytokines, such as interleukin (IL)-4, IL-5 and IL-13, which drive IgE antibody production and promote the proliferation and differentiation of multiple effector cells, for example mast cells, eosinophils and basophils (27,28). This results in the secretion of multiple key inflammatory mediators, such as leukotrienes and histamine (29-31). In addition to these well-characterized Th2 cytokines, other mediators that are primarily released by epithelial cells, which constitute the first line of defense against environmental stimuli, have also attracted attention owing to their determinant roles in initiating the inflammatory responses. For example, epithelium-derived IL-33 has been indicated to induce allergic airway inflammation (32,33). Previous studies have also reported that during allergen-driven inflammation, IL-33 was expressed by a variety of immune cells, including important immune cell types such as eosinophils (32,33).

IL-33 is a member of the IL-1 family of cytokines (34,35). The role of IL-33 in bronchial asthma has been extensively studied (34,35). SNPs in the genes encoding IL-33 and its receptor ST2, which is also referred to as IL1 receptor-like (RL1), have been indicated to be associated with allergic asthma and atopic dermatitis (35-37). In addition, clinical studies have demonstrated that the serum levels of IL-33 and the soluble form of ST2 (sST2) were indicators of asthma progression and exacerbation (38,39). Functional studies have additionally verified the impact of IL-33 and ST2 on allergic airway inflammation. For example, IL-33 has been indicated to induce the production of Th2 cytokines by polarized Th2 cells in vitro (40), and increase the expression of Th2 cytokines and the serum levels of IgE in vivo (40). In murine models of allergic airway inflammation, allergen exposure has been revealed to increase the endogenous levels of ST2 and Th2-mediated airway inflammation (41), while blockade of ST2 using a recombinant IgG fusion protein has been indicated to inhibit the allergic inflammation (42,43). Notably, ST2 has been indicated to be highly expressed on mast cells, except for Th2 cells, (42,44,45), which suggests a potential direct impact of IL-33 on a number of hypersensitive and allergic reactions that are mediated by these cells.

Previous studies have examined the potential association of IL-33 with AR, as IL-33 has been indicated to exhibit a well-characterized regulatory impact on the initiation and exacerbation of allergic airway responses (46-48). In patients with intermittent AR who are sensitive to trees and/or grass pollens, serum IL-33 has been revealed to be increased compared with normal controls and positively correlate with disease severity (49). The serum levels of IL-33 have also been reported to be higher in patients with Japanese cedar (JC) pollinosis compared with normal controls (50), and two IL-33 SNPs have been associated with the disease (50). Notably, IL-33 in the serum of patients with AR can be decreased following immunotherapy (51). Intranasal administration of anti-tumor necrosis factor-α IgY has been indicated to reduce both IL-33 and ovalbumin-specific IgE antibodies in the serum and nasal lavage fluids of patients (52). Although IL-33 is not always detectable in serum, increased levels of IL-33 in sinus mucosa have also been reported in patients with JC pollinosis and house dust mite (HDM)-sensitized AR (53). A previous study has demonstrated that an anti-IL-33 antibody exhibited a therapeutic potential in experimental AR (54). Moreover, IL-33 receptor ST2 has been indicated to be highly expressed in the nasal epithelium of patients with AR or chronic rhinosinusitis (55-57), and the concentration of sST2 in the nasal lavage fluids of patients with AR have been revealed to increase when patients experience pollen-induced allergic reactions (58). Taken together, these data suggest that IL-33 likely facilitates the pathogenesis of AR. Therefore, additional studies on this cytokine may provide novel insights on potential preventive strategies against AR.

As aforementioned, the prevalence of AR has increased in recent years in the Chinese population; however little is known about the susceptibility factors to AR, especially the genetic basis, in this ethnic group. Therefore, the aim of the present study was to perform genetic analyses of the IL-33 variations in the Han Chinese population to reveal whether variations in this gene confer an inheritable risk of AR.

Materials and methods

Subjects. A total of 769 patients with AR (453 men and 316 women) aged between 10 and 65 years (mean ± standard deviation, 37.74±16.52) were recruited between January 2014 and July 2019. The diagnosis of AR was performed following the criteria of Allergic Rhinitis and its Impact on Asthma (1). Subjects presenting > two common symptoms of AR (for example nasal congestion, rhinorrhea, nasal itching and sneezing) for >4 days/week for at least 3 weeks during the past 12 months were diagnosed with AR. All patients received anti-histaminic and steroids at The Second Hospital of Jingzhou and Shishou People’s Hospital (Jingzhou, China). Specific allergens as potential cause of AR were determined for each patient via skin prick tests (SPT; Allergopharma GmbH & Co. KG) in accordance with the recommendations by the Subcommittee on Allergen Standardization and Skin Tests of the European Academy of Allergy and Clinical Immunology (59). A total of 18 inhaled allergens, including HDM, grass, tree, mold, food and weed panel allergens, were examined. The presence of a wheal ≥ one half of the diameter of the histamine control and >3 mm of the negative control was defined positive in the SPT. Patients with any other systemic diseases were excluded from the study.

A total of 769 healthy volunteers (442 men and 327 women) aged between 10 and 65 years (mean ± standard deviation, 37.01±15.94) that tested negative for allergens in the skin prick tests were recruited as control subjects in the current study. The controls were also recruited at The Second Hospital of Jingzhou and Shishou People’s Hospital (Jingzhou, China) between January 2014 and July 2019. In addition, the control
subjects did not exhibit any clinical diagnosis or family history of allergy, and did not experience an upper respiratory tract infection within the last 4 weeks prior to the study. All patients and controls were Han Chinese.

**SNP selection.** SNPs were selected based on previous studies and the genotype data from the 1,000 Genomes Project (60). Based on the high comorbidity rate of AR and asthma, genome-wide association studies (GWAS) on asthma were firstly screened to select genome-wide significant variants spanning IL-33. Briefly, four SNPs (rs928413, rs9775039, rs992969 and rs2381416) in IL-33 have been indicated to exhibit genome-wide significant associations with asthma in previous GWAS on European and Asian populations (24,61–64). According to the 1,000 Genomes Project, three IL-33 SNPs (rs928413, rs992969 and rs2381416) have been indicated to be polymorphic in the Han Chinese population, and therefore were selected in the present study. The aforementioned study in subjects with JC pollinosis was also reviewed (50), and two additional SNPs that have been associated with this disease were selected (rs1929992 and rs10975519; Fig. 1). The linkage disequilibrium (LD) pattern of IL-33 in the Han Chinese population was subsequently examined based on data from the 1,000 Genomes Project, and nine representative tagging SNPs (rs1475658, rs10815374, rs1891385, rs16924144, rs10435816, rs16924159, rs16924161, rs117414011 and rs78100995) spanning the IL-33 gene were selected (Fig. 1). In summary, a total of 14 SNPs (Table I) were selected for subsequent investigation in the current study.

**SNP genotyping.** Saliva was collected from the participants, and genomic DNA was extracted according to the manufacturer's protocol (MagMAX gDNA Saliva Isolation Kit; Thermo Fisher Scientific, Inc.). The extracted DNA was dissolved in 50 ml Tris-EDTA buffer (10 mM Tris, pH 7.8; 1 mM EDTA). The concentration of the DNA samples was quantified via measuring the optical density at 260 nm using NanoDrop.

A multiplex PCR was performed to simultaneously amplify multiple SNPs in one reaction in 96-well plates, and each well contained 20 ng genomic DNA, 10 mmol/l primers (the primer sequences are presented in Table SI), 2 µl dNTP, 2 µl 10X PCR buffer, 1 µl 25 mmol/l MgCl2, and 0.4 units Taq polymerase (Takara Biotechnology Co., Ltd.). The multiplex PCR reaction was conducted on an ABI 9700 cycler (Applied Biosystems; Thermo Fisher Scientific, Inc.) using the following thermocycling conditions: Denaturation for 5 min at 95˚C; 40 cycles of 30 sec at 95˚C, 30 sec at 60˚C and 30 sec at 72˚C; and extension for 10 min at 72˚C. The PCR products were subsequently used to genotype candidate SNPs with the SNaPshot method. In brief, the PCR products were purified via shrimp alkaline phosphatase (cat. no. M0371L; New England Biolabs) and exonuclease I (cat. no. M0293L; BioLabs) treatment at 37˚C for 1 h. Specifically designed SNaPshot primers (the primer sequences are presented in Table SII) were used to amplify the SNP target sites for one base extension using SNaPshot Multiplex Ready Reaction Mix (cat. no. 4323166; Applied Biosystems; Thermo Fisher Scientific, Inc.), following which the reaction was terminated and the products were loaded on an ABI 3730 automated sequencer (Applied Biosystems; Thermo Fisher Scientific, Inc.). The SNP genotype calling results were automatically retrieved using GeneMarker version 2.2.0 software (SoftGenetics) and were manually verified. All genotypes were called blind to sample identity and affection status and were independently reviewed and confirmed by two authors. A total of 5% of the samples were randomly selected for re-analysis to ensure 100% concordance.

**Statistical analyses.** Haploview software (version 4.1; Broad Institute of Massachusetts Institute of Technology and Harvard) (65) and the online program SHEsis (http://analysis.bio-x.cn/myAnalysis.php) (66,67) were used to calculate the Hardy-Weinberg equilibrium (HWE) of each SNP in cases and controls, and the LD between paired SNPs was defined using the r2 algorithm. SHEsis was also used to compare the allelic and genotypic distributions of each SNP between cases and controls, and Pearson’s χ2 test was applied to calculate the P-values, odds ratios (ORs) and 95% confidence intervals (66,67). Haplotype frequencies and association signals were estimated with the online program SHEsis using the χ2 test (66,67). The haplotypes with a frequency <0.005 in both cases and controls were discarded, and nine haplotypes remained for subsequent analyses.

**Results**

**Demographics of the participants.** No significant differences were observed between cases and controls in terms of their
According to the SPT results of 769 patients with AR, 352 (45.8%) were sensitized to HDM, 147 (19.1%) were sensitized to tree pollen and 270 (35.1%) were sensitized to multiple allergens (data not shown).

LD of the IL-33 gene. A total of 14 SNPs in IL-33, with a frequency >0.01, were selected to examine their associations with AR. Their locations within or close to the IL-33 gene and the LD map in the Han Chinese population are presented in Table I.

<table>
<thead>
<tr>
<th>SNP</th>
<th>Genotype, n (frequency)</th>
<th>HWE</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>rs2381416</td>
<td>A/A 674 (0.880) A/C 87 (0.114) C/C 5 (0.007)</td>
<td>0.24</td>
<td>0.009</td>
</tr>
<tr>
<td>Controls</td>
<td>A/A 710 (0.926) A/T 55 (0.072) T/T 2 (0.003)</td>
<td>0.40</td>
<td></td>
</tr>
<tr>
<td>rs1475658</td>
<td>A/A 627 (0.826) A/T 125 (0.165) T/T 7 (0.009)</td>
<td>0.78</td>
<td>0.013</td>
</tr>
<tr>
<td>Controls</td>
<td>A/A 665 (0.880) A/T 87 (0.115) T/T 4 (0.005)</td>
<td>0.53</td>
<td></td>
</tr>
<tr>
<td>rs992969</td>
<td>A/A 3 (0.004) A/G 77 (0.100) G/G 689 (0.896)</td>
<td>0.59</td>
<td>0.0023</td>
</tr>
<tr>
<td>Controls</td>
<td>A/A 2 (0.003) A/G 41 (0.053) G/G 726 (0.944)</td>
<td>0.09</td>
<td></td>
</tr>
<tr>
<td>rs928413</td>
<td>A/A 651 (0.849) A/G 114 (0.149) G/G 2 (0.003)</td>
<td>0.20</td>
<td>0.0012</td>
</tr>
<tr>
<td>Controls</td>
<td>A/A 690 (0.903) A/G 69 (0.090) G/G 5 (0.007)</td>
<td>0.03</td>
<td></td>
</tr>
<tr>
<td>rs10815374</td>
<td>A/A 5 (0.007) A/G 102 (0.133) G/G 662 (0.861)</td>
<td>0.62</td>
<td>0.02</td>
</tr>
<tr>
<td>Controls</td>
<td>A/A 4 (0.005) A/G 68 (0.088) G/G 697 (0.906)</td>
<td>0.10</td>
<td></td>
</tr>
<tr>
<td>rs1891385</td>
<td>A/A 401 (0.533) A/C 306 (0.407) C/C 45 (0.060)</td>
<td>0.18</td>
<td>0.74</td>
</tr>
<tr>
<td>Controls</td>
<td>A/A 383 (0.515) A/C 312 (0.419) C/C 49 (0.066)</td>
<td>0.17</td>
<td></td>
</tr>
<tr>
<td>rs16924144</td>
<td>C/C 91 (0.118) C/T 346 (0.450) T/T 332 (0.432)</td>
<td>0.95</td>
<td>0.2</td>
</tr>
<tr>
<td>Controls</td>
<td>C/C 71 (0.092) C/T 369 (0.480) T/T 329 (0.428)</td>
<td>0.02</td>
<td></td>
</tr>
<tr>
<td>rs10435816</td>
<td>A/A 273 (0.355) A/G 377 (0.490) G/G 119 (0.155)</td>
<td>0.55</td>
<td>0.81</td>
</tr>
<tr>
<td>Controls</td>
<td>A/A 282 (0.367) A/G 376 (0.489) G/G 111 (0.144)</td>
<td>0.43</td>
<td></td>
</tr>
<tr>
<td>rs16924159</td>
<td>A/A 70 (0.091) A/G 333 (0.433) G/G 366 (0.476)</td>
<td>0.64</td>
<td>0.98</td>
</tr>
<tr>
<td>Controls</td>
<td>A/A 68 (0.088) A/G 333 (0.433) G/G 368 (0.479)</td>
<td>0.55</td>
<td></td>
</tr>
<tr>
<td>rs16924161</td>
<td>C/C 98 (0.127) C/T 351 (0.456) T/T 320 (0.416)</td>
<td>0.91</td>
<td>0.97</td>
</tr>
<tr>
<td>Controls</td>
<td>C/C 101 (0.131) C/T 352 (0.458) T/T 316 (0.411)</td>
<td>0.85</td>
<td></td>
</tr>
<tr>
<td>rs117414011</td>
<td>C/C 3 (0.004) C/T 67 (0.087) T/T 699 (0.909)</td>
<td>0.31</td>
<td>0.60</td>
</tr>
<tr>
<td>Controls</td>
<td>C/C 1 (0.001) C/T 68 (0.088) T/T 700 (0.910)</td>
<td>0.62</td>
<td></td>
</tr>
<tr>
<td>rs78100995</td>
<td>C/C 412 (0.537) C/G 293 (0.382) G/G 62 (0.081)</td>
<td>0.33</td>
<td>0.54</td>
</tr>
<tr>
<td>Controls</td>
<td>C/C 428 (0.557) C/G 288 (0.375) G/G 52 (0.068)</td>
<td>0.71</td>
<td></td>
</tr>
<tr>
<td>rs1929992</td>
<td>C/C 233 (0.304) C/T 385 (0.502) T/T 149 (0.194)</td>
<td>0.66</td>
<td>0.90</td>
</tr>
<tr>
<td>Controls</td>
<td>C/C 240 (0.314) C/T 376 (0.492) T/T 148 (0.194)</td>
<td>0.97</td>
<td></td>
</tr>
<tr>
<td>rs10975519</td>
<td>C/C 168 (0.218) C/T 388 (0.505) T/T 213 (0.277)</td>
<td>0.73</td>
<td>0.97</td>
</tr>
<tr>
<td>Controls</td>
<td>C/C 165 (0.215) C/T 387 (0.503) T/T 217 (0.282)</td>
<td>0.76</td>
<td></td>
</tr>
</tbody>
</table>

SNP, single-nucleotide polymorphism; HWE, Hardy-Weinberg equilibrium.
Association of polymorphisms in the IL-33 gene with the risk of allergic rhinitis.

Allergic rhinitis (AR) is a common allergic disease exhibiting important comorbidity with a number of atopic diseases, for example, asthma. In the present study, we examined the associations between the 5'-flanking region of the IL-33 gene and allergic rhinitis. In summary, we conducted the allelic analyses (Table I). The LD maps of the 14 IL-33 SNPs in the Han Chinese case and control samples were constructed, and it was indicated that the LD patterns of cases and controls were similar (Fig. 2). The LD map that was constructed using genotype data from the 1,000 Genomes Project was also indicated to be similar to the LD maps of the cases and controls of the current study (Fig. 1).

Association of haplotypes in the IL-33 gene with the risk of AR. As all five SNPs in the 5'-flanking region of IL-33 exhibited a significant association with the risk of AR, the association of their haplotypes with the risk of AR was subsequently examined. A protective haplotype that exhibited a significant association with AR was identified (A-A-G-A-G; P=0.0001; Table III). This haplotype contained the protective alleles of all five SNPs, and its frequency was significantly lower in AR cases compared with controls (frequency=0.884 in cases vs. 0.925 in controls; P=0.0001; OR=0.61). The minor allele frequencies of these five SNPs in the Han Chinese population were relatively low (MAF <0.1; Table II), and the frequency of their haplotypes was even lower compared with MAFs (Table III). Nevertheless, two minor haplotypes exhibited a significant association with increased risk of AR in the present study (C-T-A-G-A frequency =0.041 in cases vs. 0.026 in controls, P=0.0023; OR=1.60; C-T-A-G-A frequency =0.044 in cases vs. 0.028 in controls, P=0.0013; OR=1.60). Global analysis also indicated a significant association of these haplotypes with the risk of AR (global P=0.00064). Collectively, the haplotype analysis additionally demonstrated that IL-33 may harbor risk variants for AR.

Discussion

Allergic rhinitis (AR) is a common allergic disease exhibiting important comorbidity with a number of atopic diseases, for example, asthma. In the present study, we examined the associations between the 5'-flanking region of the IL-33 gene and allergic rhinitis. In summary, we conducted the allelic analyses (Table I). The LD maps of the 14 IL-33 SNPs in the Han Chinese case and control samples were constructed, and it was indicated that the LD patterns of cases and controls were similar (Fig. 2). The LD map that was constructed using genotype data from the 1,000 Genomes Project was also indicated to be similar to the LD maps of the cases and controls of the current study (Fig. 1).

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### Table II. Allelic association analysis of 14 interleukin-33 SNPs with the risk of allergic rhinitis.

<table>
<thead>
<tr>
<th>SNP</th>
<th>Position</th>
<th>A1/A2</th>
<th>Frequency of A1</th>
</tr>
</thead>
<tbody>
<tr>
<td>rs2381416</td>
<td>9:6193455</td>
<td>C/A</td>
<td>0.0633, 0.0385</td>
</tr>
<tr>
<td>rs1475658</td>
<td>9:6201574</td>
<td>T/A</td>
<td>0.0916, 0.0628</td>
</tr>
<tr>
<td>rs992969</td>
<td>9:6209697</td>
<td>A/G</td>
<td>0.0540, 0.0293</td>
</tr>
<tr>
<td>rs928413</td>
<td>9:6213387</td>
<td>G/A</td>
<td>0.0769, 0.0517</td>
</tr>
<tr>
<td>rs10815374</td>
<td>9:6213705</td>
<td>A/G</td>
<td>0.0728, 0.0494</td>
</tr>
<tr>
<td>rs1891385</td>
<td>9:6219845</td>
<td>C/A</td>
<td>0.263, 0.276</td>
</tr>
<tr>
<td>rs16924144</td>
<td>9:6221246</td>
<td>C/T</td>
<td>0.343, 0.332</td>
</tr>
<tr>
<td>rs10435816</td>
<td>9:6225535</td>
<td>G/A</td>
<td>0.400, 0.389</td>
</tr>
<tr>
<td>rs16924159</td>
<td>9:6229417</td>
<td>A/G</td>
<td>0.308, 0.305</td>
</tr>
<tr>
<td>rs16924161</td>
<td>9:6230912</td>
<td>C/T</td>
<td>0.356, 0.360</td>
</tr>
<tr>
<td>rs117414011</td>
<td>9:6242939</td>
<td>C/T</td>
<td>0.0475, 0.0455</td>
</tr>
<tr>
<td>rs78100995</td>
<td>9:6248007</td>
<td>G/C</td>
<td>0.272, 0.255</td>
</tr>
<tr>
<td>rs1929992</td>
<td>9:6251588</td>
<td>T/C</td>
<td>0.445, 0.440</td>
</tr>
<tr>
<td>rs10975519</td>
<td>9:6253571</td>
<td>C/T</td>
<td>0.471, 0.466</td>
</tr>
</tbody>
</table>

SNP, single-nucleotide polymorphism; A1/A2, allele 1/allele 2; OR, odds ratio; CI, confidence interval.
example allergic asthma. Although Th2-mediated allergic inflammation has been indicated to facilitate AR pathogenesis, the associated genetic components remain to be fully determined (1). As >80% of patients with asthma exhibit rhinitis, and 10-40% of patients with rhinitis develop asthma, these two diseases are considered to share risk factors and mechanisms (1). Therefore, it is of interest to examine the association between the GWAS risk loci of asthma and AR. IL-33 has been highlighted as an important initiating factor for allergic asthma, and a recent study on whole blood gene expression profiles has revealed a consistent overexpression of the IL1RL1 gene, which encodes IL-33 receptor ST2, in patients diagnosed with multiple morbidities for asthma, dermatitis and rhinitis compared with normal controls (68). Therefore, the present study aimed to explore whether polymorphisms in IL-33 also affect the susceptibility to AR. Using the SNaPshot method, the current study indicated that SNPs in the IL-33 gene, which have been previously associated with asthma (61,62,64), were also associated with AR. This observation supported the shared pathological mechanism between asthma and AR, and provided insights into inflammatory responses associated with AR (69). Previous studies have also reported that asthma susceptibility loci exhibited an association with AR, and several other risk variants have also been highlighted conferring a risk of both asthma and AR (70-73).

The SNPs that were significantly associated with AR in the present study were localized in the promoter region of IL-33, and may have potential regulatory impact on this gene. For example, the A-allele of rs992969, which is associated with an increased risk of AR in the present study, was also significantly associated with higher IL-33 expression in bronchial epithelial cells (P=1.3x10^−6) in a previous study (74). A recent study has indicated that the A-allele of rs992969 was significantly associated with a higher mRNA level of IL-33 in bronchial epithelial cells (P=8.3x10^−12) (75). In addition, a functional study has revealed that the G-allele of rs928413 was associated with higher IL-33 promoter activity compared with the A-allele in lung cancer cells, and the G-allele of rs928413 included a binding site for cyclic AMP-responsive element-binding protein 1, which is likely an activator of IL-33.

### Table III. Haplotypic association analysis of five IL-33 SNPs (rs2381416, rs1475658, rs992969, rs928413 and rs10815374) with the risk of allergic rhinitis.

<table>
<thead>
<tr>
<th>Haplotype</th>
<th>Case, frequency</th>
<th>Control, frequency</th>
<th>(\chi^2) value</th>
<th>P-value</th>
<th>OR</th>
<th>95% CI</th>
</tr>
</thead>
<tbody>
<tr>
<td>C-T-A-G-A</td>
<td>0.041</td>
<td>0.026</td>
<td>5.180</td>
<td>0.0230</td>
<td>1.60</td>
<td>1.06-2.40</td>
</tr>
<tr>
<td>C-T-A-A-G</td>
<td>0.009</td>
<td>0.000</td>
<td>13.210</td>
<td>0.0003</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>C-T-A-G-G</td>
<td>0.005</td>
<td>0.003</td>
<td>0.570</td>
<td>0.4500</td>
<td>1.54</td>
<td>0.50-4.74</td>
</tr>
<tr>
<td>C-T-G-A-G</td>
<td>0.009</td>
<td>0.009</td>
<td>0.047</td>
<td>0.8300</td>
<td>0.92</td>
<td>0.43-1.96</td>
</tr>
<tr>
<td>A-T-G-A-G</td>
<td>0.012</td>
<td>0.013</td>
<td>0.007</td>
<td>0.9400</td>
<td>0.97</td>
<td>0.51-1.86</td>
</tr>
<tr>
<td>A-A-G-A-A</td>
<td>0.006</td>
<td>0.003</td>
<td>1.210</td>
<td>0.2700</td>
<td>1.82</td>
<td>0.61-5.42</td>
</tr>
<tr>
<td>A-T-G-A-A</td>
<td>0.013</td>
<td>0.007</td>
<td>2.700</td>
<td>0.1000</td>
<td>1.84</td>
<td>0.88-3.83</td>
</tr>
<tr>
<td>A-A-G-G-G</td>
<td>0.018</td>
<td>0.010</td>
<td>3.230</td>
<td>0.0700</td>
<td>1.78</td>
<td>0.94-3.35</td>
</tr>
<tr>
<td>A-A-G-A-G</td>
<td>0.884</td>
<td>0.925</td>
<td>15.000</td>
<td>0.0001</td>
<td>0.61</td>
<td>0.47-0.79</td>
</tr>
</tbody>
</table>

Global \(\chi^2=27.2\); degree of freedom=8; P=0.00064. OR, odds ratio; CI, confidence interval.

Figure 2. LD maps of 14 IL-33 SNPs examined in case and control subjects in the present study. The LD maps were generated using Haploview software. SNP, single-nucleotide polymorphism; LD, linkage disequilibrium; IL, interleukin.
transcription in the presence of this allele (76). These studies have demonstrated the potential regulatory effects of these SNPs on the transcription of IL-33.

Although a number of atopic conditions are considered to arise from altered Th2 immune responses against environmental allergens (77,78), the involvement of IL-33 in this process has attracted attention in recent years. In addition to targeting altered Th2 immune responses and their downstream effectors, it has been recognized that the successful prevention of allergic airway diseases is important to alleviate public health and economic burdens (79-81). Therefore, pathways mediating the initiation of allergic airway responses have been extensively studied, and IL-33 has been indicated to be a potential target molecule for the early control and prevention of allergic airway inflammation (82-84). In the current study, the association between IL-33 and AR was demonstrated, and novel data regarding the genetic basis of AR in the Han Chinese population were analyzed. Subsequent analyses validating these findings may reveal further genetic susceptibility to AR. Moreover, GWAS are still required to determine the shared and unique mechanisms of AR compared with other atopic diseases.

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Availability of data and materials

All data generated or analyzed during this study are included in this published article.

Authors’ contributions

HR and HX conceived the present study, conducted the primary experiments and analyses. XZ, LG and SL performed additional experiments. HR and XZ collected clinical samples. HR and HX drafted the manuscript. All authors read and approved the final manuscript.

Ethics approval and consent to participate

The present study was approved by the Ethics Committees of The Second Hospital of Jingzhou, Hubei College of Chinese Medicine and Shishou People's Hospital. All investigation procedures were performed in agreement with the Declaration of Helsinki. Oral informed consent was provided by the participants or their guardians.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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