

Puerarin protects vascular smooth muscle cells from oxidized low-density lipoprotein-induced reductions in viability via inhibition of the p38 MAPK and JNK signaling pathways

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Abstract. Puerarin belongs to the family of flavonoids extracted from *Pueraria lobata* (Wild.) Ohwi, which exhibits anti-oxidative, anti-inflammatory, anti-hyperglycemic, antitumor, anti-hypertensive and anti-atherosclerotic activities. In the present study, the effects of puerarin on oxidized low-density lipoprotein (ox-LDL)-stimulated vascular smooth muscle cells (VSMCs) were explored to understand the mechanisms underlying the anti-atherosclerotic effects of puerarin. VSMCs were treated with various concentrations of puerarin (0, 20, 40 and 80 μ M) prior to stimulation with ox-LDL (50 μ g/ml). VSMC viability was evaluated by performing MTT and Cell Counting Kit-8 assays. Moreover, superoxide dismutase (SOD) and malondialdehyde (MDA) levels were measured by performing ELISAs. The mRNA expression levels of interleukin-6 (IL-6) and tumor necrosis factor- α (TNF- α) were determined via reverse transcription-quantitative PCR. Western blotting was conducted to assess the levels of p38-MAPK and JNK phosphorylation. The results indicated that puerarin inhibited ox-LDL-induced VSMC viability. Moreover, puerarin significantly decreased the mRNA expression levels of IL-6 and TNF- α , significantly reduced the production of MDA and significantly increased SOD activity in ox-LDL-stimulated VSMCs. Puerarin also inhibited ox-LDL-induced phosphorylation of p38 and JNK in VSMCs. The results suggested that puerarin reduced ox-LDL-induced VSMC viability via inhibition of the p38 MAPK and JNK signaling pathways. The present study provided theoretical evidence that puerarin may

serve as a therapeutic agent to reduce the development of atherosclerosis.

Introduction

Atherosclerosis is an important pathological manifestation of cardiovascular diseases and chronic inflammatory disease (1), which is the result of a build-up of fatty materials, such as cholesterol and lipids (2-4). Atherosclerosis affects arterial blood vessels, resulting in thickening of the vascular wall and a narrowing of the lumen (5). Previous studies have indicated that cells of the arterial wall, including T cells, monocyte-derived macrophages, endothelial cells and vascular smooth muscle cells (VSMCs), are associated with the development of atherosclerosis (1,6). Although the mechanism is not fully understood, it is widely accepted that the abnormal proliferation of VSMCs located in the arterial intima leads to intimal thickening of the aorta, serving an important role in the pathogenesis and progression of atherosclerosis (7,8). Therefore, inhibiting VSMC proliferation may serve as a useful therapeutic approach for atherosclerosis.

Pueraria lobata (Wild.) Ohwi, is widely used in traditional Chinese medicine as a treatment for cardiovascular diseases, diabetes and liver diseases (9-11). Puerarin (4'-7'-dihydroxy-8- β -D-glucosylisoflavone, C₂₁H₂₀O₁₀; Fig. 1), one of the major isoflavonoid compounds isolated from the root of *Pueraria lobata*, is considered as one of the main pharmacologically active constituents of this treatment (12). Moreover, puerarin has been reported to display various pharmacological properties, including anti-oxidative (13), anti-inflammatory (14), antitumor (15), anti-hypercholesterolemic (9), anti-hyperglycemic (16) and anti-hypertensive activities (13). Furthermore, puerarin has been reported to have beneficial effects in the treatment of atherosclerosis (12,17,18). However, the role and mechanism underlying puerarin activity on oxidized low-density lipoprotein (ox-LDL)-stimulated VSMCs has not been previously reported. Ox-LDL-induced VSMC proliferation in the intima of the arterial wall serves a critical role in the progress of atherosclerosis (19), therefore inhibiting ox-LDL-induced proliferation may serve as a

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potential therapeutic strategy for atherosclerosis. The present study aimed to evaluate the potential protective effect of puerarin on ox-LDL-stimulated VSMCs and to further identify the underlying mechanisms of its action.

Materials and methods

Cell culture and treatments. Human VSMCs were obtained from The Cell Bank of Type Culture Collection of the Chinese Academy of Sciences. VSMCs were seeded (1×10^4 cells/well) into 96-well microplates and cultured in DMEM (Gibco; Thermo Fisher Scientific, Inc.) supplemented with 10% heat-inactivated FBS (Hangzhou Sijiqing Biological Engineering Materials Co., Ltd.) and 1% penicillin/streptomycin (Sigma-Aldrich; Merck KGaA) at 37°C with 5% CO₂. Following pretreatment with 0, 20, 40 or 80 μM puerarin (purity, ≥98%; Sichuan Weikeyi Biological Technology Co., Ltd.) for 24 h at 37°C, cells were treated with 50 μg/ml ox-LDL (Guangzhou Yiyuan Biological Technology Co., Ltd.) for 24 h at 37°C (20).

Cell viability assay. Cell viability was evaluated by performing an MTT assay (Beijing Solarbio Science & Technology Co., Ltd.). Cells were seeded (1×10^4 cells/well) into 96-well microplates and cultured in DMEM overnight at 37°C. Subsequently, cells were incubated with DMEM containing 0.01% DMSO or puerarin (20, 40 or 80 μM) at 37°C for 24 h. Cells were then stimulated with ox-LDL (50 μg/ml) for 24 h at 37°C. Subsequently, 5 mg/ml MTT solution was added to each well for 4 h at 37°C. DMSO (150 μl) was added to each well for 15 min to dissolve the purple formazan. Absorbance was measured at a wavelength of 490 nm using a microplate reader (Thermo Fisher Scientific, Inc.).

The Cell Counting Kit-8 (CCK-8) assay (Sigma-Aldrich; Merck KGaA) was conducted to determine cell viability according to the manufacturer's protocol. Briefly, following treatment, 10 μl CCK-8 reagent was added into each well for 2 h at 37°C. Absorbance was measured at a wavelength of 450 nm using a microplate reader (Thermo Fisher Scientific, Inc.). Cell viability was calculated according to the following formula: Cell viability (%) = absorbance of test sample / absorbance of control $\times 100$.

Detection of oxidative stress biomarkers, superoxide dismutase (SOD) and malondialdehyde (MDA). Cells were seeded (1×10^4 cells/well) into 6-well microplates and cultured in DMEM overnight at 37°C. Subsequently, cells were incubated with DMEM containing 0.01% DMSO or puerarin (20, 40 and 80 μM) for 24 h at 37°C. Cells were then stimulated with 50 μg/ml ox-LDL for 24 h at 37°C. Cells were collected and lysed on ice using RIPA lysis buffer (Beyotime Institute of Biotechnology). Following centrifugation at 13,000 \times g for 5 min at 4°C, the supernatant was collected to determine SOD (cat. no. 20190602) activity and MDA (cat. no. 20190508) content using commercial kits (Nanjing Jiancheng Institute of Biological Engineering Co., Ltd.) according to the manufacturer's protocol.

Western blotting. Cells were seeded (1×10^4 cells/well) into 6-well microplates and cultured in DMEM overnight at 37°C.

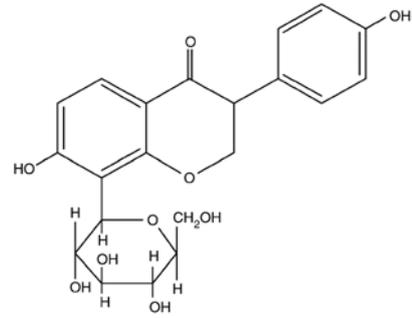


Figure 1. Chemical structure of puerarin.

Cells were incubated with DMEM containing 0.01% DMSO or puerarin (20, 40 and 80 μM) for 24 h at 37°C. Cells were then stimulated with 50 μg/ml ox-LDL for 24 h at 37°C. Cells were collected and lysed on ice with RIPA lysis buffer (Beyotime Institute of Biotechnology). Total protein was quantified using a bicinchoninic acid protein assay kit (Beyotime Institute of Biotechnology). Equal amounts of protein (50 μg) were separated via 10% SDS-PAGE (120 V for 1.5 h) and transferred onto PVDF membranes (100 V for 1 h), which were blocked with 5% skimmed milk in TBST (0.05% Tween-20) for 1 h at room temperature. After blocking, the membranes were incubated overnight at 4°C with the following primary antibodies: Anti-p38 (1:1,000; cat. no. sc-7281; Santa Cruz Biotechnology, Inc.), anti-p-p38 (1:1,000; cat. no. sc-7937; Santa Cruz Biotechnology, Inc.), JNK (1:1,000; cat. no. sc-6531; Santa Cruz Biotechnology, Inc.), p-JNK (1:1,000; cat. no. sc-3824; Santa Cruz Biotechnology, Inc.), anti-GAPDH (1:1,000; cat. no. sc-6341; Santa Cruz Biotechnology, Inc.). Following incubation, the membranes were washed three times in TBST, before the membranes were incubated with an appropriate horseradish peroxidase-conjugated secondary antibody for 1 h at room temperature (1:5,000; cat. no. sc-5203; Santa Cruz Biotechnology, Inc.). Protein bands were visualized using enhanced chemiluminescence reagent (Amersham; Cytiva). Protein expression levels were semi-quantified using ImageJ software (version 1.46; National Institutes of Health) with GAPDH as the loading control.

Reverse transcription-quantitative PCR (RT-qPCR). Cells were seeded (1×10^4 cells/well) into 6-well microplates and cultured in DMEM overnight at 37°C. Cells were incubated with DMEM containing 0.01% DMSO or puerarin (20, 40 and 80 μM) for 24 h at 37°C. Cells were then exposed to 50 μg/ml ox-LDL for 24 h at 37°C. Subsequently, total RNA was extracted from the cells using TRIzol® (Takara Biomedical Technology Co., Ltd., China) according to the manufacturer's protocol. Total RNA was reverse transcribed using the PrimeScript RT Master Mix kit (Takara Biotechnology Co., Ltd.) at 37°C for 15 min and 85°C for 5 min. Subsequently, qPCR was performed using SYBR® Premix Ex Taq (TransGen Biotech Co., Ltd.) with the following thermocycling conditions: Initial denaturation at 95°C for 5 min; followed by 40 cycles of denaturation at 95°C for 10 sec, annealing at 60°C for 15 sec and extension at 72°C for 30 sec; followed by a final extension at 72°C for 10 min. The following primers were used: IL-6 forward, 5'-CTCTCCGCAAGAGACTTCCA-3' and reverse,

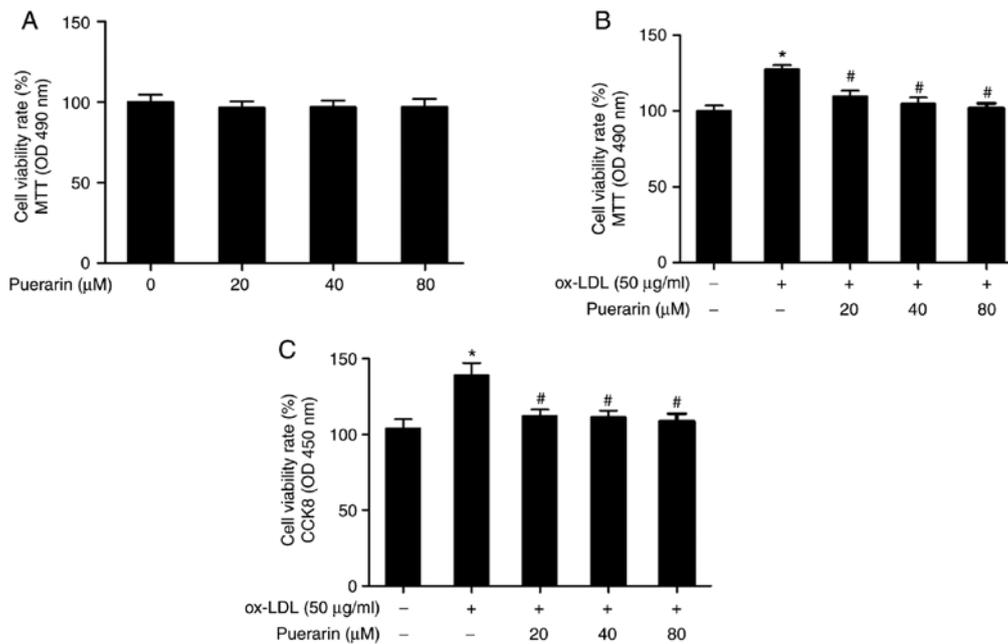


Figure 2. Puerarin inhibited cell viability in ox-LDL-stimulated VSMCs. The MTT assay was performed to assess the effect of puerarin on cell viability in (A) VSMCs and (B) ox-LDL-induced VSMCs. (C) The CCK-8 assay was performed to assess the effect of puerarin on cell viability in VSMCs. * $P < 0.05$ vs. control VSMCs; # $P < 0.05$ vs. ox-LDL-stimulated VSMCs. ox-LDL, oxidized low-density lipoprotein; VSMC, vascular smooth muscle cell; CCK-8, Cell Counting Kit-8; OD, optical density.

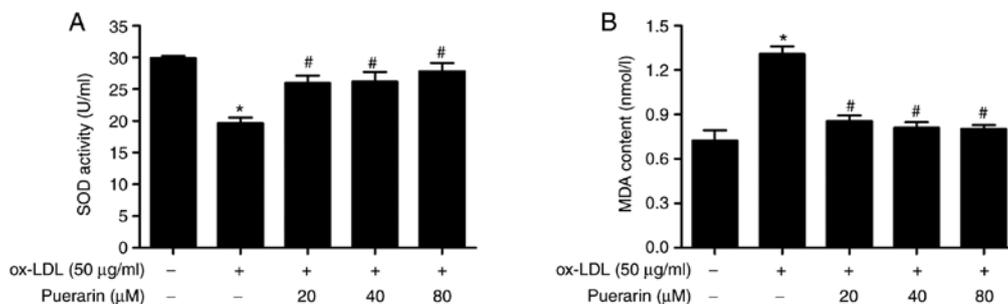


Figure 3. Puerarin regulates the production of oxidative stress-related makers in ox-LDL-induced VSMCs. Effect of puerarin on (A) SOD activity and (B) MDA content in ox-LDL-induced VSMCs. * $P < 0.05$ vs. control VSMCs; # $P < 0.05$ vs. ox-LDL-stimulated VSMCs. ox-LDL, oxidized low-density lipoprotein; VSMC, vascular smooth muscle cell; SOD, superoxide dismutase; MDA, malondialdehyde.

5'-TGGTCTTCTGGAGTTCGGTT-3'; TNF- α forward, 5'-TCTCATCAGTTCATGGCCC-3' and reverse, 5'-GGGAGTAGACAAGGTACAAC-3'; GAPDH forward, 5'-GTTACCAGGCTGCCTTCTC-3' and reverse, 5'-GATGGTGATGGTTTCCCGT-3'. mRNA expression levels were quantified using the $2^{-\Delta\Delta C_t}$ method (21) and normalized to the internal reference gene GAPDH.

Statistical analysis. Statistical analyses were performed using SPSS (version 19.0; IBM Corp.). All experiments were performed in triplicate. Comparisons among multiple groups were analyzed using one-way ANOVA followed by Tukey's post hoc test. Data are presented as the mean \pm SEM. $P < 0.05$ was considered to indicate a statistically significant difference.

Results

Puerarin inhibits cell viability in ox-LDL-stimulated VSMCs. To evaluate the cytotoxicity of puerarin on VSMCs, cells were

treated with puerarin (0, 20, 40 and 80 μ M) for 24 h. The MTT assay results suggested that puerarin did not significantly alter VSMC viability compared with the control group (Fig. 2A). Following incubation with different concentrations of puerarin for 24 h, cells were stimulated with 50 μ g/ml ox-LDL for 24 h to evaluate the effect of puerarin on cell viability in ox-LDL-induced VSMCs. Cell viability was significantly increased in ox-LDL-induced VSMCs compared with the control group (Fig. 2B). However, pretreatment with puerarin significantly inhibited ox-LDL-induced cell viability. Similar results were observed in the CCK-8 assay (Fig. 2C).

Puerarin regulates the production of oxidative stress-related markers in ox-LDL-stimulated VSMCs. To investigate the mechanism underlying the inhibitory effect of puerarin on ox-LDL-induced cell proliferation, the effect of puerarin on SOD activity and MDA content was determined. ox-LDL significantly increased the content of MDA in VSMCs compared with the control group (Fig. 3B). Puerarin

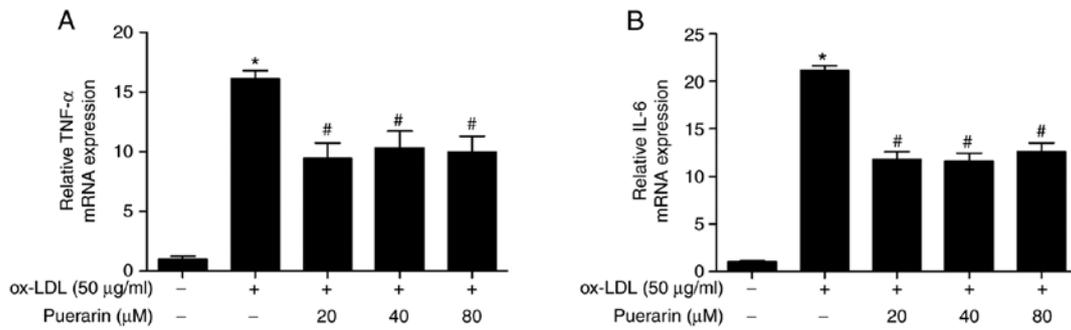


Figure 4. Puerarin inhibits proinflammatory cytokines in ox-LDL-induced VSMCs. Effect of puerarin on the mRNA expression levels of (A) TNF- α and (B) IL-6 in ox-LDL-induced VSMCs. * $P < 0.05$ vs. control VSMCs; # $P < 0.05$ vs. ox-LDL-stimulated VSMCs. ox-LDL, oxidized low-density lipoprotein; VSMC, vascular smooth muscle cell.

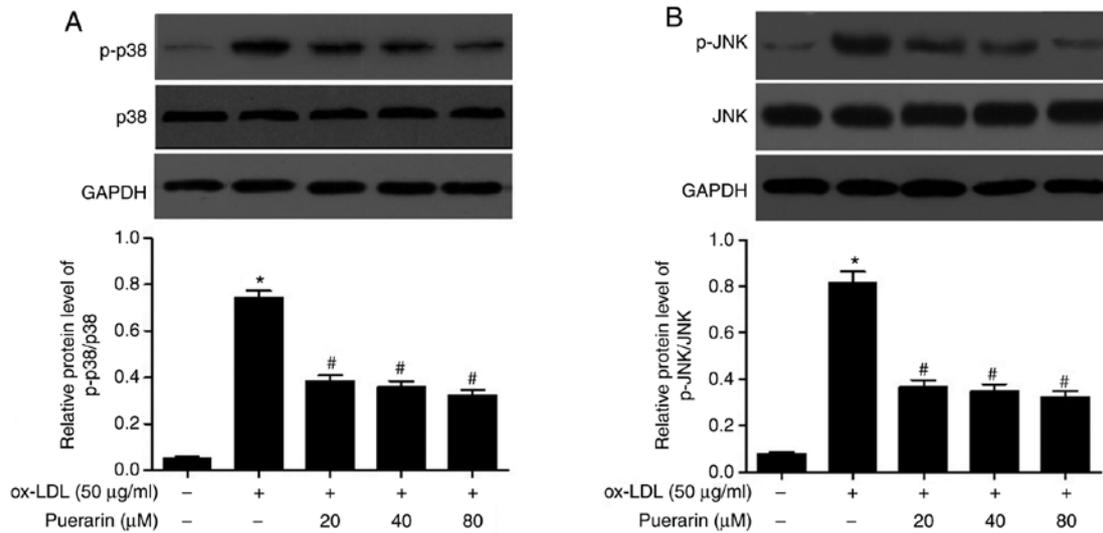


Figure 5. Puerarin prevents the activation of p38 MAPK and JNK signaling pathways in ox-LDL-induced VSMCs. The protein expression ratios of (A) p-38/p38 and (B) p-JNK/JNK were determined via western blotting. * $P < 0.05$ vs. control VSMCs; # $P < 0.05$ vs. ox-LDL-stimulated VSMCs. ox-LDL, oxidized low-density lipoprotein; VSMC, vascular smooth muscle cell; p, phosphorylated.

pretreatment significantly attenuated ox-LDL-induced MDA levels. In addition, compared with the control group, SOD activity was significantly decreased by ox-LDL in VSMCs, but pretreatment with puerarin diminished ox-LDL-mediated effects on SOD activity (Fig. 3B). The results suggested that puerarin significantly inhibited ox-LDL-induced oxidative stress in VSMCs.

Puerarin inhibits proinflammatory cytokines in ox-LDL-stimulated VSMCs. The effect of puerarin on IL-6 and TNF- α mRNA expression levels in ox-LDL-stimulated VSMCs was examined. ox-LDL significantly increased the mRNA expression levels of IL-6 and TNF- α in VSMCs compared with the control group (Fig. 4A). However, puerarin pretreatment significantly decreased the mRNA expression levels of IL-6 and TNF- α in ox-LDL-stimulated VSMCs (Fig. 4B). The results suggested that puerarin inhibited proinflammatory cytokines in ox-LDL-stimulated VSMCs.

Puerarin prevents the activation of p38 MAPK and JNK signaling pathways in ox-LDL-stimulated VSMCs. To further elucidate whether the p38 MAPK and JNK signaling pathways

were involved in the effect of puerarin in ox-LDL-stimulated VSMCs, the phosphorylation levels of p38 and JNK were analyzed via western blotting. The expression levels of p-p38 and p-JNK were significantly increased by ox-LDL in VSMCs compared with the control group (Fig. 5A and B). Puerarin pretreatment significantly decreased the expression levels of p-p38 and p-JNK in ox-LDL-stimulated VSMCs. The results indicated that puerarin inhibited the activation of p38 MAPK and JNK signaling pathways in ox-LDL-induced VSMCs.

Discussion

During the progression of atherosclerosis, abnormal VSMC proliferation and migration serve key roles in causing stenosis and intimal thickening (22). A number of factors affect VSMC proliferation and migration, including hypertension, dyslipidemia and oxidative stress (4). As a well-established index of oxidative stress, ox-LDL is involved in the generation of atherosclerotic lesions (4). Numerous studies have indicated that ox-LDL can induce VSMC proliferation and migration, which are pathological events that are crucial for neointima

formation (23) and stimulate activation of the MAPK signaling pathway (24,25). Therefore, if a substance can effectively prevent ox-LDL-induced VSMC proliferation, it should inhibit the development of atherosclerosis.

Puerarin, a major isoflavone isolated from the root of *Pueraria lobata*, has been reported to treat various cardiovascular diseases, including arterial hypertension, angina, myocardial infarction and arrhythmia (26-28). Puerarin has anti-atherosclerotic activity in diet-induced atherosclerosis in rabbits and rats (17,18). Puerarin prevents the pathogenesis of atherosclerosis, which may be mediated by its anti-inflammatory and antioxidative stress actions (28,29). A key finding of the present study was that puerarin significantly inhibited ox-LDL-induced VSMC viability. In the present study, whether cell viability was inhibited by puerarin when VSMCs were exposed to ox-LDL was investigated. The results demonstrated that ox-LDL enhanced VSMC viability compared with the control group, but pretreatment with puerarin significantly reversed ox-LDL-mediated effects in VSMCs. Moreover, compared with the control group, ox-LDL significantly decreased SOD activity and significantly increased MDA content, suggesting that the level of oxidative stress in VSMCs induced by ox-LDL was higher compared with control conditions. The antioxidant activity of puerarin was further indicated by its ability to reverse ox-LDL-mediated alterations in SOD activity and MDA content.

For the clinical development of puerarin as a treatment, understanding the mechanism underlying how it inhibits VSMC proliferation is important. The present study focused on the MAPK signaling pathway in VSMCs. Previous studies demonstrated that ox-LDL activated the MAPK signaling pathway, including p38, ERK and JNK in VSMCs, which promoted cell proliferation (30,31). Puerarin prevents ox-LDL-induced proliferation and inhibits the phosphorylation of ERK1/2 in VSMCs (32). Puerarin inhibits VSMC proliferation induced by fine particulate matter via reducing the elevated expression levels of p-p38 MAPK (33). Moreover, puerarin markedly stimulates bone marrow stromal cell differentiation towards an osteogenic phenotype via the ERK1/2 and p38-MAPK signaling pathways (34). The aforementioned study results indicated that puerarin displays protective effects in different diseases by regulating MAPK signaling pathways. In the present study, puerarin inhibited the activation of the p38 MAPK and JNK signaling pathways in ox-LDL-induced VSMCs.

Certain studies have speculated that the p38 MAPK signaling pathway is associated with inflammasome activation (5,35,36). Activation of the p38 MAPK signaling pathway increases the expression of TNF- α and IL-6 (36,37). In the present study, similar results were obtained. Although the present study did not further investigate the mechanism of MAPK signaling pathway and inflammasome activation, relative indices were observed and it was suggested that the MAPK signaling pathway was involved in the activation of inflammation in ox-LDL-induced VSMCs. The results of the present study indicated that the phosphorylation levels of p38 and JNK were significantly increased in ox-LDL-induced VSMCs compared with the control group. Similarly, the mRNA expression levels of IL-6 and TNF- α were significantly increased in ox-LDL-induced VSMCs compared

with control VSMCs. However, puerarin significantly down-regulated the expression levels of p-p38 and p-JNK and reduced the mRNA expression levels of IL-6 and TNF- α in ox-LDL-stimulated VSMCs. The effect of puerarin on IL-6 and TNF- α did not display a concentration gradient trend, which indicated that the effect of puerarin was not closely related to the concentration. The effect of puerarin on other proinflammatory cytokines requires further investigation. The results of the present study suggested that alterations in proinflammatory cytokines were closely related to alterations in the expression levels of p-p38 MAPK and p-JNK, indicating that puerarin altered the expression of TNF- α and IL-6 by regulating the p38 MAPK and JNK signaling pathways in ox-LDL-induced VSMCs. However, cell networks are complex, thus, whether puerarin exerts its function via other signaling pathways in ox-LDL-induced VSMCs requires further investigation.

In conclusion, the results of the present study suggest that puerarin inhibited ox-LDL-induced VSMC viability, and that the antiproliferative effects of puerarin were partly associated with inactivation of the p-p38 MAPK and p-JNK signaling pathways, which was mediated via suppression of the expression levels of TNF- α and IL-6. Therefore, the results of the present study suggest that puerarin may inhibit the development of atherosclerosis.

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Availability of data and materials

The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

YH performed the experiments and wrote the manuscript. RL, ZW and WY performed the cell experiments. HL conceived the study and revised the manuscript. WQ designed the study and analysed the data. All authors read and approved the final manuscript.

Ethics approval and consent to participate

Not applicable.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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