

# Analysis of crucial genes, pathways and construction of the molecular regulatory networks in vascular smooth muscle cell calcification

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**Abstract.** Vascular calcification (VC) accompanies the trans-differentiation of vascular smooth muscle cells (VSMCs) into osteo/chondrocyte-like cells and resembles physiological bone mineralization. However, the molecular mechanisms underlying VC initiation and progression have remained largely elusive. The aim of the present study was to identify the genes and pathways common to VSMC and osteoblast calcification and construct a regulatory network of non-coding RNAs and transcription factors (TFs). To this end, the Gene Expression Omnibus dataset GSE37558 including mRNA microarray data of calcifying VSMCs (CVSMCs) and calcifying osteoblasts (COs) was analyzed. The differentially expressed genes (DEGs) were screened and functionally annotated and the microRNA (miRNA/mRNA)-mRNA, TF-miRNA and long non-coding RNA (lncRNA)-TF regulatory networks were constructed. A total of 318 DEGs were identified in the CVSMCs relative to the non-calcified VSMCs, of which 43 were shared with the COs. The CVSMC-related DEGs were mainly enriched in the functional terms cell cycle, extracellular matrix (ECM), inflammation and chemotaxis-mediated signaling pathways, of which ECM was enriched by the DEGs for the COs as well. The protein-protein interaction network of CVSMCs consisted of 281 genes and 3,650 edges. There were 30 hub genes in this network, including maternal embryonic leucine zipper kinase (MELK), which potentially regulates the differentially expressed TF (DETF) forkhead box (FOX)M1

and is a potential target gene of *Homo sapiens* miR-485-3p and miR-181d. The TF-miRNA network included 251 TFs and 60 miRNAs, including 10 DETFs such as FOXO1 and snail family transcriptional repressor 2 (SNAI2). Furthermore, the lncRNAs H19 imprinted maternally expressed transcript (H19) and differentiation antagonizing non-protein coding RNA (DANCR) were predicted as the upstream regulators of FOXO1 and SNAI2 in the lncRNA-TF regulatory network. DANCR, MELK and FOXM1 were downregulated, and H19, FOXO1 and SNAI2 were upregulated in the CVSMCs. Taken together, the CVSMCs and COs exhibited similar molecular changes in the ECM. In addition, the MELK-FOXM1, H19/DANCR-FOXO1 and SNAI2 regulatory pathways likely mediate VSMC calcification.

## Introduction

Vascular calcification (VC) refers to the ectopic deposition of calcium phosphate crystals or hydroxyapatite on the vascular walls. It is frequently observed during ageing, as well as in degenerative diseases such as chronic kidney disease (CKD), diabetes and atherosclerosis, and significantly increases the risk of cardiovascular disease (CVD) and mortality (1-3). Contrary to the long-held surmise that calcium and phosphorus are passively deposited on the vascular walls, recent studies have indicated that VC is an active and cell-regulated process similar to the mineralization of osteo/chondrocyte-like cells during bone formation (4). The incidence of coronary artery calcification in CKD patients is >50% in the absence of dialysis and significantly higher at 70-90% among patients undergoing dialysis (5). In addition, increased calcium burden of the thoracic aorta elevates the risk of CVD by 3.7-fold (6), whereas abdominal aorta calcification increases the relative risk of coronary, cerebrovascular and cardiovascular events as well as mortality rates (7). VC is a significant risk factor of CVD in 90% of males and 67% of females above the age of 70 years (8).

Vascular smooth muscle cells (VSMCs) may differentiate into the major cell types in the vessel wall in response to suitable environmental stimuli. There is evidence that VSMCs lose their contractile phenotype and trans-differentiate into osteoblast-like cells expressing osteogenic transcription factors and proteins. These cells may initiate calcification of the vascular

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*Abbreviations:* VC, vascular calcification; CVSMCs, calcifying vascular smooth muscle cells; CO, calcifying osteoblast; DEGs, differentially expressed genes; GO, Gene Ontology

*Key words:* VSMC calcification, osteoblast, lncRNA, miRNA, transcription factor, bioinformatics

wall by secreting calcium and phosphorous-loaded exosomes into the extracellular matrix (ECM) (9-11). However, little is known regarding the specific pathways and molecular mechanisms underlying VC, which markedly limits the development of effective drugs.

Non-coding RNAs are a class of transcripts that regulate the expression of protein-encoding genes through various mechanisms. MicroRNAs (miRNAs/miRs) are 18-22 nucleotides in length and regulate target gene expression at the post-transcriptional level through binding at the 3' untranslated region. Long non-coding RNAs (lncRNAs) are >200 nucleotides long and exert their regulatory effects through more complex mechanisms (12). Several lncRNAs have been identified in recent years that are involved in the progression of various pathological conditions, including CVD (13-15). Furthermore, specific miRNAs and lncRNAs have been implicated in VSMC calcification. For instance, downregulation of miR-204, miR-29b or miR-30e trigger the osteogenic differentiation and calcification of VSMCs both *in vitro* and *in vivo*, whereas upregulation of miRNA-128 accelerates cardiovascular calcification (16-20). Lin *et al* (21) demonstrated that lncRNA-ES3 enhanced hyperglycemia-induced calcification of VSMCs by suppressing miR-34c-5p expression as a sponge. In addition, Jeong *et al* (22) identified numerous differentially expressed lncRNAs in calcified rat VSMCs, of which leucine rich repeat containing 75a-antisense (AS)1 was significantly downregulated and its ectopic expression attenuated calcium accumulation in VSMCs cultured with inorganic phosphate. However, the exact mechanistic roles of non-coding RNAs in VSMC calcification have remained to be elucidated.

In the present study, the differentially expressed genes (DEGs) and pathways in VSMCs exposed to high and normal calcium levels for varying durations were identified using bioinformatics. Given the physiological similarities between VCs and bone mineralization, the DEGs and pathways common to both calcifying VSMCs (CVSMCs) and osteoblasts (COs) were also screened and certain potentially crucial genes were experimentally validated. The putative regulatory networks of non-coding RNAs and transcription factors (TFs) in CVSMCs were also predicted. The present results provide novel insight into the molecular basis of the pathogenesis of VC.

## Materials and methods

**Microarray data and identification of DEGs.** The microarray dataset GSE37558 profiled on the GPL6947 platform (Illumina HumanHT-12 V3.0 expression beadchip) was downloaded from the National Center for Biotechnology Information (NCBI)-Gene Expression Omnibus database (<https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi>). It includes the mRNA expression data of 32 VSMCs and osteoblasts samples cultured for 0, 2, 8, 12 or 25 days (3-4 replicates per time-point) in calcified medium containing 1.8 mM Ca<sup>2+</sup> (23). The raw data were integrated and the DEGs between the control and calcified VSMCs were identified using the GEO2R tool, which is a GEO tool (<https://www.ncbi.nlm.nih.gov/geo/geo2r/?acc=GSE37558>) and Morpheus website (<https://software.broadinstitute.org>) using an adjusted P<0.05 with  $\log_2$  fold change >1 as the thresholds. The DEGs common to VSMCs calcified for varying durations were also identified

Table I. Primer sequences or numbers for PCR.

Gene symbol	Primer sequence (5'-3') or catalogue number
DANCR	Forward: CAGCTGACCCTTACCCTGAA Reverse: GACCCTGGGGTTGTTAGTCA
H19	Forward: CAGAGTCCGTGGCCAAGG Reverse: CGCCTTCAGTGACTGGCA
MELK	Forward: ACTGCCCTGGAGGAGAGCT Reverse: AGCCCTGGCTGTGCACATAA
FOXO1	Forward: GGAGGAAATGCCACACTTAGCG Reverse: TAGGACTTCTTGGGTCTTGGGGTG
FOXO1	Forward: TACGCCGACCTCATCACCAAG' Reverse: GCACGCTCTTCACCATCCACT'
SNAI2	Forward: CACCTCCTCCAAGGACCA' Reverse: GGCCAGCCCAGAAAAAGT
GAPDH	Forward: GTCAGCCGCATCTTCTTT' Reverse: CGCCCAATACGACCAAAT'
U6	RQP086799
Hsa-miR-485-3p	HmiRQP0521
Hsa- miR-181d-5p	HmiRQP0237

Hsa, *Homo sapiens*; miR, microRNA; FOX, forkhead box; DANCR, differentiation antagonizing non-protein coding RNA; H19, H19 imprinted maternally expressed transcript; MELK, maternal embryonic leucine zipper kinase; SNAI2, snail family transcriptional repressor 2.

using the same criteria. The DEGs in osteoblasts were similarly screened, and the shared DEGs between CVSMCs and COs were also defined using the Morpheus website.

**Gene ontology (GO) and pathway enrichment analysis.** The DEGs were functionally annotated by GO, Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway and reactome analyses using the Database for Annotation, Visualization and Integrated Discovery version 6.8 (<https://david.ncifcrf.gov/>). The significantly enriched genes or pathways were screened on the basis of P<0.05.

**Protein-protein interaction (PPI) network.** The PPI networks of the DEGs were constructed using a Search Tool for the Retrieval of Interacting Genes and proteins (STRING) database (<http://string.embl.de/>; accessed April, 2019) (24) in order to identify the interacting and hub genes. Cytoscape software (25) was used for visualizing the networks and analyzing the degree of connectivity of nodes. Finally, module analysis of the PPI network was performed using the molecular complex detection application of Cytoscape software. The module genes were functionally annotated as described above.

**Construction of regulatory networks.** The target miRNAs of calcification-related DEGs were predicted using the miRWalk (<http://zmf.umm.uni-heidelberg.de/>;



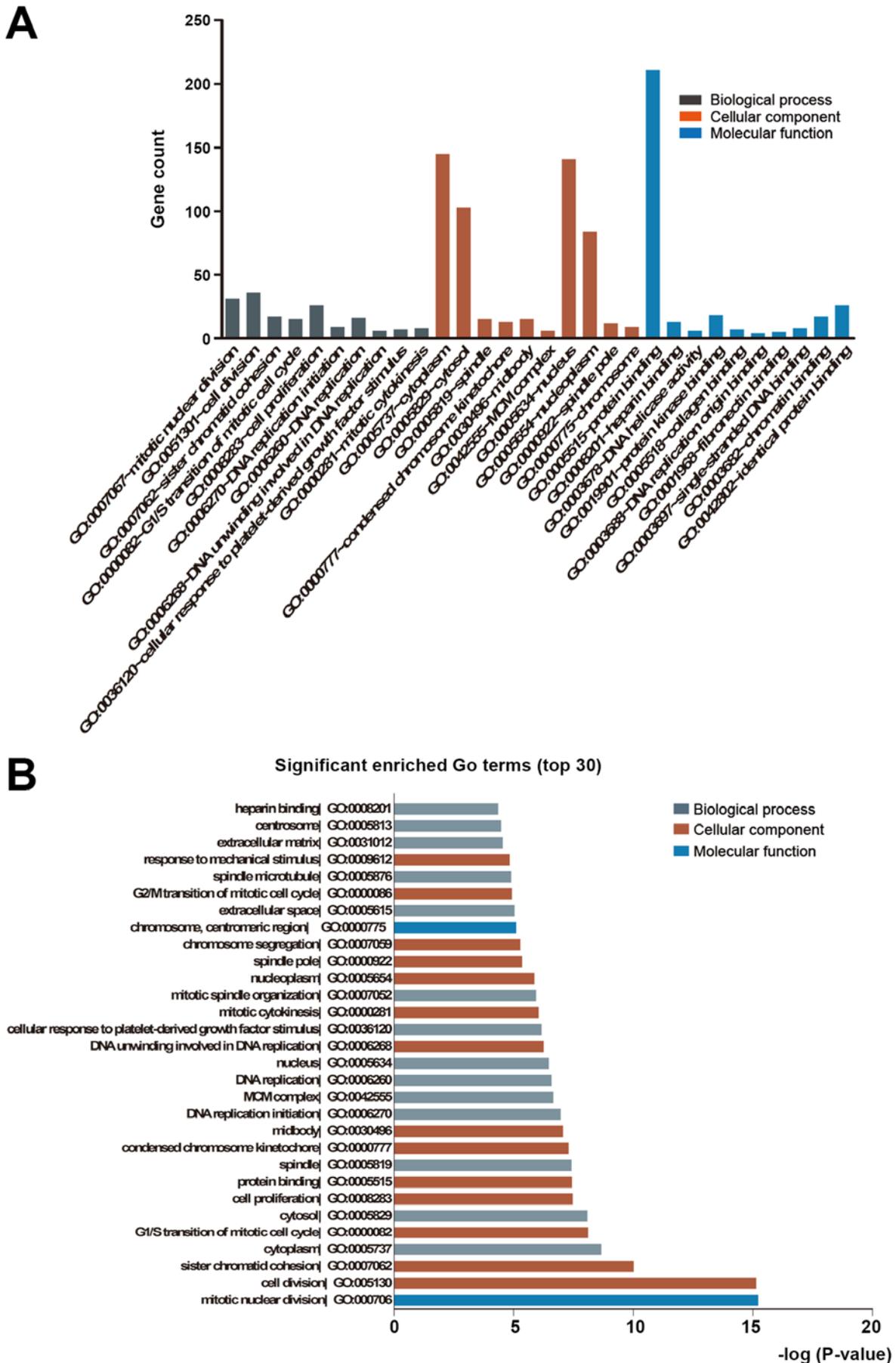


Figure 2. GO term enrichment analysis of DEGs from CVSMCs. (A) GO terms related to molecular function, biological process and cellular component enriched in the DEGs from CVSMCs. (B) Top 30 GO terms for DEGs from CVSMCs. CVSMCs, calcifying vascular smooth muscle cells; DEGs, differentially expressed genes; GO, Gene Ontology.

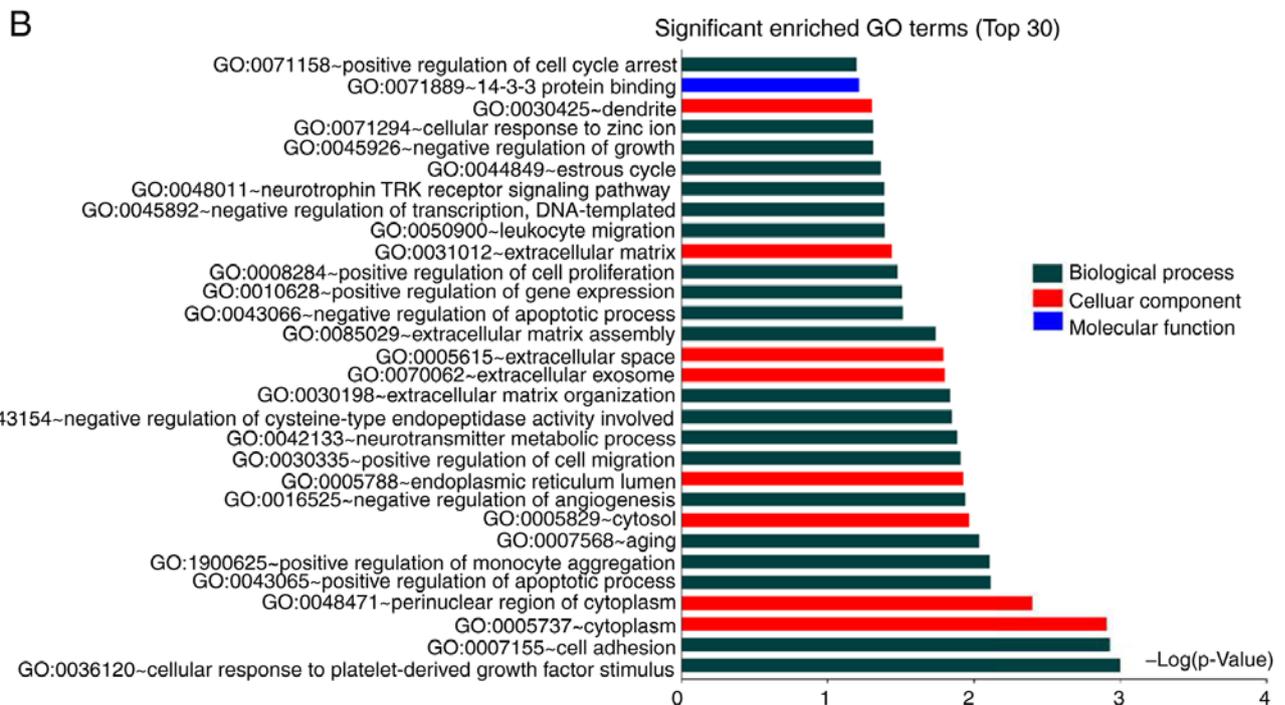
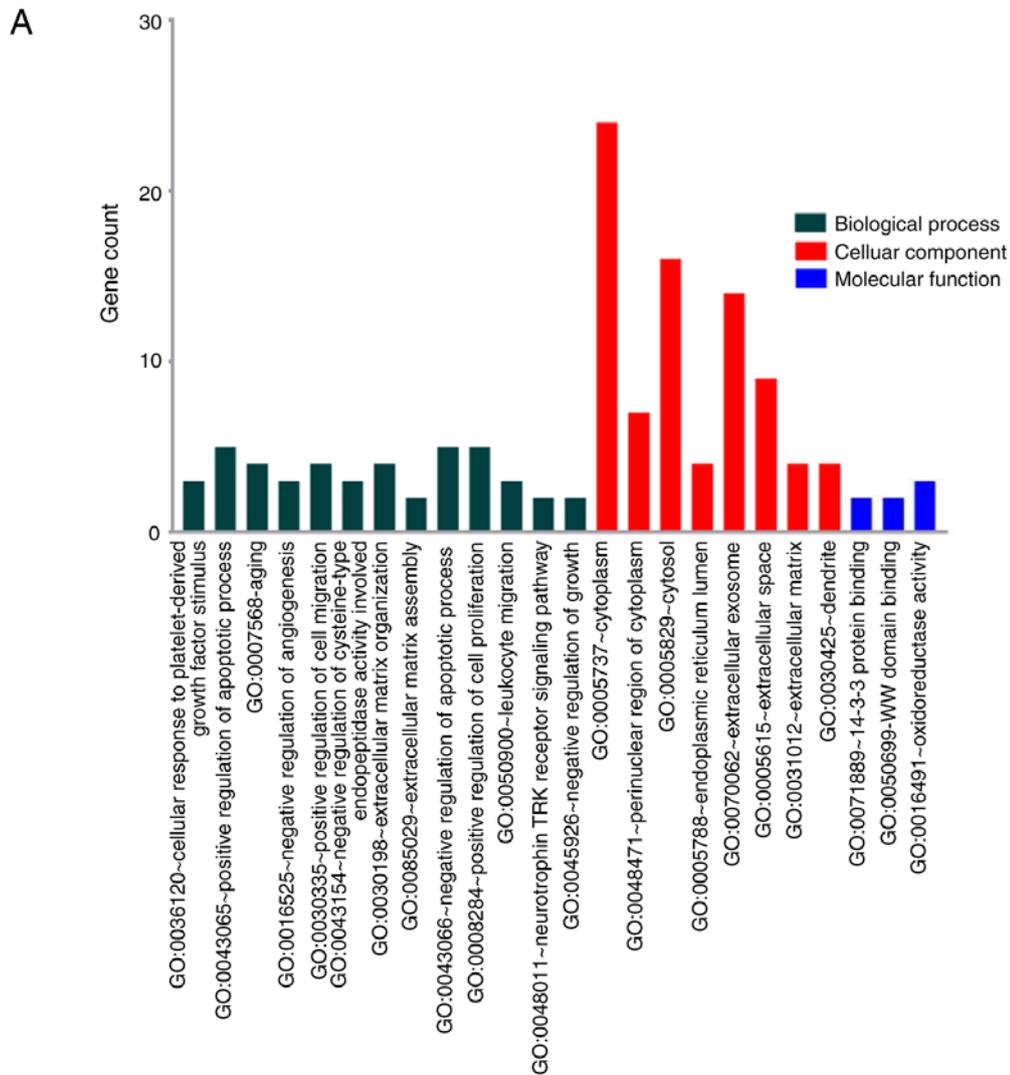


Figure 3. GO analysis of DEGs shared between CVSMCs and CO. (A) GO terms of DEGs in CVSMCs and CO according to molecular function, biological process and cellular component. (B) Top 30 GO terms for DEGs common to CVSMCs and COs. CVSMCs, calcifying vascular smooth muscle cells; CO, calcifying osteoblast; DEGs, differentially expressed genes; GO, Gene Ontology.

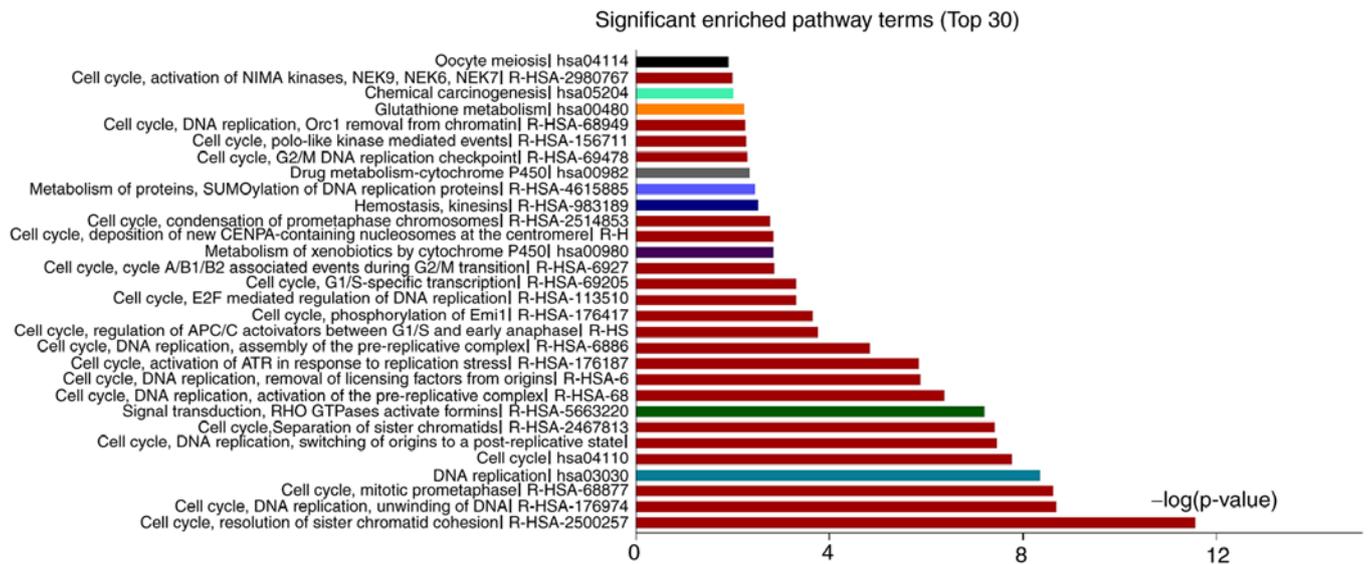


Figure 4. Top 30 signaling pathways for calcification-related differentially expressed genes in calcifying vascular smooth muscle cells determined using Kyoto Encyclopedia of Genes and Genomes pathway analysis and Reactome online dataset. Hsa, *Homo sapiens*.

**RNA extraction and reverse transcription-quantitative (RT-q) PCR analysis.** TRIzol® (Invitrogen; Thermo Fisher Scientific, Inc.) was used to extract total RNA from the cultured VSMCs according to the manufacturer's protocol. The integrity of the isolated RNA was determined by measuring the absorbance at 260 nm (A260) and the purity in terms of the A260/A280 ratio. Total RNA was reverse-transcribed into complementary (c)DNA using the TransScript kit (Takara Biotechnology, Inc.). qPCR was performed with 2  $\mu$ l first-strand cDNA as the template and the FastStart SYBR Green kit (Roche). The primers were purchased from GeneCopoeia Biotechnology Company and their sequences are presented in Table I. The thermocycling conditions of qPCR were as follows: 2 min at 50°C and 10 min at 95°C, followed by 40 cycles of 15 sec at 95°C, 30 sec 60°C and 30 sec at 72°C (ABI 7500 Fast; Applied Biosystems; Thermo Fisher Scientific, Inc.). GAPDH and U6 were used as the internal controls for mRNAs and miRNAs, respectively. The relative expression of these genes was determined using the comparative CT ( $2^{-\Delta\Delta C_t}$ ) method (28).

**Statistical analysis.** The *in vitro* experiments were performed in triplicate and were independently repeated three times. SPSS version 21.0 (IBM Corp.) was used for statistical analysis. Data were expressed as the mean  $\pm$  standard deviation and compared using an unpaired t-test.  $P < 0.05$  was considered to indicate statistical significance.

## Results

**Identification of calcification-related DEGs in the CVSMCs and COs.** To identify the genes affected by short- and long-term high  $Ca^{2+}$  exposure, VSMCs were cultured in the calcification medium for varying durations and DEGs were detected (GSE37558). As presented in Fig. 1A, VSMCs cultured for 2, 8, 12 and 24 h had 393, 684, 892 and 799 DEGs relative to the normal cells (0-day control), respectively. Furthermore, 318 genes were consistently differentially expressed across all

time-points, including 147 upregulated and 185 downregulated genes (Fig. 1A, Tables SI and SII). Furthermore, there were 206, 441, 565 and 681 DEGs in the osteoblasts cultured for 2, 8, 12 and 25 days, respectively, in the calcification medium compared to normal osteoblasts, of which 120 DEGs were common to all time-points (Fig. 1B, Tables SIII and SIV). Furthermore, 43 DEGs were common to the CVSMCs and COs, including 34 upregulated and 9 downregulated genes (Fig. 1C).

**Functional annotation of calcification-related DEGs.** The common DEGs (see above) were functionally annotated in GO terms and KEGG pathways. The upregulated genes were significantly enriched in biological process terms of ECM organization, cell adhesion, positive regulation of apoptotic process and positive regulation of gene expression, whereas the downregulated DEGs were significantly enriched in cell division, mitotic nuclear division, cell proliferation and cell cycle. The significant molecular function terms for upregulated genes were integrin, heparin, receptor and fibronectin binding, while those for downregulated genes were protein, ATP, protein kinase and chromatin binding. Finally, in the category cellular component, the upregulated genes were mainly enriched in terms such as cytoplasm, extracellular space, extracellular exosome, ECM and extracellular region, while the downregulated genes were mainly related to the nucleoplasm, nucleus, cytosol and cytoplasm (Fig. 2A and B, Table SV). GO analysis of the shared DEGs between CVSMCs and CO revealed cytoplasm, ECM/exosome/space and tyrosine metabolism as the enriched terms (Fig. 3A and B, Table SVI). These results indicate that the functional clusters of these DEGs are closely related to the cell cycle, ECM and cell binding. Furthermore, the ECM has an important role in both VSMC and osteoblast calcification.

The significantly enriched pathways of upregulated genes were platelet-derived growth factor, mineral absorption, vascular smooth muscle contraction, focal adhesion and drug

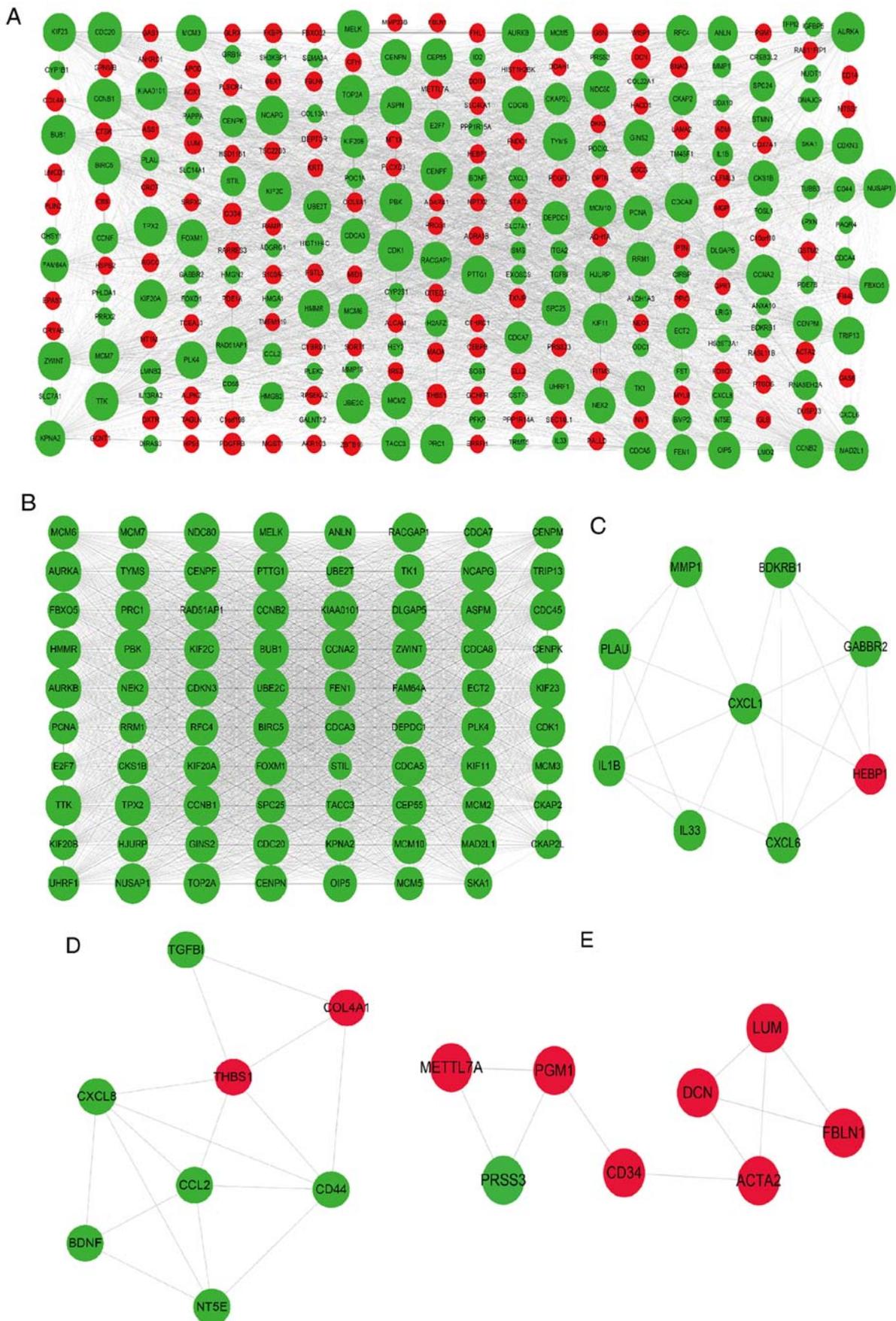


Figure 5. PPI networks and modular analysis of DEGs. (A) PPI network of DEGs in calcifying vascular smooth muscle cells, consisting of 281 gene nodes and 3,650 edges. The red and green gene nodes represent the upregulated and downregulated genes, respectively. The volume of gene nodes is proportional to the degree of connectivity. (B) Module 1 consisted of 79 gene nodes and 2,770 edges. (C) Module 2 consisted of 9 gene nodes and 20 edges. (D) Module 3 consisted of 8 gene nodes and 16 edges. (E) Module 4 consisted of 8 gene nodes and 10 edges. Module analysis utilized the following cut-off criteria: Degree cutoff, 2; node score cutoff, 0.2; K-core cutoff, 2; and max depth, 100. DEGs, differentially expressed genes; PPI, protein-protein interaction.

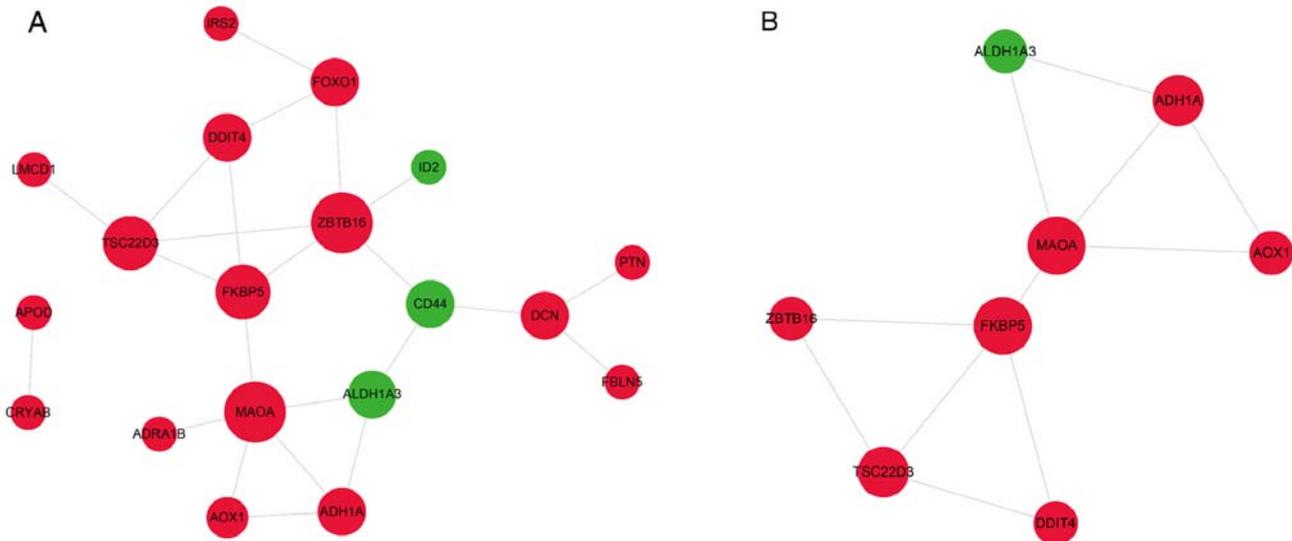


Figure 6. Protein-protein interaction of shared DEGs between calcifying vascular smooth muscle cells and calcifying osteoblasts. (A) Module 1 of the shared DEGs consisted of 19 gene nodes and 23 edges. (B) Module 2 of the shared DEGs consisted of 8 gene nodes and 11 edges. The red and green nodes represent upregulated and downregulated genes, respectively. The volume of gene nodes is proportional to the degree of connectivity. Module analysis utilized the following cut-off criteria: Degree cutoff, 2; node score cutoff, 0.2; K-core cutoff, 2; and max depth, 100. DEGs, differentially expressed genes.

metabolism-cytochrome P450, and those for downregulated DEGs were resolution of sister chromatid cohesion, mitotic prometaphase, cell cycle and separation of sister chromatids (Fig. 4, Table SVII). Finally, the DEGs common to CVMCs and COs were enriched in pathways of amino acid metabolism and metallothionein-binding metals (Table SVIII). Thus, genes related to distinct pathways were affected during calcification of VSMCs and the downregulated genes in particular were strongly associated with cell cycle and proliferation.

*Identification of hub genes involved in VSMC calcification and osteoblast mineralization.* The hub genes involved in the calcification of VSMCs were next identified by constructing a PPI network of the 318 common DEGs. As presented in Fig. 5A, the PPI network consisted of 281 genes, including 143 upregulated and 175 downregulated genes, and 3,650 edges. A total of 30 genes with a degree of connectivity of >80 in the PPI network were designated as the hub genes, including cyclin dependent kinase 1, mitotic arrest deficient 2 like 1 (MAD2L1), kinesin family member 11 (KIF11), maternal embryonic leucine zipper kinase (MELK), non-SMC condensin I complex subunit G (NCAPG), PDZ binding kinase (PBK), kinesin family member 23 (KIF23) and cyclin A2. Module analysis of the PPI network using the MCODE app further revealed 4 modules (Fig. 5B-E). Module 1 with 79 nodes and 2,770 edges was closely associated with the cell cycle, cell division and separation of sister chromatid pathways. Module 2 consisted of 9 nodes and 20 edges and was significantly enriched in the pathways of inflammatory response, immune response, chemokine-mediated signaling and positive regulation of interleukin (IL)-6 production (Fig. 7A and Table SIX). In addition, module 3 was comprised of 8 nodes and 16 edges and was mainly involved in ECM organization, cellular response to fibroblast growth factor stimulus, cell adhesion, chemokine-mediated signaling and NOD-like receptor signaling pathways. Finally, module 4

had 8 nodes and 10 edges, and was associated with extracellular space and ECM pathways (Fig. 7B and Table SIX). These results suggested that signaling pathways related to the cell cycle, immune response, ECM, chemotaxis and inflammatory response have a key role in VSMC calcification.

VSMCs are able to differentiate into osteoblast-like cells expressing osteogenic proteins in response to dysregulated calcium-phosphate metabolism and thus contribute to VC (9,10). Therefore, the PPI network of the 43 DEGs shared by CVMCs and COs was also established in order to identify signaling pathways involved in VSMCs and osteoblast mineralization. As presented in Fig. 6A, the PPI network contained 19 gene nodes and 23 edges. One module including 8 nodes and 11 edges (Fig. 6B) was identified, which was associated with amino acid metabolism, drug metabolism-cytochrome P450, metabolic pathways and oxidoreductase activity (Fig. 7C and Table SX).

*VSMC calcification-related regulatory networks.* To further explore the regulatory mechanisms involved in VSMC calcification, miRNAs targeting the CVMC-related DEGs were predicted by the miRanda, miRDB, miRWalk, RNA22 and TargetScan databases. A total of 76 putative miRNAs and 140 miRNA-target pairs were identified. A miRNA-target regulatory network was constructed with 76 miRNAs, 53 DEGs and 140 edges, and included *Homo sapiens* (hsa)-miR-20a-PBK, hsa-miR-15a- kinesin family member 23 (KIF23), hsa-miR-511-snail family transcriptional repressor 2 (SNAI2), hsa-miR-507-SNAI2, hsa-miR-181d- maternal embryonic leucine zipper kinase (MELK) and hsa-miR-485-3p-MELK regulatory pairs (Fig. 8 and Table SXI). Similarly, lncRNA and TF-mediated regulatory networks were also constructed. A total of 251 TFs were predicted and the TF-miRNA regulatory network consisted of 754 miRNA-target pairs, 251 TFs and 60 target DEGs (Fig. 9A and Table SXII). The TFs FOS like 1, AP-1

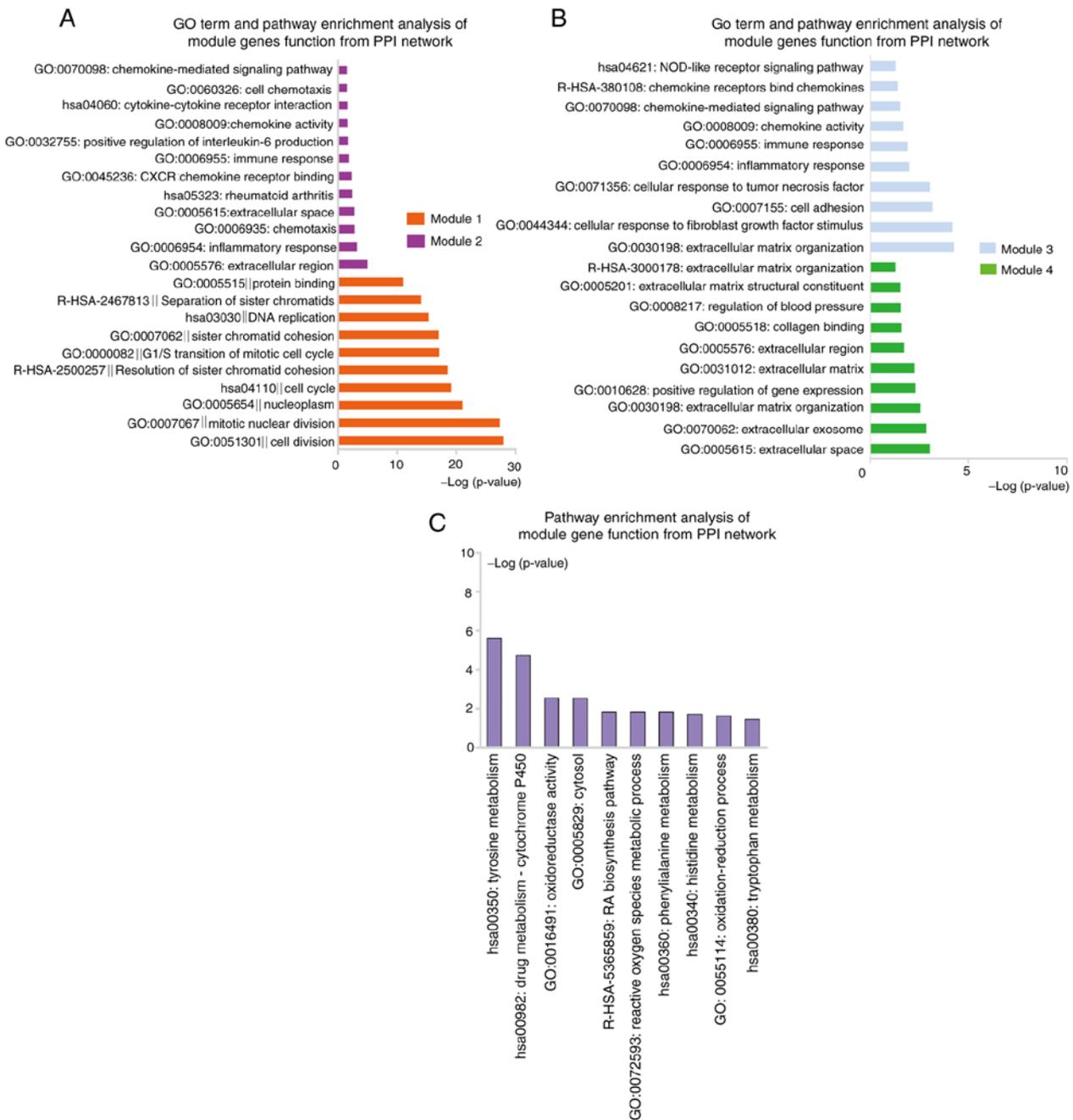


Figure 7. GO terms and pathways enrichment analysis of module genes from the PPI network. (A and B) GO terms and pathways enriched in the module genes in CVSMCs. (C) GO terms and pathways enriched in module genes between CVSMCs and calcifying osteoblasts. CVSMCs, calcifying vascular smooth muscle cells; PPI, protein-protein interaction; GO, Gene Ontology; Hsa, *Homo sapiens*.

transcription factor subunit, SNAI2, zinc finger and BTB domain containing 16, high mobility group AT-hook 1, CCAAT enhancer binding protein beta (CEBPB), E2F transcription factor 7 (E2F7), IL1B, forkhead box (FOX)M1, endothelial PAS domain protein 1 and forkhead box O1 (FOXO1) were differentially expressed and were used to construct a DETF-miRNA-target network, including 29 DETF-miRNAs and 27 miRNAs-target DEG pairs, such as FOXO1-hsa-miR-145, SNAI2-hsa-miR-221/222, SNAI2-hsa-miR-200a and SNAI2-hsa-miR-145 (Fig. 9B and Table SXIII). The hub genes in the PPI network that were potential targets of the predicted miRNAs, such as MAD2L1, KIF11, NCAPG, PBK, KIF23 and MELK, were incorporated into the TF-miRNA-hub

gene network. The latter included 135 TF-miRNA and 13 miRNA-hub gene pairs, such as CEBPB-hsa-miR-20a-PBK, E2F7-hsa-miR-15a-KIF23 and RUNX family transcription factor 1 (RUNX1)-hsa-miR-181d-MELK (Fig. 9C and Table SXIV). Since CEBPB and E2F7 were identified as DETFs, the regulatory lncRNAs were next predicted. The putative CEBPB-targeting lncRNAs were metastasis associated lung adenocarcinoma transcript 1, chondrogenesis-associated transcript, RAD51 antisense RNA 1 and negative regulator of antiviral response, whereas FOXF1, tensin 1, non-coding RNA activated by DNA damage and CDK6 antisense RNA 1 were predicted to regulate E2F7. In addition, lncRNAs regulating FOXO1 and SNAI2 were



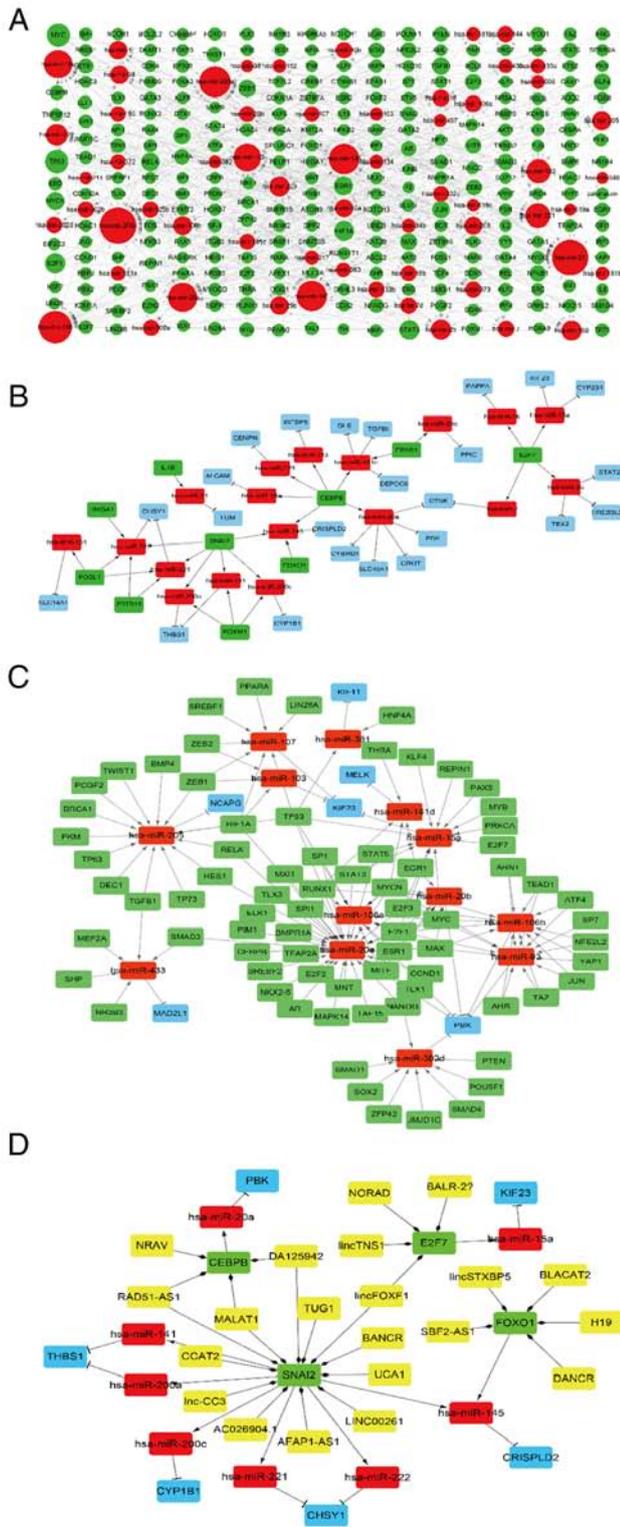


Figure 9. LncRNA-TF-miRNA-target gene regulatory network. (A) TF-miRNA regulatory network. Red and green nodes represent the miRNAs and TFs, respectively. The volume of gene nodes is proportional to the degree of connectivity of the gene nodes. (B) DETF-miRNA-target gene regulatory network. Red, green and blue nodes represent miRNAs, DETFs and DEGs, respectively. (C) TF-target miRNA-target hub gene regulatory network. Red, green and blue nodes represent miRNAs, TFs and target hub genes from the PPI network, respectively. (D) LncRNA-DETF-miRNA-target gene regulatory network. Yellow, green, red and blue nodes represent LncRNAs, DETFs, miRNAs and DEGs, respectively. Arrows, diamonds and 'T' respectively indicate the TF-miRNA, lncRNA-target DETF and miRNA-target gene relationships. PPI, protein-protein interaction; TF, transcription factor; DETF, differentially expressed TF; lncRNA, long non-coding RNA; miRNA, microRNA; DEG, differentially expressed gene.

collagen XIV and cartilage oligomeric matrix protein were all able to inhibit VSMC calcification and osteogenic differentiation through different signaling pathways (35,36,40,41).

Several hub genes were identified in the PPI networks, several of which may regulate VSMC calcification. For instance, inhibition of MELK and its downstream target genes with the specific inhibitor OTSSP167 was reported to enhance osteoblast formation and matrix mineralization (42). In line with this, in the present study, MELK was downregulated in the CVSMCs, indicating that it is likely inhibited during VSMC calcification. The TFs FOXM1 and EZH2 are the downstream targets of MELK (43), of which FOXM1 was significantly downregulated in the present study. Dioscin-mediated inhibition of FOXM1 reduced VSMC proliferation and migration *in vitro*, as well as intimal thickening in a rat model with carotid artery balloon injury (44). EZH2 is also known to suppress osteogenic differentiation of mesenchymal cells and its inhibition promoted osteoblast differentiation (45,46). Therefore, the MELK-FOXM1/EZH2 axis negatively regulates osteogenic differentiation and mineralization. MELK was also predicted as a target gene of hsa-miR-485-3p and hsa-miR-181d miRWalk (<http://zmf.umm.uni-heidelberg.de>; accessed April, 2019), indicating a novel has-485-3p/miR-181d-MELK-FOXM1/EZH2 axis (Fig. 11) in VSMC calcification that is worth exploring.

Other DETFs identified in the present study were FOXO1 and SNAI2. FOXO1 was previously reported to inhibit the osteogenic TF RUNX2 in prostate cancer cells (47,48) and its inhibition in VSMCs prevented RUNX2 ubiquitination, which increased RUNX2 levels and calcification (49). However, FOXO1 was determined to be upregulated in the present study, suggesting that it may also facilitate VSMC calcification. Consistent with this finding, FOXO1 levels were previously indicated to be increased in calcified femoropopliteal arteries from human subjects. Furthermore, five lncRNAs, including H19, DANCR, SBF2 antisense RNA 1, long intergenic non-protein coding syntaxin binding protein 5 and long intergenic non-protein coding RNA 958, were predicted as regulators of FOXO1 in this study. H19 downregulated FOXO1 in bovine skeletal muscle satellite cells, induced the osteogenic phenotype in valve interstitial cells and promoted aortic valve calcification by upregulating RUNX2 and bone morphogenetic protein (BMP)2, and inhibiting notch receptor 1 (50,51). It is also upregulated during osteogenesis of human mesenchymal stem cells and promotes osteoblast differentiation via the Wnt/ $\beta$ -catenin pathway (52). However, the role of the H19-FOXO1 axis in VSMC calcification and differentiation has remained elusive. DANCR regulates FOXO1 expression by affecting its ubiquitination (53). Zhu and Xu (54) and Jia *et al* (55) indicated that downregulation of DANCR promoted osteogenic differentiation of human fetal osteoblastic cells and periodontal ligament stem cells, respectively. Therefore, it may be hypothesized that the DANCR-FOXO1 axis potentially regulates osteogenic differentiation of VSMCs. In addition, hsa-miR-145 is a potential biomarker of VC in chronic kidney disease (56) and was predicted as a target miRNA of FOXO1 in the DETF-miRNA-target regulatory network. Taken together, the H19/DANCR-FOXO1-miRNA-145-target gene axis has a crucial role during VSMC calcification and should be experimentally validated (Fig. 11). SNAI2 was predicted as

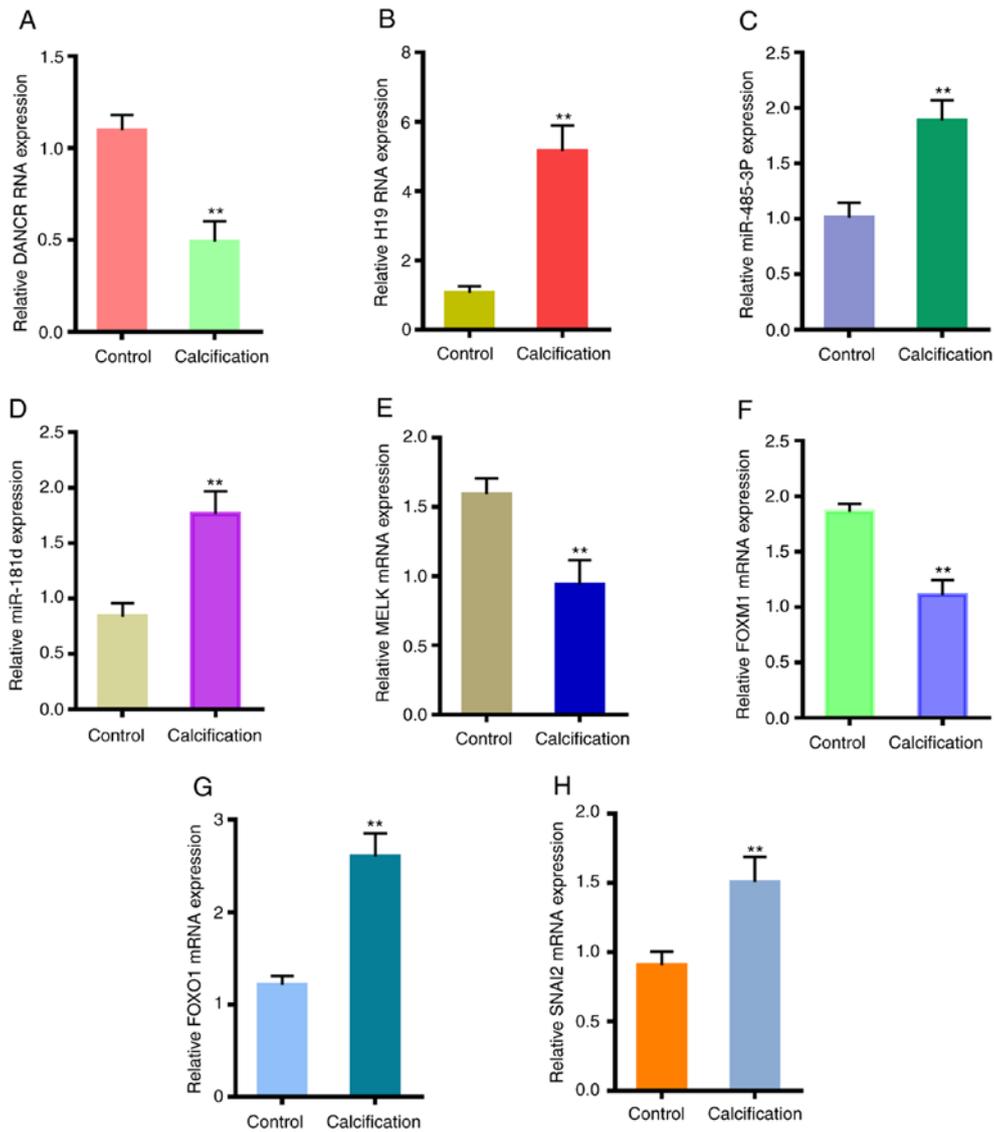


Figure 10. Validation of crucial genes in CVSMCs. Expression levels of (A) DANCR, (B) H19, (C) miR-485-3p, (D) miR-181d, (E) MELK, (F) FOXM1, (G) FOXO1 and (H) SNAI2 in the normal VSMCs and CVSMCs. \*\*P<0.05 vs. control. CVSMCs, calcifying vascular smooth muscle cells; miR, microRNA; FOX, forkhead box; DANCR, differentiation antagonizing non-protein coding RNA; H19, H19 imprinted maternally expressed transcript; MELK, maternal embryonic leucine zipper kinase; SNAI2, snail family transcriptional repressor 2.

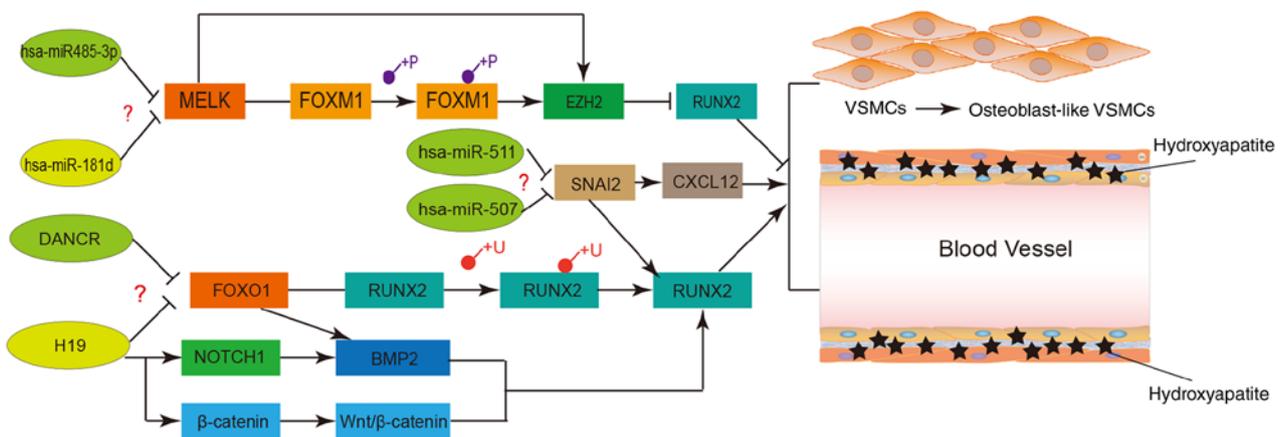


Figure 11. Potential regulatory networks and pathways of certain crucial long non-coding RNAs, miRNAs and proteins involved in VSMC calcification. VSMCs, vascular smooth muscle cells; miRNA/miR, microRNA; FOX, forkhead box; DANCR, differentiation antagonizing non-protein coding RNA; H19, H19 imprinted maternally expressed transcript; MELK, maternal embryonic leucine zipper kinase; SNAI2, snail family transcriptional repressor 2. RUNX2, RUNX family transcription factor 2; CXCL12, C-X-C motif chemokine ligand 12; NOTCH1, notch receptor 1; EZH2, enhancer of zeste 2 polycomb repressive complex 2 subunit; BMP2, bone morphogenetic protein 2; β-catenin, catenin beta interacting protein 1.

a target of six miRNAs. Previous studies indicated that SNAI2 promotes osteoblast maturation by upregulating RUNX2, as well as osteoblast mineralization through C-X-C motif chemokine ligand 12 signaling (57,58). In addition, SNAI2 also mediated BMP-dependent transdifferentiation of mouse non-ciliated aortic endothelial cells into mineralizing osteogenic cells and promoted atherosclerosis and VC *in vivo* (59). In line with this, in the present study, the upregulation of SNAI2 suggested its stimulatory effect on VSMC calcification. Furthermore, hsa-miR-511 and hsa-miR-507, as well as 12 lncRNAs, were predicted to regulate SNAI2. Most of these lncRNAs were identified in tumors. Therefore, it was hypothesized that the hsa-miR-511/hsa-miR-507-SNAI2 axis and the lncRNAs-SNAI2 axis potentially regulate VSMC calcification and should be explored further (Fig. 11).

In conclusion, the present study identified several potential regulatory mechanisms of VC and the DEGs and signaling pathways associated with the calcification of VSMCs. They involve changes in the inflammatory response, chemotaxis and ECM, and the latter is characteristic of osteoblast mineralization as well. Mechanistically, the hsa-485-3p/miR-181d-MELK-FOXO1/EZH2, H19/DANCR-FOXO1 and SNAI2-mediated pathways possibly regulate VC. These mechanisms require to be experimentally validated in future studies.

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#### Availability of data and materials

The data sets used and/or analyzed during the current study are available from the corresponding author on reasonable request. In addition, the dataset GSE37558 may be obtained from the GEO database (<https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE37558>).

#### Authors' contributions

XW, YS, QL, ZZ and PH contributed to the study conception and design. PH, XW and YS checked the associated databases and analyzed raw data for bioinformatics analysis, cell culture, PCR. XW, QL and ZZ wrote and revised the manuscript. All of the authors read and approved the final manuscript. PH and XW checked and approved the authenticity of the raw data. All authors read and approved the final manuscript.

#### Ethics approval and consent to participate

Not applicable.

#### Patient consent for publication

Not applicable.

#### Competing interests

The authors declare that they have no competing interests.

#### References

- Lanzer P, Boehm M, Sorribas V, Thiriet M, Janzen J, Zeller T, St Hilaire C and Shanahan C: Medial vascular calcification revisited: Review and perspectives. *Eur Heart J* 35: 1515-1525, 2014.
- Kovacic JC and Fuster V: Vascular calcification, diabetes, and cardiovascular disease: Connecting the dots. *JACC Cardiovasc Imaging* 5: 367-369, 2012.
- Chen NX and Moe SM: Pathophysiology of vascular calcification. *Curr Osteoporos Rep* 13: 372-380, 2015.
- Leopold JA: Vascular calcification: Mechanisms of vascular smooth muscle cell calcification. *Trends Cardiovasc Med* 25: 267-274, 2015.
- Block GA, Spiegel DM, Ehrlich J, Mehta R, Lindbergh J, Dreisbach A and Raggi P: Effects of sevelamer and calcium on coronary artery calcification in patients new to hemodialysis. *Kidney Int* 68: 1815-1824, 2005.
- Blomberg BA, de Jong PA, Thomassen A, Lam MGE, Vach W, Olsen MH, Mali WPTM, Narula J, Alavi A and Højlund-Carlsen PF: Thoracic aorta calcification but not inflammation is associated with increased cardiovascular disease risk: Results of the CAMONA study. *Eur J Nucl Med Mol Imaging* 44: 249-258, 2017.
- Bastos Gonçalves F, Voûte MT, Hoeks SE, Chonchol MB, Boersma EE, Stölker RJ and Verhagen HJ: Calcification of the abdominal aorta as an independent predictor of cardiovascular events: A meta-analysis. *Heart* 98: 988-994, 2012.
- Bild DE, Detrano R, Peterson D, Guerci A, Liu K, Shahar E, Ouyang P, Jackson S and Saad MF: Ethnic differences in coronary calcification: The Multi-Ethnic Study of Atherosclerosis (MESA). *Circulation* 111: 1313-1320, 2005.
- Zhang MJ, Zhou Y, Chen L, Wang YQ, Wang X, Pi Y, Gao CY, Li JC and Zhang LL: An overview of potential molecular mechanisms involved in VSMC phenotypic modulation. *Histochem Cell Biol* 145: 119-130, 2016.
- Gomez D and Owens GK: Smooth muscle cell phenotypic switching in atherosclerosis. *Cardiovasc Res* 95: 156-164, 2012.
- Kapustin AN and Shanahan CM: Calcium regulation of vascular smooth muscle cell-derived matrix vesicles. *Trends Cardiovasc Med* 22: 133-137, 2012.
- Yao RW, Wang Y and Chen LL: Cellular functions of long noncoding RNAs. *Nat Cell Biol* 21: 542-551, 2019.
- Bar C, Chatterjee S and Thum T: Long noncoding RNAs in cardiovascular pathology, diagnosis, and therapy. *Circulation* 134: 1484-1499, 2016.
- Barwari T, Joshi A and Mayr M: MicroRNAs in cardiovascular disease. *J Am Coll Cardiol* 68: 2577-2584, 2016.
- Haemmig S, Simion V, Yang D, Deng Y and Feinberg MW: Long noncoding RNAs in cardiovascular disease, diagnosis, and therapy. *Curr Opin Cardiol* 32: 776-783, 2017.
- Jiang W, Zhang Z, Yang H, Lin Q, Han C and Qin X: The involvement of miR-29b-3p in arterial calcification by targeting matrix metalloproteinase-2. *Biomed Res Int* 2017: 6713606, 2017.
- Cui RR, Li SJ, Liu LJ, Yi L, Liang QH, Zhu X, Liu GY, Liu Y, Wu SS, Liao XB, *et al*: MicroRNA-204 regulates vascular smooth muscle cell calcification in vitro and in vivo. *Cardiovasc Res* 96: 320-329, 2012.
- Ding W, Li J, Singh J, Alif R, Vazquez-Padron RI, Gomes SA, Hare JM and Shehadeh LA: miR-30e targets IGF2-regulated osteogenesis in bone marrow-derived mesenchymal stem cells, aortic smooth muscle cells, and ApoE 2/2 mice. *Cardiovasc Res* 106: 131-142, 2015.
- Du Y, Gao C, Liu Z, Wang L, Liu B, He F, Zhang T, Wang Y, Wang X, Xu M, *et al*: Upregulation of a disintegrin and metalloproteinase with thrombospondin motifs-7 by miR-29 repression mediates vascular smooth muscle calcification. *Arterioscler Thromb Vasc Biol* 32: 2580-2588, 2012.
- Wang XY, Zhang XZ, Li F and Ji QR: MiR-128-3p accelerates cardiovascular calcification and insulin resistance through ISL1-dependent Wnt pathway in type 2 diabetes mellitus rats. *J Cell Physiol* 234: 4997-5010, 2019.

21. Lin X, Zhan JK, Zhong JY, Wang YJ, Wang Y, Li S, He JY, Tan P, Chen YY, Liu XB, *et al.*: lncRNA-ES3/miR-34c-5p/BMF axis is involved in regulating high-glucose-induced calcification/senescence of VSMCs. *Aging (Albany NY)* 11: 523-535, 2019.
22. Jeong G, Kwon DH, Shin S, Choe N, Ryu J, Lim YH, Kim J, Park WJ, Kook H and Kim YK: Long noncoding RNAs in vascular smooth muscle cells regulate vascular calcification. *Sci Rep* 9: 5848, 2019.
23. Alves RD, Eijken M, van de Peppel J and van Leeuwen JP: Calcifying vascular smooth muscle cells and osteoblasts: Independent cell types exhibiting extracellular matrix and biomineralization-related mimics. *BMC Genomics* 15: 965, 2014.
24. Karagoz K, Sevimglu T and Arga KY: Integration of multiple biological features yields high confidence human protein interactions. *J Theor Biol* 403: 85-96, 2016.
25. Shannon P, Markiel A, Ozier O, Baliga NS, Wang JT, Ramage D, Amin N, Schwikowski B and Ideker T: Cytoscape: A software environment for integrated models of biomolecular interaction networks. *Genome Res* 13: 2498-2504, 2003.
26. Tong Z, Cui Q, Wang J and Zhou Y: TransmiR v2.0: An updated transcription factor-microRNA regulation database. *Nucleic Acids Res* 47: D253-D258, 2019.
27. Cheng L, Wang P, Tian R, Wang S, Guo Q, Luo M, Zhou W, Liu G, Jiang H and Jiang Q: LncRNA2Target v2.0: A comprehensive database for target genes of lncRNAs in human and mouse. *Nucleic Acids Res* 47: D140-D144, 2019.
28. Livak KJ and Schmittgen TD: Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta C(T)) method. *Method* 25: 402-408, 2001.
29. Ohukainen P, Syväranta S, Näpänkangas J, Rajamäki K, Taskinen P, Peltonen T, Helske-Suihko S, Kovanen PT, Ruskoaho H and Rysä J: MicroRNA-125b and chemokine CCL4 expression are associated with calcific aortic valve disease. *Ann Med* 47: 423-429, 2015.
30. Matsumoto K, Maniwa T, Tanaka T, Satoh K, Okunishi H and Oda T: Proteomic analysis of calcified abdominal and thoracic aortic aneurysms. *Int J Mol Med* 30: 417-429, 2012.
31. Tanaka K, Sata M, Fukuda D, Suematsu Y, Motomura N, Takamoto S, Hirata Y and Nagai R: Age-associated aortic stenosis in apolipoprotein E-deficient mice. *J Am Coll Cardiol* 46: 134-141, 2005.
32. Hodroge A, Trécherel E, Cornu M, Darwiche W, Mansour A, Ait-Mohand K, Verissimo T, Gomila C, Schembri C, Da Nascimento S, *et al.*: Oligogalacturonic acid inhibits vascular calcification by two mechanisms: inhibition of vascular smooth muscle cell osteogenic conversion and interaction with collagen. *Arterioscler Thromb Vasc Biol* 37: 1391-1401, 2017.
33. Shanahan CM, Cary NR, Salisbury JR, Proudfoot D, Weissberg PL and Edmonds ME: Medial localization of mineralization-regulating proteins in association with Mönckeberg's sclerosis. *Circulation* 100: 2168-2176, 1999.
34. Tyson KL, Reynolds JL, McNair R, Zhang Q, Weissberg PL and Shanahan CM: Osteo/chondrocytic transcription factors and their target genes exhibit distinct patterns of expression in human arterial calcification. *Arterioscler Thromb Vasc Biol* 23: 489-494, 2003.
35. Watson KE, Parhami F, Shin V and Demer LL: Fibronectin and collagen I matrices promote calcification of vascular cells in vitro, whereas collagen IV matrix is inhibitory. *Arterioscler Thromb Vasc Biol* 18: 1964-1971, 1998.
36. Freise C, Bobb V and Querfeld U: Collagen XIV and a related recombinant fragment protect human vascular smooth muscle cells from calcium-/phosphate-induced osteochondrocytic trans-differentiation. *Exp Cell Res* 358: 242-252, 2017.
37. Lei Y, Sinha A, Nosoudi N, Grover A and Vyavahare N: Hydroxyapatite and calcified elastin induce osteoblast-like differentiation in rat aortic smooth muscle cells. *Exp Cell Res* 323: 198-208, 2014.
38. Sinha A and Vyavahare NR: High-glucose levels and elastin degradation products accelerate osteogenesis in vascular smooth muscle cells. *Diabet Vasc Dis Res* 10: 410-419, 2013.
39. Urry DW: Neutral sites for calcium ion binding to elastin and collagen: A charge neutralization theory for calcification and its relationship to atherosclerosis. *Proc Natl Acad Sci USA* 68: 810-814, 1971.
40. Wang L, Zheng J, Du Y, Huang Y, Li J, Liu B, Liu CJ, Zhu Y, Gao Y, Xu Q, *et al.*: Cartilage oligomeric matrix protein maintains the contractile phenotype of vascular smooth muscle cells by interacting with alpha(7)beta(1) integrin. *Circ Res* 106: 514-525, 2010.
41. Du Y, Wang Y, Wang L, Liu B, Tian Q, Liu CJ, Zhang T, Xu Q, Zhu Y, Ake O, *et al.*: Cartilage oligomeric matrix protein inhibits vascular smooth muscle calcification by interacting with bone morphogenetic protein-2. *Circ Res* 108: 917-928, 2011.
42. Muller J, Bolomsky A, Dubois S, Duray E, Stangelberger K, Plougonven E, Lejeune M, Léonard A, Marty C, Hempel U, *et al.*: Maternal embryonic leucine zipper kinase inhibitor OTSSP167 has preclinical activity in multiple myeloma bone disease. *Haematologica* 103: 1359-1368, 2018.
43. Kim SH, Joshi K, Ezhilarasan R, Myers TR, Siu J, Gu C, Nakano-Okuno M, Taylor D, Minata M, Sulman EP, *et al.*: EZH2 protects glioma stem cells from radiation-induced cell death in a MELK/FOXMI-dependent manner. *Stem Cell Reports* 4: 226-238, 2015.
44. Fan T, He J, Yin Y, Wen K, Kang Y, Zhao H, Chen S and Li X: Dioscin inhibits intimal hyperplasia in rat carotid artery balloon injury model through inhibition of the MAPK-FoxM1 pathway. *Eur J Pharmacol* 854: 213-223, 2019.
45. Dudakovic A, Camilleri ET, Xu F, Riester SM, McGee-Lawrence ME, Bradley EW, Paradise CR, Lewallen EA, Thaler R, Deyle DR, *et al.*: Epigenetic control of skeletal development by the histone methyltransferase Ezh2. *J Biol Chem* 290: 27604-27617, 2015.
46. Adamik J, Jin S, Sun Q, Zhang P, Weiss KR, Anderson JL, Silbermann R, Roodman GD and Galson DL: EZH2 or HDAC1 inhibition reverses multiple myeloma-induced epigenetic suppression of osteoblast differentiation. *Mol Cancer Res* 15: 405-417, 2017.
47. Zhang H, Pan Y, Zheng L, Choe C, Lindgren B, Jensen ED, Westendorf JJ, Cheng L and Huang H: FOXO1 inhibits Runx2 transcriptional activity and prostate cancer cell migration and invasion. *Cancer Res* 71: 3257-3267, 2011.
48. Sun Y, Byon CH, Yuan K, Chen J, Mao X, Heath JM, Javed A, Zhang K, Anderson PG and Chen Y: Smooth muscle cell-specific Runx2 deficiency inhibits vascular calcification. *Circ Res* 111: 543-552, 2012.
49. Deng L, Huang L, Sun Y, Heath JM, Wu H and Chen Y: Inhibition of FOXO1/3 promotes vascular calcification. *Arterioscler Thromb Vasc Biol* 35: 175-183, 2015.
50. Xu X, Ji S, Li W, Yi B, Li H, Zhang H and Ma W: LncRNA H19 promotes the differentiation of bovine skeletal muscle satellite cells by suppressing Sirt1/FoxO1. *Cell Mol Biol Lett* 22: 10, 2017.
51. Hadji F, Boulanger MC, Guay SP, Gaudreault N, Amellah S, Mkannez G, Bouchareb R, Marchand JT, Nsaibia MJ, Gouaque-Olarte S, *et al.*: Altered DNA methylation of long noncoding RNA H19 in calcific aortic valve disease promotes mineralization by silencing NOTCH1. *Circulation* 134: 1848-1862, 2016.
52. Liang WC, Fu WM, Wang YB, Sun YX, Xu LL, Wong CW, Chan KM, Li G, Waye MM and Zhang JF: H19 activates Wnt signaling and promotes osteoblast differentiation by functioning as a competing endogenous RNA. *Sci Rep* 6: 20121, 2016.
53. Tang Z, Gong Z and Sun X: LncRNA DANCR involved osteolysis after total hip arthroplasty by regulating FOXO1 expression to inhibit osteoblast differentiation. *J Biomed Sci* 25: 4, 2018.
54. Zhu L and Xu PC: Downregulated LncRNA-ANCR promotes osteoblast differentiation by targeting EZH2 and regulating Runx2 expression. *Biochem Biophys Res Commun* 432: 612-617, 2013.
55. Jia Q, Jiang W and Ni L: Down-regulated non-coding RNA (lncRNA-ANCR) promotes osteogenic differentiation of periodontal ligament stem cells. *Arch Oral Biol* 60: 234-241, 2015.
56. Masy ZA, Metzinger-Le Meuth V and Metzinger L: MicroRNAs are associated with uremic toxicity, cardiovascular calcification, and disease. *Contrib Nephrol* 189: 160-168, 2017.
57. Lambertini E, Lisignoli G, Torreggiani E, Manferdini C, Gabusi E, Franceschetti T, Penolazzi L, Gambari R, Facchini A and Piva R: Slug gene expression supports human osteoblast maturation. *Cell Mol Life Sci* 66: 3641-3653, 2009.
58. Piva R, Manferdini C, Lambertini E, Torreggiani E, Penolazzi L, Gambari R, Pastore A, Pelucchi S, Gabusi E, Piacentini A, *et al.*: Slug contributes to the regulation of CXCL12 expression in human osteoblasts. *Exp Cell Res* 317: 1159-1168, 2011.
59. Sanchez-Duffhues G, de Vinuesa AG, Lindeman JH, Mulder-Stapel A, DeRuiter MC, Van Munsteren C, Goumans MJ, Hierck BP and Ten Dijke P: SLUG is expressed in endothelial cells lacking primary cilia to promote cellular calcification. *Arterioscler Thromb Vasc Biol* 35: 616-627, 2015.

