

lncRNA ZFAS1 promotes the ox-LDL induced proliferation, invasion and migration of vascular smooth muscle cells

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Abstract. Atherosclerosis is a chronic progressive inflammatory vascular disease. The dysfunction of vascular smooth muscle cells (VSMCs) induced by oxidized low-density lipoprotein (ox-LDL) contributes to the formation of atherosclerotic lesions. Additionally, upregulation of the long non-coding RNA zinc finger antisense 1 (ZFAS1) was observed in the plaques of patients with atherosclerosis. The aim of the present study was to explore the functional role of ZFAS1 in atherosclerosis progression. Reverse transcription-quantitative PCR was performed to analyze ZFAS1 mRNA expression, and western blotting was performed to determine the protein expression levels of Ki67, proliferating cell nuclear antigen (PCNA), matrix metalloproteinase (MMP)2 and MMP9. The Cell Counting Kit-8 assay was used to test cell viability. Finally, wound healing and Transwell chamber assays were performed to evaluate cell migration and invasion, respectively. The current findings demonstrated that ZFAS1 expression was upregulated by ox-LDL stimulation in VSMCs. Moreover, ZFAS1 overexpression promoted the ox-LDL-induced proliferation, migration and invasion of VSMCs, and upregulated the expression levels of proteins associated with cellular proliferation (Ki67 and PCNA), migration and invasion (MMP2 and 9). By contrast, ZFAS1-knockdown inhibited the proliferation, migration and invasion of VSMCs, and suppressed cell proliferation-, migration- and invasion-associated protein expression. In conclusion, ZFAS1 promoted the ox-LDL-induced proliferation, invasion and

migration of VSMCs. Thus, ZFAS1 may represent a novel biomarker for dysfunction of VSMCs in the pathological condition of atherosclerosis.

Introduction

Atherosclerosis is a chronic progressive inflammatory vascular disease, which poses a major threat to human health and has been attracting increasing attention (1). Thrombotic events associated with acute rupture or erosion of an unstable plaque, rather than gradual narrowing of the lumen, have been shown to be responsible for the majority of the clinical consequences of atherosclerosis (2). It is well established that vascular smooth muscle cells (VSMCs) serve an important role in the pathogenesis of atherosclerosis (2,3). VSMCs, as one of the key components of the plaques, are derived from the medial layer of the vessel wall, which acts as a regulator of the atherosclerotic plaque (4-6). Furthermore, excess proliferation or dysfunction of VSMCs contributes to atherogenesis as a response to vascular injury, inflammation and lipoprotein accumulation during disease progression (1,7). Oxidized low-density lipoprotein (ox-LDL) contributes to the atherosclerotic lesion through several mechanisms, such as the dysfunction of endothelial cells, and the excess migration and proliferation of VSMCs, as well as contributing to plaque instability (8). Therefore, in the present study VSMCs were treated with ox-LDL and used as the cell model for atherosclerosis in order to investigate the specific mechanisms underlying the pathogenesis of atherosclerosis.

Previous studies have revealed that long non-coding RNAs (lncRNAs), a class of non-coding RNAs >200 nucleotides in length, serve an important role in the onset and development of multiple human diseases, including cancer, diabetes, inflammatory diseases and cardiovascular diseases (9-11). The expression levels of the lncRNA zinc finger antisense 1 (ZFAS1) were revealed to be upregulated in the plaques of patients with atherosclerosis compared with in controls, as well as in atherosclerosis rat models (12,13), indicating that ZFAS1 is closely associated with the progression of atherosclerosis. A recent study reported that ZFAS1 upregulation was observed in the cytoplasm and sarcoplasmic reticulum of mouse

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cardiomyocytes challenged with hypoxic stimulation, and that it impaired cardiac function in a mouse model of acute myocardial infarction, and these effects were readily reversed by ZFAS1-knockdown (14). Furthermore, lncRNA ZFAS1 promotes the proliferation, invasion and migration of various cancer cells, including nasopharyngeal carcinoma (15), cervical carcinoma (16) and colorectal cancer (17). Additionally, ZFAS1 may promote the proliferation and migration of chondrocytes, and suppress apoptosis and matrix synthesis in osteoarthritis (18). Overall, the aforementioned findings indicate that ZFAS1 serves a key role in promoting cell proliferation and invasion. Thus, it was hypothesized that ZFAS1 may serve as a potential biomarker in atherosclerosis induced by ox-LDL, and may promote the proliferation and invasion of VSMCs under pathological conditions.

In the present study, VSMCs were treated with multiple doses of ox-LDL to induce a cell model of atherosclerosis, in order to investigate whether ZFAS1 expression was upregulated by ox-LDL treatment in a dose-dependent manner and to determine whether ZFAS1 may be of value as a novel biomarker for dysfunction of VSMCs in the pathological condition of atherosclerosis.

Materials and methods

Cell culture and transfection. The VSMC cell line was obtained from the China Infrastructure of Cell Line Resources, Institute of Basic Medical Sciences (Chinese Academy of Sciences). The cells were cultured in DMEM (HyClone; Cytiva) with 10% fetal bovine serum (FBS; Gibco; Thermo Fisher Scientific, Inc.), 1% penicillin (100 U/ml) and 1% streptomycin (100 mg/ml) (Beyotime Institute of Biotechnology) at 37°C in a humidified incubator with 5% CO₂. VSMCs were cultured in 6-well plates for 12 h, and then transfected with indicated plasmids. ZFAS1-1 small interfering (si)RNA (5'-CTGGCTGAACCAGTTCACAAGGTT-3'), ZFAS1-2 siRNA (5'-TAC TTCTCCTAGTTGCAGTCAGG-3') and the scramble negative control siRNA (si-NC; 5'-ACGTGACACGTTTCGGAGAATT-3') were obtained from Shanghai GenePharma Co., Ltd., and 100 nM of each siRNA was transfected into VSMCs for ZFAS1-knockdown using Lipofectamine® 2000 (Invitrogen; Thermo Fisher Scientific, Inc.) following the manufacturer's instructions. In addition, ZFAS1 transcript cDNA (5'-UGCGUGCCAAGCGCGACAUGGCGCGGAAGCCGAGAAGCCCCGGAGGCC-3') was inserted into the pCDNA3.1 vector [Jieman Biotechnology (Shanghai) Co., Ltd.] and constructed by Biotech Integrated Solutions, and then 2 mg/l ZFAS1 pcDNA3.1 or empty pcDNA3.1 was transfected into VSMCs to achieve ZFAS1 overexpression (oe-ZFAS1) using Lipofectamine® 2000. After transfection for 8 h at 37°C, the VSMCs were exposed to ox-LDL (25, 50 and 100 mg/l; Beijing Solarbio Science & Technology Co., Ltd.) for 12, 24 and 48 h at 37°C.

RNA isolation and reverse transcription-quantitative (RT-q) PCR. Total RNA from VSMCs was isolated using TRIzol® reagent (Invitrogen; Thermo Fisher Scientific, Inc.) following the manufacturer's instructions. RNA (1 µg) was reverse transcribed into first-strand cDNA using the Reverse Transcriptase kit according to the manufacturer's protocol (TransGen Biotech Co., Ltd.).

qPCR was performed using SYBR Green Mixture (Takara Bio, Inc.) in the ABI Prism 7300 Sequence Detection System (Applied Biosystems; Thermo Fisher Scientific, Inc.). The thermocycling conditions were: Initial denaturation for 10 min at 94°C, followed by 40 of cycles of denaturation for 30 sec at 94°C, annealing for 30 sec at 55°C and extension for 30 sec at 72°C. The 2^{-ΔΔC_q} method (19) was applied to determine the relative target gene expression. The sequences for the qPCR primers were as follows: ZFAS1 forward, 5'-AGCGTTTGCTTT GTTCCC-3' and reverse, 5'-CTCCCTCGATGCCCTTCT-3'; GAPDH forward, 5'-GGTCTCCTCTGACTTCAACA-3' and reverse, 5'-AGCCAAATTTCGTTGTCATAC-3'. GAPDH was employed as an internal control.

Cell Counting Kit-8 (CCK-8) assay. The cell viability of VSMCs was determined using the CCK-8 assay (Dojindo Molecular Technologies, Inc.). Briefly, VSMCs transfected with or without oe-ZFAS1 or si-ZFAS1 were seeded at the density of 2x10³ cells/well into 96-well plates and treated with ox-LDL. After transfection for 12, 24 and 48 h, CCK-8 reagent (10 µl) was added into each well and incubated with VSMCs for another 2 h. The absorbance at 450 nm was measured using an ELISA plate reader (Bio-Rad Laboratories, Inc.).

Western blotting. After treatment, proteins were isolated from VSMCs transfected with or without oe-ZFAS1 or si-ZFAS1 using RIPA lysis buffer (Beyotime Institute of Biotechnology) and quantified using a BCA assay kit (Thermo Fisher Scientific, Inc.). Protein samples (25 µg/lane) were loaded and separated via 10% SDS-PAGE and transferred onto PVDF membranes (EMD Millipore). After blocking with 5% skimmed milk for 1 h at room temperature, the membrane was incubated with primary antibodies against Ki67 (cat. no. ab21700; 1:1,000; Abcam), proliferating cell nuclear antigen [PCNA; cat. no. 13110; 1:1,000; Cell Signaling Technology, Inc. (CST)], matrix metalloproteinase (MMP)2 (cat. no. 4022; 1:1,000; CST), MMP9 (cat. no. 3852; 1:1,000; CST) and GAPDH (cat. no. 8884; 1:2,000; CST) overnight at 4°C. After washing with PBS, the membrane was incubated with horseradish peroxidase-conjugated secondary antibody (cat. no. ab97080; 1:10,000; Abcam) for 2 h at room temperature. ECL Reagent was used to develop color of protein bands (SuperSignal West Atto; Thermo Fisher Scientific, Inc.). The gray values of the protein bands were determined using ImageJ software (version 1.48; National Institutes of Health).

Wound healing assay. Wound healing assay was performed to assess the migration of VSMCs. Cells were plated in 6-well plates to generate a confluent monolayer. A scratch was created using a 200-µl sterile pipette tip, followed by washing with PBS three times. Subsequently, VSMCs were cultured with fresh serum-free DMEM for 24 h at 37°C. Finally, the width of the scratch wound was observed, and images were captured at x100 magnification using a fluorescence microscope (Olympus IX53; Olympus Corporation). The recovered wound area (%) at the indicated time point (24 h) was calculated according to the following formula: [(wound width at 0 h) - (wound width at 24 h)] / wound width at 0 h.

Transwell chamber assay. The Transwell chamber assay was used to determine the invasive ability of VSMCs. Briefly,

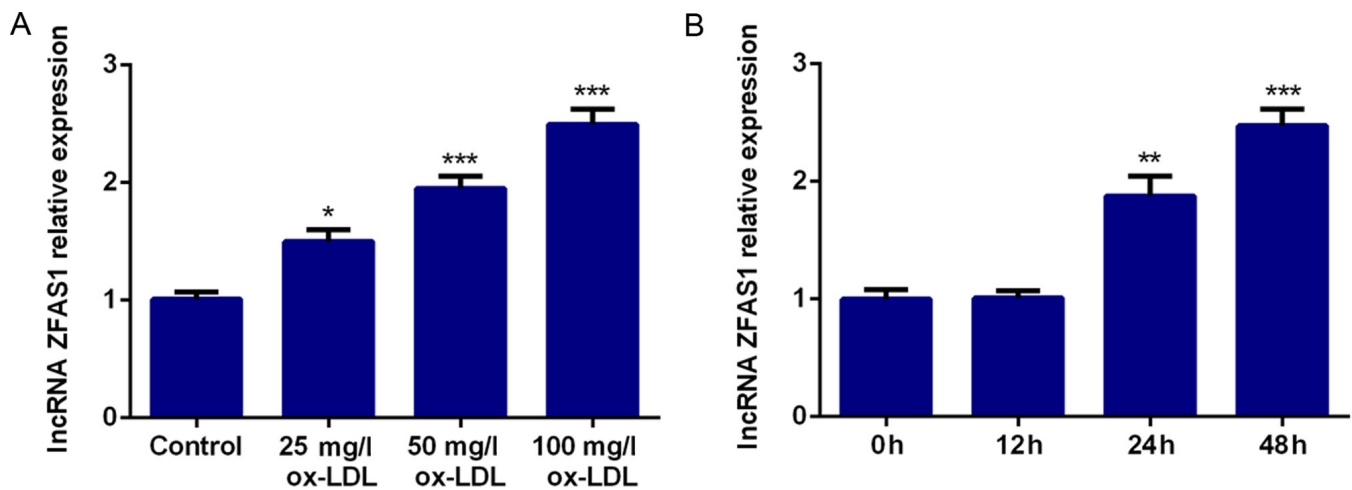


Figure 1. ZFAS1 expression is upregulated by ox-LDL treatment in VSMCs. Relative ZFAS1 mRNA expression was quantified via reverse transcription-quantitative PCR in VSMCs treated with (A) different concentrations of ox-LDL and (B) 100 mg/l ox-LDL for 12, 24 or 48 h. The data are expressed as the mean \pm SEM from three independent experiments. * $P<0.05$, ** $P<0.01$ and *** $P<0.001$ vs. Control or 0 h. ZFAS1, zinc finger antisense 1; ox-LDL, oxidized low-density lipoprotein; VSMCs, vascular smooth muscle cells.

after transfection with or without oe-ZFAS1 or si-ZFAS1, the VSMCs were resuspended in 200 μ l serum-free DMEM, and 4×10^4 cells were loaded into the upper chambers of Transwell plates precoated with Matrigel Mix for 5 h at 37°C (BD Biosciences), and the lower chambers were filled with 500 μ l DMEM with 10% FBS as a chemoattractant. After incubation for 24 h at 37°C, the membrane was fixed with 4% paraformaldehyde for 25 min at room temperature, and the cells on the lower surface of the membrane were stained with 0.1% crystal violet solution for 30 min at room temperature, and finally examined under a fluorescence microscope at $\times 100$ magnification (Olympus IX53; Olympus Corporation). The invasion rate was calculated according to the following formula: Number of cells in tested group / number of cells in control group.

Statistical analysis. Statistical analyses were performed using SPSS 19.0 (IBM Corp.), and data are presented as the mean \pm SEM from at least three repeated experiments. Differences between multiple groups were analyzed by one-way ANOVA followed by Tukey's post hoc test. $P<0.05$ was considered to indicate a statistically significant difference.

Results

ZFAS1 expression is upregulated by ox-LDL treatment in VSMCs. To verify the role of ZFAS1 in atherosclerosis, VSMCs were incubated with ox-LDL to simulate the high blood lipid environment, and ZFAS1 mRNA expression was detected by RT-qPCR. The results revealed that ZFAS1 mRNA expression was significantly higher with increasing doses of ox-LDL compared with the control group, and the increase was dose-dependent (Fig. 1A). Hence, 100 mg/l ox-LDL was selected for further experimentation. Moreover, VSMCs were stimulated with ox-LDL (100 mg/l) for 12, 24 and 48 h. The results of RT-qPCR revealed that ZFAS1 mRNA expression was significantly increased by

ox-LDL stimulation at 24 and 48 h in a time-dependent manner (Fig. 1B). Thus, VSMCs treated with 100 mg/l ox-LDL for 48 h were selected for subsequent experiments.

ZFAS1-knockdown inhibits the ox-LDL-induced excessive proliferation of VSMCs. To explore the effect of ZFAS1 expression on cellular behaviors, ZFAS1 expression was knocked down, and the proliferation of VSMCs was analyzed. First, si-ZFAS1 was used to achieve ZFAS1-knockdown. The RT-qPCR results demonstrated that ZFAS1 mRNA expression in the si-ZFAS1-1 and si-ZFAS1-2 groups was significantly lower compared with that in the control group, particularly in the si-ZFAS1-2 group (Fig. 2A). Hence, si-ZFAS1-2 was selected for further experiments. The CCK-8 assay results indicated that ox-LDL treatment significantly increased the viability of VSMCs at 24 and 48 h compared with the control group, while ZFAS1-knockdown significantly decreased VSMC viability when compared with the ox-LDL group (Fig. 2B). Finally, the expression levels of proteins associated with cell proliferation was quantified by western blotting. PCNA and Ki67 were identified as proliferation markers. As shown in Fig. 2C, ox-LDL stimulation caused significant upregulation of Ki67 and PCNA expression. Notably, ZFAS1-knockdown in ox-LDL-treated cells led to a significant decrease in Ki67 and PCNA expression (Fig. 2C). Hence, these results indicated that ZFAS1-knockdown inhibited the ox-LDL-induced excessive proliferation of VSMCs.

ZFAS1 overexpression promotes the proliferation of VSMCs stimulated by ox-LDL. To explore the effect of ZFAS1 expression on cell proliferation, VSMC proliferation was determined following ZFAS1 overexpression. The RT-qPCR results revealed that ZFAS1 mRNA expression in VSMCs was significantly increased by oe-ZFAS1 transfection (Fig. 3A). The CCK-8 assay results revealed that ZFAS1 overexpression significantly increased the viability of VSMCs induced by ox-LDL (Fig. 3B). Finally, the western blotting results demonstrated that ZFAS1 overexpression significantly upregulated Ki67 and PCNA expression in ox-LDL-induced VSMCs

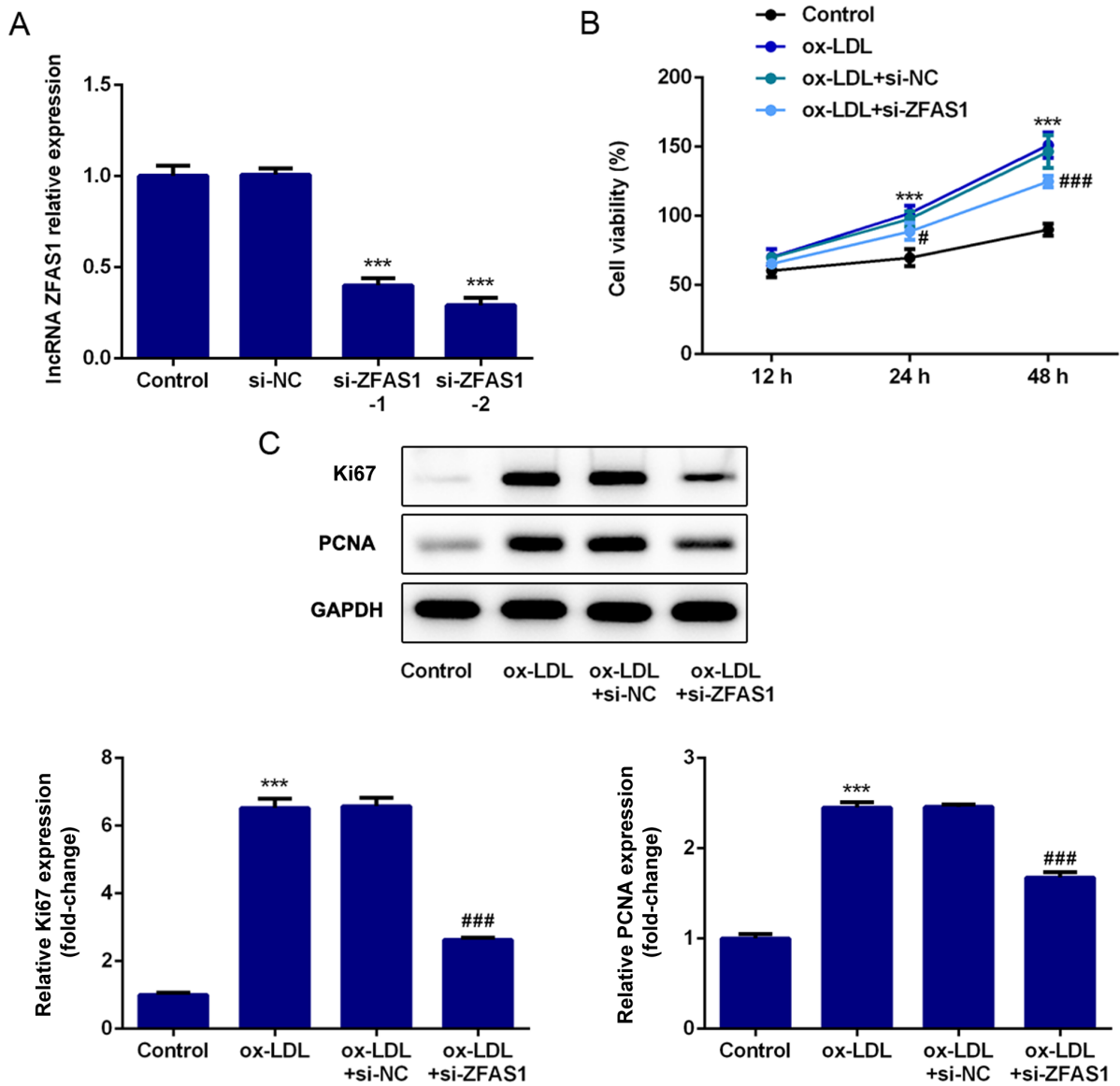


Figure 2. ZFAS1-knockdown inhibits the ox-LDL-induced excessive proliferation of VSMCs. (A) Relative mRNA ZFAS1 expression was quantified via reverse transcription-quantitative PCR following ZFAS1-knockdown. (B) Viability of VSMCs was examined using the Cell Counting Kit-8 assay. (C) Protein expression levels of Ki67 and PCNA were determined by western blotting. The data are expressed as the mean \pm SEM from three independent experiments. *** P <0.001 vs. Control; # P <0.05 and ### P <0.001 vs. ox-LDL + si-NC. ZFAS1, zinc finger antisense 1; ox-LDL, oxidized low-density lipoprotein; VSMCs, vascular smooth muscle cells; PCNA, proliferating cell nuclear antigen; si, small interfering RNA; NC, negative control.

(Fig. 3C). Thus, ZFAS1 overexpression promoted the proliferation of VSMCs stimulated by ox-LDL.

ZFAS1-knockdown suppresses the excessive migration and invasion of VSMCs induced by ox-LDL. To investigate the effect of ZFAS1 expression on cellular behavior, ZFAS1 expression was inhibited, and the migration and invasion of VSMCs were analyzed. The results of the wound healing assay revealed that ox-LDL treatment significantly promoted the migration of VSMCs, while ZFAS1-knockdown significantly suppressed the migration of ox-LDL-induced VSMCs (Fig. 4A and C). Furthermore, the results of the Transwell chamber assay demonstrated that ox-LDL stimulation significantly promoted the invasion of VSMCs, whereas ZFAS1-knockdown significantly inhibited the invasion of ox-LDL-treated VSMCs (Fig. 4B and D). Additionally,

the expression levels of proteins associated with migration and invasion were quantified. MMP2 and MMP9 are involved in cell migration and invasion. The western blotting results revealed that the expression levels of MMP2 and MMP9 in the ox-LDL and ox-LDL + si-NC groups were significantly higher compared with that in the control group (Fig. 5). Notably, ZFAS1-knockdown significantly inhibited the expression levels of MMP2 and MMP9 following ox-LDL stimulation (Fig. 5). These results indicated that ZFAS1-knockdown suppressed the excessive migration and invasion of VSMCs induced by ox-LDL.

ZFAS1 overexpression promotes the migration and invasion of VSMCs induced by ox-LDL. To investigate the effect of ZFAS1 expression on cell migration and invasion, VSMC migration and invasion were determined following ZFAS1

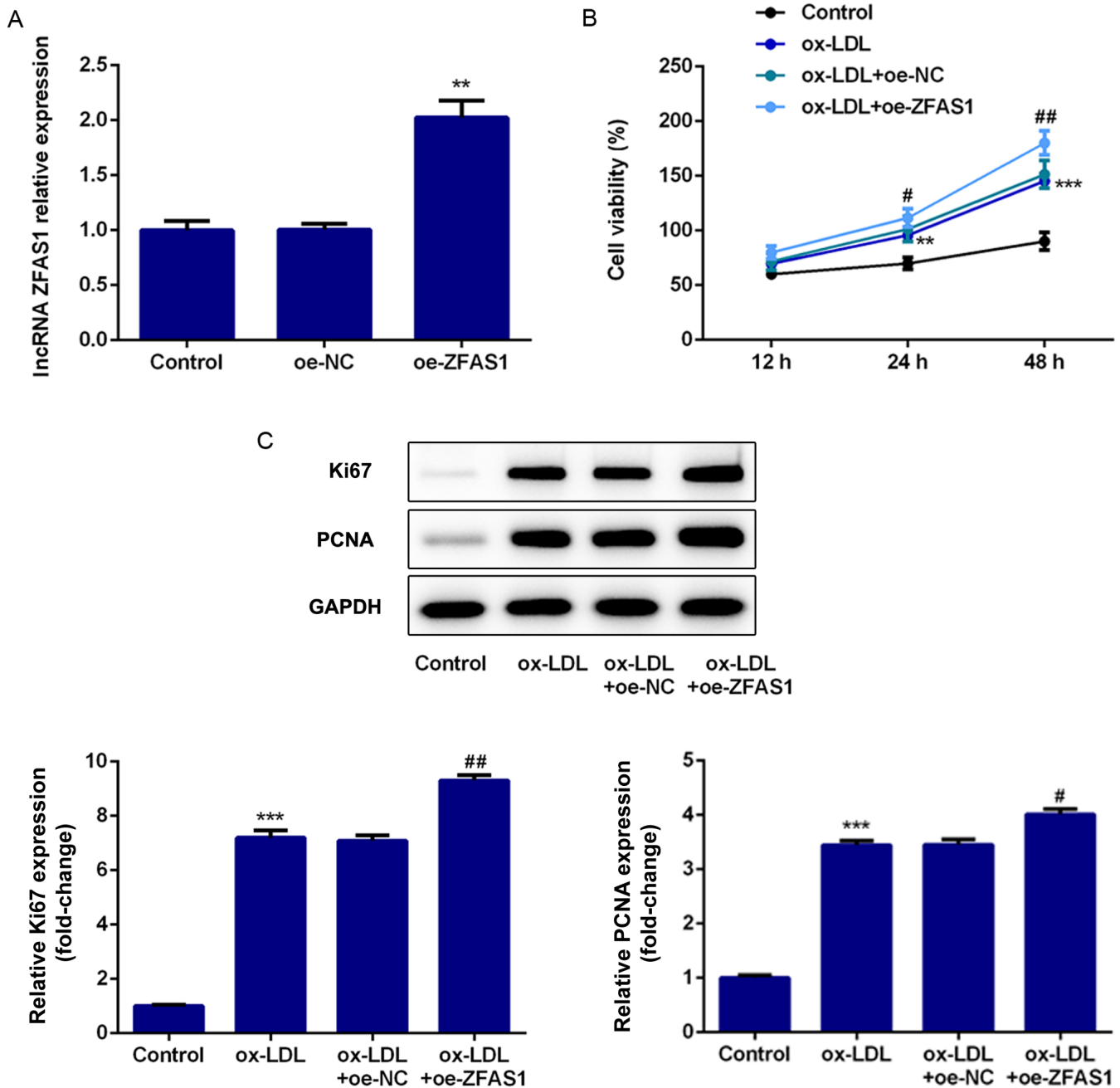


Figure 3. ZFAS1 overexpression promotes the proliferation of VSMCs stimulated by ox-LDL. (A) Relative mRNA ZFAS1 expression was quantified via reverse transcription-quantitative PCR following ZGAS1 overexpression. (B) Viability of VSMCs was examined using the Cell Counting Kit-8 assay. (C) Protein expression levels of Ki67 and PCNA were determined by western blotting. The data are expressed as the mean \pm SEM from three independent experiments. ** $P < 0.01$ and *** $P < 0.001$ vs. Control; # $P < 0.05$ and ## $P < 0.01$ vs. ox-LDL + oe-NC. ZFAS1, zinc finger antisense 1; ox-LDL, oxidized low-density lipoprotein; VSMCs, vascular smooth muscle cells; PCNA, proliferating cell nuclear antigen; oe, overexpression; NC, negative control.

overexpression. As shown in Fig. 6, ZFAS1 overexpression significantly promoted the migration and invasion of ox-LDL-induced VSMCs. Moreover, ZFAS1 overexpression significantly increased MMP2 and MMP9 expression (Fig. 7). These results suggested that ZFAS1 overexpression promoted the migration and invasion of VSMCs induced by ox-LDL.

Discussion

Atherosclerosis is characterized by endothelium dysfunction, accumulation of ox-LDL and intimal hyperplasia (20).

A previous study has reported that the excessive proliferation and migration of VSMCs in response to vascular injury, inflammation and lipoprotein accumulation mainly leads to the initiation of intimal hyperplasia (21). Hence, VSMCs were treated with ox-LDL to simulate the high blood lipid environment in the present study, which focused on the functional role of ZFAS1 in the behavior of ox-LDL-induced VSMCs. The current findings demonstrated that ox-LDL treatment significantly increased ZFAS1 mRNA expression in VSMCs in a dose- and time-dependent manner, suggesting that ZFAS1 expression is closely associated with the pathogenesis

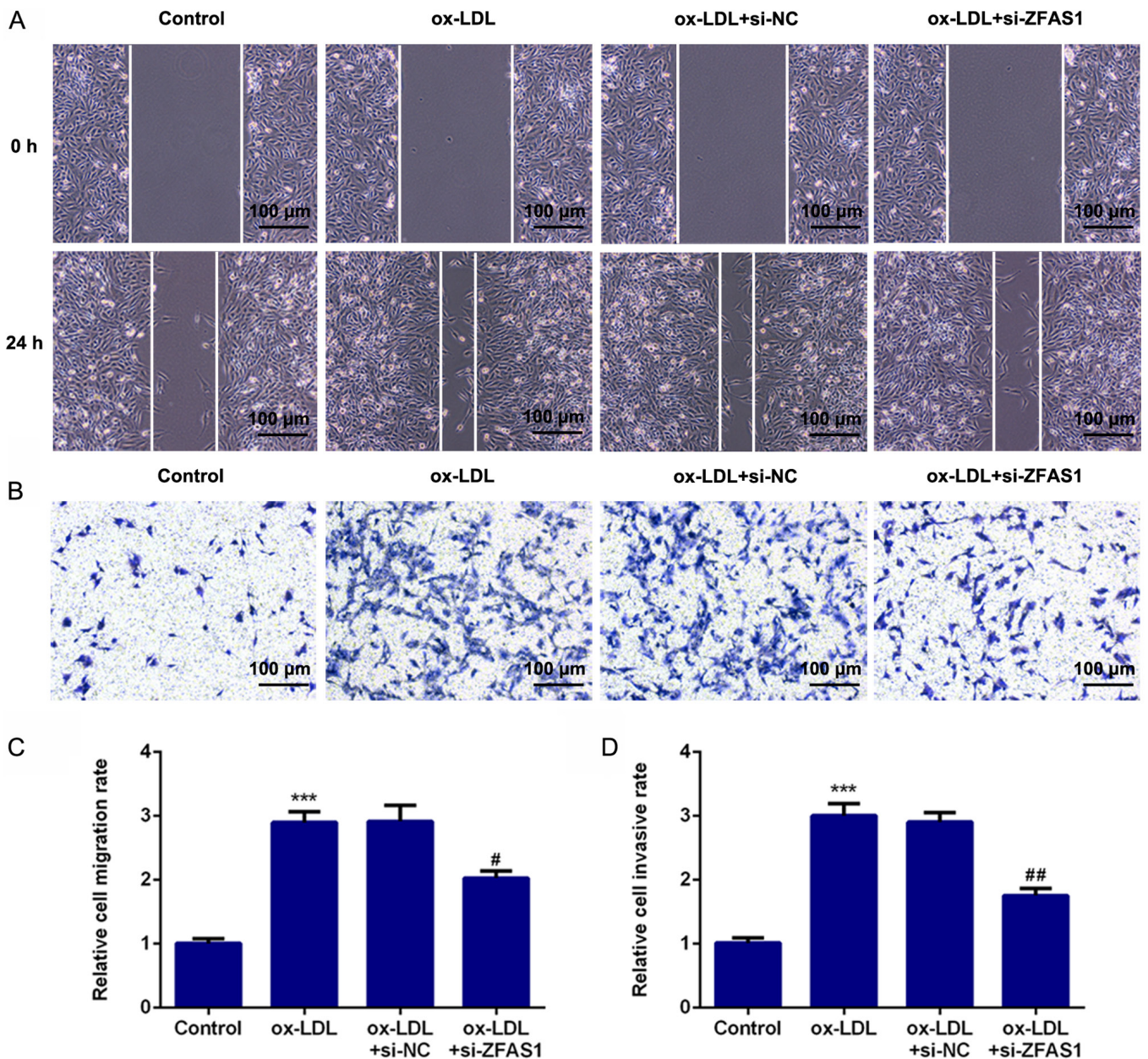


Figure 4. ZFAS1-knockdown suppresses the excessive migration and invasion of VSMCs induced by ox-LDL. (A) Migration of VSMCs was determined by wound healing assay (scale bar, 100 μ m). (B) Invasion of VSMCs was determined by Transwell chamber assay (scale bar, 100 μ m). (C) Quantification of cell migration. (D) Quantification of cell invasion. The data are expressed as the mean \pm SEM from three independent experiments. *** P <0.001 vs. Control; # P <0.05 and ## P <0.01 vs. ox-LDL + si-NC. ZFAS1, zinc finger antisense 1; ox-LDL, oxidized low-density lipoprotein; VSMCs, vascular smooth muscle cells; si, small interfering RNA; NC, negative control.

of atherosclerosis. The aim of the present study was to verify the effect of ZFAS1 on the behavior of ox-LDL-induced VSMCs.

Multiple lncRNAs have recently emerged as regulators of different processes in cardiovascular diseases (22-24). ZFAS1 has been reported as a key lncRNA in atherosclerosis (12). Moreover, ZFAS1 attenuates the rate of cholesterol efflux and facilitates inflammatory responses in atherosclerosis (25). Additionally, ZFAS1 has been found to be dysregulated in various types of cancer and to serve an oncogenic role in the onset and development of malignant tumors, such as ovarian cancer and glioma, by promoting cancer metastasis, growth and epithelial-to-mesenchymal transition (26-28). However,

the functional effect of ZFAS1 on cell migration and the proliferation of VSMCs has yet to be fully elucidated. PCNA and Ki67 are identified as cell proliferation markers (29,30). Moreover, it has been reported that MMP2 and MMP9 are associated with cancer metastasis, angiogenesis and invasion (31,32). In the present study, it was observed that ZFAS1 overexpression promoted the ox-LDL-induced proliferation, migration and invasion of VSMCs, and upregulated the expression levels of proteins associated with cell proliferation, migration and invasion (Ki67, PCNA, MMP2 and MMP9). Notably, ZFAS1-knockdown partly reversed the effect of ox-LDL treatment on the proliferation, migration and invasion of VSMCs, and the expression levels of proteins

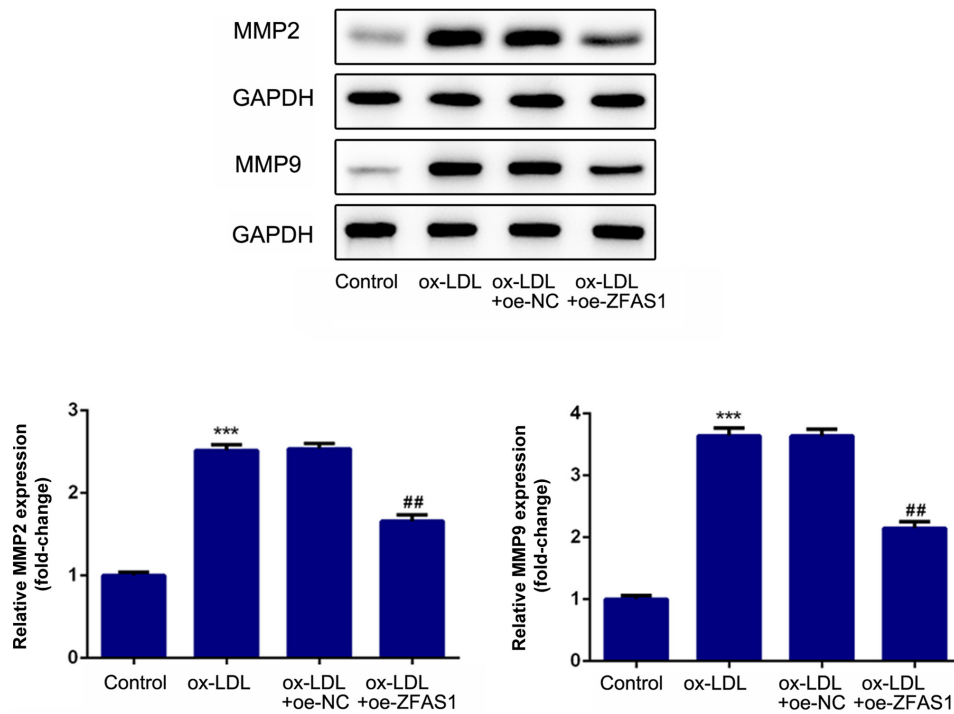


Figure 5. ZFAS1-knockdown suppresses the expression levels of proteins associated with migration and invasion in vascular smooth muscle cells induced by ox-LDL. Protein expression levels of MMP2 and MMP9 were detected by western blotting. The data are expressed as the mean \pm SEM from three independent experiments. *** P <0.001 vs. Control; ** P <0.01 vs. ox-LDL + si-NC. ZFAS1, zinc finger antisense 1; ox-LDL, oxidized low-density lipoprotein; MMP, matrix metalloproteinase; si, small interfering RNA; NC, negative control.

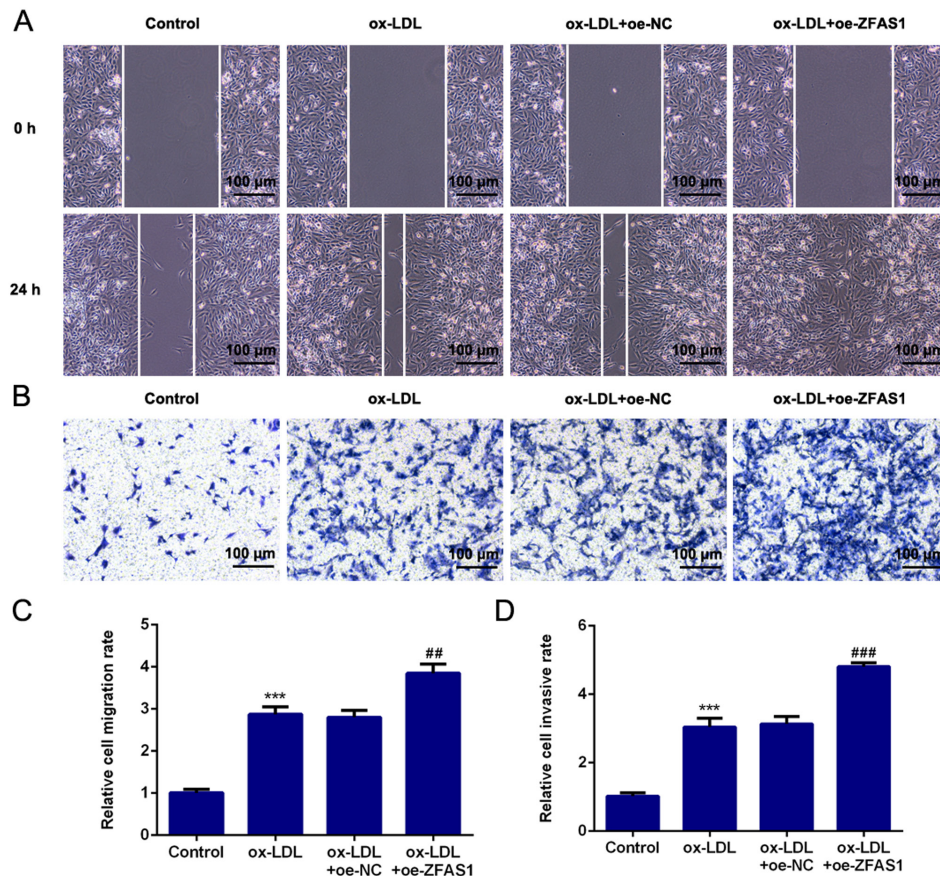


Figure 6. ZFAS1 overexpression promotes the migration and invasion of VSMCs induced by ox-LDL. (A) Migration of VSMCs was determined by the wound healing assay (scale bar, 100 μ m). (B) Invasion of VSMCs was determined by the Transwell chamber assay (scale bar, 100 μ m). (C) Quantification of cell migration. (D) Quantification of cell invasion. The data are expressed as the mean \pm SEM from three independent experiments. *** P <0.001 vs. Control; ** P <0.01 and *** P <0.001 vs. ox-LDL + oe-NC. ZFAS1, zinc finger antisense 1; ox-LDL, oxidized low-density lipoprotein; VSMCs, vascular smooth muscle cells; oe, overexpression; NC, negative control.

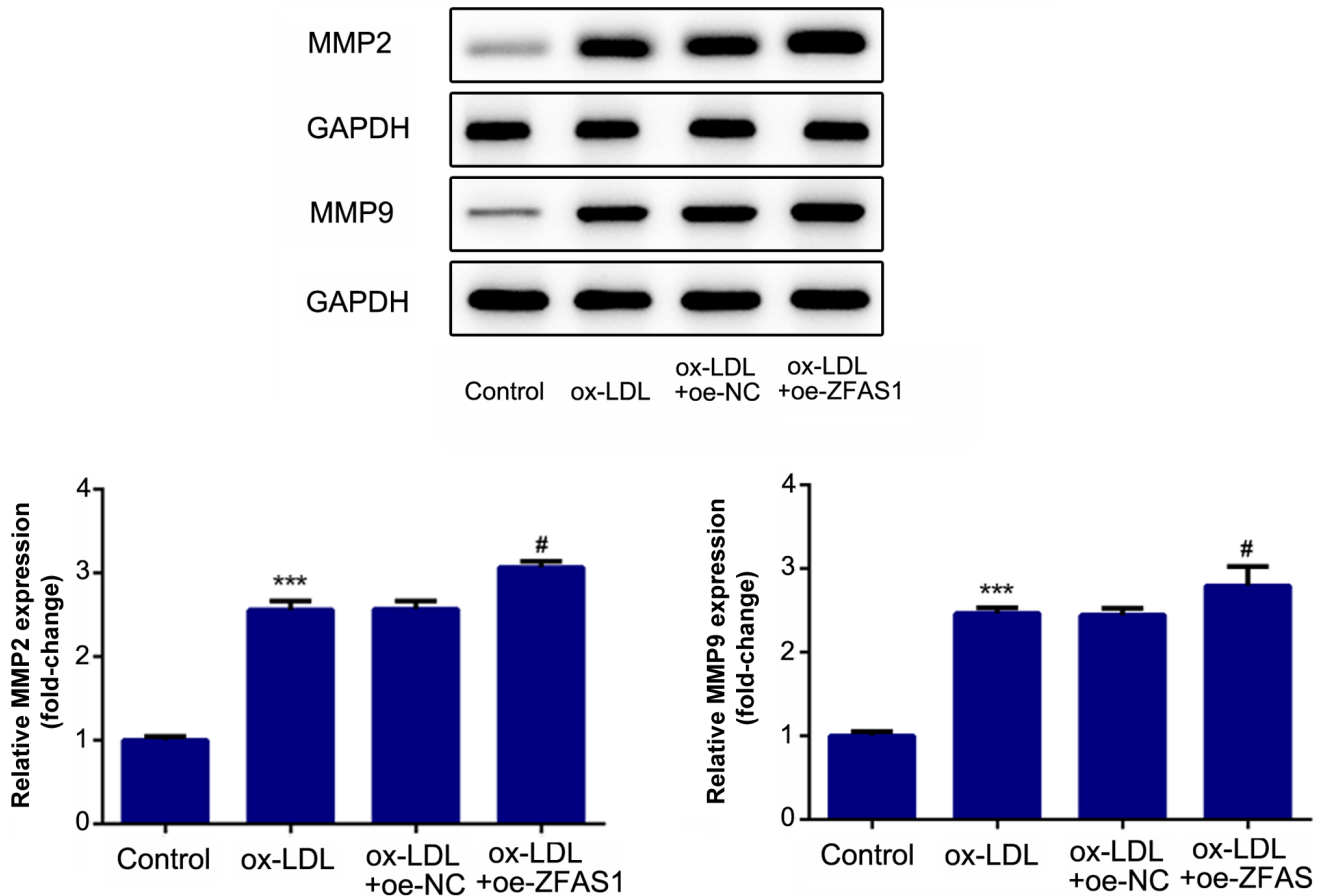


Figure 7. ZFAS1-knockdown increases the expression levels of proteins associated with migration and invasion in vascular smooth muscle cells induced by ox-LDL. Protein expression levels of MMP2 and MMP9 were detected by western blotting. The data are expressed as the mean \pm SEM from three independent experiments. *** $P < 0.001$ vs. Control; # $P < 0.05$ vs. ox-LDL + oe-NC. ZFAS1, zinc finger antisense 1; ox-LDL, oxidized low-density lipoprotein; MMP, matrix metalloproteinase; oe, overexpression; NC, negative control.

associated with cell proliferation, migration and invasion. Previous studies have demonstrated that the phenotype of VSMCs can be switched from contractile to proliferative and migratory during the process of atherosclerosis (33,34). Furthermore, inhibiting cell proliferation, migration and invasion may serve as an effective therapeutic strategy for preventing cardiovascular disease (35). Thus, the present findings indicated that ZFAS1 may serve an important regulatory role in the phenotypic transition of ox-LDL-induced VSMCs. Overall, the aforementioned findings indicated that ZFAS1 promoted the ox-LDL-induced proliferation, invasion and migration of VSMCs, and may serve as a potential biomarker for the dysfunction of VSMCs in the pathological condition of atherosclerosis. However, there are some limitations in the present study, since the conclusion was only from the results of *in vitro* experiments. It is necessary to further investigate the role of ZFAS1 *in vivo* to confirm the findings of the present study. Hence, further studies should be performed to confirm whether ZFAS1 may be used as a biomarker and therapeutic target of atherosclerosis.

In conclusion, the findings of the present study suggested that ZFAS1 expression was upregulated by ox-LDL stimulation in VSMCs. Moreover, ZFAS1 promoted the ox-LDL-induced proliferation, invasion and migration of VSMCs, as well as the expression levels of Ki67, PCNA,

MMP2 and MMP9, and may represent a novel biomarker for dysfunction of VSMCs in the pathological condition of atherosclerosis.

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

HW, HH, JM, YJ and RC conceived and designed the study, collected, analyzed and interpreted the data, and revised the manuscript. HW wrote the manuscript. HW and RC confirmed the authenticity of the raw data. All authors read and approved the final manuscript.

Ethics approval and consent to participate

Not applicable.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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