

Knockdown of DLGAP5 suppresses cell proliferation, induces G₂/M phase arrest and apoptosis in ovarian cancer

HUIJUN ZHANG^{1*}, YUAN LIU^{2*}, SHENGCHUN TANG³, XIAOMIN QIN¹,
LIN LI¹, JINTING ZHOU¹, JING ZHANG¹ and BO LIU²

Departments of ¹Obstetrics and Gynecology and ²Oncology, Xiangyang Central Hospital, Affiliated Hospital of Hubei University of Arts and Science, Xiangyang, Hubei 441000; ³Department of Physical Examination, Xiangyang City Center for Disease Control and Prevention, Xiangyang, Hubei 441021, P.R. China

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Abstract. Discs large-associated protein 5 (DLGAP5) is a microtubule-associated protein and is reported to exert oncogenic role in tumorigenesis, including lung cancer and hepatocellular carcinoma. However, the prognostic value and biological function of DLGAP5 in ovarian cancer (OC) still remain unclear. The present study investigated the expression pattern of DLGAP5 by searching the Oncomine microarray database. The correlation between DLGAP5 and survival prognosis of OC patients was analyzed by the online tool KM-plotter. Knockdown of DLGAP5 was achieved by transfection with small interfering RNA targeting DLGAP5 in two OC cell lines (SKOV3 and CAOV3). Cell proliferation was assessed by Cell Counting Kit-8 assay and colony-formation assay. Flow cytometry was utilized to determine the effects of DLGAP5 on cell cycle distribution and apoptosis. The present study data showed that DLGAP5 was significantly upregulated in OC and its higher expression was associated with poor survival prognosis. Knockdown of DLGAP5 significantly suppressed cell proliferation, induced cell cycle G₂/M phase arrest and apoptosis. Western blot analysis further demonstrated that DLGAP5 knockdown downregulated the expression of CDK1, Cyclin B1 and Bcl-2, but upregulated Bax expression. Collectively, these data demonstrate that

DLGAP5 might be a promising prognostic therapeutic target for OC treatment.

Introduction

Ovarian cancer (OC) is one of the leading causes of tumor-associated deaths and most common gynecological malignancies (1). Current standard treatments, including surgery and adjuvant chemotherapy or radiotherapy, have made significant improvements in diagnosing and treating OC (2). However, the prognosis is still unfavorable, especially for patients with OC at advanced stage when first diagnosed. It has been widely accepted that OC pathogenesis is accompanied by malignant uncontrolled proliferation, extensive invasion and lymphatic metastasis, which is regulated by aberrantly expressed genes (3,4). Consequently, there is a great need for research to elucidate the molecular mechanisms associated with OC pathogenesis to help improve the patient outcomes.

The family of discs large-associated protein (DLGAP) composed of five members (DLGAP1-5) were originally found in rats, which have three key domains, including a 14-amino-acid repeat domain (5), a dynein light chain domain (6) and a guanylate kinase-associated protein homology domain (7). DLGAP1-4 have been reported to participate in a variety of neurological disorders, such as cerebellar ataxia, obsessive compulsive disorder and autism spectrum disease (8-10). In the present study, DLGAP5 (also named as HURP or KIAA0008) is of great interest for it is mainly associated with to various types of cancer (11-13). For instance, DLGAP5 was identified as a cycle cycle-related gene associated with long-term *in vitro* proliferation in patients with acute myeloid leukemia (14). Bioinformatics analysis by different investigators indicated that DLGAP5 was correlated with poor prognosis in anaplastic thyroid carcinoma (15), non-small cell lung cancer (16,17), pancreatic carcinoma (18) and glioblastoma (19). Loss-of-functional experiments showed that DLGAP5 could promote cell growth, proliferation and migration in hepatocellular carcinoma (20,21). Notably, Liu *et al* (22) and Shen *et al* (23) consistently showed that DLGAP5 was aberrantly expressed and correlated with prognosis in OC. Nevertheless, the biological function of DLGAP5 in OC still has not been reported yet.

Correspondence to: Mrs. Jing Zhang, Department of Obstetrics and Gynecology, Xiangyang Central Hospital, Affiliated Hospital of Hubei University of Arts and Science, 136 Jingzhou Street, Xiangcheng, Xiangyang, Hubei 441000, P.R. China
E-mail: 59714336@qq.com

Mr. Bo Liu, Department of Oncology, Xiangyang Central Hospital, Affiliated Hospital of Hubei University of Arts and Science, 136 Jingzhou Street, Xiangyang, Hubei 441000, P.R. China
E-mail: liubo1306@163.com

*Contributed equally

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Therefore, the present study first searched the Oncomine microarray database (<http://www.oncomine.org>) and Kaplan-Meier Plotter (<http://kmplot.com/analysis/>) to investigate the expression pattern and prognostic value of DLGAP5 in OC. By performing loss-of-functional assays, the effects of DLGAP5 on cell proliferation, cell cycle progression and apoptosis was further explored in OC cells. The findings of the present study might provide evidence that targeting DLGAP5 could be a promising therapeutic strategy in OC.

Materials and methods

Oncomine gene expression analysis. The expression of DLGAP5 in OC was analyzed using publicly available Oncomine microarray database (<http://www.oncomine.org>) by searching the following key words: Gene name, DLGAP5; primary filter, 'Cancer vs. Normal Analysis'; and cancer type 'Ovarian cancer'. A total of six datasets, including Yoshihara Ovarian (24), Welsh Ovarian (25), Bonome Ovarian (26), The Cancer Genome Atlas (TCGA, <http://tcga-data.nci.nih.gov/tcga/>), Adib Ovarian (27) and Lu Ovarian (28) were screened to compare the expression pattern of DLGAP5 between OC tissues with normal tissues. The DLGAP5 expression data were log-transformed and median-centered per array with normalized standard deviation (SD).

Kaplan-Meier plotter analysis. The prognostic value of DLGAP5 was further evaluated in OC using Kaplan-Meier Plotter (<http://kmplot.com/analysis/>), a database that contains information on 22,277 genes and their effect on survival in 2,977 breast, 1,464 ovarian and 1,715 patients with lung cancer. In the present study, patients with OC and complete overall survival information were classified into high and low DLGAP5 expression groups. The effects of DLGAP5 on overall survival rate were displayed using a Kaplan-Meier survival plot. Meanwhile, the hazard ratio was calculated with 95% confidence intervals, log rank P-value and median survival time.

Cell culture and transfection. Two human OC cell lines, including SKOV3 and CAOV3 were purchased from the American Type Culture Collection and routinely cultured in RPMI-1640 medium (Sigma-Aldrich; Merck KGaA) supplemented with 10% FBS (Sigma-Aldrich; Merck KGaA) at 37°C in a humidified incubator containing 5% CO₂. For DLGAP5 knockdown, SKOV3 and CAOV3 cell were seeded into six-well plates at a density of 2x10⁵ cells per well and transfected with 50 nM small interfering (si)RNA targeting DLGAP5 (si-DLGAP5-1: 5'-ATTCAAACCATCAGCC TGC-3' or si-DLGAP5-2: 5'-TTGACCGGAGAAGATAGA GGA-3') or negative control (si-NC: 5'-GAAACACAGCCT GTACGCCTG-3') (all chemically synthesized from Shanghai GenePharma Co., Ltd.) using Lipofectamine® 2000 reagent (Invitrogen; Thermo Fisher Scientific, Inc.) according to the manufacturer's protocol. Subsequent experiments were performed 48 h post transfection.

Cell counting kit-8 (CCK-8) assay. Transfected cells were seeded in a 96-well plate at a density of 3,000 cells per and cultured overnight. At 24, 48 and 72 h, cells in each well were

incubated with 10 µl CCK-8 solution (Biosharp Life Sciences) for 2 h at 37°C. Absorbance at 450 nm was recorded using a microplate reader. Each sample was performed in triplicate.

Colony-formation assay. Transfected cells were plated in six-well culture plates at 500 cells per well and cultured to attach for 24 h. After incubation for 14 consecutive days at 37°C, naturally formed colonies were fixed for 30 min on ice with 70% ethanol and stained with 0.1% crystal violet for 15 min at room temperature. Images of the colonies (each colony contains at least 50 cells) were captured and counted in five randomly selected fields under a light microscope (magnification, x25). Each sample was performed in triplicate.

Flow cytometry analysis. Transfected cells were harvested by trypsinization, washed with cold PBS and fixed with 70% cold ethanol for 30 min at ice. For cell cycle analysis, fixed cells were washed and stained with propidium iodide (PI)/RNase A solution (BD Biosciences) in darkness for 30 min at 37°C. Stained cells were applied for cell cycle distribution analysis using a FACSCalibur flow cytometer (BD Biosciences). For apoptosis analysis, fixed cells were stained with Annexin V-fluorescein isothiocyanate/PI apoptosis detection (BD Biosciences), followed by flow cytometry analysis according to the manufacturer's instructions. Data were analyzed using FlowJo™ v.10.8 software (FlowJo, LLC).

Western blot analysis. All protein samples were extracted and the corresponding protein concentration were determined according to the instructions of RIPA lysis buffer and BCA protein assay provided by Beyotime Institute of Biotechnology, respectively. Next, protein samples (30 µg/lane) were separated on 12% SDS-PAGE gel and transferred onto polyvinylidene fluoride membranes. After conventional blocking with 5% skimmed non-fat dry milk (Cell Signaling Technology, Inc.) for 2 h at room temperature, the membranes were incubated overnight at 4°C with primary antibodies against CDK1 (1:1,000; cat. no. ab131450; Abcam), Cyclin B1 (1:1,000; cat. no. 4138; Cell Signaling Technology, Inc.), Bax (1:1,000; cat. no. 2772; Cell Signaling Technology, Inc.), Bcl-2 (1:1,000; cat. no. sc-7382; Santa Cruz Biotechnology Inc.) and GAPDH (1:1,000; cat. no. G9545; Sigma-Aldrich; Merck KGaA) at 4°C, followed by incubation with secondary antibody conjugated to horseradish peroxidase (1:5,000; cat. no. sc-2357; Santa Cruz Biotechnology, Inc.) for 2 h at room temperature. The protein bands were visualized with an enhanced chemiluminescence substrate (Beyotime Institute of Biotechnology). Results of western blots were analyzed using ImageLab software version 4.1 (Bio-Rad Laboratories, Inc.).

Statistical analysis. Data were analyzed using GraphPad Prism 6.0 (GraphPad Software, Inc.) and expressed as mean ± SD of three independent experiments. One way ANOVA followed by the post hoc Tukey's HSD test was used to assess the differences among different groups. The Renyi test was used for analyzing survival curves. P<0.05 was used to indicate a statistically significant difference.

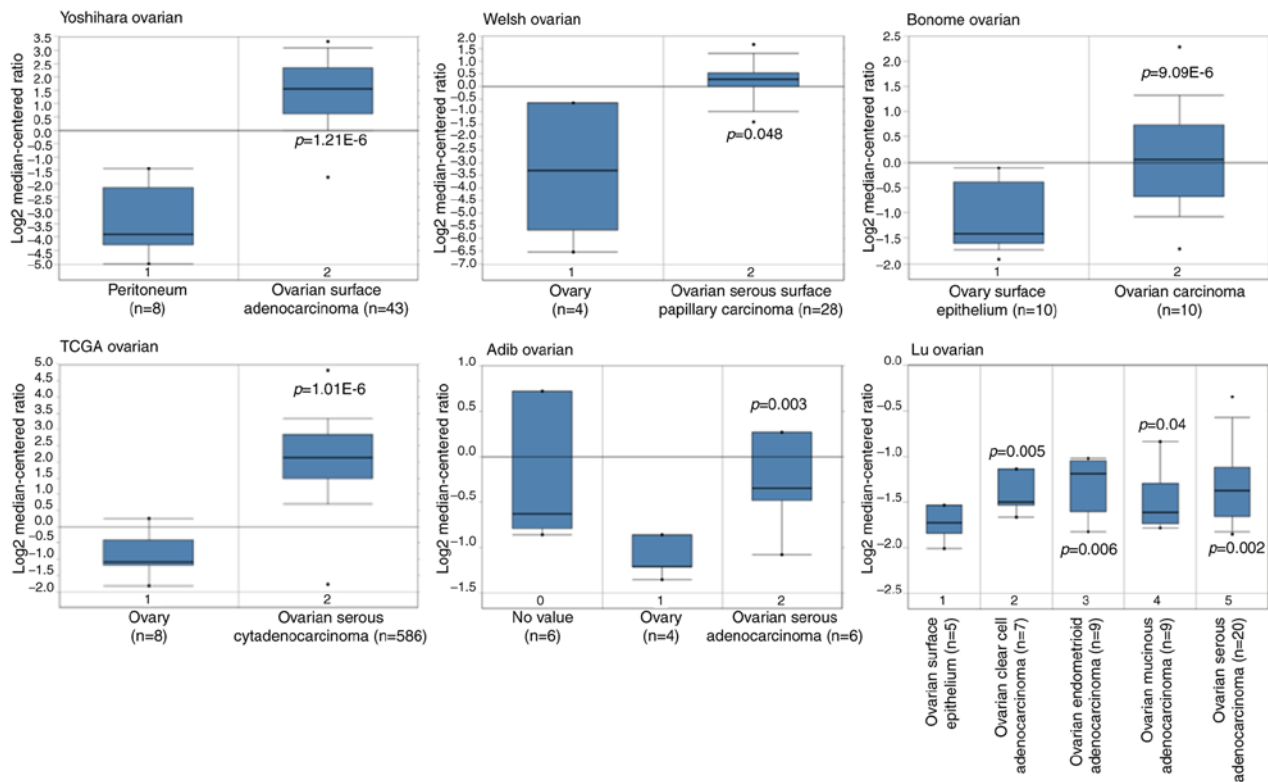


Figure 1. DLGAP5 expression is upregulated in OC tissues. The expression of DLGAP5 mRNA in Oncomine datasets including Yoshihara Ovarian, Welsh Ovarian, Bonome Ovarian, The Cancer Genome Atlas Ovarian, Adib Ovarian and Lu Ovarian is upregulated in OC tissues compared with the normal control tissues. OC, ovarian cancer; DLGAP5, disc large homolog associated protein 5.

Results

DLGAP5 expression in human OC. In order to obtain the expression pattern of DLGAP5 in OC, multiple cancer microarray data sets available from Oncomine database were downloaded to determine the differential mRNA expression of DLGAP5 between OC tissues and normal tissues. As shown in Fig. 1, the mRNA expression of DLGAP5 was markedly increased in OC tissues, including epithelium OC (for example, serous, endometrioid, mucinous and clear cell subtypes) vs. normal tissues in the Yoshihara Ovarian dataset ($P=1.21 \times 10^{-6}$), Welsh Ovarian dataset ($P=0.048$), Bonome Ovarian dataset ($P=9.09 \times 10^{-6}$), TCGA database, ($P=1.01 \times 10^{-6}$), Adib Ovarian dataset ($P=0.003$) and Lu Ovarian dataset ($P<0.05$). The aforementioned observations highlighted the overexpression of DLGAP5 in the development of OC.

Prognostic value of DLGAP5 in OC. Next, the focus was on the survival information of patients with OC to analyze the prognostic value of DLGAP5 using online Kaplan-Meier analysis. As depicted in Fig. 2, 1,656 patients with OC were divided into high DLGAP5 expression group ($n=600$) and low DLGAP5 expression group ($n=1,056$) according to the median value of DLGAP5 expression. Kaplan-Meier survival curve analysis showed that high expression of DLGAP5 was negatively associated with overall survival (HR=1.20, 95% CI: 1.05-1.37, $P=0.0071$). The median survival in the low expression group and high expression group was 48 and 39.93 months, respectively.

Knockdown of DLGAP5 significantly suppresses cell proliferation in OC cells. Considering the overexpression of DLGAP5 in OC tissues, loss-of-functional experiments were subsequently performed to investigate the function of DLGAP5 in OC cells. Firstly, two OC cell lines (SKOV3 and CAOV3) were transfected with two different siRNAs targeting DLGAP5. As demonstrated by western blot analysis, both si-DLGAP5-1 and si-DLGAP5-2 caused a notable down-regulation in DLGAP5 expression in SKOV3 and CAOV3 cells compared with si-NC transfection, while there was no significant change in DLGAP5 expression between control and si-NC group (Fig. 3A). Furthermore, si-DLGAP5-2 transfection caused stronger suppressive effects on DLGAP5 protein expression compared with si-DLGAP5-1 transfection. CCK-8 assay showed that significant inhibition of cell proliferation was observed in SKOV3 (at 48 and 72 h) and CAOV3 (at 24, 48 and 72 h) cells after transfection with either si-DLGAP5-1 or si-DLGAP5-2 compared with si-NC transfection, while there was no significant change in cell proliferation ability between control and si-NC group (Fig. 3B). si-DLGAP5-2 transfection had stronger suppressive effects on the proliferation ability of SKOV3 and CAOV3 cells compared with the effects of si-DLGAP5-1. Thus, si-DLGAP5-2 transfection was selected for the subsequent experiments. Consistent with CCK-8 assay, the number of colonies was significantly decreased in the si-DLGAP5-2 transfection group compared with the si-NC group in both SKOV3 and CAOV3 cells (Fig. 3C and D). The aforementioned data indicated that DLGAP5 might be an oncogene by promoting cell proliferation in OC.

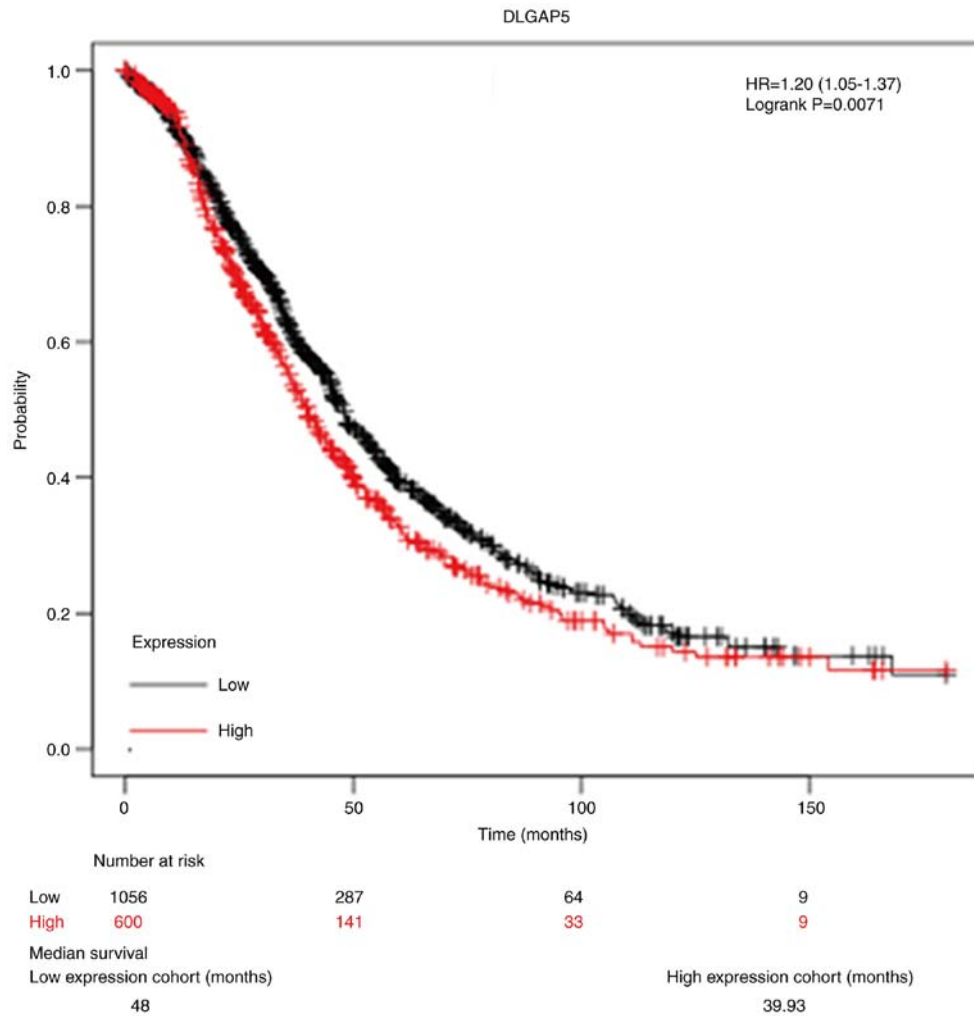


Figure 2. DLGAP5 gene in ovarian cancer (Kaplan-Meier Plotter). Kaplan-Meier plots showing overall survival in ovarian cancer. In red, patients with expression above the median; and in black, patients with expression below the median. Patients with high DLGAP5 expression levels exhibited a poor prognosis with respect to overall survival ($P=0.0071$). DLGAP5, disc large homolog associated protein 5.

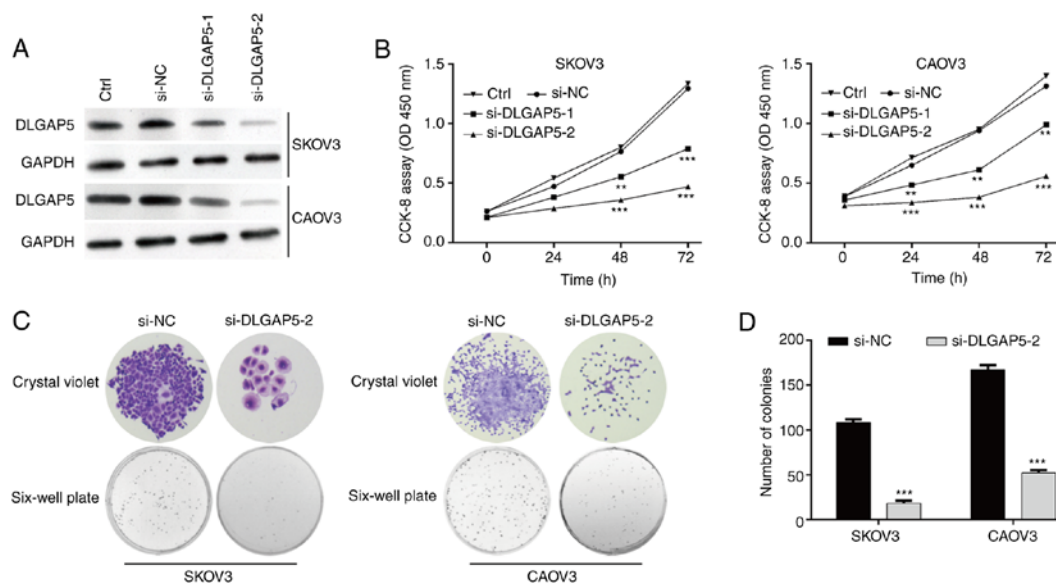


Figure 3. Knockdown of DLGAP5 significantly suppresses cell proliferation in ovarian cancer cells. (A) The expression of DLGAP5 protein was measured in SKOV3 and CAOV3 cells in control (Ctrl), si-NC, si-DLGAP5-1 or si-DLGAP5-2 group. (B) Cell Counting Kit-8 assay was performed to analyze cell proliferation in SKOV3 and CAOV3 cells in Ctrl, si-NC, si-DLGAP5-1 or si-DLGAP5-2 group. (C) Representative images of colonies in SKOV3 and CAOV3 cells after transfection with si-DLGAP5-2 or si-NC are displayed in left panel (magnification, x50) and (D) quantification of colonies is shown in right panel. Data are expressed as mean \pm SD of three independent experiments. $^{**}P<0.01$, $^{***}P<0.001$, vs. si-NC. si-, small interfering RNA; Ctrl, control; NC, negative control.

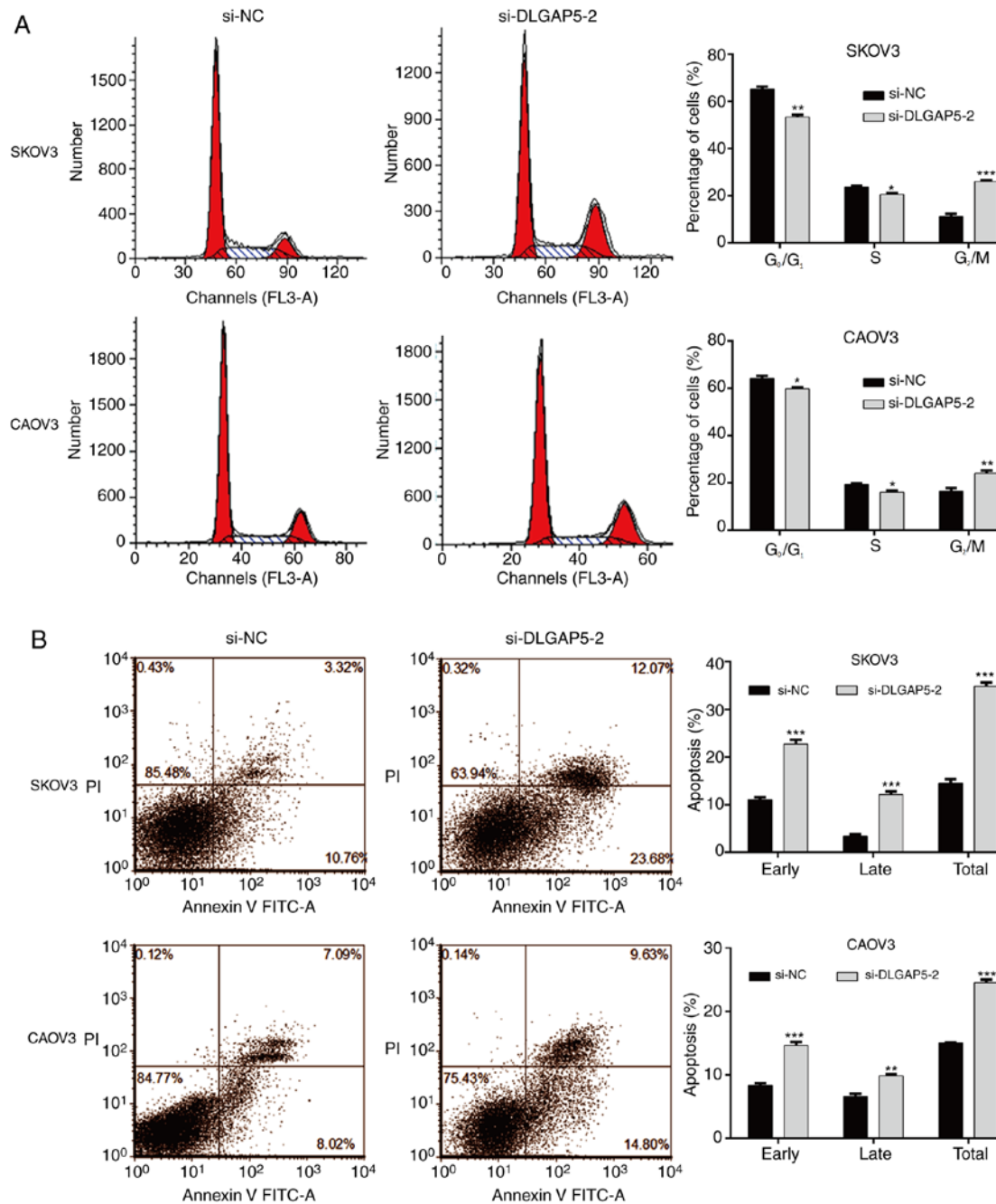


Figure 4. Knockdown of DLGAP5 causes G₂/M phase arrest and apoptosis in ovarian cancer cells. (A) Cell cycle distribution was analyzed in SKOV3 and CAOV3 cells after transfection with si-DLGAP5-2 or si-NC with PI staining, followed by flow cytometry analysis. (B) Apoptotic cells were determined in SKOV3 and CAOV3 cells after transfection with si-DLGAP5-2 or si-NC with double Annexin V/PI staining, followed by flow cytometry analysis. Data are expressed as mean \pm SD of three independent experiments. *P<0.05, **P<0.01, ***P<0.001, vs. si-NC. si-, small interfering RNA; NC, negative control.

Knockdown of DLGAP5 causes G₂/M phase arrest and apoptosis in OC cells. Subsequently, the effects of DLGAP5 knockdown on cell cycle progression and apoptosis was studied by flow cytometry analysis. As shown in Fig. 4A, the percentage of cells at G₀/G₁ and S phases was significantly decreased, accordingly accompanied with increased cell proportion at G₂/M phase in si-DLGAP5-2 group compared with the si-NC group in SKOV3 and CAOV3 cells, which indicated that knockdown of DLGAP5 caused cell cycle G₂/M phase arrest. Moreover, it was found that knockdown of DLGAP5 remarkably increased the early, late and overall apoptotic cells in both SKOV3 and CAOV3 cells (Fig. 4B).

Knockdown of DLGAP5 regulates the protein expression levels associated with G₂/M transition and apoptosis. To explore the mechanisms underlying DLGAP5 regulating cell cycle progression and apoptosis, the expression levels of major cell cycle-promoting factors and apoptosis regulatory factors were analyzed. As shown in Fig. 5, the expression of CDK1 and Cyclin B1, associated with the G₂/M checkpoint, was notably downregulated after DLGAP5 knockdown in both SKOV3 and CAOV3 cells. Meanwhile, DLGAP5 knockdown activated the pro-apoptotic Bax, while suppressing the anti-apoptotic Bcl-2 expression in both SKOV3 and CAOV3 cells.

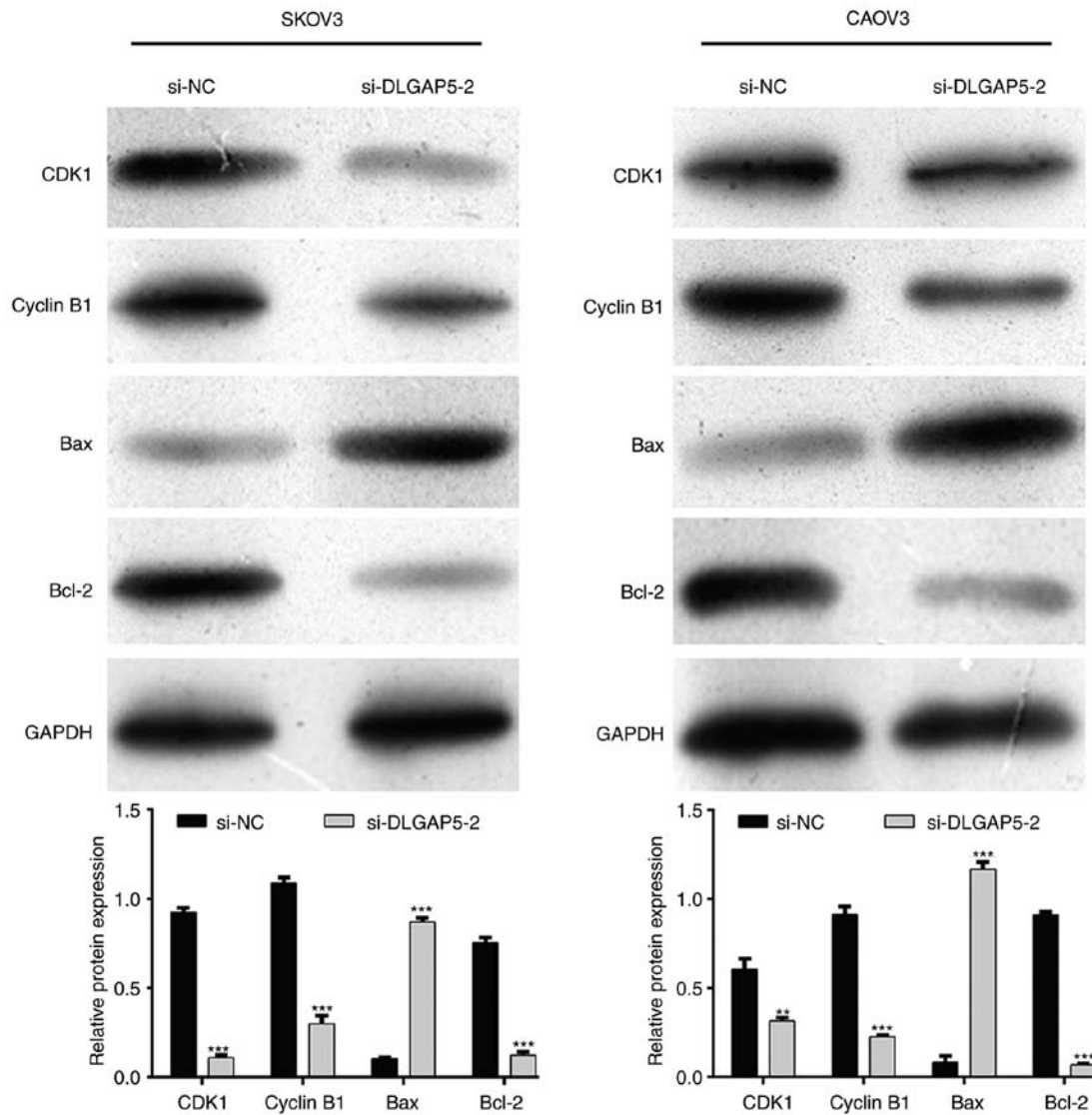


Figure 5. Knockdown of DLGAP5 regulates the protein expression levels associated with G₂/M transition and apoptosis. The protein expression of CDK1, Cyclin B1, Bax and Bcl-2 was measured in SKOV3 and CAOV3 cells after transfection with si-DLGAP5-2 or si-NC. Data are expressed as mean \pm SD of three independent experiments. **P<0.01, ***P<0.001, vs. si-NC. si-, small interfering RNA; NC, negative control.

Discussion

In the present study, DLGAP5 expression was found to be significantly overexpressed in OC tissues and negatively associated with overall survival in patients with OC by bioinformatics analysis. The Lu Ovarian dataset in Fig. 1 demonstrated that the mRNA expression of DLGAP5 was different between mucinous OC and serous, endometrioid and clear cell OC, of which endometrioid OC presented higher DLGAP5 mRNA expression. The present study focused on the expression of DLGAP5 or prognosis in tumor tissues; however, more clinical samples should be collected to confirm this. In fact, DLGAP5 has been reported to be upregulated in hepatocellular cancer (13,29), urinary bladder transitional cell carcinoma (30) and transitional cell carcinoma (31). Consistent with the bioinformatic analysis, DLGAP5 was identified and validated as one of the most upregulated genes regulating cell division and mitotic spindle formation in ovarian granulosa cells with cancer-like characteristics (32). DLGAP5 was also screened as a hub

gene and was associated with the prognosis of endometrial carcinoma (33). Interestingly, the Gene Expression Omnibus database showed that DLGAP5 was one of the core gene with more than ten degrees in epithelial OC (22). GEPIA database analysis indicated that DLGAP5 was highly expressed in OC specimens and was associated with worse prognosis (23). Furthermore, DLGAP5 was overexpressed and associated with worse overall survival in distinct molecular colorectal cancer subtypes (34). The aforementioned evidences suggested that DLGAP5 might be candidate target for the diagnosis and treatment of OC.

Next, two OC cell lines (SKOV3 and CAOV3) were selected to investigate the biological function of DLGAP5, by successfully constructing DLGAP5-silenced OC cell lines. Two different siRNAs targeting DLGAP5 (si-DLGAP5-1 and si-DLGAP5-2) were designed to knockdown DLGAP5 expression and it was found that si-DLGAP5-2 transfection caused stronger suppressive effects on DLGAP5 protein expression compared with si-DLGAP5-1 transfection. Notably, si-DLGAP5-2 transfection had stronger suppressive

effects on the proliferation ability of SKOV3 and CAOV3 cells compared with si-DLGAP5-1 transfection. The differences in knockdown efficiency between si-DLGAP5-2 and si-DLGAP5-1 are ascribed to the different sequences designed for targeting DLGAP5 and different response of OC cells to the designed sequences. It was further found that knockdown of DLGAP5 significantly suppressed colony formation, induced cell cycle G₂/M phase arrest and apoptosis in OC cells. At the molecular level, it was further demonstrated that knockdown of DLGAP5 down-regulated the expression of CDK1, Cyclin B1 and Bcl-2, while upregulating Bax expression in both SKOV3 and CAOV3 cells. To the best of our knowledge, normal cell cycle regulation comprised of G₀/G₁, S, G₂/M stages requires the balance of growth factors and growth inhibitor factors, while its deregulation often causes uncontrolled malignant proliferation involved in the pathogenesis of tumors (35,36). DLGAP5, as a microtubule-associated protein, promotes spindle formation (37), novel tubulin sheet formation (38) and the capture of spindle by kinetochore (39). Associated studies indicated that DLGAP5 is periodically changed as the cells progress from M to G₀/G phase, whose degradation could be modulated by CDK1/Cyclin B (40,41). Through GSEA analysis, Zhang *et al* (42) found that DLGAP5 is involved in the G₁/S phase transition and M/G₁ transition. In addition, DLGAP5 participated in the NUSAP1 knock-down downregulating the expression of genes associated with G₂/M phase, including CDK1 expression in invasive breast cancer cells (42). Liao *et al* (20) also reported that the silencing of DLGAP5 gene expression by RNA interference significantly suppressed cell growth and colony formation *in vitro* in hepatocellular carcinoma cells. Based on the aforementioned facts, one might speculate that knockdown of DLGAP5 inhibited cell proliferation by inducing G₂/M phase arrest via downregulating CDK1/Cyclin B1 expression in OC cells. Meanwhile, it was found that the knockdown of DLGAP5 promoted apoptotic cells by regulating Bax and Bcl-2 expression levels. Similarly, Kuo *et al* (21) indicated that knockdown of DLGAP5 sensitized SK-Hep-1 cells to cisplatin and resulted in smaller tumors in nude mice via promoting apoptosis in hepatocellular carcinoma. The over-expression of DLGAP5 suppressed γ -irradiation-induced apoptosis in prostate cancer cells (43).

In conclusion, it was revealed that DLGAP5 was overexpressed in OC tissues and predicted poor prognosis in patients with OC. The potential mechanisms of DLGAP5 in facilitating OC cell proliferation via regulating G₂/M phase and apoptosis was also demonstrated, suggesting that DLGAP5 might be a potential therapeutic target of OC. In addition, some limitations of the present study were as follows: A non-cancerous cell line as a normal/negative control was not used in the present study. DLGAP5 expression will be analyzed in clinical samples by immunohistochemistry, and the association between DLGAP5 expression and the prognosis of ovarian cancer will be determined in subsequent investigations.

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

HZ designed the study, performed the experiments and made substantial contributions to the conception of this work. YL performed the experiments and assessed the literature. ST and XQ collected the data and were involved in drafting the manuscript or revising it critically for important intellectual content. LL and JTZ analyzed the data. JZ and BL performed the experiments and gave suggestions for the revision of this manuscript. HZ and JZ confirmed the authenticity of all the raw data and designed this work. All authors read and approved the final manuscript.

Ethics approval and consent to participate

Not applicable.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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