

Long non-coding RNA XIST promotes the proliferation of cardiac fibroblasts and the accumulation of extracellular matrix by sponging microRNA-155-5p

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Abstract. Acute myocardial infarction (AMI) is characterized by cardiomyocyte death followed by myocardial fibrosis, eventually leading to heart failure. Long non-coding (lnc)RNA X-inactive specific transcript (XIST) serves a vital role in the regulation of fibrosis. The aim of the present study was to determine whether myocardial fibrosis may be regulated by XIST and to elucidate the underlying mechanism. The relative mRNA expression levels of the target genes were evaluated using reverse transcription-quantitative polymerase chain reaction. Cell viability and apoptosis were determined using a Cell Counting Kit-8 assay and flow cytometry, respectively. The apoptosis and fibrosis-related protein expression levels were detected using western blot analysis. Finally, the interaction between XIST and microRNA (miR)-155-5p was analyzed using a luciferase reporter assay. XIST-overexpression increased proliferation and the expression level of the fibrosis-related proteins in the human cardiac fibroblast cells (HCFs). XIST directly targeted miR-155-5p and downregulated its expression, while miR-155-5p downregulation abolished the effect of XIST-silencing on cell viability and the expression level of the fibrosis-related proteins in the HCFs. XIST promoted cell proliferation and the expression level of fibrosis-related proteins by sponging miR-155-5p. Therefore, XIST may represent a novel effective target for AMI treatment.

Introduction

Acute myocardial infarction (AMI) is associated with adverse cardiac remodeling and impaired ventricular function, which is a leading cause of mortality worldwide (1,2). Myocardial

fibrosis following myocardial infarction is a crucial mechanism of cardiac insufficiency in patients with AMI. A previous study reported that myocardial fibrosis was characterized by the abnormal proliferation of fibroblasts and the accumulation of extracellular matrix (ECM) in the heart, which affected the diastolic and contractile function of the heart, eventually leading to heart failure (HF) (3,4). Due to the adverse impact of AMI on the survival rate of patients, developing novel therapeutic strategies to improve cardiac performance by reducing the number of myocardial fibroblasts following AMI are desirable.

Long non-coding (lnc)RNAs are a type of RNA transcript, longer than 200 nucleotides in length and with no open reading frame (5). A previous study suggested that lncRNAs serve important roles in diverse biological processes, and their vital role in the regulation of fibrosis is increasingly being elucidated (6). X-inactive specific transcript (XIST) was highly expressed in hypertrophic mouse hearts and promoted the progression of cardiac hypertrophy by targeting microRNA(miR)-101 to upregulate TLR2 expression (7). However, the biological roles of lncRNA XIST in myocardial injury and fibrosis have not been widely reported. McKiernan *et al* (8) reported that XIST was aberrantly expressed in cystic fibrosis bronchial epithelium *in vivo*, which may be associated with inflammation. Furthermore, XIST promoted bleomycin-induced pulmonary fibrosis by inhibiting miR-139 to enhance the proliferative ability of human/mouse fibroblasts and ECM protein expression (9). Notably, XIST-silencing may protect against renal interstitial fibrosis in diabetic nephropathy by miR-93-5p-dependent CDKN1A inhibition (10). Therefore, we hypothesized that XIST may contribute toward myocardial fibroblasts following AMI.

Angiotensin II (Ang II) is the main active protein in the rennin-angiotensin-aldosterone system (RAAS), which serves a crucial role in myocardial interstitial fibrosis and cardiac remodeling (11,12). Under pathological conditions, Ang II stimulates the proliferation of cardiac fibroblast cells (CFs) by autocrine and paracrine signaling, and promotes the synthesis of collagen (Col) I to repair injury in myocardial tissues (13-15). However, the continuous abnormal synthesis of Col I, caused by the excessive proliferation of CFs, may lead toward myocardial interstitial fibrosis and decreased compliance of the ventricle, which ultimately affects the diastolic and contractile function of the heart.

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In the present study, human cardiac fibroblast cells (HCFs) were used and treated with Ang II, while pcDNA3.1 XIST and short hairpin (sh)RNA-XIST was used to induce XIST overexpression and silencing in cells, respectively. The present study was designed to determine whether myocardial fibrosis could be regulated by XIST and to elucidate the related mechanism. The results demonstrated that XIST promoted the proliferation of myocardial fibroblasts and the accumulation of ECM, suggesting that XIST may be a novel potential target for AMI treatment.

Materials and methods

Cell culture and transfection. Primary HCFs were obtained from the American Type Culture Collection and were maintained in Dulbecco's modified Eagle's medium (DMEM, Hyclone; Cytival), supplemented with 10% FBS (Invitrogen; Thermo Fisher Scientific, Inc.) and 1% penicillin and 1% streptomycin (Gibco; Thermo Fisher Scientific, Inc.) at 37°C for 48 h in a humidified incubator with 5% CO₂, followed by Ang II (Sigma-Aldrich; Merck KGaA) treatment (0.01 mM) at 37°C for 24 h to induce the myocardial fibrosis phenotype.

At 70–80% confluence, HCFs were seeded onto a corresponding culture plate and starved for 12 h in serum-free DMEM (Hyclone; Cytival) prior to treatment. The plasmids, including 1 µg/well pcDNA3.1 XIST (forward, 5'-CCAAGC TTTGCACACGGCCTATCTCATC-3' and reverse, 5'-CCG CTCGAGTGAAAAGAGGTGGGGCATCC-3') inserted into pcDNA3.1, 100 pmol of short hairpin RNA (shRNA)-XIST (shRNA-XIST-1, 5'-TTCTCCGAACGTGTCACGT-3'; 100 pmol of shRNA-XIST-2, 5'-GCTGCTAGTTTCCCAATG ATA-3'), 100 pmol of shRNA-NC (5'-TTCTCCGAACGTGTC ACGT-3'), 20 nM of miR-155-5p mimic (forward, 5'-UUAUUG CUAUUCGUGAUAGGGGU-3' and reverse, 5'-CCCUAU CACGAUUAAGCAUUAUU-3'), 20 nM of mimic-negative control (NC; forward, 5'-UUCUCCGAACGUGUCACGUT T-3' and reverse, 5'-ACGUGACACGUUCGGAGAATT-3'), 100 nM of miR-155-5p inhibitor (5'-ACCCCUAUCACGAU AGCAUUA-3') and 100 nM of inhibitor-NC (5'-CAGUAC UUUUGUGUAGUACAA-3') were purchased from Shanghai GenePharma Co., Ltd., and transfected into the HCFs using Lipofectamine® 3000 (Invitrogen; Thermo Fisher Scientific, Inc.) according to the manufacturer's protocols. The interaction between XIST and miR-155-5p was predicted using the Starbase website (<http://starbase.sysu.edu.cn/index.php>). After 48 h, the HCFs were used for further experiments and reverse transcription-quantitative polymerase chain reaction (RT-qPCR) was performed to examine transfection efficiency.

RT-qPCR. Total RNA was extracted from the HCFs using TRIzol® (Invitrogen; Thermo Fisher Scientific, Inc.) according to the manufacturer's protocols. First strand cDNA was synthesized at 42°C for 60 min and 95°C for 5 min, using a PrimeScript RT reagent kit. qPCR was performed using a SYBR® Green Real-Time PCR Master Mix (Takara Bio, Inc.). qPCR amplification parameters were: Annealing at 25°C for 10 min, extension at 42°C for 30 min; inactivation at 85°C for 10 min, after extension of the detection of fluorescence signals, a total of 40 cycles. The 2^{-ΔΔCq} method (16) was used to quantify the relative mRNA expression of the target and

housekeeping genes. The following primers were used: XIST forward, 5'-ACGCTGCATGTGTCCTTAG-3' and reverse, 5'-GAGCCTCTTATAAGCTGTTT-3'; miR-155-5p forward, 5'-GAGGGTTAATGCTAATCGTGATAGG-3' and reverse, 5'-GCACAGAATCAACACGACTCACTAT-3'; GAPDH forward, 5'-CTGGGCTACACTGAGCACC-3' and reverse, 5'-AAGTGGTCGTTGAGGGCAATG-3'; and U6 forward, 5'-GAGGGTTAATGCTAATCGTGATAGG-3' and reverse, 5'-GCACAGAATCAACACGACTCACTAT-3'.

Cell proliferation. After the HCFs (4×10³/well) from different experimental groups, were plated onto 96-well plates and cultured for 24 h, the Cell Counting Kit-8 (CCK-8; Dojindo Molecular Technologies, Inc.) was added (10 µl/well) and incubated with the HCFs for another 2 h according to manufacturer's protocol. A microplate reader was used to detect the absorbance at 450 nm. The experiment was repeated independently three times.

Flow cytometry. The Annexin V-FITC/PI apoptosis detection kit (Beyotime Institute of Biotechnology) was used according to the manufacturer's protocols. The harvested cells (1×10⁵ cells/ml) were resuspended in 500 µl of 1X binding buffer (Beyotime Institute of Biotechnology). Next, the cells were incubated with 5 µl Annexin V-FITC followed by 10 µl PI staining solution. Following incubation in the dark, at room temperature for 20 min, the fluorescence was detected using a flow cytometer (FACScan™; BD Biosciences) and ModFit software (version 3.2; Verity Software House, Inc.) to analyze the apoptotic rate of HCFs.

Western blot analysis. The HCFs in each group were lysed with RIPA lysis buffer (Beyotime Institute of Biotechnology). Total protein was determined using a bicinchoninic acid assay (Thermo Fisher Scientific, Inc.). Next, the protein samples (25 µg) were separated using 10% SDS-PAGE, transferred to polyvinylidene difluoride membranes, and blocked with 5% skimmed milk at room temperature for 1 h. The membranes were incubated overnight at 4°C with the primary antibodies against Bcl-2 (1:1,000; cat. no. 2875; Cell Signaling Technology, Inc.), Bax (1:1,000; cat. no. 2772, Cell Signaling Technology, Inc.), cleaved-caspase-9 (1:500; cat. no. 9509; Cell Signaling Technology, Inc.), cleaved caspase-3 (1:500; cat. no. 9654; Cell Signaling Technology, Inc.), caspase-3 (1:1,000; cat. no. 9668; Cell Signaling Technology, Inc.), caspase-9 (1:1,000; cat. no. 9502; Cell Signaling Technology, Inc.), Col I (1:1,000; ab34710; Abcam), Col III (1:1,000; ab6310; Abcam), α-smooth muscle actin (α-SMA; 1:1,000; cat. no. 68463; Cell Signaling Technology, Inc.), cytochrome C (1:1,000; cat. no. 12959; Cell Signaling Technology, Inc.), cleaved PARP (1:500; cat. no. 9185; Cell Signaling Technology, Inc.), and PARP (1:1,000; cat. no. 9542; Cell Signaling Technology, Inc.). Next, the membranes were incubated with horseradish peroxidase-conjugated horseradish peroxidase-conjugated anti-rabbit IgG secondary antibodies (1:1,000; cat. no. 7074; Cell Signaling Technology, Inc.) at room temperature for 2 h and visualized using a Tanon-5200 chemiluminescence imager (Tanon Science and Technology Co., Ltd) using an enhanced chemiluminescence kit and the intensity of the bands was normalized to GAPDH expression.

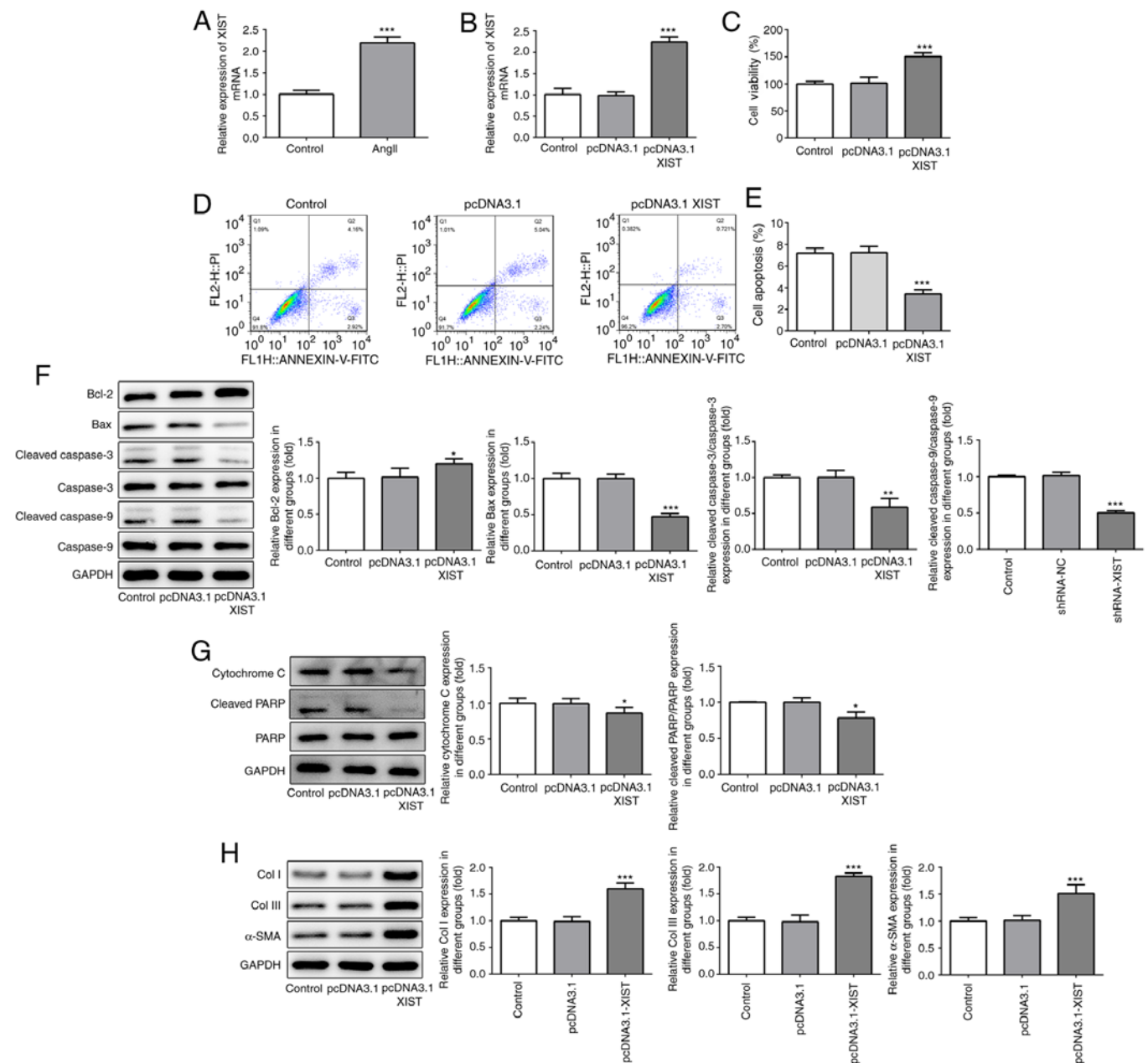


Figure 1. XIST-overexpression promoted cell proliferation and increased the expression of fibrosis-related proteins in the HCFs. (A and B) The relative mRNA expression level of XIST was determined using reverse transcription-quantitative polymerase chain reaction. (C) The viability of the HCFs was analyzed using a Cell Counting Kit-8 assay. (D and E) The apoptotic rates of the HCFs transfected with or without pcDNA3.1 XIST were determined using flow cytometry. The protein expression of (F) Bcl-2, Bax, cleaved caspase-3, caspase-3, cleaved caspase-9, caspase-9; (G) cytochrome *c*, cleaved PARP and PARP; and (H) Col I, Col III and α -SMA were detected using western blot analysis. The data are expressed as the mean \pm standard error of the mean, from three independent experiments. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$ vs. control. Col, collagen; α -SMA, α smooth muscle actin; HCFs, human cardiac fibroblast cells.

Luciferase reporter assay. The HCFs (1×10^5) were seeded onto 24-well plates and cultured for 24 h. The luciferase reporter vector for the wild-type (WT) or mutant (MUT) type 3'-untranslated region (UTR) of XIST, which contained the binding sites between miR-155-5p was cloned into the pMIR-reporter luciferase system (Thermo Fisher Scientific, Inc.). The MUT 3'-UTR of XIST was constructed using the QuikChange Site-Directed Mutagenesis kit (Agilent Technologies). miR-155-5p mimic or mimic-NC combined with the vector expressing firefly luciferase reporter fused with WT or MUT 3'-UTR of XIST was co-transfected into the HCFs using Lipofectamine[®] 2000 (Invitrogen; Thermo Fisher

Scientific, Inc.). Following incubation for 24 h, the HCFs were collected and the luciferase activities were determined using the dual-luciferase reporter assay system (Promega Corporation) and the firefly luciferase activity was normalized to *Renilla* luciferase activity.

Statistical analysis. The data are presented as the mean \pm standard error of the mean and analyzed using GraphPad Prism v5.0 (GraphPad Software, Inc.). The difference between two groups was determined using a two-tailed Student's t-test, while a one-way analysis of variance, followed by Bonferroni's post hoc test, was used for multiple

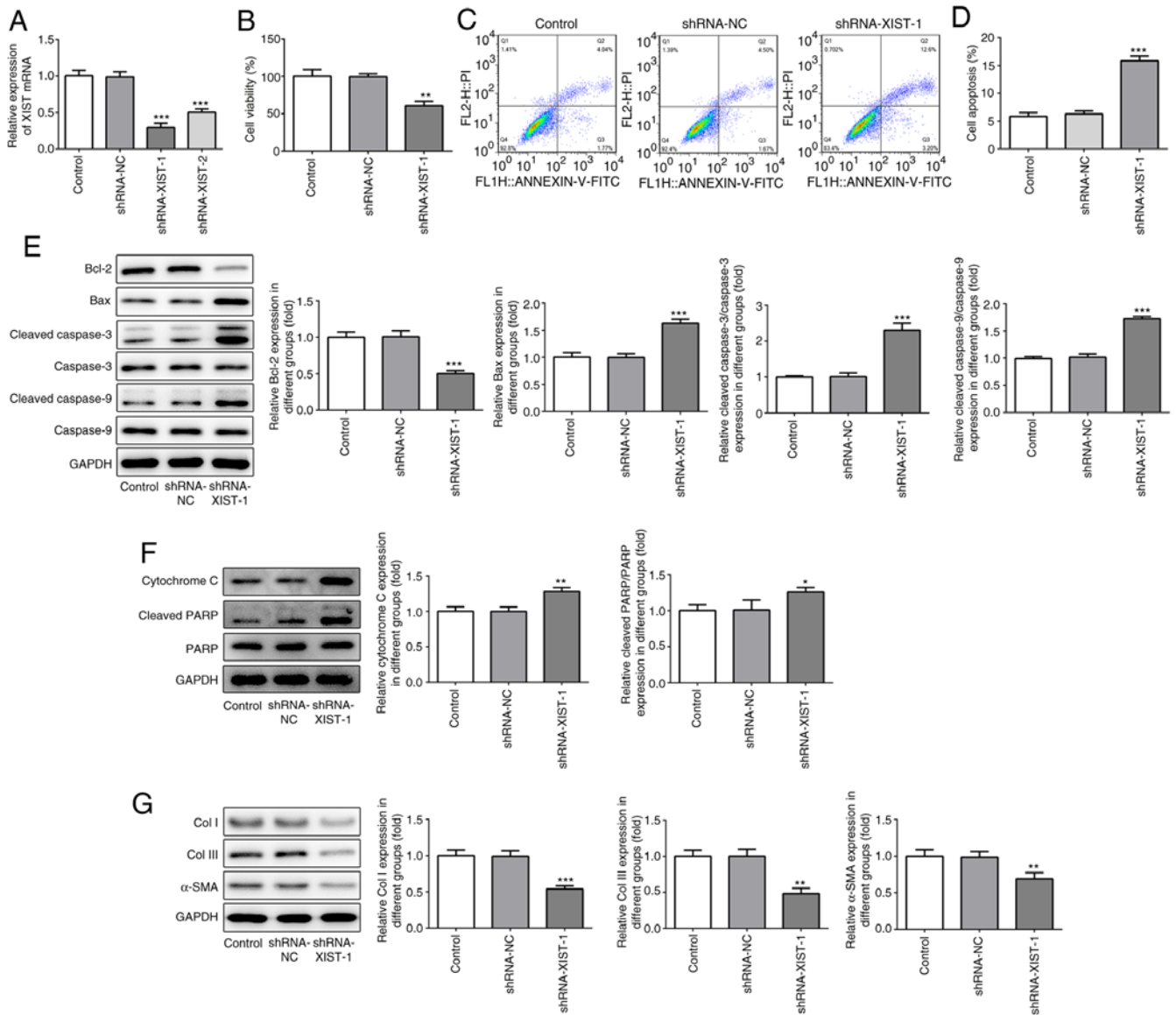


Figure 2. XIST-silencing inhibited the proliferation and expression of fibrosis-related proteins in the HCFs. (A) The relative mRNA expression level of XIST was determined using reverse transcription-quantitative polymerase chain reaction. (B) The viability of the HCFs was analyzed using Cell Counting Kit-8 assay. (C) The apoptotic rate of the HCFs transfected with or without shRNA-XIST-1 was determined using flow cytometry and was (D) subsequently quantified. The protein expression of (E) Bcl-2, Bax, cleaved caspase-3, caspase-3, cleaved caspase-9, caspase-9; (F) cytochrome *c*, cleaved PARP and PARP; and (G) Col I, Col III and α -SMA was detected using western blot analysis. The data are presented as the mean \pm standard error of the mean, from three independent experiments. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$ vs. control. Col, collagen; α -SMA, α smooth muscle actin; sh, short hairpin; NC, negative control; HCFs, human cardiac fibroblast cells.

groups. $P < 0.05$ was considered to indicate a statistically significant difference.

Results

XIST overexpression promotes proliferation and the protein expression level of fibrosis-related proteins in HCFs. To investigate the association between XIST and myocardial fibrosis, RT-qPCR was performed to detect XIST mRNA expression following treatment of the HCFs with Ang II. As shown in Fig. 1A, Ang II treatment significantly increased the relative mRNA expression of XIST, compared with that in the control group. Subsequently, the pcDNA3.1-XIST plasmid was used to induce XIST overexpression, which was confirmed by the RT-qPCR results (Fig. 1B). The CCK-8 assay

was performed to analyze the cell viability of the HCFs. As shown in Fig. 1C, the HCFs in the pcDNA3.1-XIST group had a higher rate of viability, compared with that in the control and pcDNA3.1 groups. Furthermore, cell apoptosis was detected using flow cytometry and the protein expression level of the apoptosis-related proteins were quantified using western blot analysis. As shown in Fig. 1D and E, the apoptotic rate was significantly decreased in the pcDNA3.1-XIST group, compared with that in the control group. In addition, XIST-overexpression induced upregulation of Bcl-2, while Bax, cleaved caspase-3 and -9, cytochrome *c* and cleaved PARP were downregulated (Fig. 1F and G). Finally, the protein expression levels of Col I, Col III and α -SMA were also determined using western blot analysis and the results revealed that the expression of these proteins was significantly increased by

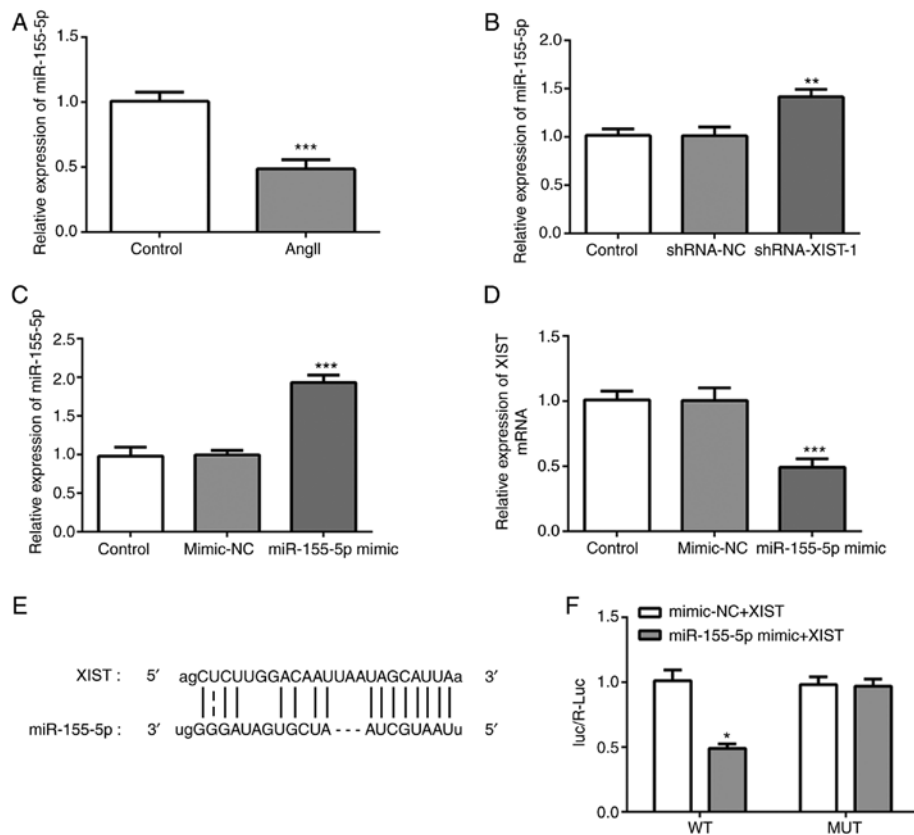


Figure 3. XIST directly targets miR-155-5p and downregulates its expression. (A-C) Relative mRNA expression of miR-155-5p was determined using RT-qPCR. (D) Relative mRNA expression of XIST was determined using RT-qPCR. (E) The binding sites between XIST and miR-155-5p. (F) The relative luciferase activities in the human cardiac fibroblast cells were evaluated using a luciferase reporter assay. The data are presented as the mean \pm standard error of the mean, from three independent experiments. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$ vs. control. miR, microRNA; RT-qPCR, reverse transcription-quantitative polymerase chain reaction; WT, wild-type; MUT, mutant; NC, negative control; Ang II, angiotensin II; sh, short hairpin.

XIST overexpression (Fig. 1H). These results suggested that XIST-overexpression promoted the proliferation of HCFs and the expression of the fibrosis-related proteins *in vitro*.

XIST-silencing inhibits proliferation and decreases the expression of fibrosis-related proteins in HCFs. To confirm the role of XIST in myocardial fibrosis, shRNA-XIST-1 and shRNA-XIST-2 were used to induce XIST-silencing. As shown in Fig. 2A, the RT-qPCR results demonstrated that a lower mRNA expression level of XIST was observed in the HCFs in the shRNA-XIST-1 group, compared with that in the shRNA-XIST-2 group; therefore, shRNA-XIST-1 was selected in the following experiments. The CCK-8 assay results demonstrated that XIST-silencing significantly suppressed cell viability of the HCFs (Fig. 2B). The flow cytometry results revealed that XIST-silencing significantly enhanced the apoptotic rate of the HCFs (Fig. 2C and D). In addition, XIST-silencing increased the protein expression level of Bax, cleaved caspase-3 and -9, cytochrome *c* and cleaved PARP, while the protein expression level of Bcl-2, Col I, Col III and α -SMA was decreased (Fig. 2E-G). These results indicated that XIST-silencing inhibited proliferation of the HCFs and the protein expression level of fibrosis-related proteins *in vitro*.

XIST directly targets miR-155-5p and downregulates its mRNA expression level. To investigate the specific mechanism of XIST in myocardial fibrosis, RT-qPCR was performed

to detect miR-155-5p mRNA expression, following treatment with Ang II. As shown in Fig. 3A, Ang II treatment significantly decreased the relative mRNA expression of miR-155-5p. In addition, the RT-qPCR results demonstrated that the mRNA expression of miR-155-5p could be increased by XIST-silencing (Fig. 3B). Subsequently, miR-155-5p mimic was used to induce miR-155-5p-overexpression. The RT-qPCR results demonstrated that miR-155-5p mimic was transfected into the cells successfully and upregulated miR-155-5p mRNA expression (Fig. 3C). Notably, XIST expression was significantly decreased by miR-155-5p-overexpression (Fig. 3D). The interaction between XIST and miR-155-5p was predicted using the Starbase website (<http://starbase.sysu.edu.cn/index.php>), as presented in Fig. 3E. Subsequently, the luciferase reporter assay was performed to confirm whether miR-155-5p was a direct target of XIST. As shown in Fig. 3F, the relative luciferase activity was significantly lower in the HCFs co-transfected with WT-XIST and miR-155-5p mimic than that in the control group. These results indicated that XIST specifically bound to the 3'-UTR of miR-155-5p to decrease its expression.

miR-155-5p-downregulation abolishes the effect of XIST-silencing on cell viability and the protein expression level of the fibrosis-related proteins in the HCFs. To further investigate the molecular mechanism of XIST in myocardial fibrosis, the HCFs were transfected with an miR-155-5p inhibitor. As

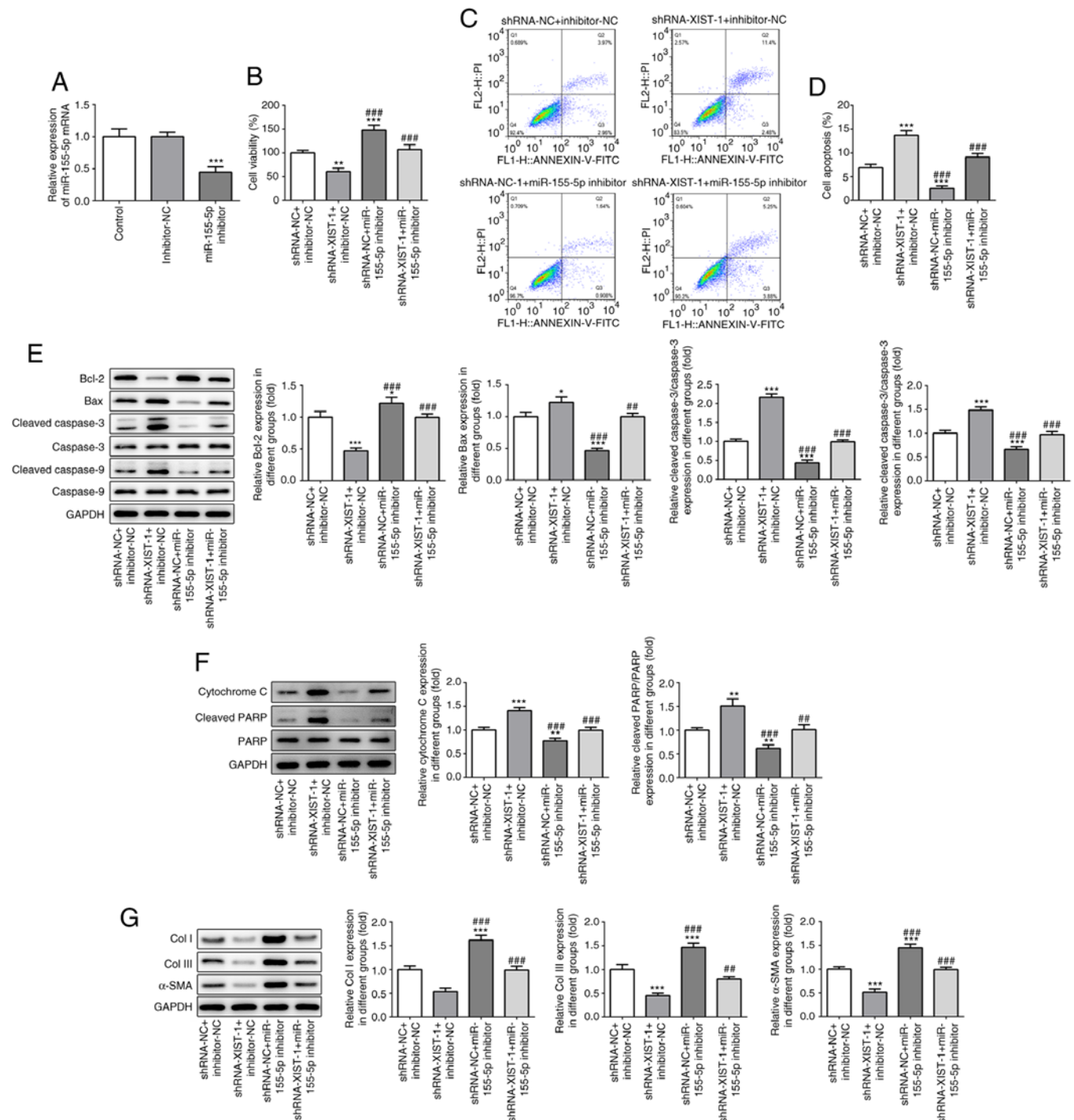


Figure 4. miR-155-5p-downregulation abolished the effect of XIST-silencing on cell viability and the expression of fibrosis-related proteins in the HCFs. (A) The relative mRNA of miR-155-5p was determined by reverse transcription-quantitative polymerase chain reaction. (B) The viability of the HCFs was analyzed using a Cell Counting Kit-8 assay. (C) The apoptotic rate of the HCFs in each group was determined using flow cytometry and was (D) subsequently quantified. The protein expression of (E) Bcl-2, Bax, cleaved caspase-3, caspase-3, cleaved-caspase-9, caspase-9; (F) cytochrome *c*, cleaved PARP and PARP; and (G) Col I, Col III and α -SMA was detected using western blot analysis. The data are presented as the mean \pm standard error of the mean, from three independent experiments. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$ vs. shRNA-NC+inhibitor-NC. # $P < 0.05$, ### $P < 0.001$ vs. shRNA-XIST-1+inhibitor-NC. miR, microRNA; HCFs, human cardiac fibroblast cells; sh, short hairpin; NC, negative control; Col, collagen; α -SMA, α smooth muscle actin.

shown in Fig. 4A, miR-155-5p-downregulation was achieved via miR-155-5p inhibitor transfection. The CCK-8 assay results demonstrated that miR-155-5p inhibitor abolished the suppressive effects of XIST-silencing on cell viability (Fig. 4B). The flow cytometry results demonstrated that XIST-silencing promoted apoptosis in the HCFs, which was partly reversed

by miR-155-5p downregulation (Fig. 4C and D). The western blot result showed that miR-155-5p downregulation abolished the regulatory effect of XIST on the protein expression level of the apoptosis-related proteins (Fig. 4E and F), as well as the fibrosis-related proteins (Fig. 4G). These results suggested that XIST promoted the proliferation of the HCFs and the

expression level of the fibrosis-related proteins by sponging miR-155-5p.

Discussion

AMI is one of the diseases with the highest morbidity and mortality rates in the world (17), and it may induce cardiac remodeling and HF (18). Myocardial fibrosis has been reported to be associated with sparseness of cardiac microvessels, and found to contribute toward HF, which is caused by myocardial fibroblast accumulation and imbalance of synthesis, metabolism and degradation of collagen in ECM deposition (19). CFCs are the primary cells that secrete the ECM of the myocardium. In the present study, HCFs were treated with Ang II to induce the myocardial fibrosis phenotype. Notably, it was observed that XIST was highly expressed in the HCFs treated with Ang II, suggesting that XIST was associated with the onset and development of myocardial fibrosis.

A recent study demonstrated that XIST was upregulated in ethanol-induced human hepatic stellate cells and contributed toward liver fibrogenesis (20), which was consistent with the results of the present study. Furthermore, the results of the present study suggested that XIST-overexpression promoted proliferation and enhanced the protein expression of fibrosis-related proteins in HCFs, while XIST-silencing had the opposite effect. These data indicated that XIST may promote the development of myocardial fibrosis.

To investigate the specific mechanisms underlying the role of XIST in myocardial fibrosis, the interaction between XIST and miR-155-5p was predicted using the Starbase website. It was found that miR-155-5p was one of the potential targets of XIST. In the present study, Ang II treatment led to a decrease in miR-155-5p mRNA expression, whereas XIST downregulation significantly increased miR-155-5p mRNA expression. These data suggested that XIST may directly target miR-155-5p and downregulate its mRNA expression level. A recent study has demonstrated that miR-155-5p suppressed the proliferation, migration and invasion of vascular smooth muscle cells (VSMCs) and human umbilical vein endothelial cells in atherosclerosis by regulating Akt (21). Chen *et al* (21) reported that miR-155-5p inhibition prevented the development of abdominal aortic aneurysms in mice by regulating macrophage-mediated inflammation (22). It was reported that miR-155-overexpression decreased cellular proliferation in VSMCs, and the expression of miR-155 was decreased and negatively correlated with calcification in the aorta of chronic kidney disease rats (23). Additionally, a previous study reported that miR-155-5p mediated the regulatory network for myocardial infarction (24), suggesting that miR-155-5p may be a putative therapeutic target for AMI. In addition, miR-155-5p-upregulation decreased vascular thickening and fibrosis in a vascular calcification model (25). In the present study, miR-155-5p-downregulation promoted the proliferation of the HCFs and suppressed cell apoptosis. It is well-known that collagen, an insoluble fibrous protein, is the main component of the ECM (26,27). miR-155-5p downregulation abolished the inhibitory effect of XIST-silencing on the protein expression level of Col I, Col III and α -SMA in

the HCFs, suggesting that XIST promoted the proliferation of the HCFs and the protein expression of fibrosis-related proteins by sponging miR-155-5p, as shown in Fig. S1.

In summary, XIST-silencing inhibited the proliferation of the HCFs and collagen expression, which was partially reversed by miR-155-5p-downregulation; therefore, XIST promoted cell proliferation and the protein expression of the fibrosis-related proteins by sponging miR-155-5p. Therefore, XIST may be a novel effective target for AMI treatment.

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

JZ designed the study. HZ and JM were responsible for the data collection and analysis. HZ and FL collaborated to perform the data analysis. All authors collaborated to interpret results and develop the manuscript. All authors read and approved the final version of the manuscript.

Ethics approval and consent for participation

Not applicable.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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