

TRIM21-a potential biomarker for the prognosis of thyroid cancer

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Abstract. Thyroid cancer (THCA) is one of the commonest malignancies associated with increased recurrence. Therefore, identifying the putative molecular markers and therapeutic targets to improve the treatment of THCA is essential. The present study analyzed the potential role of tripartite motif-containing 21 (TRIM21), a member of the TRIM family belonging to the subfamily of E3 ubiquitin ligases, in the progression of THCA. Using bioinformatics analysis and immunohistochemistry of THCA tissues, it was observed that TRIM21 is overexpressed in THCA tissues. The present study also found that TRIM21 is associated with lymph node metastasis and high-risk recurrence of THCA. Furthermore, it identified a promotional role of TRIM21 in THCA cell migration and invasion. In addition, the present study analyzed TRIM21-enriched pathways and co-expressed genes in THCA. The present study suggested that TRIM21 may serve as a potential biomarker for THCA prognosis.

Introduction

Thyroid cancer (THCA) is one of the commonest malignancies associated with increased recurrence (1). Generally, THCA shows a good overall prognosis and low fatality rate in most cases; however, due to its aggressive characteristics and

metastasis, and poor prognosis can be seen in some patients with THCA (2). Identifying available molecular markers and therapeutic targets is urgently required to improve the treatment outcome.

The tripartite motif (TRIM) family of proteins belongs to the subfamily of E3 ubiquitin ligases and participates in various biological and pathophysiological processes, including tumor progression (3-5). TRIMs share similar domains in their protein structure, including the N-terminal RING domain with E3 ubiquitin ligase activity, the B-box domain, and the coiled-coil domain (6). Several members of the TRIM family are associated with tumorigenesis and disease progression of THCA. TRIM14 has been reported as an oncogene in THCA (7). TRIM44 knockdown suppresses the tumor progression of THCA by inhibiting the Wnt/ β -catenin signaling pathway (8). TRIM8 serves as a target for miR-182 in promoting tumor growth and increasing chemoresistance in human THCA (9). However, the roles of other TRIMs in THCA remain to be elucidated. The authors of the present study aim to investigate the roles of other TRIMs in THCA and so far, TRIM21 is the one which has been elucidated. Therefore, the present study reported TRIM21. TRIM21 is a member of the TRIM family involved in innate immunity and the development of diseases such as systemic lupus erythematosus, and Sjögren's syndrome (10). TRIM21 may play opposing roles in tumorigenesis and its progression (6,11). TRIM21 suppresses hepatocellular carcinoma cell invasion (12); it inhibits renal cancer tumorigenesis and metastasis by mediating hypoxia-inducible factor-1 α (HIF-1 α) degradation (13). TRIM21 promotes glioma progression by regulating cell proliferation and migration (14). By contrast, TRIM21 is related to the therapeutic sensitivity of several tumors: By suppressing EZH1 stability, TRIM21 improves the sensitivity of gastric cancer to apatinib (15). In squamous cell carcinoma of the head and neck, high TRIM21 expression is associated with a shorter overall survival rate (16). However, the participation of TRIM21 in THCA regulation remains unelucidated.

The present study aimed to identify the role of TRIM21 in THCA and to analyze the functional networks related to TRIM21 using public databases such as The Cancer Genome Atlas (TCGA) database. The function of TRIM21 in the proliferation, migration and invasion of THCA cells was evaluated.

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Key words: thyroid cancer, tripartite motif-containing 21, bioinformatical analysis, clinicopathologic features, migration and invasion

Materials and methods

Clinical sample collection. Paraffin tumor tissue samples and paraffin para-tumor normal tissues 1 cm away from the tumor tissues were collected from 120 patients diagnosed with papillary thyroid carcinoma and who underwent surgical resection in Liaocheng People's Hospital between 2018 and 2020. The patients were aged from 21-70. The clinical information of all cases was collected and is given in Table I. All patients were free of other malignancies or a history of chemoradiotherapy. Written informed consent was obtained from all the participants. The experiment was approved by the Ethics committee of Liaocheng People's Hospital (approval no. LC2021059).

Immunohistochemical staining and scoring. Immunohistochemical staining (IHC) was performed to detect the expression of TRIM21 in papillary thyroid carcinoma tissues. Endogenous peroxidase activity was blocked using 3% hydrogen peroxide (cat. no. 88597; Merck KGaA) after routine dewaxing, hydration, and antigen retrieval. Permeabilization of samples was performed using 0.1% Triton X-100 (cat. no. ST797; Beyotime Institute of Biotechnology) and blocked with 5% bovine serum albumin (BSA) (cat. no. ST025; Beyotime Institute of Biotechnology). Tissue sections (10 μ m thick) were incubated with TRIM21 antibodies (1:200 dilution; ProteinTech; cat. no. 12108-1-AP) at 4°C for 12 h. After washing with phosphate-buffered saline (PBS), the sections were incubated with horseradish peroxidase (HRP)-conjugated goat anti-rabbit IgG (1:1,000 dilution; cat. no. A0208; Beyotime Institute of Biotechnology) for 1 h at 20°C. TRIM21 expression was visualized using 3,3'-diaminobenzidine (DAB; cat. no. P0202; Beyotime Institute of Biotechnology) staining at 20°C for 1 min. A blind evaluation was performed by two pathologists. Brown-yellow staining indicated a positive expression. Staining score = staining intensity score \times staining-positive area score. The staining intensity was scored as 0 (negative), 1 (weakly positive), 2 (moderately positive) and 3 (strongly positive). The score of the staining-positive area was recorded according to the proportion of positive cells: 0 (<5%), 1 (5-25%), 2 (26-50%), 3 (51-75%) and 4 points (>75%). A staining score of <3 was classified as low TRIM21 expression, and those \geq 3 were classified as high TRIM21 expression.

Reverse-transcription quantitative (RT-q) PCR. TRIzol® reagent (cat. no. 15596-026; Thermo Fisher Scientific, Inc) was used for total RNA isolation according to the manufacturer's protocols. To quantify TRIM21 expression, the total RNA was reverse-transcribed into cDNA using a PrimeScript RT Reagent kit (cat. no. RR037A; Takara Bio, Inc.) according to the manufacturer's protocols, which, in turn, was subjected to qPCR analysis with a SYBR Premix Ex Taq kit (cat. no. DRR041A; Takara Bio, Inc.) according to the manufacturer's protocols. The PCR conditions were as follows: 95°C, 10 min; (95°C, 15 sec; 60°C, 60 sec) \times 40 cycles. The primer sequences were as follows: TRIM21, 5'-CCATGTGCCAGGGCTGAAGAAG-3' (forward), 5'-AGGTATGCTCTGCTGGGTGCTC-3' (reverse); β -actin, 5'-CATGGAGTCCTGTGGCATC-3' (forward), 5'-CAGGGCAGTGATCTCCTTCT-3' (reverse). All the primers were synthesized in Sangon Biotech Co., Ltd. Relative gene expression was

calculated using the $2^{-\Delta\Delta C_q}$ method (17). These experiments were replicated three times.

Public database data sources. THCA transcriptome data were downloaded from The Cancer Genome Atlas (TCGA) database using the UCSC Xena tool (<https://xena.ucsc.edu/>). TRIM21 expression levels in all types of cancer obtained from the cBioPortal database (<https://www.cbioportal.org/>) (18) were analyzed using R software version 3.6.1 (<http://www.R-project.org/>) (19). An unpaired Student's t-test was applied to compare TRIM21 expression in various cancers with that in normal tissues.

Gene set enrichment analysis (GSEA) of TRIM21-related cancer pathways. THCA transcriptome expression profiles were obtained from the TCGA database (<https://gdc-portal.nci.nih.gov/>). A total of 510 THCA samples and 59 normal tissues derived from healthy individuals were included in the present study. The correlations between the Kyoto Encyclopedia Of Genes And Genomes (KEGG) signaling pathways of TRIM21 and co-expressed genes were explored using GSEA (<http://software.broadinstitute.org/gsea/index.jsp>) (20). The latter was performed using three enrichment statistics: Enrichment scores, normalized enrichment scores and nominal P-values. The enrichment score indicates the degree of enrichment of a functional gene set before or after a given sequence. The normalized enrichment score is the major parameter in enrichment analyses of functional gene sets. The nominal P-value indicates the statistical significance of the enrichment score of a given functional gene subset with lower P-values. KEGG enrichment was reported at a significance threshold of $P < 0.05$.

Cell culture and lentivirus infection. FTC-133 cell line (cat. no. 1101HUM-PUMC000687) was purchased from the National Infrastructure of Cell Line Resource of China. The cells were cultured in Dulbecco's modified eagle medium F-12 (DMEM-F12, cat. no. 11320-033, Gibco; Thermo Fisher Scientific, Inc.) containing 10% fetal bovine serum (FBS, cat. no. 10100147, Gibco; Thermo Fisher Scientific, Inc.) in an incubator with 5% CO₂ at 37°C.

Cultured cells were seeded into 24-well plates at 30,000 cells/well. Once the cells reached 90% confluence, lentivirus (Lv-shCon or Lv-shTRIM21) was added to the wells at a multiplicity of infection (MOI) of 10. At 48 h later, infected cells were selected using 10 μ g/ml puromycin (cat. no. ST551; Beyotime Institute of Biotechnology). Virally infected cells were observed under a fluorescence microscope (cat. no. IX73; Olympus Corporation). TRIM21 expression was determined using RT-qPCR analysis. Lv-shCon and Lv-shTRIM21 were designed and constructed at Shanghai GeneChem Co., Ltd. The targeting sequence of Lv-shCon was: 5'-AACAAAGATGAAGAGCACCAAC TCGAGTTGGTGTCTCTTCATCTTGTTG-3'. The targeting sequence of Lv-shTRIM21 was: 5'-GGAAGTCACTTCACC ATCACTCGAGTGATGGTGAAGTGACTTCCTTTTTT-3'.

Transwell assay. Migration: Cells were seeded into the upper chambers of a Transwell plate. The upper chambers contained DMEM media without FBS, whereas the lower

Table I. Relationship between TRIM21 expression and clinicopathologic features of patients with thyroid papillary carcinoma.

| Pathological clinical data | Expression of TRIM21 protein in cancer tissue | | Statistical quantity | P-value |
|--------------------------------|---|---------------------------|----------------------|---------|
| | High-expression (89 cases) | Low-expression (31 cases) | | |
| Sex | | | | |
| Male | 35 | 10 | $\chi^2=0.23$ | 0.62 |
| Female | 54 | 21 | | |
| Age (years) | | | | |
| <55 | 46 | 19 | $\chi^2=0.51$ | 0.47 |
| ≥55 | 43 | 12 | | |
| Tumor diameter (cm) | | | | |
| ~1.0-2.0 | 36 | 11 | $\chi^2=0.55$ | 0.75 |
| ~2.1-3.0 | 28 | 12 | | |
| ~3.1-4.0 | 25 | 8 | | |
| Extracranular invasion | | | | |
| Yes | 60 | 11 | $\chi^2=2.78$ | 0.09 |
| No | 29 | 21 | | |
| Lymph node metastasis (pieces) | | | | |
| No | 11 | 10 | $\chi^2=12.63$ | 0.002 |
| ~1-3 | 30 | 15 | | |
| ≥4 | 48 | 6 | | |

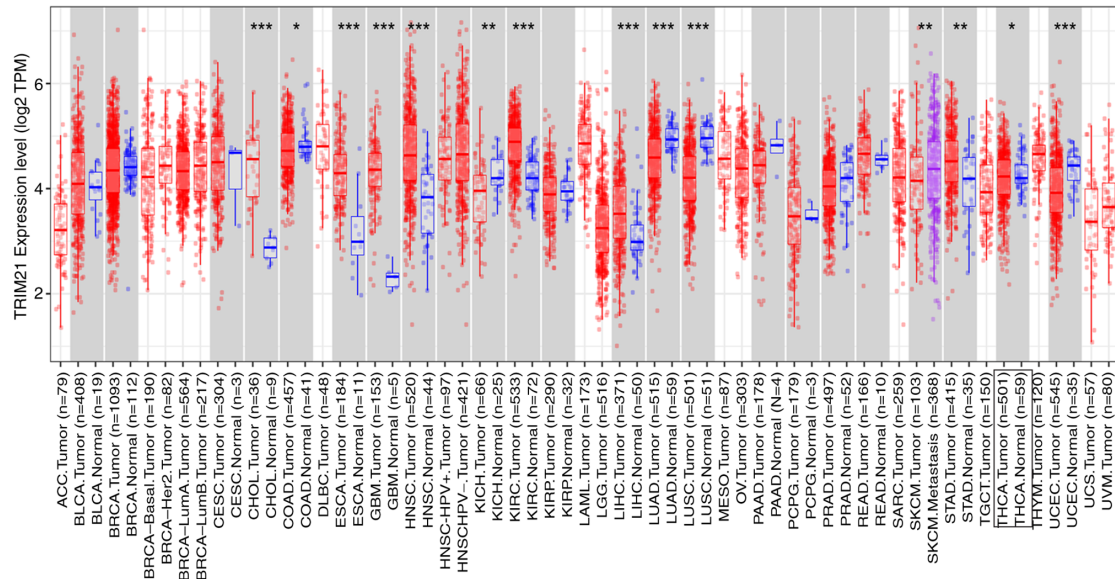


Figure 1. TRIM21 expression levels in different types of tumors. * $P<0.05$; ** $P<0.01$; *** $P<0.001$. ACC, adrenocortical carcinoma; BLCA, bladder carcinoma; BRCA, breast carcinoma; CESC, cervical squamous cell carcinoma; CHOL, cholangiocarcinoma; COAD, colon adenocarcinoma; DLBC, lymphoid neoplasm diffuse large B-cell lymphoma; ESCA, esophageal carcinoma; GBM, glioblastoma multiforme; HNSC, head and neck squamous carcinoma; KICH, kidney chromophobe; KIRC, kidney renal clear cell carcinoma; KIRC, kidney renal clear cell carcinoma; LAML, acute myeloid leukemia; LIHC, liver hepatocellular carcinoma; LUAD, lung adenocarcinoma; LUSC, lung squamous cell carcinoma; MESO, mesothelioma; OV, ovarian serous cystadenocarcinoma; PAAD, pancreatic adenocarcinoma; PCPG, pheochromocytoma and paraganglioma; PRAD, rectum adenocarcinoma; READ, rectum adenocarcinoma; SARC, sarcoma; SKCM, skin cutaneous melanoma; STAD, stomach adenocarcinoma; TGCT, testis germ cell tumors; THCA, thyroid carcinoma; THYM, thymoma; UCEC, uterine corpus endometrial carcinoma; UCS, uterine carcinosarcoma; UVM, uveal melanoma.

chambers contained DMEM media with 10% FBS. The cells were cultured for 24 h, and then the remaining cells in the upper chamber were wiped away with a swab. The cells

passed through the membrane were stained with crystal violet (cat. no. C0121; Beyotime Institute of Biotechnology). The cells in three randomly selected visual fields were

Table II. Expression levels of TRIM in various cancers vs. normal tissue included in TCGA.

| Tumor | Normal | P-value |
|---------------------|---------------------|------------------------|
| KIRC.Tumor (n=533) | KIRC.Normal (n=72) | 2.08×10^{-20} |
| LUSC.Tumor (n=501) | LUSC.Normal (n=51) | 1.22×10^{-18} |
| HNSC.Tumor (n=520) | HNSC.Normal (n=44) | 2.37×10^{-11} |
| LUAD.Tumor (n=515) | LUAD.Normal (n=59) | 8.63×10^{-9} |
| CHOL.Tumor (n=36) | CHOL.Normal (n=9) | 6.77×10^{-8} |
| LIHC.Tumor (n=371) | LIHC.Normal (n=50) | 8.37×10^{-6} |
| UCEC.Tumor (n=545) | UCEC.Normal (n=35) | 1.55×10^{-5} |
| ESCA.Tumor (n=184) | ESCA.Normal (n=11) | 6.16×10^{-5} |
| GBM.Tumor (n=153) | GBM.Normal (n=5) | 1.96×10^{-4} |
| STAD.Tumor (n=415) | STAD.Normal (n=35) | 2.54×10^{-3} |
| KICH.Tumor (n=66) | KICH.Normal (n=25) | 2.94×10^{-3} |
| THCA.Tumor (n=501) | THCA.Normal (n=59) | 1.55×10^{-2} |
| COAD.Tumor (n=457) | COAD.Normal (n=41) | 4.65×10^{-2} |
| PRAD.Tumor (n=497) | PRAD.Normal (n=52) | 9.42×10^{-2} |
| PAAD.Tumor (n=178) | PAAD.Normal (n=4) | 9.59×10^{-2} |
| BRCA.Tumor (n=1093) | BRCA.Normal (n=112) | 1.59×10^{-1} |
| KIRP.Tumor (n=290) | KIRP.Normal (n=32) | 3.10×10^{-1} |
| READ.Tumor (n=166) | READ.Normal (n=10) | 3.69×10^{-1} |
| BLCA.Tumor (n=408) | BLCA.Normal (n=19) | 5.75×10^{-1} |
| CESC.Tumor (n=304) | CESC.Normal (n=3) | 6.83×10^{-1} |
| PCPG.Tumor (n=179) | PCPG.Normal (n=3) | 9.38×10^{-1} |

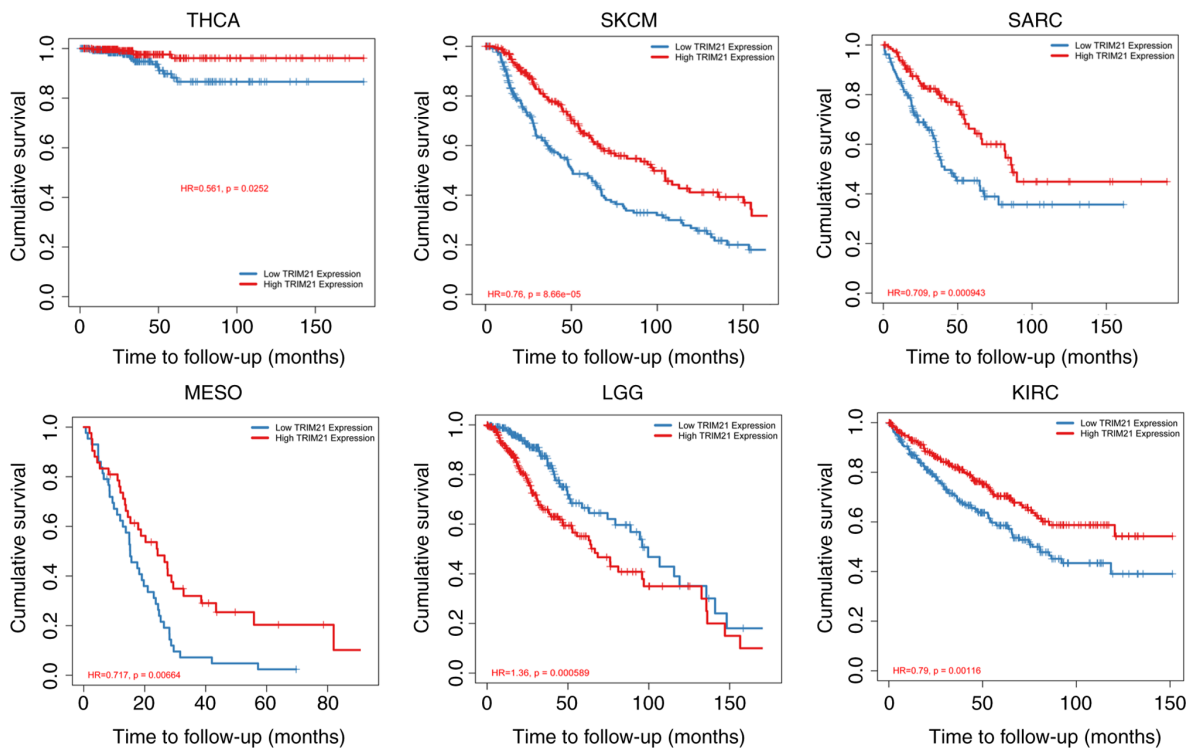


Figure 2. The Kaplan-Meier curve of prognosis is related TRIM21 expression level in different tumor samples. The blue and red curves represent the high and low TRIM21 expression sample groups, respectively. TRIM21, tripartite motif-containing 21; THCA, thyroid carcinoma; SKCM, skin cutaneous melanoma; SARC, sarcoma; MESO, mesothelioma; LGG, Low-grade glioma; KIRC, kidney renal clear cell carcinoma.

counted under a fluorescence microscope (cat. no. IX73; Olympus Corporation).

Invasion: The invasion assay was performed with the similar procedures as the migration assay, excepting that the wells were

pre-coated with 20 μ g Matrigel at 37°C for 2 h (MilliporeSigma).

CCK-8 assay. Cells were seeded into a 96 well plate at 5,000 cell/well. After the cells were cultured for 0, 24 and 48 h in an incubator with 5% CO₂ at 37°C, 10 μ l of CCK-8 reagent (cat. no. C0037; Beyotime Institute of Biotechnology) was added. The cells were then incubated for 1 h at 37°C and the absorbance at 450 nm wavelength was measured using a Multiskan GO microplate reader (Thermo Fisher Scientific, Inc.).

Statistical analysis. All pathological and experimental data were analyzed using SPSS software (version 25.0; IBM Corp.). The measurement data conformed to a normal distribution and are presented as mean \pm standard deviation (SD). The χ^2 test was performed to analyze the association between TRIM21 expression and clinicopathologic features of patients with thyroid papillary carcinoma. Considering the normal tissues derived from different patients to those who donated the cancer tissues, the expression of TRIM21 in 14 different types of tumors was evaluated using an unpaired Student's t-test. TRIM21 expression in 120 pairs THCA and the corresponding adjacent normal tissues was evaluated using paired Student's t-test. P-value was obtained from a two-tailed Student's t-test. The survival time of the patients was calculated using the Kaplan-Meier method. Log-rank test was performed to analyze the Kaplan-Meier survival curves. P<0.05 was considered to indicate a statistically significant difference.

Results

TRIM21 expression in multiple tumor sites. TRIM21 expression was assessed in various cancers based on TCGA and GTEX databases. TRIM21 expression was dysregulated in the 14 tumor types compared with the corresponding normal tissues, including kidney renal clear cell carcinoma (KIRC), lung squamous cell carcinoma (LUSC), head and neck squamous carcinoma (HNSC), lung adenocarcinoma (LUAD), cholangiocarcinoma (CHOL), liver hepatocellular carcinoma (LIHC), uterine corpus endometrial carcinoma (UCEC), esophageal carcinoma (ESCA), glioblastoma multiforme (GBM), stomach adenocarcinoma (STAD), kidney chromophobe (KICH), skin cutaneous melanoma (SKCM) tumor and THCA. (Fig. 1 and Table II).

Association of TRIM21 with the survival of patients with THCA. In the TCGA data set, according to TRIM21 expression, tumors were divided into Low TRIM21 group and High TRIM21 group. Fig. 2 and Table I show the survival analysis of the two groups. In low grade gliomas, the low TRIM21 group showed a better prognosis than that in the High TRIM21 group. In contrast, in SKCM, mesothelioma (MESO), sarcoma (SARC), KIRC, and THCA, the patients in the high TRIM21 group showed a better prognosis compared with that in the low TRIM21 group.

TRIM21 expression in THCA and normal tissues. The expression of TRIM21 in 120 THCA and corresponding para-cancer normal tissues was evaluated using IHC. As shown in Fig. 3A, the staining intensity of TRIM21 in THCA

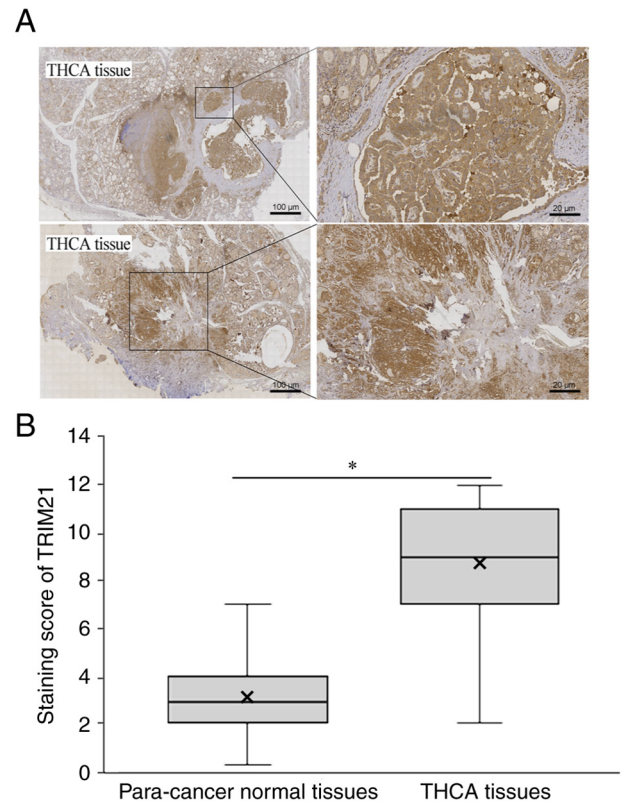


Figure 3. Expression of TRIM21 in THCA. (A) IHC staining of TRIM21 in THCA and para-cancer normal tissues. Magnification: Left, x40; Right, x200. (B) The IHC staining score of TRIM21 protein in THCA and para-cancer normal tissues. *P<0.05. TRIM21, tripartite motif-containing 21; THCA, thyroid carcinoma; IHC, immunohistochemistry.

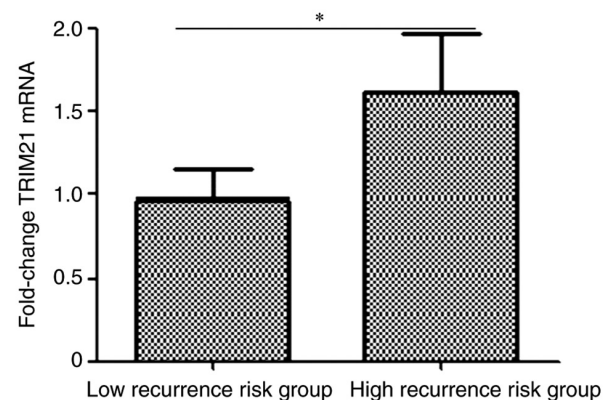


Figure 4. Expression of TRIM21 mRNA in high- and low recurrence-risk group. *P<0.05. TRIM21, tripartite motif-containing 21.

tissues was significantly higher than that in para-cancer normal tissues. Simultaneously, the staining score of TRIM21 protein was calculated, and the score in THCA tissues was significantly higher than that in para-cancer normal tissues (Fig. 3B). These results indicate that TRIM21 is overexpressed in THCA.

Relevance of TRIM21 in clinicopathological features of THCA cases. According to TRIM21 expression, 120 THCA cases were divided into TRIM21 high- and low-expression groups. Subsequently, the relevance of TRIM21 to the clinicopathologic features of THCA was evaluated. As shown in

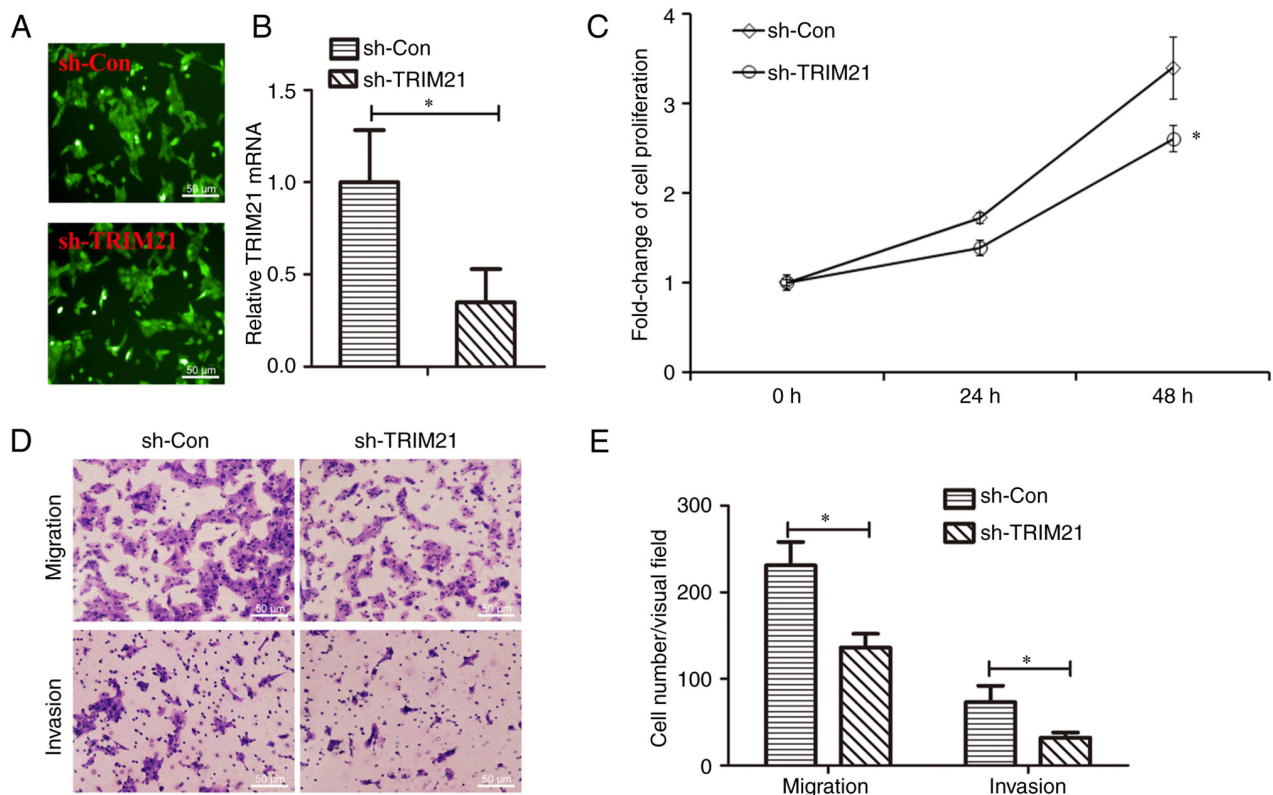


Figure 5. Knockdown of TRIM21 induces inhibition of cell migration and invasion of THCA cells. (A) After lentivirus infection, cells were observed under an inverted fluorescence microscope (magnification, x100). (B) TRIM21 mRNA were detected using reverse transcription-quantitative PCR, * $P < 0.05$ vs. sh-Con. (C) Cell proliferation was measured by CCK-8 assay. (D) Cells were determined using a Transwell plate and observed under a microscope (magnification, x100). (E) Cells counts obtained from three random visual fields, * $P < 0.05$ vs. sh-Con. TRIM21, tripartite motif-containing 21; THCA, thyroid carcinoma; sh-Con, short hairpin control.

Table I, the expression of TRIM21 showed no relevance to patients' gender, age, tumor diameter, and extra-granular invasion; however, TRIM21 was significantly associated with lymph node metastasis. According to the recurrence risk based on the American Thyroid Association (ATA) guidelines 2021 (21), 120 patients were divided into high- and low recurrence-risk groups. As shown in Fig. 4, the expression of TRIM21 was measured using RT-qPCR and the expression of TRIM21 in the high recurrence-risk group was 1.69 folds of that in the low recurrence-risk group ($P = 0.0242$).

Knockdown of TRIM21 induced inhibition of cell proliferation, migration and invasion of THCA cells. As TRIM21 was overexpressed in THCA and was associated with lymph node metastasis, the role of TRIM21 in THCA cell migration and invasion was further examined. TRIM21 was knocked down in lentivirus-infected FTC-133 cells. As shown in Fig. 5A, green fluorescent labeling indicated that the cells were infected with lentivirus. Fig. 5B demonstrated that TRIM21 expression was reduced ~62.88% in Lv-shTRIM21 infected cells, indicating that the efficiency of TRIM21 knockdown was 37.12%. Cell migration and invasion capacities were measured using Transwell assays. The migration and invasion capacities of Lv-shTRIM21-infected cells were inhibited compared to those infected with Lv-shCon (Fig. 5C, D and E). These results indicate that TRIM21 knockdown inhibits proliferation, migration and invasion of THCA cells.

KEGG analysis of TRIM21 and TRIM21 co-expression genes in THCA. The potential biological functions of TRIM21 with high or low expression in THCA were investigated using GSEA, and the genes were significantly enriched in 38 KEGG pathways, including 'butanoate metabolism', 'oxidative phosphorylation', and 'valine leucine and isoleucine degradation' (Fig. 6).

After excluding the genes that met the condition of false discovery rate < 0.05 and with a correlation coefficient > 0.6 were included. A total of 252 TRIM21 co-expression genes were identified, including SP110, APOL2, and UBE2L6. Furthermore, the KEGG database was used to screen for TRIM21 co-expression gene enrichment pathways in thyroid carcinoma. As shown in Fig. 7A, the genes were significantly enriched in seven KEGG pathways. The top three were 'antigen processing and presentation', 'autoimmune thyroid disease', and 'cell adhesion molecules'. The enrichment of co-expression genes in the transcription factor (TF) and kinase datasets were further investigated. As shown in Fig. 7B, a total of 14 TFs of TRIM21 co-expressed genes were identified, including NFKAPPAB, NFKB, and IRF2.

Discussion

THCA is one of the most common endocrine malignancies worldwide. Dysregulation of TRIM21 is responsible for the progression of various diseases, including tumors. However,

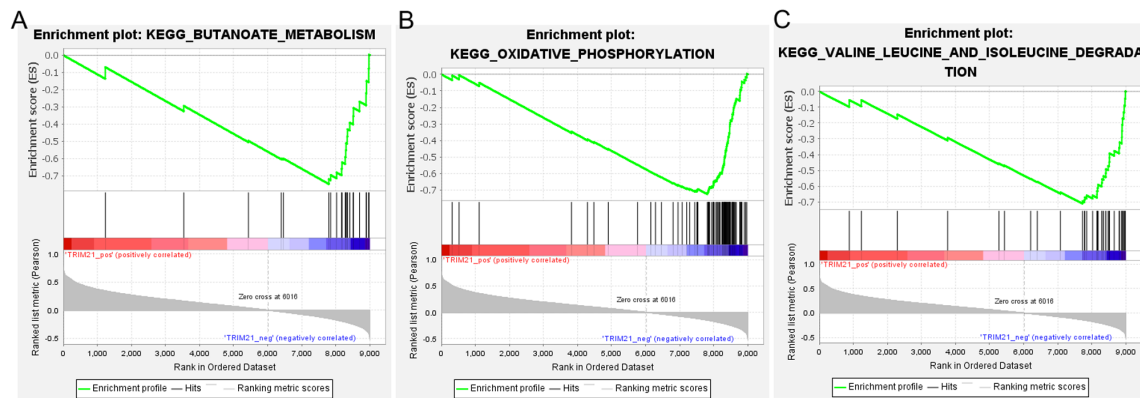


Figure 6. ES-plots of significantly related KEGG signalling pathways. (A) 'butanoate metabolism'; (B) 'oxidative phosphorylation'; and (C) 'valine leucine and isoleucine degradation'.

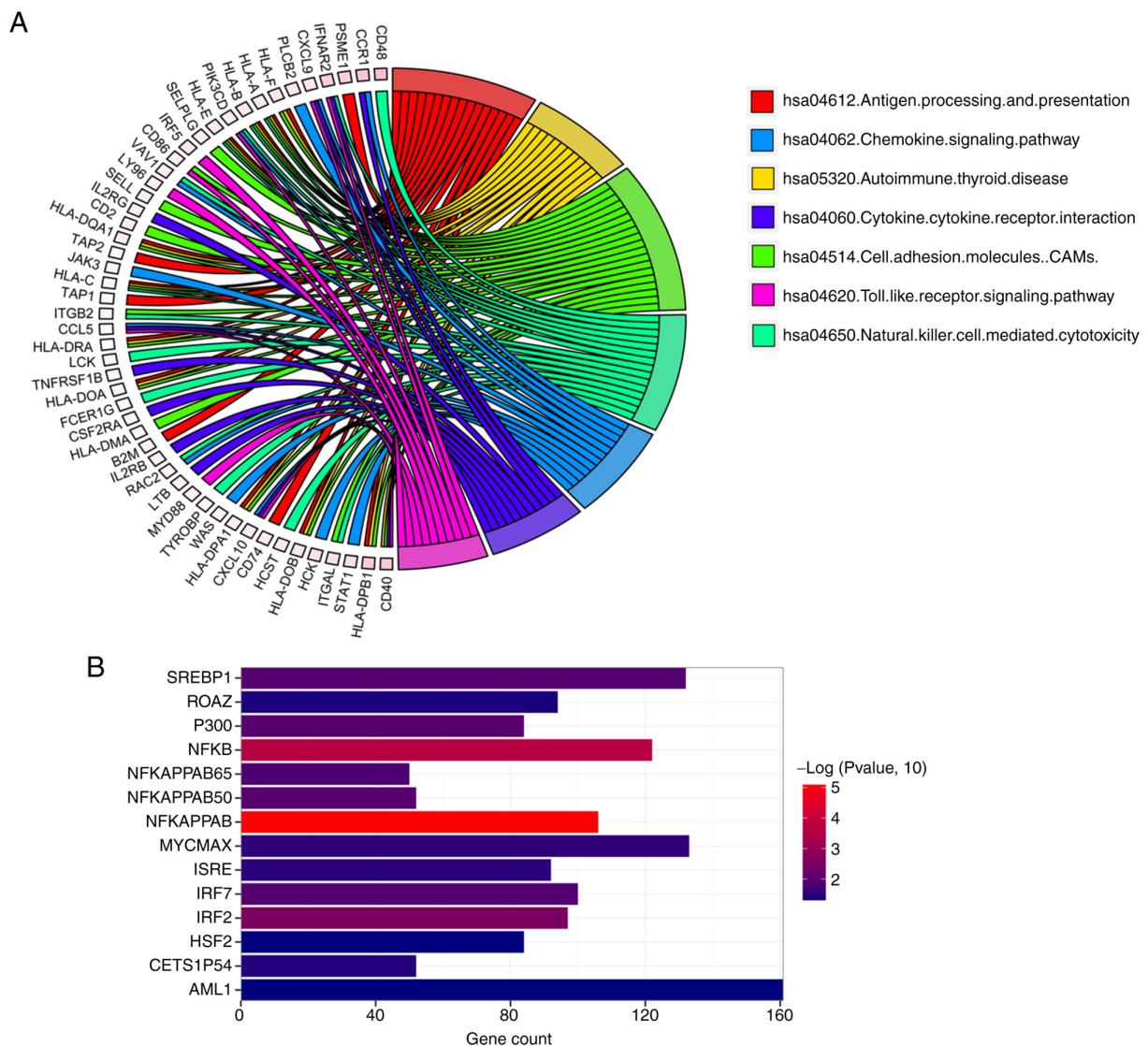


Figure 7. Significantly enriched KEGG signaling pathways and TF of genes associated with TRIM21 expression. (A) Significantly enriched KEGG signalling pathway group of genes that are significantly associated with TRIM21 expression. (B) TF of genes significantly correlated with TRIM21 expression. The number of genes is indicated on the x-axis, and the TF is shown on the y-axis. The significance level is indicated with colour codes, with red representing the highest significance. KEGG, Kyoto Encyclopedia of Genes and Genomes; TF, transcription factors; TRIM21, tripartite motif-containing 21.

limited information is available regarding the potential contribution of TRIM21 to THCA.

The current study found, using bioinformatics analysis, that TRIM21 was upregulated in THCA. In addition, the

results of the bioinformatics analyses were verified by measuring TRIM21 expression in THCA and matched adjacent normal tissues. Higher TRIM21 expression was observed in THCA tissues compared with matched adjacent normal tissues. Furthermore, a high TRIM21 level was associated with a high risk of recurrence and lymph node metastasis. The results indicated that TRIM21 may be a potential biological marker to distinguish tumor recurrence rates.

TRIM21 expression and its role in various cancers have been previously investigated. The effects of TRIM21 on tumor progression differed in different types of cancer. Zhao *et al* (14) observed TRIM21 upregulation in gliomas and confirmed its role in tumor proliferation, migration, and drug resistance. By contrast, TRIM21 is downregulated in breast cancer, associated with tumor size and clinical stage, and is considered an important factor for overall survival (22). In patients with colitis-associated colorectal cancer, decreased TRIM21 expression causes dysregulation of epithelial cell proliferation, angiogenesis and pro-inflammatory responses, resulting in intestinal epithelial carcinogenesis (23). The present study investigated the role of TRIM21 in THCA progression *in vitro*. It was observed that TRIM21 knock-down inhibited THCA cell proliferation, migration and invasion. This may be one of the biological involvements of TRIM21 in the high recurrence risk and lymph node metastasis of THCA.

TRIM21 may destabilize the tumor suppressor protein p53, the disruption of which often leads to cancer development (22). TRIM21 can degrade p27 and enable cells to enter the S phase, leading to tumor progression (24). By contrast, TRIM21 negatively regulates anti-apoptotic proteins and inactivates the glycogen synthase kinase-3 β (GSK3 β)-NF- κ B pathway to initiate cell apoptosis (25). In addition, increased TRIM21 expression increases the activation of caspase-8 and enhances the death receptor-mediated apoptosis (26). In the present study, 252 TRIM21 co-expressed genes, including SP110, APOL2, and UBE2L6, were identified. These significantly enriched genes were associated with the 'antigen processing and presentation', 'autoimmune thyroid disease' and 'cell adhesion molecules' pathways. The expression of TRIM21 co-expression genes in THCA may be influenced by 14 TFs, including NFKAPPAB, NF- κ B, and IRF2. However, the mechanism by which TRIM21 regulates THCA progression remains to be elucidated.

There are several members of the TRIM family. TRIM14 has been reported as an oncogene in THCA (7). TRIM44 knockdown suppresses the tumor progression of THCA by inhibiting the Wnt/ β -catenin signaling pathway (8). Although others are also of concern, the current study focused on TRIM21. Further studies on the expression and function of other members of the TRIM family in THCA progression are required to establish the regulation network of the TRIM family in THCA.

In conclusion, using bioinformatics analysis and an *in vitro* study, the present study revealed that TRIM21 promoted tumor progression, indicating that TRIM21 may be a potential biomarker and therapeutic target for THCA. In the future, the mechanism by which TRIM21 regulates THCA progression will be further investigated.

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Availability of data and materials

The datasets generated and/or analyzed during the current study are available in the Figshare repository, https://figshare.com/articles/figure/Untitled_Item/19501369.

Authors' contributions

ZW and JF designed the study and wrote the manuscript. ZW and YW participated in performing the experiments. ZM was responsible for data acquisition and the interpretation of data. ZY was responsible for statistical analysis and the literature search. WD participated in collecting the tissue samples, performing the RT-qPCR experiments and revising the manuscript. All authors read and approved the final manuscript. ZW and ZM confirm the authenticity of all the raw data.

Ethics approval and consent to participate

This study has been checked and approved by the Ethics committee of Liaocheng People's Hospital (approval no. LC2021059).

Patient consent for publication

Not applicable.

Competing interests

All the authors declare that they have no competing interests.

References

1. Du L, Zhao Z, Zheng R, Li H, Zhang S, Li R, Wei W and He J: Epidemiology of thyroid cancer: Incidence and mortality in China, 2015. *Front Oncol* 10: 1702, 2020.
2. Takami H, Ito Y, Okamoto T and Yoshida A: Therapeutic strategy for differentiated thyroid carcinoma in Japan based on a newly established guideline managed by Japanese society of thyroid surgeons and Japanese association of endocrine surgeons. *World J Surg* 35: 111-121, 2011.
3. Hatakeyama S: TRIM family proteins: Roles in autophagy, immunity, and carcinogenesis. *Trends Biochem Sci* 42: 297-311, 2017.
4. Vunjak M and Versteeg GA: TRIM proteins. *Curr Biol* 29: R42-R44, 2019.
5. Valletti A, Marzano F, Pesole G, Sbisà E and Tullo A: Targeting chemoresistant tumors: Could TRIM proteins-p53 axis be a possible answer? *Int J Mol Sci* 20: 1776, 2019.

6. Alomari M: TRIM21-A potential novel therapeutic target in cancer. *Pharmacol Res* 165: 105443, 2021.
7. Sun W, Wang Y, Li D, Wu Y, Ji Q and Sun T: Tripartite motif containing 14: An oncogene in papillary thyroid carcinoma. *Biochem Biophys Res Commun* 521: 360-367, 2020.
8. Zhou Z, Liu Y, Ma M and Chang L: Knockdown of TRIM44 inhibits the proliferation and invasion in papillary thyroid cancer cells through suppressing the Wnt/ β -catenin signaling pathway. *Biomed Pharmacother* 96: 98-103, 2017.
9. Liu Y, Zhang B, Shi T and Qin H: miR-182 promotes tumor growth and increases chemoresistance of human anaplastic thyroid cancer by targeting tripartite motif 8. *OncoTargets Ther* 10: 1115-1122, 2017.
10. Oke V and Wahren-Herlenius M: The immunobiology of Ro52 (TRIM21) in autoimmunity: A critical review. *J Autoimmun* 39: 77-82, 2012.
11. Simoes Eugénio M, Faurez F, Kara-Ali GH, Lagarrigue M, Uhart P, Bonnet MC, Gallais I, Com E, Pineau C, Samson M, *et al*: TRIM21, a new component of the TRAIL-induced endogenous necrosome complex. *Front Mol Biosci* 8: 645134, 2021.
12. Zhang Z, Zhu Z, Sheng H, Sun J and Cao C: TRIM21 suppresses invasion of hepatocellular carcinoma cells by promoting beta-catenin ubiquitylation and degradation. *Nan Fang Yi Ke Da Xue Xue Bao* 42: 55-62, 2022 (In Chinese).
13. Chen X, Li Z, Yong H, Wang W, Wang D, Chu S, Li M, Hou P, Zheng J and Bai J: Trim21-mediated HIF-1 α degradation attenuates aerobic glycolysis to inhibit renal cancer tumorigenesis and metastasis. *Cancer Lett* 508: 115-126, 2021.
14. Zhao Z, Wang Y, Yun D, Huang Q, Meng D, Li Q, Zhang P, Wang C, Chen H and Lu D: TRIM21 overexpression promotes tumor progression by regulating cell proliferation, cell migration and cell senescence in human glioma. *Am J Cancer Res* 10: 114-130, 2020.
15. Ping M, Wang S, Guo Y and Jia J: TRIM21 improves apatinib treatment in gastric cancer through suppressing EZH1 stability. *Biochem Biophys Res Commun* 586: 177-184, 2022.
16. Chuang CY, Chien YC, Lin CW, Chou CH, Chen SC, Liu CL, Bai LY, Yang SF and Yu YL: TRIM21 polymorphisms are associated with susceptibility and clinical status of oral squamous cell carcinoma patients. *Int J Med Sci* 18: 2997-3003, 2021.
17. Livak KJ and Schmittgen TD: Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta C(T)) method. *Methods* 25: 402-408, 2001.
18. Gao J, Aksoy BA, Dogrusoz U, Dresdner G, Gross B, Sumer SO, Sun Y, Jacobsen A, Sinha R, Larsson E, *et al*: Integrative analysis of complex cancer genomics and clinical profiles using the cBioPortal. *Sci Signal* 6: pii, 2013.
19. Shim SR, Kim SJ, Lee J and Rucker G: Network meta-analysis: Application and practice using R software. *Epidemiol Health* 41: e2019013, 2019.
20. Subramanian A, Tamayo P, Mootha VK, Mukherjee S, Ebert BL, Gillette MA, Paulovich A, Pomeroy SL, Golub TR, Lander ES and Mesirov JP: Gene set enrichment analysis: A knowledge-based approach for interpreting genome-wide expression profiles. *Proc Natl Acad Sci USA* 102: 15545-15550, 2005.
21. Bible KC, Kebebew E, Brierley J, Brito JP, Cabanillas ME, Clark TJ Jr, Di Cristofano A, Foote R, Giordano T, Kasperbauer J, *et al*: 2021 American thyroid association guidelines for management of patients with anaplastic thyroid cancer. *Thyroid* 31: 337-386, 2021.
22. Guha A, Ahuja D, Das Mandal S, Parasar B, Deyasi K, Roy D, Sharma V, Willard B, Ghosh A and Ray PS: Integrated regulation of HuR by translation repression and protein degradation determines pulsatile expression of p53 under DNA damage. *iScience* 15: 342-359, 2019.
23. Zhou G, Wu H, Lin J, Lin R, Feng B and Liu Z: TRIM21 is decreased in Colitis-associated cancer and negatively regulates epithelial carcinogenesis. *Inflamm Bowel Dis* 27: 458-468, 2021.
24. Sabile A, Meyer AM, Wirbelauer C, Hess D, Kogel U, Scheffner M and Krek W: Regulation of p27 degradation and S-phase progression by Ro52 RING finger protein. *Mol Cell Biol* 26: 5994-6004, 2006.
25. Gao X, Xu F, Zhang HT, Chen M, Huang W, Zhang Q, Zeng Q and Liu L: PKC α -GSK3 β -NF- κ B signaling pathway and the possible involvement of TRIM21 in TRAIL-induced apoptosis. *Biochem Cell Biol* 94: 256-264, 2016.
26. Zhang J, Fang L, Zhu X, Qiao Y, Yu M, Wang L, Chen Y, Yin W and Hua ZC: Ro52/SSA sensitizes cells to death receptor-induced apoptosis by down-regulating c-FLIP(L). *Cell Biol Int* 36: 463-468, 2012.



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