

# Methylation levels in the *GRIN2B* promoter region are associated with bipolar disorder and its anxiety and insomnia symptoms

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**Abstract.** This study investigated the potential role of promoter-region DNA methylation in *GRIN2B* in bipolar disorder (BD) and its association with anxiety and insomnia symptoms among patients with BD. The DNA methylation levels in the *GRIN2B* promoter region were quantified using MassARRAY technology in peripheral blood samples from 31 patients with BD and 32 healthy controls. To assess associations, binary logistic regression and partial correlation analyses were performed to examine potential links between *GRIN2B* methylation, BD diagnosis and symptom severity. The exploratory analysis revealed six differentially methylated CpG sites between groups, four of which exhibited hypermethylation in patients with BD compared with the controls. Significant positive correlations were observed between Hamilton Anxiety Rating Scale scores and CpG9 methylation ( $r=0.408$ ,  $P=0.038$ ), as well as between Pittsburgh Sleep Quality Index scores and CpG8 methylation ( $r=0.419$ ,  $P=0.033$ ). These findings provide preliminary evidence supporting an association between *GRIN2B* methylation levels and BD, as well as related clinical symptoms. However, given the small sample size and the exploratory nature of this study, these results should be interpreted with caution. Further replication in larger, longitudinal cohorts is warranted to confirm these observations and evaluate their clinical implications.

## Introduction

Bipolar disorder (BD) is associated with severe mental disability and an increased risk of suicide, significantly contributing to the societal burden (1,2). Insomnia and anxiety, two common concomitant symptoms of BD, often persist throughout the

course of the illness. Approximately 45% of individuals with BD will experience co-morbid anxiety disorders over their lifetime (3). This comorbidity further exacerbates the risk of substance dependence and suicide (4), worsening social functioning and quality of life in individuals with BD (5). Crucially, anxiety symptoms have become a key target for early intervention in individuals at risk of BD (6), anxiety in BD is associated with an increased risk of subjective sleep disturbances (7). Sleep, as a key regulator of both metabolic homeostasis and oxidative stress, plays a particularly significant role; when disrupted, it can contribute to the onset and progression of BD. Given their role as cross-diagnostic precursors, insomnia and anxiety represent high-priority therapeutic targets for preventing BD episodes (8,9).

The N-methyl-D-aspartate receptor subunit 2B (*GRIN2B*) gene, which is located at 12p13.1, spans 419 kilobases and consists of 15 exons. N-methyl-D-aspartate receptors (NMDARs), a class of ionotropic glutamate receptors, are involved in regulating neuronal activity, synaptic plasticity and excitatory transmission, with substantial implications for the pathophysiology and treatment of affective disorders (10). The *GRIN2B* gene encodes the NMDAR subunit 2B (NR2B), which is crucial for determining both the structure and functional dynamics of NMDARs; overexpression of NR2B leads to long-lasting enhancement and increased synaptic efficacy (11). Light, which is the primary driver of sleep rhythm regulation (12), triggers numerous intracellular cascades through NMDARs in the suprachiasmatic nucleus (SCN), ultimately affecting the expression of clock genes (13) studies have demonstrated that circadian rhythm disturbances in mice correlate with reduced expression of the NR2B subunit of NMDARs in the SCN (14,15). Additionally, rats subjected to maternal separation showed a significant increase in anxiety, accompanied by upregulated expression of the *GRIN2B* gene (16), while ethanol-exposed mice displayed anxiety-like behaviors with elevated levels of *GRIN2B* mRNA expression in the cerebellum (17).

DNA methylation, one of the most stable epigenetic mechanisms, is increasingly recognized as a potential biomarker for numerous neuropsychiatric disorders (18) and various malignancies (19). It plays a significant role in the pathophysiology of BD (20) and may serve as a biomarker for variability in BD treatment response (21). Numerous studies have identified significant DNA methylation changes in patients with BD, particularly in regions such as the prefrontal cortex and

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peripheral leukocytes (22-24), with potential links to suicidal behavior (25), cognitive impairment (26) and substance dependence (27). Emerging evidence from cancer epigenetics demonstrates that methylation patterns can effectively stratify disease subtypes and predict clinical outcomes (28,29), suggesting similar precision medicine applications could be explored in BD. However, few studies have specifically examined the associations of DNA methylation with anxiety and insomnia in BD, despite established methodological frameworks for epigenetic analysis in other neuropsychiatric conditions, such as depression (30) and schizophrenia (31).

Based on these findings, *GRIN2B* epigenetics appears to play a significant role in the development of BD and related anxiety and insomnia symptoms. However, the role of *GRIN2B* in BD remains controversial; a study detected no disease-associated variants in glutamatergic genes (including *GRIN2B*) through targeted sequencing in patients with BD (32), whereas another study reported associations between *GRIN2B* genotypes and both psychotic symptoms and disease relapse in BD (33). Research on *GRIN2B* gene inheritance as it relates to anxiety and insomnia has been largely restricted to animal models (34,35). Therefore, to further clarify the relationship between *GRIN2B* gene inheritance (particularly DNA methylation) and BD, as well as its potential role in anxiety and insomnia among patients with BD, DNA methylation levels were measured in the *GRIN2B* gene's promoter region in peripheral blood leukocytes from patients with BD and healthy controls via the MassARRAY method and the possibility of an association was analyzed.

## Patients and methods

**Participants and procedure.** A total of 31 patients diagnosed with BD (in the depressive phase) were recruited from the inpatient units of the Clinical Psychology Department at the People's Hospital of Xinjiang Uygur Autonomous Region (Urumqi, China) between April and December 2023, while 32 healthy controls (HCs) were concurrently enrolled from hospital staff and students at the affiliated Medical University during the same study period. All participants were aged 18-55 years. The upper age limit was set to 55 years to minimize age-related confounding factors. This threshold aligns with evidence that epigenetic drift accelerates after age 55 (36), age-associated methylation changes increasingly interact with inflammatory/metabolic pathways beyond age 50 (37) and neurodegenerative comorbidities in older populations may introduce spurious associations (38). BD diagnosis was confirmed using the Diagnostic and Statistical Manual of Mental Disorders, Fourth Edition, Text Revision (DSM-IV-TR) criteria (39), based on structured clinical interviews conducted by two senior psychiatrists. Eligible patients met the following criteria: i) No medication, psychotherapy or physical therapy for  $\geq 6$  months before recruitment; ii) Hamilton Depression Rating Scale (HAMD)-24 [specifying the 24-item Hamilton Depression Rating Scale (HAMD-24); total score range: 0-76] to distinguish clinical severity thresholds from the 17-item version (HAMD-17), total score  $>20$  (40); and iii) Young Mania Rating Scale (YMRS) total score  $<7$  (41). The exclusion criteria (applied to all participants) were as follows: i) Comorbid psychiatric disorders (DSM-IV-TR axis I/II) (42);

ii) history of organic brain disease or severe systemic illness; iii) pregnancy or lactation (women); and iv) substance abuse/dependence (drugs, alcohol or psychoactive substances). This study was reviewed and approved by the Ethics Committee of the People's Hospital of Xinjiang Uygur Autonomous Region (Urumqi, China; approval no. KY2023020968). All subjects volunteered to participate in the study and provided written informed consent prior to the study.

**Data collection.** Demographic data, including age, gender, education level and history of smoking and alcohol use, were collected from all participants. Clinical data, such as age at onset, illness duration, subtype, HAMD-24 score, depression severity, YMRS score, 14-item Hamilton Anxiety Scale (HAMA-14) (43) score and Pittsburgh Sleep Quality Index (PSQI) (44) score, were obtained from patients only.

**DNA extraction and bisulfite conversion.** Peripheral fasting venous blood samples (~5 ml) were collected from patients with BD using EDTA anticoagulant tubes (BD Biosciences) within 24 h of hospital admission, prior to the initiation of any new pharmacotherapy. Healthy control samples were obtained the following morning after enrollment. Genomic DNA was subsequently extracted using the solution-based Genenode DNA extraction kit (Wuhan Genenode Biotech) according to the manufacturer's protocol and stored at  $-80^{\circ}\text{C}$ . Quality control assessments included spectrophotometric analysis for determining DNA concentration and agarose gel electrophoresis to verify DNA integrity. Following extraction, the isolated DNA underwent modification and purification using the EZ DNA Methylation-Gold™ Kit (Zymo Research Corp.) according to the manufacturer's protocol. During this process, cytosine residues that were non-methylated were deaminated to uracil, while methylated cytosine residues remained stable.

### DNA methylation assay

**Primer design.** The CpG island prediction website was utilized to predict potential CpG islands in the *GRIN2B* promoter region gene sequences, identifying two CpG islands (Fig. 1A). The Agena EpiDesigner program (<http://www.epidesigner.com>) was used to design primers for the target *GRIN2B* sequences (Fig. 1B), and a fragment with a high level of CpG methylation (#14 in Fig. 2A) was selected as the primer sequence (Fig. 2A and B). Details of the primer sequences are provided in Table I.

**Methylation assay of *GRIN2B*.** The bisulfite-converted *GRIN2B* DNA was amplified by PCR using the primers mentioned above. Optimal amplification conditions were achieved through digestion with shrimp alkaline phosphatase, transcriptional cleavage and resin-based purification. The purified PCR product was then transferred to a 384-well SpectroCHIP® bioarray (Axygen; Corning, Inc.) for precise spot sampling using the Agena Nanodispenser RS1000 spotter (Agena Bioscience). The spotted bioarray was analyzed by matrix-assisted laser desorption/ionization time-of-flight mass spectrometry with the MassARRAY Analyzer 4.0 (Axygen; Corning, Inc.) to generate mass spectra. Methylation levels were then quantified from these spectra using EpiTyper 1.2 software (Sequenom).



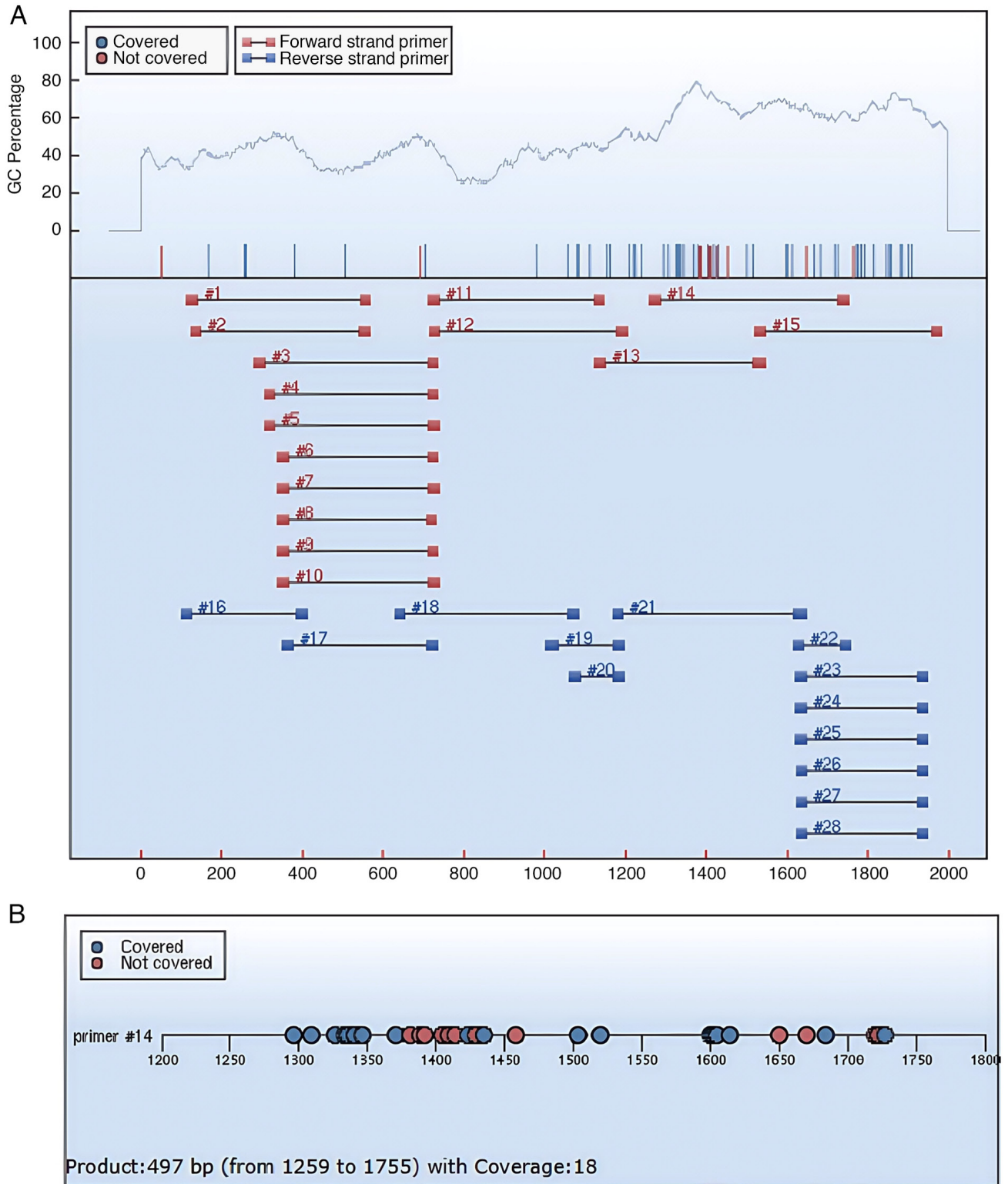


Figure 2. Target gene fragments. (A) Candidate Programs for Targeted Regions at a Glance. (B) Program 14 information: The sequences shown in the figure are forward sequences. Dots indicate CpG sites, with blue-marked CpG sites representing those that can be detected. Red-labeled CpG sites, however, cannot be detected due to issues with the sequence.

were presented. The BD group included 31 participants [8 male (25.8%), 23 female (74.2%)] with a median age of 23 years [interquartile range (IQR), 20-39], while the HC group comprised 32 individuals [11 male (34.4%), 21 female (65.6%)] with a median age of 26.5 years (IQR, 22.0-37.5). No statistically significant differences were observed between the

groups in terms of age, years of education, gender or smoking and drinking history ( $P > 0.05$ ).

#### *GRIN2B* DNAm and BD

*Altered GRIN2B DNAm patterns in patients with BD.* In the present study, the DNA methylation levels of 12 CpG sites in

Table I. Primer information.

Primer	5' end primer sequence	3' end primer sequence
Start, bp	1,259	1,755
Size, bp	28	27
Tm, °C	58.14	57.06
GC%	25	22
Cs	4	5
Original sequence	CTGATTTATGGAAAATACAGCAAGGGTC	TCTGGATTTGGGTCTCACACTCAAAAA
Methylation sequence	TTGATTTATGGAAAATATAGTAAGGGTT	TCTAAATTTAAATCTCACACTCAAAAA
Insert sequence	aggaagagag	cagtaatcagactcactatagggagaaggct

Product size, 497; no. of CpGs, 30; coverage, 18. Tm, melting temperature.

Table II. Demographic and clinical information of BD and HC groups.

Item	BD (n=31)	HC (n=32)	$\chi^2/t/Z$	P-value
Age, years	23.0 (20.0-39.0)	26.5 (22.0-37.5)	-1.343	0.179
Education, years	16.0 (15.0-16.0)	16.0 (16.0-16.0)	-0.748	0.455
Sex, male/female	(8,23)	(11,21)	0.207	0.649
Smoking, yes/no	(6,25)	(8,24)	0.290	0.590
Drinking, yes/no	(6,25)	(7,25)	0.061	0.805
Age of onset, years	20 (16.58-31.00)	-	-	-
Disease duration, years	4 (1.42-8.00)	-	-	-
HAMD-24	32 (23-36)	-	-	-
YMRS	2 (2-3)	-	-	-
HAMA-14	19 (15-29)	-	-	-
PSQI	14.32±4.05	-	-	-
Subtype (BDI/BDII)	(13,18)	-	-	-
Depression severity (moderate/severe)	(18,13)	-	-	-

Age, education, age of onset, disease duration, HAMD-24, YMRS, HAMA-14 and PSQI are expressed as the median (interquartile range). HAMD-24 (total score range: 0-76 to distinguish clinical severity thresholds from the HAMD-17) severity criteria: 20-34, moderate (n=18); ≥35, severe (n=13). BD, bipolar disorder; HC, healthy controls; HAMD, Hamilton Depression Rating Scale; YMRS, Young Mania Rating Scale; HAMA, Hamilton anxiety scale; PSQI, Pittsburgh Sleep Quality Index scale.

the *GRIN2B* promoter region were compared between patients with BD and HCs. An independent-samples t-test showed that 6 out of the 12 CpG sites (50%) had statistically significant differences in DNA methylation levels between the groups (Fig. 3). Specifically, CpG3, CpG5, CpG10 and CpG12 showed hypermethylation, while CpG1 and CpG7 showed hypomethylation in patients with BD compared to HCs. Among these, CpG3 (BD: 52.52±10.00%; HC: 32.60±25.70%) and CpG1 (BD: 36.03±13.94%; HC: 19.15±12.33%) exhibited the most significant differences in methylation levels between the two groups (P<0.001).

*Association of BD with GRIN2B methylation.* The predictive role of *GRIN2B* DNA methylation in the development of BD was analyzed using a binary logistic regression model with stepwise forward selection. Age, gender and years of education were included as covariates and the final model chosen through stepwise selection yielded a chi-square test

value of  $\chi^2=44.49$  (P<0.001), indicating model significance. The Hosmer-Lemeshow test result was  $\chi^2=3.896$  (P=0.866), suggesting a good fit. The model identified four variables associated with the likelihood of BD development, specifically the hypermethylation of CpG3, CpG5, CpG10 and CpG12. Among these, CpG3 [P=0.002, odds ratio (OR)=1.079, 95% CI: 1.029-1.132] and CpG10 (P=0.002, OR=1.113, 95% CI: 1.042-1.190) demonstrated a significant predictive role (Table III).

*Association of GRIN2B DNA methylation with anxiety and insomnia in BD.* Partial correlation analysis (Table IV) revealed that the HAMA-14 score was positively associated with methylation levels of CpG9 (r=0.408, P=0.038) after controlling for age, gender and years of education (r=0.419, P=0.033). Similarly, the PSQI score showed a positive correlation with methylation levels of CpG8 at the *GRIN2B* locus.

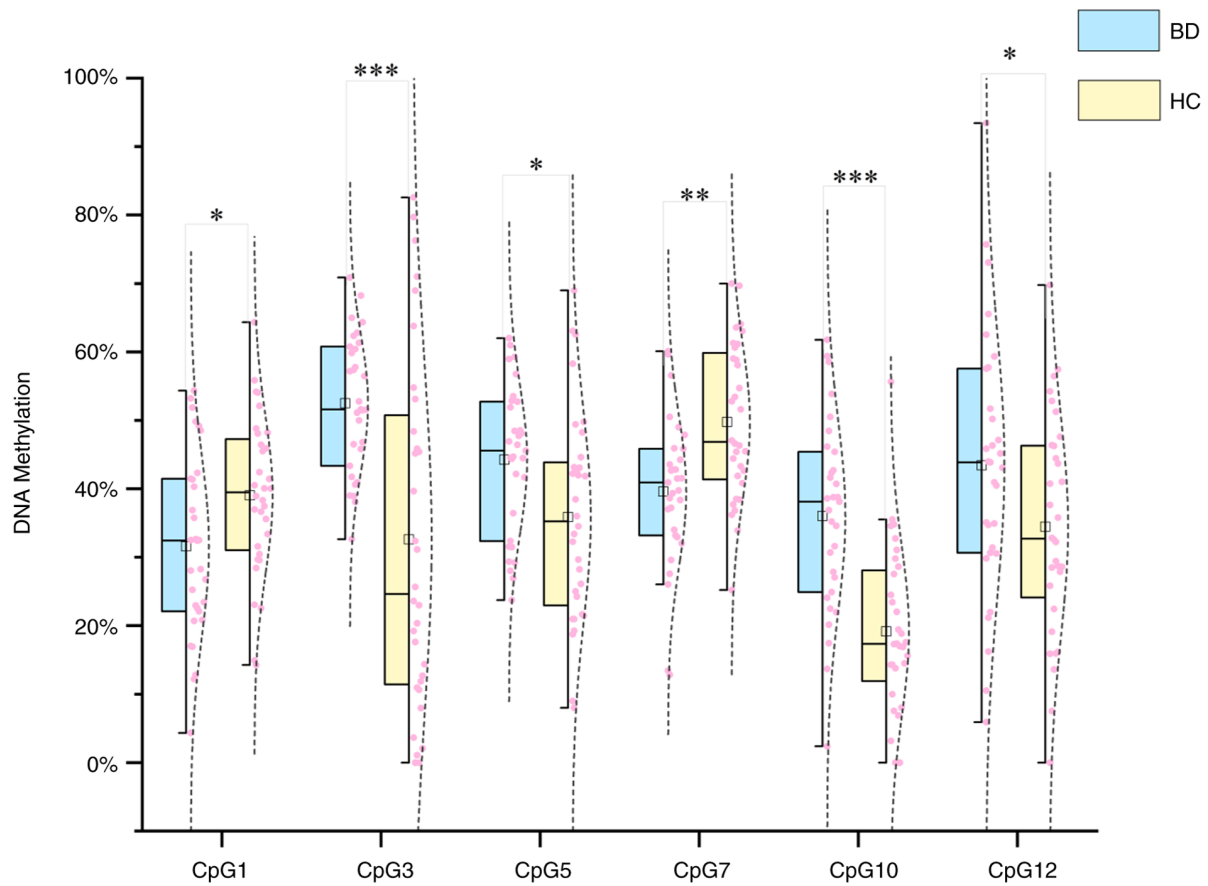


Figure 3. Comparative analysis of *GRIN2B* methylation profiles in BD and HC cohorts. Boxplots display the median (central line), mean (black square), interquartile range (boxes), theoretical normal distribution (black dashed curve), individual measurements (pink dots) and error bars represent the standard deviation. Significance gradient (group comparisons by independent samples t-test): \* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.001$  (two-tailed). BD, bipolar disorder; HC, healthy controls.

No significant correlations were observed between other CpG sites and either the HAMA or PSQI scores ( $P > 0.05$ ).

## Discussion

To our knowledge, this study provides the first evidence of differential DNA methylation patterns in the *GRIN2B* promoter region among patients with BD. The present findings not only demonstrate the association between *GRIN2B* methylation status and BD diagnosis but also reveal clinically meaningful correlations with comorbid anxiety symptoms and sleep disturbances in this population.

The present analysis revealed differential DNA methylation patterns at specific CpG sites within the *GRIN2B* promoter region between patients with BD and HCs. Among the 12 analyzed CpG loci, six exhibited methylation alterations: CpG3, CpG5, CpG10 and CpG12 showed hypermethylation, whereas CpG1 and CpG7 displayed hypomethylation. Notably, after adjusting for covariates through logistic regression, only the hypermethylation patterns at CpG3, CpG5, CpG10 and CpG12 maintained statistical significance, suggesting that site-specific epigenetic modifications may be particularly relevant to BD pathophysiology. The *GRIN2B* gene encodes a critical subunit of the NMDAR complex and DNA methylation-dependent transcriptional modulation may alter NMDAR subunit composition, potentially disrupting

synaptic plasticity in prefrontal-temporal-limbic circuits. The functional impact of BD-associated *GRIN2B* polymorphisms identified in German-Jewish (45) and Chinese Han populations (46) could be modulated by the local methylation status. Notably, the clinical relevance of these mechanisms is underscored by prior findings that the *GRIN2B* genotype predicts psychotic symptom severity and relapse frequency in patients with BD (33), suggesting methylation-mediated expression changes may similarly impact the disease course. Intriguingly, the therapeutic efficacy of quetiapine in BD may involve *GRIN2B*-related pathways, as molecular docking studies suggest its interaction with neuroactive ligand-receptor systems regulated by *GRIN2B* (47). Hypermethylation-induced *GRIN2B* downregulation could reduce NMDAR density in key brain regions (e.g., prefrontal cortex and hippocampus) (48) creating a neurobiological state amenable to mood stabilizer modulation. While the present findings align with gene-environment interaction models proposing methylation as a dynamic interface between genetic predisposition and environmental stressors, three critical considerations emerge: First, the observed methylation differences may represent either compensatory adaptations or pathogenic processes. Second, glutamatergic dysfunction (49) could serve as both precursor and consequence of neurodevelopmental alterations in BD; and third, the functional consequences of site-specific methylation require validation through allele-specific expression analyses.

Table III. Binary logistic regression analysis of the effect of *GRIN2B* DNA methylation on bipolar disorder.

CpG methylation site	B	SE	Wald	P-value	OR	95%CI
CpG3	0.076	0.024	9.759	0.002	1.079	1.029-1.132
CpG5	0.082	0.034	5.798	0.016	1.085	1.015-1.160
CpG10	0.107	0.034	10.058	0.002	1.113	1.042-1.190
CpG12	0.045	0.022	4.153	0.042	1.046	1.002-1.093

SE, standard error; OR, odds ratio.

Table IV. Correlation analysis of *GRIN2B* DNA methylation with anxiety and insomnia in bipolar disorder.

Methylation % at CpG site	HAMA score		PSQI score	
	r	P-value	r	P-value
CpG1	-0.178	0.383	-0.172	0.402
CpG2	0.098	0.635	-0.033	0.874
CpG3	-0.036	0.862	-0.061	0.767
CpG4	0.107	0.602	0.194	0.342
CpG5	-0.043	0.833	-0.286	0.157
CpG6	0.169	0.408	-0.029	0.887
CpG7	-0.240	0.237	-0.198	0.332
CpG8	0.159	0.439	0.419	0.033
CpG9	0.408	0.038	0.312	0.121
CpG10	0.144	0.484	0.251	0.215
CpG11	-0.081	0.696	0.057	0.783
CpG12	-0.313	0.119	-0.021	0.921

Control variables: Age, gender, education level and HAMD. HAMD, Hamilton Depression Rating Scale; HAMA, Hamilton Anxiety Scale; PSQI, Pittsburgh Sleep Quality Index Scale.

In the present study, it was also found that DNA methylation levels of *GRIN2B* were associated with anxiety and insomnia symptoms in BD, independent of age, gender, education or depression severity. This epigenetic alteration may contribute to BD-related psychopathology by suppressing *GRIN2B* transcription, thereby reducing the availability of functional NR2B subunits—a critical component of NMDARs implicated in synaptic plasticity and glutamatergic signaling (50), diminished NR2B activity could impair glutamate-mediated neurotransmission in limbic circuits (e.g., prefrontal cortex-amygdala connectivity and hippocampal networks), which are essential for emotion regulation and stress adaptation (34,51-53).

The association between *GRIN2B* methylation levels and anxiety symptoms may stem from dysregulated NMDAR-dependent excitatory-inhibitory balance. Specifically, hippocampal NMDARs are critical to behavioral inhibitory systems (40), and NR2B subunit deficiency-induced functional impairment could exacerbate anxiety-related neural hyperactivity. For instance, preclinical studies demonstrate that environmental stressors [e.g., benzo (a) pyrene exposure

or high-fat diets] induce anxiety-like behaviors in tandem with elevated *GRIN2B* methylation in brain regions governing fear responses (34,54) epigenetic silencing likely attenuates NMDAR-mediated synaptic potentiation in the ventral hippocampus and anterior cingulate cortex—key regions where NR2B modulates anxiety-like phenotypes (52,53,55), ketamine's rapid anxiolytic effects—mediated by NMDAR antagonism (56)—further support the hypothesis that *GRIN2B* methylation-driven NMDAR hypofunction could perpetuate anxiety states in BD.

Insomnia is a hallmark symptom of anti-NMDAR encephalitis, with clinical evidence underscoring the critical role of NMDAR dysfunction in sleep architecture disruption (57), aberrant *GRIN2B* methylation may perturb sleep-wake regulation through two interconnected pathways: i) Diminished NR2B expression in the hypothalamic SCN, where NR2B-containing NMDARs are essential for circadian rhythm entrainment (15,58); and ii) impaired Non-Rapid Eye Movement (NREM) sleep homeostasis caused by reduced NMDAR-mediated excitability in the lateral preoptic hypothalamus, a key region for sleep initiation and maintenance (59). The findings of the present study align with chronic sleep deprivation studies in young mice demonstrating reduced NR2B levels (35), suggesting that *GRIN2B* aberrant methylation may exacerbate insomnia by impairing NMDAR-dependent synaptic adaptation to sleep pressure. Additionally, selective NR2B antagonism disrupts cortical gamma oscillations during REM sleep (60), indicating that *GRIN2B* epigenetic silencing could destabilize sleep architecture through dysregulated glutamatergic neurotransmitter interactions.

While the present study yielded promising results, several critical limitations must be acknowledged. First, the conclusions may be constrained by the relatively small sample size, which reduces statistical power to detect robust epigenetic associations. Given the established clinical and biological heterogeneity of BD, the limited sample of the present study may not adequately represent the full spectrum of disease subtypes. These factors, combined with the cross-sectional design, preclude causal inferences; it cannot be determined whether the observed *GRIN2B* methylation alterations are causes or consequences of BD pathophysiology. Second, the reliance on peripheral blood methylation profiles raises questions about their biological relevance to brain processes. While blood-based biomarkers offer clinical practicality, it remains elusive whether these patterns mirror those in the central nervous system. Future validation should incorporate postmortem brain tissue analyses or emerging cerebrospinal

fluid-based epigenetic profiling techniques. Third, the present study relied on retrospective data collection and self-reported medical histories, which may have resulted in the under-reporting of comorbidities. This could lead to unintentional inclusion of patients with undiagnosed conditions that met exclusion criteria, potentially confounding the observed methylation patterns. Fourth, while cohort homogeneity was improved by exclusively recruiting patients with BD in the depressive phase, this design limits the generalizability of findings to other disease phases (e.g., manic or euthymic states). Epigenetic markers may exhibit phase-dependent fluctuations, implying the results could reflect state-specific alterations rather than trait characteristics of BD. Fifth, the lack of integrated single-nucleotide polymorphism analyses and gene expression data limits our ability to elucidate gene-environment interactions underlying the observed methylation changes. Most importantly, these preliminary findings require replication in larger, independent cohorts with longitudinal designs to establish clinical generalizability. Future investigations should prioritize multicenter collaborations, incorporate pharmaco-epigenetic analyses and employ multi-omics approaches to comprehensively characterize *GRIN2B* methylation dynamics in BD progression, and explicitly address phase-specific biomarker variations through cross-state comparisons.

In conclusion, the present study demonstrated for the first time that DNA hypermethylation in the *GRIN2B* promoter region is associated with BD and may play a role in mediating the development of anxiety and insomnia symptoms in patients with BD. These findings underscore the importance of gene-environment interactions in BD and advance a better understanding of its complex etiology. Early intervention strategies targeting these mechanisms could improve timely diagnosis, symptom management and long-term outcomes in BD. Further studies are warranted to validate *GRIN2B* methylation as both a diagnostic biomarker and therapeutic target for BD.

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### Availability of data and materials

The datasets generated and/or analyzed during the current study are not publicly available due to ethical issues involving the participants' data and privacy but may be requested from the corresponding author.

### Authors' contributions

SZ and HY conducted and designed the study. HY and YW collected the data. HY analyzed the data and drafted the

manuscript. SZ revised the manuscript. SZ and YW have independently checked and verified the authenticity of the raw data. All authors have read and approved the final manuscript.

### Ethics approval and consent to participate

This study was approved by the Human Research and Ethics Committee of Xinjiang Uygur Autonomous Region People's Hospital (Urumqi, China; approval no. KY2023020968). This study adhered to the guidance listed in the latest version of the Declaration of Helsinki. All subjects volunteered to participate in the study provided written informed consent prior to the study. The participants were also informed that they could withdraw from the study at any time without any reason or consequence.

### Patient consent for publication

Not applicable.

### Competing interests

The authors declare that they have no competing interests.

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