Abstract. Herbal medicine is used by millions of diabetic patients due to economic and cultural factors. The current study investigates the antidiabetic potential of fenugreek (Trigonella foenum-graecum) seed extract at a dose of 100 mg/kg in a Streptozocin-induced diabetic model. Male Sprague-Dawley rats received either intraperitoneal fenugreek [daily (ED) or every other day (EOD)] or oral fenugreek supplement daily, for four weeks. Results show that fenugreek significantly reduced blood glucose. Urea levels were reduced after daily intraperitoneal injection, and creatinine levels dropped after oral treatment, respectively. AST and ALT levels were reduced following fenugreek treatment, while protein levels significantly increased. High-density lipoprotein (HDL) increased after daily injections, while triglycerides decreased significantly in all groups. Glutathione S-transferase and catalase increased with treatment, while peroxidase antioxidant enzyme levels were reduced. Glutathione peroxidase levels increased only after daily injection. Histologically, fenugreek mildly protected hepatic, renal, and pancreatic tissues. In conclusion, the current study shows some potential benefits of fenugreek use. Oral and injectable fenugreek showed improvement in blood glucose, renal and liver functions. Although triglyceride levels decreased significantly, no significant changes in cholesterol levels were seen after fenugreek use. Higher doses and longer fenugreek treatment duration are recommended for the optimum protection of the liver, kidneys, and pancreatic tissues.

Introduction

Diabetes is a metabolic disorder characterized by hyperglycemia resulting from abnormalities in insulin secretion and action (1). Chronic hyperglycemia leads to microvascular (e.g. neuropathy, and nephropathy) and macrovascular (mainly cardiovascular) complications that arise due to the increased reactive oxygen species production and reduced antioxidants (2).

Herbal products have been widely used throughout history for the treatment of several diseases. Since the characterization and exact mechanisms of action of these natural products remain unclear, researchers are trying to evaluate their beneficial effects on human health as well as their possible adverse effects (3). Despite the presence of several treatments for diabetes such as insulin analogs, sulphonylureas, biguanides, dipeptidyl peptidase-4 inhibitors, thiazolidine, and α-glucosidase inhibitors, patients prefer to use botanicals due to the increased cost and side effects of these medications (2).

Many plants, such as cinnamon and ginseng reduce glucose and lipid levels through the stimulation of insulin secretion, delay of gastric emptying, inhibiting glucosidase activity, increasing GLUT4 expression (4), and the activation of AMP-activated protein pathway (5), and inhibiting gluconeogenesis (6). Several medicinal plants are used for the treatment of diabetes, such as cinnamon and ginseng (7,8).

Fenugreek (Trigonella foenum-graecum), is a historically used herbal medicinal plant that is popular in Africa, India, South, and Central Asia (9). It is traditionally used to treat several conditions, such as diabetes and obesity. It possesses antioxidant, antihyperlipidemic, antibacterial, anti fungal, anti-inflammatory, and galactagogic properties (10).

Fenugreek's pharmacological effects are attributed to a range of bioactive compounds such as polyphenols, steroids, lipids, alkaloids, saponins, flavonoids, hydrocarbons, carbohydrates,
galactomannan fiber, and amino acids. Several scientific groups examined its antidiabetic effect. A previous study showed that fenugreek increased glucose uptake in HepG2 cells is due to the overexpression of the glucose transporter (GLUT-2) (12) and sterol regulatory element-binding protein (SREBP1c) mRNA levels (11). Another report by Pradeep and Srinivasan (13), demonstrated that when combined with 3% onion, better fenugreek antidiabetic results were seen. A potential fenugreek-based drug (Fenfuro®) was compared to Metformin in a clinical trial. Results showed that Fenfuro combined with Metformin gave better results than Metformin alone (14).

Diosgenin saponin is considered the most bioactive substance of fenugreek. It has antioxidative effects and plays a pivotal role in improving the diabetic status by several mechanisms (1,15). The mechanisms include β-cell renewal and insulin secretion stimulation. Besides, diosgenin elevates the mRNA transcription levels of CCAAT/enhancer-binding protein (C/EBPβ) and peroxisome proliferator-activated receptor-γ (PPAR-γ) (10,12).

Other components in fenugreek include; 4-hydroxyisoleucine, which is an amino acid that enhances insulin secretion, decrease plasma triglycerides, and total cholesterol levels (1). Galactomannan is a carbohydrate that represents 45-60% of the seed of fenugreek. It has been shown to block the carbohydrate and lipid hydrolyzing enzymes in the digestive system, resulting in lowering the postprandial glucose level (1).

Although the detailed mechanisms of action of the fenugreek antidiabetic activity are yet to be identified, many studies suggest that antioxidant activity plays a significant role in hepatoprotection. Another possibility would be that fenugreek reverses protein glycation caused by hyperglycemia (16). Further investigations into the molecular mechanisms of actions and active components of the plant are needed.

In many parts of the world, fenugreek is commonly consumed as a drink. In the current study, we attempt to compare different routes of administration of fenugreek at a clinically feasible dose (100 mg/kg) in a diabetic rat model induced by streptozocin (STZ).

Materials and methods

Animals. All experiments were conducted in compliance with the guidelines established by the NIH for Animal Care and Use and were approved by the Institutional Animal Care and Use Committee (IACUC) of the October University for Modern Sciences and Arts (2018). Male Sprague Dawley rats weighing (175-200 g), were obtained from Theodore Bilarz Research Institute (Cairo, Egypt). Rats were randomly divided into the following groups: Group 1: diabetic (DM) rats receiving 100 mg/kg every other day (EOD) of fenugreek extract (HERB-PHARM). Group 2: DM rats receiving daily (ED) fenugreek 100 mg/kg IP. Group 3: DM rats receiving oral fenugreek 100 mg/kg daily. Group 4: untreated diabetic group. Group 5: healthy nondiabetic rat group.

Diabetes model. Animals received Streptozocin (STZ; 75 mg/kg in sterile citrate buffer) intraperitoneally. Diabetes was confirmed one week following STZ injection, by blood glucose levels. Rats showing fasting glucose levels at or above 270 mg/dl (>15 mmol/l) were included in the study (3). At the end of the experiment, the histological examination of the rat pancreatic tissues confirmed DM.

Sample collection. At the end of the 4 weeks of treatment, rats were anesthetized by ketamine/xylazine (ketamine 80-100 mg/kg, xylazine 10-12.5 mg/kg IP). Blood was collected by cardiac puncture, and rats were dissected and tissue samples (pancreas, kidney and liver) were collected for biochemical and histological analysis. There were no sample size differences; animals were added to replace lost animals due to mortality.

Biochemical analysis. The collected serum was divided into aliquots to assess the liver, and kidney functions as well as serum glucose and the lipid profile, as previously described (3).

Antioxidant activity assays. Frozen liver tissue samples (0.2 g) were homogenized in phosphate-buffered saline. The suspension was centrifuged at 4400 rpm, and the supernatants collected and tested for the antioxidant enzyme levels.

Catalase enzyme activity. Catalase enzyme activity was measured according to the method originally described by Aebi (17). Briefly, the assay is based on catalase reaction with a known quantity of hydrogen peroxide (H₂O₂), and the reaction is stopped after 1 min by a catalase inhibitor. The remaining H₂O₂ reacts with 4-aminophenazone and 3,5-dichloro-2-hydroxy-benzene sulfonic acid to form a chromophore. The absorbance is measured at 510 nm using a spectrophotometer.

Glutathione peroxidase. Glutathione peroxidase was measured based on the method described initially by Paglia and Valentine (18). The assay principle is based on the indirect measurement of the activity of cellular glutathione peroxidase enzyme. Oxidized glutathione (GSSG) is produced by reduction of an organic peroxide by cellular glutathione. The rate of decrease in the absorbance at 340 is directly proportional to the glutathione peroxidase activity in the sample.

Glutathione S-transferase (GST) enzymatic activity. The measured enzyme activity is based on the method designed by Habig et al (19). The action of GST enzyme is to catalyze the
conjugation of reduced glutathione (GSH) with 1-chloro 2,4-dinitrobenzene (CDNB) via the -SH group of glutathione. This results in the production of the conjugate, S-(2,4-dinitrophenyl)-L-glutathione, which can be detected. The absorbance is read at 340 nm.

**Peroxidase activity.** Peroxidase activity is based on the enzyme inhibition (20) by the addition of sulfite so that it is inactive when the hydrogen peroxide is added. Freshly prepared 1% o-phenylenediamine and 0.3% hydrogen peroxide are added to liver tissue homogenate. The reaction is stopped after 5 min by adding sodium bisulfite. The absorbance was measured at 430 nm. The enzyme activity is expressed as the change in absorbance at 430 nm (∆OD430)/min/mg protein.

**Histological analysis.** Formaldehyde fixed tissue samples were paraffin-embedded, and 5 µm sections were cut and stained with hematoxylin and eosin. Slides were examined and photographed using a BX51 light microscope with an Olympus digital camera (DP20) (Olympus).

**Statistical analysis.** The statistical analyses were performed using GraphPad Prism software, version 7.04 (GraphPad, Inc.). Data are presented as mean ± standard deviation. One-way ANOVA and Tukey test were used. P<0.05 was considered to indicate a statistically significant difference.

**Results**

**Blood glucose levels.** Fasting blood glucose levels of rats treated with fenugreek extract were significantly reduced after treatment compared to the diabetic control group in all treated groups (Fig. 1). There was no significant difference between treatment groups.

**Renal function.** Urea levels increased in diabetic rats (Fig. 2). No significant reduction was seen after fenugreek treatment except with the daily injection group (Fig. 2). Creatinine levels also increased in untreated DM rats. Fenugreek injection
did not reduce creatinine levels. Only after oral treatment, a significant decrease in creatinine was seen (Fig. 2).

Liver functions and lipid profile. Alanine aminotransferase (ALT) and aspartate aminotransferase (AST) levels increased in diabetic rats. Fenugreek treatment in both the oral and daily injection (ED) groups significantly decreased both AST and ALT levels compared to the diabetic control group. Protein levels significantly increased in all treated groups compared to untreated diabetic rats. ED injections showed the highest value and significance compared to the diabetic control (Fig. 3).

High-density lipoprotein (HDL) levels did not increase in diabetic rats compared to normal. Only after daily fenugreek Figure 4. Effect of fenugreek treatment on lipid profile: (A) High-density lipoprotein (HDL). (B) Cholesterol levels were not increased in diabetic rats compared to normal, only after ED injection of fenugreek a significant increase was seen in HDL levels. (C) Triglycerides were significantly increased in diabetic rats. All treatment groups showed a significant decrease in triglycerides. Fenugreek treatment did not cause any significant change in cholesterol levels. Data are shown as mean ± SD, n=5, ED, daily.

Figure 5. Effect of fenugreek treatment on liver antioxidant enzymes. (A) Catalase enzyme levels were not significantly different between normal and diabetic rats. These levels increased significantly in all treated groups. (B) Peroxidase levels were increased in diabetic rats compared to normal. All treated groups showed a significant reduction in peroxidase levels compared to diabetic untreated rats. (C) Glutathione peroxidase levels were not changed in diabetic untreated or EOD and oral treated rats. The only group showing a significant increase was the ED injection treated group. (D) Glutathione S-transferase enzyme was significantly reduced in diabetic untreated rats. No significant change was seen with the oral administration of fenugreek, whereas both EOD and ED injected groups showed a significant increase in the enzyme levels compared to the diabetic control group. Data are shown as mean ± SD, n=5, ED, daily, EOD, every other day.
Injection, a significant increase was seen. Triglycerides, on the other hand, significantly increased in diabetic rats. All treatment groups showed a significant decrease in triglyceride levels compared to diabetic rats injected daily, which showed the highest significance. Cholesterol levels increased following induction of diabetic and were not decreased after fenugreek treatment (Fig. 4).

Antioxidant enzymes. Catalase enzyme in the liver tissue of untreated diabetic rats was slightly lower compared to the normal rats. Catalase levels increased significantly in all treatment groups compared to both normal and untreated diabetic rats, with oral fenugreek showing the highest levels (Fig. 5A). Quantitative determination of peroxidase activity in liver tissue showed that the peroxidase antioxidant enzyme levels were increased in diabetic rats compared to normal. All treated groups showed a significant reduction in peroxidase levels compared to diabetic untreated rats. The daily injection group showed the highest and most significant reduction (Fig. 5B).

Glutathione peroxidase levels were changed slightly in diabetic untreated rats. Only following daily injections, there was a significant increase in glutathione peroxidase levels (Fig. 5C). Glutathione s-transferase enzyme was significantly reduced in untreated diabetic rats. Both EOD and daily injections treated groups showed a significant increase in the enzyme levels compared to the diabetic group with the daily injection group significantly higher than the EOD injection group (Fig. 5D).

Histological changes in the pancreas. Normal rats showed a typical healthy architecture of the pancreatic islets of Langerhans, with no signs of cell injury observed. Untreated DM rats showed severe alteration in both the acini and islets of the pancreas, most of the cell nuclei were thickened and appeared larger, indicating karyo-pyknosis of the cells; also, numerous vacuoles were observed (Fig. 6A and B).

In rats receiving oral fenugreek extract, the pancreas showed no signs of protection, with destructive and degenerative changes in the pancreatic islets in almost all tissue sections, with many cells demonstrating pyknotic nuclei with variable records of cytoplasmic vacuolation (Fig. 6C). Treatment with fenugreek injections (ED and EOD) showed no histological signs of improvement with changes in cellular components of pancreatic islets with numerous cells demonstrating pyknotic nuclei, and cytoplasmic vacuolation.

Histological changes in the kidneys. Normal rats demonstrated normal histological features of the renal cortex and medulla with intact corpuscles, while diabetic rats showed wide areas of damage with intraluminal cast formation and many degenerated tubular cells and congested inter-tubular blood vessels. (Fig. 7A and B). Oral fenugreek extract showed focal areas of degenerative vacuolar changes in lining cells

![Image of histological changes in the pancreas](image-url)
and or pyknotic nuclei (Fig. 7C). Fenugreek injections showed pronounced damage of renal corpuscles, severe vacuolar changes in mesangial cells, and severe necrotic changes in tubular lining cells, with most cells losing cellular details (Fig. 7D and E).

**Histological changes in the liver.** Hepatic tissue samples from normal rats showed normal hepatocytes, blood vessels, and sinusoids, as shown in Fig. 8A. Untreated diabetic rats showed high numbers of degenerated hepatocytes with karyopyknosis, and dilatation and congestion of blood vessels and
sinusoids (Fig. 8B). Oral fenugreek extract treatment showed degenerating hepatocytes with shrunken pyknotic nuclei and missing cellular details (Fig. 8C). Fenugreek injections showed degenerating hepatocytes with shrunken pyknotic nuclei and loss of cellular details (Fig. 8D and E).

**Discussion**

In the current study, we used an STZ model to compare different routes of administration of fenugreek (10). STZ have been used extensively in diabetes research especially for the study of various therapeutic approaches, including the use of plant extracts as supplements in diabetes care (21-23).

Fenugreek is a plant widely consumed in different parts of the world. Here we provide a comprehensive evaluation of fenugreek seed extract treatment on the pancreas, liver, and kidneys both biochemically and histologically. Induction of diabetes was confirmed biochemically with elevated blood glucose levels, as well as histologically with islets of the pancreas showing signs of destruction on histological examination.

We attempted to test glycosylated hemoglobin (HbA1c); unfortunately, the results obtained were unreliable (data not shown). This might be due to the difficulty of measuring HbA1C in Sprague-Dawley rats, as reported by other researchers (24,25). Some researchers suggest that HbA1C should be estimated using other tools such as ELISA, or high-performance liquid chromatography (HPLC) (26).

Fenugreek seed extract administration reduced blood glucose levels, possibly due to the high content of alkaloid trigonelline and steroidal saponins in fenugreek, especially the 4-hydroxyisoleucine compound that is said to be insulinotropic (27). The current data confirm previous reports (28) that showed a dose of 50 mg/kg of orally administered fenugreek for 4 weeks in STZ diabetic rabbits, produced similar effects.

The architecture of the pancreatic tissue correlates with the stage of diabetes and its severity (29). In the current study after STZ injection, diabetic rats showed damage to the pancreatic islet cells and severe pathological changes to exocrine and endocrine components, which is consistent with previous findings (30). Rats treated with fenugreek, showed protection of pancreatic tissues, possibly due to the presence of diosgenin which is postulated to have several antidiabetic effects, such as the regeneration of pancreatic β-cells and enhancement of insulin secretion in general (1,31). Diosgenin also improved blood glucose levels maintenance and preserved the pancreas, liver, and skeletal muscle tissues (1). Previous report (32) on the effects of fenugreek oil on the pancreas in an Alloxan induced diabetic rat model showed that pancreatic cell damage and renal function were slightly reversed after treatment with fenugreek oil.

Oral fenugreek showed a significant decrease in creatinine levels, in contrast to EOD and ED injections. This is consistent with previous reports (33), showing improved renal functions with fenugreek administration. Histologically better protection was achieved (33), possibly due to the longer treatment period. Other researchers also reported improvement in creatinine levels and improvement of the glomerular base membrane in the kidneys of diabetic rats when fenugreek extract was administered orally. These findings also support the present study and show the potential of fenugreek as a drug for diabetes and its renal complications (34,35).

Recent metabolomics studies on the effect of fenugreek flavonoids on STZ showed a significant impact on liver, kidney, and pancreas (36,37). The fenugreek flavonoids lowered insulin resistance, improved glycolysis, and gluconeogenesis, and protected kidneys and pancreatic islet cells from damage.

Serum levels of liver enzymes (ALT and AST) increased in untreated diabetic rats and were significantly reduced compared to diabetic rats after fenugreek treatment. This high serum level is attributed to the injuries to the liver cells (38). Fenugreek caused a significant reduction in the liver enzyme levels, indicating a protective effect of liver cells. These findings are consistent with previous data that showed a protective effect of fenugreek as a daily supplement (39-42). Other studies reported that treatment with fenugreek aqueous seed extract (43), using a dose of 25 mg/kg body weight for 60 days, significantly decreased blood glucose and liver enzyme levels. However, no histological liver protective role was reported.

The effects of fenugreek seed extract on lipid profile in the current study showed a significant reduction in triglyceride levels compared to diabetic rats. However, a significant increase in HDL levels was demonstrated only after daily injections, compared to diabetic rats, unlike other treatment groups which did not show a substantial increase in HDL levels. This suggests a better effect after the daily (ED) injection. No effects on total cholesterol levels were observed in any of the treatment groups. Cholesterol findings are somewhat in disagreement with previous reports (44), where a dose of 500 mg/kg for four weeks caused improvement in lipid profile (HDL, cholesterol, and triglycerides). This difference might be due to the lower dose used in the current study (100 mg/kg).

Catalase and glutathione-S-transferase (GST) enzymes are crucial anti-oxidative enzymes in the liver. Our results show that fenugreek treatment led to a significant increase in GST enzyme levels in EOD and ED injection groups when compared to diabetic rats, unlike the oral route, which did not show significance. Concerning catalase, a significant increase in the enzyme levels was observed in all groups compared to diabetic rats. This indicates considerable potential for fenugreek as a booster for antioxidant activity in the liver due to its high content of diosgenin (1,31). Previous studies showed similar results, therefore, confirming the antioxidant effect of fenugreek (45).

There was a significant increase in peroxidase activity within the liver tissue of untreated diabetic rats compared to healthy rats. This might be due to the high oxidative stress exerted on the liver tissue due to the induction of diabetes by STZ or as a complication of hyperglycemia (46). In all treated groups, there was a significant reduction in peroxidase activity compared to diabetic rats.

Glutathione peroxidase levels were unchanged in the present study except for the ED injection treated rats where a significant increase was observed compared to normal and diabetic rats. This suggests a better effect in this group (ED injection) over other groups (EOD injection and oral).

In the current study, a reduction in total proteins in the untreated diabetic group was reversed by fenugreek extract treatment. This increase in protein levels could be attributed to the hyperglycemic effect on the liver tissue, as a result of glycation of the antioxidant enzymes, which eventually leads to alteration and damage in cell structural proteins and enzymes caused by the reactive oxygen species (46).
Fenugreek may also improve the body's use of sugar, adjusting insulin release, and possibly lowers the absorption of glucose from the intestine (47). Oral intake of fenugreek is also associated with liver protection and a better quality of life for diabetic patients (48). Fenugreek is also useful as a digestive stimulant and has potent antibacterial and oxidant activity (49).

In conclusion, fenugreek daily injection showed better antidiabetic effects with better serum values than other groups. Although in all groups, there was an improvement of the antioxidative enzymes and other diabetic parameters, the histological structure was not fully restored in the kidney, liver or the pancreas. This can be due to the low dose tested and the relatively short test period (one month).

Fenugreek seems to be a promising anti-diabetic plant. Further work is needed to better identify the mechanism of action and the effective dose range. Longer periods are recommended to achieve histological improvement.

Acknowledgments

Not applicable.

Funding

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

Availability of data and materials

The datasets used during the present study are available from the corresponding author upon reasonable request.

Authors' contributions

MEB, TIA and HE were involved in data investigation and data curation. AMES was involved in data analysis and validation, and in the writing, reviewing and editing of the manuscript. MKS and AA were involved in the conceptualization, methodology, supervision, validation of the study, and in the reviewing and editing of the manuscript. All authors read and approved the manuscript and agree to be accountable for all aspects of the research in ensuring that the accuracy or integrity of any part of the work are appropriately investigated and resolved.

Ethics approval and consent to participate

This study was approved by the Institutional Animal Care and Use Committee (IACUC) of the October University for Modern Sciences and Arts (2018).

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Authors' information

The Orcid ID numbers for the following authors are: MTB, orcid.org/0000-0002-8570-7739; SSAZ, orcid.org/0000-0001-6069-8760; AA, orcid.org/0000-0003-0486-348X; MKS, orcid.org/0000-0002-2072-0975; AMES, orcid.org/0000-0002-5169-1214.

References


