

# Synthetic peptide F2A4-K-NS mimics fibroblast growth factor-2 *in vitro* and is angiogenic *in vivo*

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**Abstract.** A multi-domain synthetic peptide, F2A4-K-NS, mimicked the action of recombinant human FGF-2 (rhFGF-2) *in vitro* and in an *in vivo* model of angiogenesis. Like rhFGF-2, F2A4-K-NS was quantitatively shown to bind to FGF receptors in a cell-free receptor binding assay using a chimeric FGFR1 (IIIc)/Fc as monitored by surface plasmon resonance (SPR), and also shown to bind to heparin using biotinylated low-molecular weight heparin in a similar SPR assay. *In vitro*, F2A4-K-NS triggered signal transduction as monitored by the stimulation of ERK1/2 phosphorylation in human umbilical cord endothelial cells. In cell based assays, it increased cell migration, cell proliferation, and gelatinase secretion; endpoints associated with FGF-2 stimulation. Furthermore, these *in vitro* effects were mediated with quantities of F2A4-K-NS that were similar to those of rhFGF-2. *In vivo*, F2A4-K-NS was angiogenic at doses of 40 and 400 ng/implant in a subcutaneous implant assay as determined by morphologic scoring, hemoglobin content, and histology. These results support the hypothesis that F2A4-K-NS is a mimetic of FGF-2 that can substitute for FGF-2 *in vitro* and *in vivo*. A synthetic mimetic of FGF-2, such as F2A4-K-NS, could be a useful tool in studying mechanisms of cell activation and potentially in various therapeutic applications.

## Introduction

A synthetic peptide that could mimic aspects of the bioactivity of recombinant human FGF-2 (rhFGF-2) could be of considerable interest, especially if one of those actions included stimulation of angiogenesis. While FGF-2 affects many cell types in a multiplicity of ways, its effect on angiogenesis (1,2) underlies, at least in part, many of the therapeutic approaches for its use (3). Such potential uses include the treatment of ischemic heart and peripheral vascular disease, myocardial infarction, and diabetic retinopathy (3). Other potential applications for FGF-2 include preparation of artificial organ implant sites (4,5), aneurysm healing and treatment (6-8), the treatment of diabetic foot ulcers, and acceleration of bone fracture healing (9-12).

For a peptide to mimic the biological effects of FGF-2, it would be expected to bind to FGF receptor type-1 (FGFR1) (2) and in particular the 'c'-splice variants (13). Receptor binding should lead to signal transduction which, in turn, would lead to increases in cell proliferation, migration, and secretion of proteases. Each of those events would be expected to participate in angiogenesis *in vivo*. Clearly, the aforementioned actions could also participate in a range of other tissue and wound responses stimulated by FGF-2.

In the studies presented here, F2A4-K-NS, a multi-domain synthetic peptide was evaluated for its ability to mimic aspects of rhFGF-2 bioactivity including the ability to stimulate angiogenesis in an experimental model. The results support the hypotheses that F2A4-K-NS can mimic rhFGF-2 bioactivity.

## Materials and methods

**Peptide synthesis.** Peptides in milligram quantities were synthesized manually following standard Fmoc protocols using NovaSyn TGR resin (EMD BioSciences, La Jolla, CA). Fmoc-amino acids including aminohexanoic acid (Ahx) were obtained from Peptides International, Inc. (Lexington, KY).

F2A4-K-NS was synthesized as a branched peptide with the following sequence: H-YRSRKYSSWYVALKRK(H-YRSRKYSSWYVALKR)-Ahx-Ahx-Ahx-RKRLDRIAR-NH<sub>2</sub>. It was purified by RP-HPLC on a C18 column, using a linear gradient 0-60% acetonitrile/water (0.1% trifluoroacetic acid) run over 60 min at 1 ml/min flow rate (detection at 214 nm). The purified peptide generated a single uniform peak on

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**Abbreviations:** FGF, fibroblast growth factor; SPR, surface plasmon resonance; ERK, extracellular signal-regulated kinase; HUVEC, human umbilical cord endothelial cell; HPLC, high performance liquid chromatography; MALDI-TOF, matrix assisted laser desorption/ionization-time of flight; LMWH, low-molecular weight heparin; HEPES, 4-(2-hydroxyethyl)piperazine-1-ethanesulfonic acid; RMEC, rat microvascular endothelial cell; MAP, mitogen-activated protein

**Key words:** angiogenesis, F2A4-K-NS peptide, fibroblast growth factor

Table I. Chemical parameters associated with synthesis of F2A4-K-NS.

Yield from synthesis	Area under peptide peak	Molecular weight actual/predicted	mg peptide/mg lyophilized material
14%	>99%	5538/5540	0.7

analysis by RP-HPLC. Software associated with the HPLC (Shimadzu Scientific Instruments, Inc., Columbia, MD) was used to determine the % area under the peptide peak (Table I). The molecular mass of F2A4-K-NS, as determined by MALDI-TOF mass spectrometry, was 5539, in agreement with the theoretical molecular weight of 5540 Da.

F2A4-K-NS contains two copies of the receptor-targeted sequence YRSRKYSSWYVALKR derived from native FGF-2 (residues from 106 to 120) (14,15), and a heparin binding domain, RKRLDRIAR that conforms to the canonical XBBBXXBX heparin-binding motif (16). In addition, F2A4RTDS, with the receptor-targeted sequence YRSRKYSSWYVALKR of F2A4-K-NS was synthesized. A control peptide designated HBD3 was also synthesized and had the following sequence: Ahx-Ahx-Ahx-RKRLDRIAR-NH<sub>2</sub>. This peptide contained the heparin binding domain but lacked the receptor binding sequences.

**Surface plasmon resonance (SPR) analysis.** Real-time biomolecular interactions were analyzed with a BIACore 2000 system (Biacore Inc., Piscataway, NJ). F2A4-K-NS was synthesized as described above, and rhFGF-2 was obtained from the NIH/NCI Preclinical Repository, Biological Resource Branch. Soluble FGF receptor, recombinant chimera of human FGFR1 $\alpha$ (IIIc)/Fc (R&D Systems, Minneapolis, MN), was immobilized on research grade CM5 chips (Biacore Inc.). Hereafter, FGFR1 $\alpha$ (IIIc)/Fc is referred to as FGFR1. Following activation with EDC/NHS, successful immobilization of FGFR1 was achieved by injecting FGFR1 [10  $\mu$ g/ml in HBS-EP buffer (0.01 M HEPES, pH 7.4, 0.15 M NaCl, 3 mM EDTA, 0.005% Tween-20 (v/v))] onto the activated CM5 chip. To obtain kinetic data, different concentrations of analytes in HBS-EP buffer were injected over the sensor chip at a flow rate of 50  $\mu$ l/min. Peptide binding was measured in resonance units (RU). At the end of each sample injection (120 sec) HBS-EP buffer was passed over the sensor surface to monitor the dissociation phase. Reference responses from blank flow cells were subtracted from FGFR1 flow cells for each analyte injection and the kinetic parameters for each interaction were determined by globally fitting the experimental data to a 1:1 interaction with BIAEvaluation software (Biacore Inc.). The association rate constant and the dissociation rate constant ( $k_a$  and  $k_d$ , respectively) for FGFR1 interaction with F2A4RTDS can not be obtained by 1:1 Langmuir binding model, thus, the equilibrium dissociation constant ( $K_D$ ) was derived by a steady state binding model. A minimum of six different analyte concentrations were used to determine the kinetic parameters for each interaction. For each fitting, the

standard error (SE) values for the association and dissociation rate were 1-2 orders of magnitude smaller than the respective association and dissociation constant values; and the Chi-square was <10% of the Rmax in all cases.

Alternatively, heparin was immobilized on a chip as described by Sheehan *et al.* (17). Briefly, a low-molecular weight heparin (LMWH) surface was created by linking biotin-bound LMWH (Celsus, Inc., Cincinnati, OH) to a Biacore streptavidin chip using injection buffer [0.3 M NaCl, 20 mM HEPES (pH 7.4), and 0.05% Tween-20] at 10  $\mu$ l/min for 2 min. Analytes were injected at a concentration series from 800 nM to 50 nM with running buffer [0.15 M NaCl, 2 mM CaCl<sub>2</sub>, 20 mM HEPES (pH 7.4), and 0.05% Tween-20] at 50  $\mu$ l/min for 2 min. At the end of injections, running buffer was passed over the sensor surface for 2 min to monitor the dissociation. Then, the sensor surface was regenerated by injection of regeneration buffer [2 M NaCl, 20 mM HEPES (pH 7.4), and 0.05% Tween-20] at 30  $\mu$ l/min for 2 min. Peptide binding was analyzed as above.

**Cell culture.** HUVECs (Cambrex, East Rutherford, NJ) were grown in endothelial cell medium-2 (EGM-2<sup>®</sup>, Cambrex), supplemented with 5% FBS, and EGM-2 MV singlequots supplements (Cambrex), in a 37°C incubator with a humidified atmosphere containing 5% CO<sub>2</sub>. C3H10T1/2 murine multipotent stem cells were obtained from the American Type Culture Collection (Manassas, VA) and were grown in DMEM:F12 medium containing 15% newborn calf serum and antibiotics. Rat microvascular endothelial cells (RMEC) transformed by SV40 large T-antigen (18,19) were a gift from M. Goligorsky, Division of Nephrology and Hypertension, State University of New York at Stony Brook.

**Phosphorylation of ERK1/2.** HUVECs were stimulated with rhFGF-2 and F2A4-K-NS at the concentrations indicated in the figures in medium containing 1% newborn calf serum. After incubation for selected times, cells were lysed and processed for Western blot analysis on Hybond nitrocellulose membranes (Amersham Biosciences, Piscataway, NJ). The total and phosphorylated ERK1/2 were revealed by antibodies recognizing total ERK1/2 (Sigma-Aldrich, St. Louis, MO) and phosphorylated (Thr202 and Tyr204) (Cell Signaling, Beverly, MA) ERK1/2.

**Cell proliferation.** Cell proliferation was monitored using a commercially available kit (MTS) (Promega, Madison, WI) based on a tetrazolium compound 3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium, inner salt (MTS). Aliquots of 1x10<sup>3</sup> RMEC cells were seeded into wells of 96-well plates and allowed to attach. The medium was replaced with one containing 1% newborn calf serum plus F2A4-K-NS. After 3 days in culture, the relative cell number was monitored using MTS following the directions of the manufacturer.

**Migration assays.** HUVECs and C3H10T1/2 cells were used as target cells and two different methods were used in the analysis, migration across a wound margin and migration through a coated membrane (20). For studies involving migration across a simulated wound margin, the cells were

	Binding coefficients		
	$K_a(M^{-1}s^{-1}) \times 10^3$	$K_d(s^{-1}) \times 10^{-4}$	$K_D(M)$
F2A4-K-NS/ FGFR1	4.1±0.8	15.3±0.2	3.74×10 <sup>-7</sup>
FGF-2/FGFR1	108.0±1.3	107.0±2.7	9.9×10 <sup>-8</sup>
F2A4RTDS/ FGFR1			1.12×10 <sup>-6</sup>
HBD3/FGFR1			NB
F2A4-K-NS/ Heparin	17.4±0.4	12.9±0.5	7.4×10 <sup>-8</sup>
FGF-2/Heparin	7.3±0.2	2.6±0.2	3.6×10 <sup>-8</sup>

The binding coefficient  $K_a$ ,  $K_d$ , and  $K_D$  values were determined by surface plasmon resonance binding assays of test compounds to immobilized FGFR1IIIc/Fc or low-molecular weight heparin as described in Materials and methods. NB, no binding. Chi-square was <10% of Rmax in all cases.

grown *in vitro* and used when approximately 90% confluent. A simulated wound was made by scraping the cells away from the substrate. The cultures were rinsed and then incubated in DMEM:F12 medium containing 2% newborn calf serum with or without peptide (50 ng/ml). rhFGF-2 (50 ng/ml) was used as a positive control reference material. The cells were allowed to migrate for 6 h after which the cells were fixed in buffered formalin. Migration was monitored via phase contrast microscopy. Cells that had migrated across the simulated wound margin were counted. At least five fields were counted for each data point. For studies involving migration through a coated membrane, the bottoms of trans-well inserts (3  $\mu$ m pore size) were coated with a solution of growth factor reduced Matrigel® (BD Biosciences, San Jose, CA) and allowed to dry. Target cells were then placed in the upper chamber and the inserts placed into wells with medium containing rhFGF-2 or F2A4-K-NS. The cultures were maintained for 18 h after which the cells on the membrane were fixed in buffered formalin. The cells on the lower side of the insert were then stained with bis-benzamide and the cells quantitated using fluorescence microscopy.

**Gelatinase assay.** The RMECs were seeded at 1.2×10<sup>5</sup>/ml in 24-well plates and allowed to attach overnight. The medium was replaced with serum-free DMEM containing peptides. After 48 h, the medium (supernatant) was harvested and the gelatinase activity was measured with the MMP Gelatinase Activity Assay Kit (ECM700) from Chemicon (Temecula, CA). The MMP Gelatinase Activity Assay Kit utilizes a biotinylated gelatinase substrate, which is cleaved by active gelatinases (MMP-2 and MMP-9). Remaining biotinylated fragments are then added to a biotin-binding 96-well plate and detected with streptavidin-enzyme complex. Addition of

enzyme substrate results in a colored product, detectable by its optical density at 450 nm. The optical density was inversely related to the amount of gelatinase in the sample.

***In vivo angiogenesis assay.*** The angiogenesis model was based on the use of Matrigel implants in C57BL/6 mice (21). The experiments were performed under an approved IACUC protocol in accordance with the United States Department of Agriculture, Department of Health and Human Services, and the NIH policies regarding the humane care and use of laboratory animals.

Animals were randomly assigned to each treatment group. Seven implant sites were used per treatment group. Growth factor reduced-Matrigel at 4°C (liquid state) was mixed with 0.1 or 1.0  $\mu$ g/ml of rhFGF-2 or F2A4-K-NS. Aliquots of 0.4 ml of Matrigel were injected subcutaneously in the mouse dorsal surface at the paraspinal space. The Matrigel, needle and syringe were kept on ice until the time of injection to prevent gelling in the needle. After 5 days the animals were euthanized and the Matrigel plugs were dissected away from the host tissue, photographed, and then frozen in sealed tubes and weighed. Two observers reviewed the photographs and rated the angiogenic response in double-blind fashion. The angiogenic score was rated on a scale of 0-3 as described by Lucidarme *et al* (22). Plugs with no blood vessels were assigned a score of 0, those with few tiny peripheral vessels were assigned 1, and those with larger vessels with shallow penetration scored 2, and those with several large vessels with deep penetration were scored 3.

For an additional measure of vascularization, total hemoglobin levels in the Matrigel plugs were determined using aqueous extracts as described elsewhere (22). A 60  $\mu$ l aliquot of the extract was removed for analysis, in which hemoglobin was converted to hemoglobin-ferricyanate after a 5-min incubation with 200  $\mu$ l of Drabkin's reagent (Sigma-Aldrich) and quantified with a microplate reader at 525 nm using bovine hemoglobin as standard.

## Results

SPR analysis was used to investigate the interaction between F2A4-K-NS and FGFR1, the predominant receptor for FGF-2. FGF-2 was used as a reference as SPR analysis of binding to FGFR1 has been described by others (25).

The intact F2A4-K-NS molecule, its receptor targeting domain, and its heparin-binding domain (HBD3) were each evaluated with FGFR1. F2A4-K-NS and its receptor targeting domain (F2A4RTDS) both bound to FGFR1, whereas the heparin binding domain (HBD3) did not (Table II). Of interest, however, was the observation that F2A4-K-NS had an equilibrium dissociation constant ( $K_D$ ) lower than that of its receptor targeting domain (Table II). The lower  $K_D$  of F2A4-K-NS most likely arises from the presence of two receptor targeting domains in each molecule. Nonetheless, the results indicate that the interaction of F2A4-K-NS with FGFR1 was mediated by the receptor-targeted domain, not the heparin binding domain.

When compared to FGF-2, F2A4-K-NS exhibited an association rate constant ( $K_a$ ) lower than that of FGF-2, indicative of a slower on-rate with FGFR1 (Table II). On the



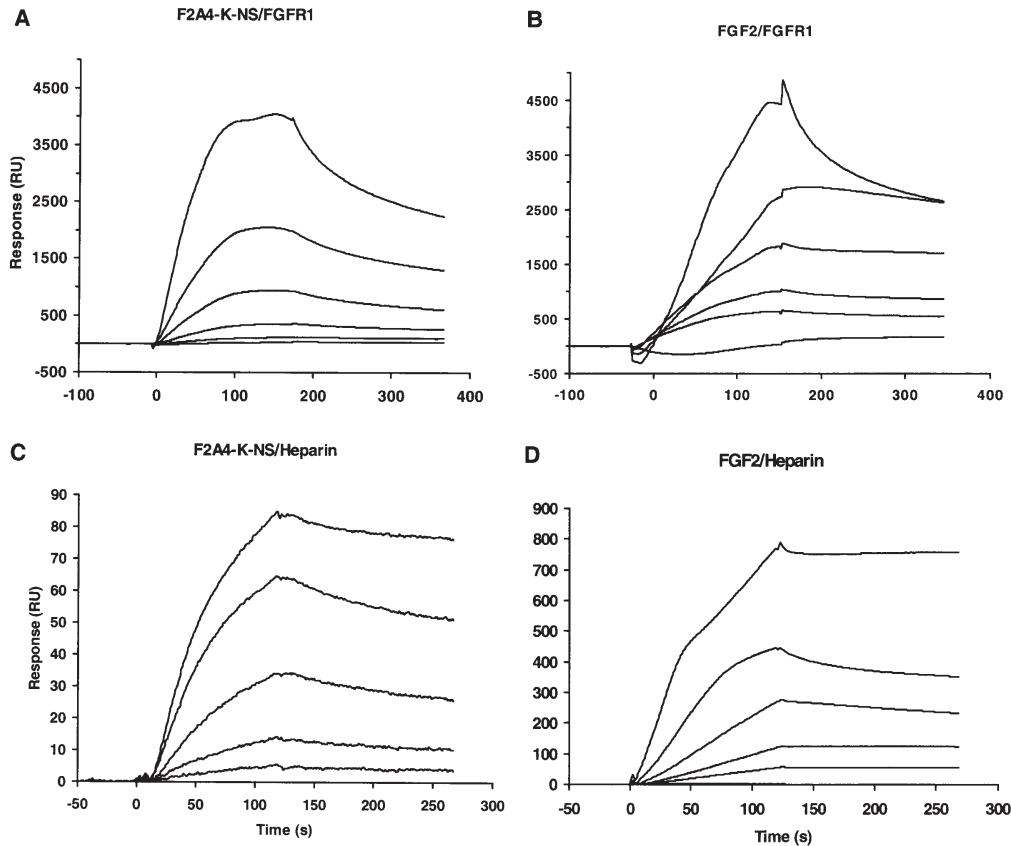


Figure 1. Sensorgrams of F2A4-K-NS binding to FGFR1. Surface plasmon resonance sensorgrams depict the interactions of F2A4-K-NS (A), FGF-2 (B) and with the FGFR1IIIc/Fc chimera. For (A) and (B), the analyte concentrations were 800, 400, 200, 100, 50, and 25 nM. Sensorgrams depict the interaction of F2A4-K-NS (C) and FGF-2 (D) with biotinylated low-molecular weight heparin. For (C) and (D), the analyte concentrations were 400, 200, 100, 50, and 25 nM. The kinetic and affinity constants calculated from these data are shown in Table II.

other hand, F2A4-K-NS also had dissociation rate constant ( $K_d$ ) lower than that of FGF-2, indicative of a slower off-rate. The estimated equilibrium dissociation constant ( $K_D$ ) for F2A4-K-NS was  $3.7 \times 10^{-7}$  M whereas that of FGF-2 was  $9.9 \times 10^{-8}$  M. The higher  $K_D$  of F2A4-K-NS may reflect the multiple receptor binding sites in FGF-2.

SPR assays were also used to establish that F2A4-K-NS could bind to heparin (Table II). F2A4-K-NS had been synthesized to contain a heparin binding domain to mimic FGF-2 (and the FGF-receptor complex) (23,24) which binds heparin a contributor to signal transduction. Relative to heparin, F2A4-K-NS had a higher  $K_a$ ,  $K_d$ , and  $K_D$  when compared to FGF-2.

As an aside, the  $K_D$  for the FGF-2/FGFR1 and FGF-2/heparin of 99 nM and 36 nM compare favorably with  $K_D$  values of 62 nM and 39 nM as reported by other investigators (25). Upon receptor binding and activation, FGF-2 induces a series of signaling events including activation of MAP kinase pathway. If F2A4-K-NS initiated signal transduction in a manner similar to FGF-2, the MAP kinase pathway would be activated with consequent phosphorylation of ERK1/2 (pERK1/2). F2A4-K-NS increased pERK1/2 in HUVEC by 30 min after addition (Fig. 2). A similar extent of phosphorylation (pERK1/2) was found in cells stimulated with either 5 ng/ml or 50 ng/ml of F2A4-K-NS. FGF-2, which was used as a reference, increased pERK1/2 at 10 min, and thereafter the pERK1/2 returned to basal levels.

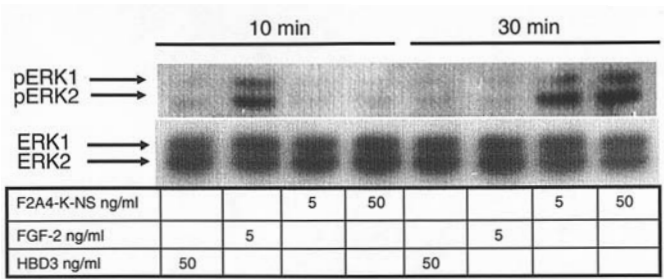


Figure 2. F2A4-K-NS induces MAP kinase ERK1/2 phosphorylation in HUVEC cells. HUVEC cells were treated with control peptide HBD3, FGF-2 or F2A4-K-N. After 10 or 30 min the cells were lysed and equal amounts of cellular proteins were fractionated by SDS-PAGE and transferred onto nitrocellulose. The Western blot was then probed with antibodies recognizing phosphorylated (Thr202 and Tyr204) ERK1/2 (upper panel) and total ERK1/2 (bottom panel).

The consequence of signal transduction by F2A4-K-NS would be expected to be expression phenotypes typically associated with FGF-2 stimulation. Such changes could include increased cell proliferation and migration, protease secretion, and angiogenesis. In the endothelial cell line RMEC, a relative increase in cell number was found for all F2A4-K-NS concentration tested between 1 and 200 ng/ml (Fig. 3). C3H10T1/2 cells also increased proliferation in response to F2A4-K-NS (data not shown).

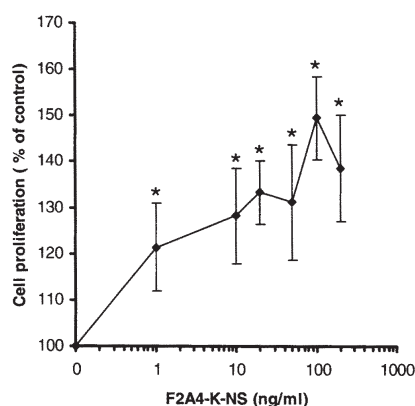


Figure 3. Effect of F2A4-K-NS on cell proliferation. REMC cells were seeded at 1000 cells per well of a 96-well plate and allowed to attach. The medium was changed to one containing low serum plus F2A4-K-NS and the cells incubated for 3 days after which MTS was added. Data are reported as the average  $\pm$  SD. Asterisks indicate  $p < 0.05$  determined by ANOVA followed by Bonferroni post-hoc multiple comparisons versus the control group.

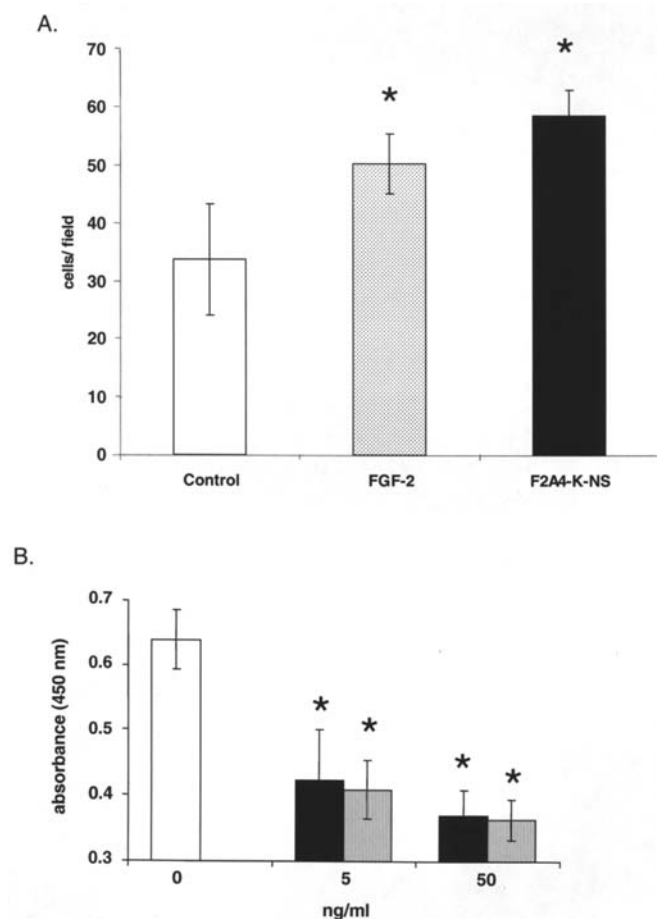


Figure 4. Phenotypic behavior of synthetic peptide F2A4-K-NS compared to recombinant FGF-2. (A) Migration of HUVECs across a simulated wound margin *in vitro*. HUVECs were grown to near confluency *in vitro*. A simulated wound was made by scraping cells away from the substrate and after 6-h migration into the gap was monitored via microscopy. (B) Induction of gelatinase activity in RMECs *in vitro*. RMECs were treated with vehicle or test compounds and after 48 h the medium was collected and gelatinase activity determined as described in Materials and methods. Open bar, control vehicle; hatched bar, FGF-2; solid bar, F2A4-K-NS. Data are reported as the average  $\pm$  SD. Asterisks indicate  $p < 0.05$  as determined by ANOVA followed by Bonferroni post-hoc multiple comparisons versus the control group.

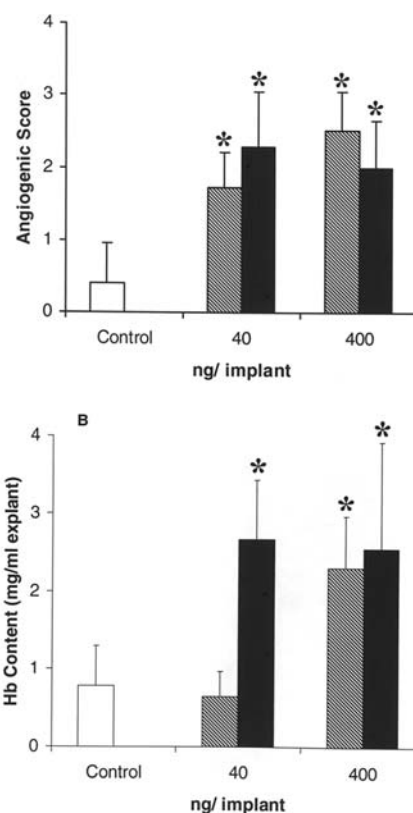


Figure 5. Angiogenesis of F2A4-K-NS in subcutaneous Matrigel implants. Open bar, control vehicle; hatched bar, FGF-2; solid bar, F2A4-K-NS. (A) Morphologic score of angiogenesis. Growth factor reduced Matrigel containing saline or test compounds was injected subcutaneously in mice and dissected out 5 days later. The gel plugs were scored on a scale of 0-3 as described in Materials and methods with 0 indicating no blood vessels. (B) Hemoglobin content of subcutaneous Matrigel implants in mice. The total hemoglobin (Hb) content of the implants was used as a quantitative index of angiogenesis. Hb was extracted from the plugs and detected colorimetrically. In all panels data are reported as the average  $\pm$  SD.

F2A4-K-NS treatment of cells also resulted in increased *in vitro* migration of HUVECs across a simulated wound margin (Fig. 4A). To ensure that migration was in fact stimulated, a different evaluation system, transwell migration, was used. In this system, migration was also increased in HUVECs and C3H10T1/2 to levels averaging 136% and 170% of control values, respectively. FGF-2 increased migration by 119% and 116%, respectively for HUVEC and C3H10T1/2 cells. As FGF-2 can influence the production of gelatinases in a number of cell lines (26-28), F2A4-K-NS was evaluated for its ability to increase gelatinase activity. F2A4-K-NS treatment (5 and 50 ng/ml) of RMEC cells resulted in an increased secretion of gelatinase (Fig. 4B). rhFGF-2, which was used as a positive control, also increased the secretion of gelatinase.

Endothelial cell migration and gelatinase secretion are two key steps that associate with angiogenesis *in vivo*. F2A4-K-NS induces cell migration and gelatinase secretion with similar efficacy as FGF-2, suggesting that it might also induce angiogenesis when introduced into animals. In this study, we used an *in vivo* Matrigel plug assay to monitor the effects of F2A4-K-NS on angiogenesis, a hallmark of FGF-2. F2A4-K-NS stimulated angiogenesis (Fig. 5A), as did rhFGF-2, which was used as a system control. Angiogenic response as

monitored by morphological scoring indicated a significant response to F2A4-K-NS at 40 and 400 ng/implant, a response that was verified by histological examination (data not shown). A quantitative assessment of the hemoglobin content of the plugs also indicated a significantly higher amount of hemoglobin in plugs that were implanted with F2A4-K-NS when compared to controls (Fig. 5B). The control peptide, HBD3 containing only the heparin binding domain (minus the receptor targeting domain) did not stimulate angiogenesis.

## Discussion

The central hypothesis of this study was that peptide F2A4-K-NS could mimic aspects of FGF-2 bioactivity, and the results support this contention. F2A4-K-NS is a multi-domain peptide with a branched design that incorporates two copies of an FGF receptor binding sequence from FGF-2, residues 106 to 120. A previous report showed that the linear version of this region, designated as FGF-(106-120) by Baird and colleagues, inhibits FGF-2 binding to FGF receptor on BHK cells and simulates thymidine incorporation in 3T3 cell when used at 500  $\mu$ g/ml, suggesting the linear peptide is a partial agonist (14). The FGF-(160-120) was also shown to be an FGF-2 weak antagonist (15). In this study, F2A4-K-NS, a branched peptide construct with two receptor binding domains, possessed 3 times higher affinity to FGFR1 than FGF-(106-120). Furthermore, F2A4-K-NS also possessed FGF-2-like activity in a similar dose range as recombinant FGF-2.

As a mimetic, F2A4-K-NS would be expected to exhibit similar, but not necessarily equivalent, binding to FGF receptors; and at least broadly result in the initiation of signal transduction, cell behavior, and phenotypes associated with FGF-2. The results from this study established that F2A4-K-NS binds to the receptor FGFR1 and also to heparin. It activated the receptor signaling cascade in cells as evidenced by increases in pERK1/2, and triggered a series of changes in cell behavior similar to that of induced by FGF-2. These changes included an increase in cell migration and cell proliferation, an increase in the secretion of gelatinase, and a stimulation of angiogenesis *in vivo*. Also, the quantities of F2A4-K-NS needed to illicit those effects were similar to that of rhFGF-2.

F2A4-K-NS bound to FGFR1 at an affinity approximately one third that of rhFGF-2. Once bound, however, it had almost a 10 times slower dissociation rate from the receptor compared to FGF-2. The slower dissociation rate could arise from the juxtaposition of the dual receptor targeted sequences of F2A4-K-NS which would tend to reduce dissociation. A slow dissociation rate of F2A4-K-NS from the receptor might benefit overall receptor activation by kinetically favoring commitment of the receptor cytoplasmic domain to the active state.

Yet, in spite of the slower dissociation rate, the net equilibrium binding constant of F2A4-K-NS to FGFR1 was lower than that of FGF-2. Nonetheless, the quantities of F2A4-K-NS needed to illicit biological effects were similar to that of rhFGF-2. This indicates that mechanisms in addition to receptor binding are likely involved in the bioactivity of F2A4-K-NS.

One factor that could contribute to the bioactivity of F2A4-K-NS is its ability to bind to heparin, and presumably, heparan sulfate found on cell surfaces. In the current study,

the equilibrium binding constant of F2A4-K-NS to heparin was approximately 2 times higher than that of FGF-2. F2A4-K-NS binding to heparin could mediate an increased biological effectiveness. This suggests a conceptual similarity to a signaling complex composed of heparin/FGF/FGFR as proposed by Ornitz (29). The signaling complex requires the assembly of a ternary complex of FGF, FGFR, and heparin (or heparan sulfate proteoglycan). Two models explain the association: the ternary complex can be formed by heparin binding two 1:1 FGF-FGFR complexes or heparin crosslinking two FGFs that in turn bring together two receptor chains (30). The binding of F2A4-K-NS to FGFR1 and also heparin would be conceptually permissive in either model.

The result of receptor dimerization, regardless of the exact mechanism, would lead to activation of the MAP kinase pathway and result in phosphorylation of ERK1/2. Treatment of cells with F2A4-K-NS resulted in an increase in ERK1/2 phosphorylation. Interestingly, the phosphorylation of ERK1/2 induced by F2A4-K-NS appeared to take longer than that of FGF-2 (30 min compared to 10 min). These observations appear consistent with the F2A4-K-NS/FGFR1 binding kinetics with a slow association rate, and also a slow dissociation rate.

F2A4-K-NS treatment of cells also resulted in increased proliferation of endothelial cells and fibroblasts, increased cellular migration, and up-regulation of gelatinase secretion. While it is unlikely that F2A4-K-NS will be established to mimic each and every action of FGF-2, the changes in cell behavior observed in this study are consistent with known actions of FGF-2. Taken together with the other data presented here the observations support the contention that F2A4-K-NS mimics FGF-2.

Cell migration, cell proliferation, and protease secretion are processes, among others, involved in FGF-2 stimulated angiogenesis. Induction of angiogenesis is a well-known outcome associated with FGF-2 and is the centerpiece of many of the experimental therapies with rhFGF-2. F2A4-K-NS stimulated angiogenesis *in vivo* in a subcutaneous assay, as did FGF-2, again strengthening the contention that F2A4-K-NS mimics FGF-2.

In summary, we demonstrated that it is possible to produce a peptide that mimics at least some of the actions of FGF-2; that is biologically active in quantities similar to the parental molecule; and that is active *in vivo*. The ability to construct synthetic molecules such as F2A4-K-NS allows for the systematic exploration of structure-function relationships, and consequent fine-tuning of the biological properties. Additionally, chemical synthesis is free of inherent biological contamination (nucleic acids, viruses, prion proteins, etc.) which may complicate purification of recombinant growth factors. Peptide synthesis uses readily available building blocks starting materials and is scalable. Also, it enables complete control over design and incorporation of non-coded elements which could improve biological activity.

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