The action of GLP-1 and exendins upon glucose transport in normal human adipocytes, and on kinase activity as compared to morbidly obese patients

VERÓNICA SANCHO¹, BERNARDO NUCHE¹, LUIS ARNÉS¹, JESÚS CANCELAS¹, NIEVES GONZÁLEZ¹, MARIANO DÍAZ-MIGUEL², ANTONIO MARTÍN-DUCE³, ISABEL VALVERDE¹ and MARÍA L. VILLANUEVA-PEÑACARRILLO¹

Departments of ¹Metabolism, Nutrition and Hormones, ²General Surgery, Fundación Jiménez Díaz, Avda. Reyes Católicos 2, 28040 Madrid; ³Department of Surgery, Hospital Príncipe de Asturias, Universidad de Alcalá de Henares, Ctra. Meco (M-121) s/n, Alcalá de Henares, 28805 Madrid, Spain

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Abstract. A role of GLP-1 (glucagon-like peptide-1) in the recovery of the metabolic conditions of morbidly obese patients after bariatric surgery has been proposed. Exendin 4 (Ex-4) and exendin 9 (Ex-9) both have GLP-1-like effects upon glucose metabolism in human myocytes. We investigated in normal human adipocytes the effect of GLP-1, Ex-4 and Ex-9, compared with insulin upon the activity of PI3K, PKB, MAPKs and p70s6 kinases, and the participation of these enzymes in their action upon 2-deoxy-D-glucose transport by using potential inhibitors. The study was extended to morbidly obese patients. In normal subjects, GLP-1, Ex-4 and insulin, but not Ex-9, increased glucose uptake. In addition, GLP-1 and Ex-4 stimulated PI3K and MAPKs, similar to insulin, but not PKB. Ex-9 only enhanced PI3K, while none affected p70s6k. Inhibition of both PI3K and MAPKs blocked the stimulatory action of GLP-1, Ex-4 and insulin upon glucose transport. In obese patients, basal PI3K, PKB and MAPK activity was, as a rule, lower than that in normal subjects, while cells maintained their normal incremental response to GLP-1, Ex-4 or insulin; Ex-9 induced a clear stimulation of p42 MAPK. In summary, in normal human adipocytes, GLP-1 and Ex-4 have a protein kinase-dependent increasing effect upon glucose transport, which is impaired in obese patients. The participation of GLP-1 in the normalization of the metabolic conditions of the obese may occur through its effects on lipid metabolism or through effects upon glucose transport and/or metabolism in the liver and muscle, which in human obesity remain to be investigated.

Introduction

GLP-1 (glucagon-like peptide-1) has glucose-dependent insulinotropic action and insulin-independent antidiabetic character (1), apart from proven insulinomimetic properties per se (2). A stimulatory effect upon both the expression of glucotransporter genes (3) and the transport and metabolism of glucose in extrapancreatic tissues (2-5) has been reported. The effect of GLP-1 in the liver (6) and muscle (7,8) is exerted through specific receptors (9-11), structurally or functionally distinct (6,7,12) from that in the pancreas (13). But in normal rat and human adipocytes, GLP-1 has, in addition, a dual effect upon the lipid metabolism (5,14-16), being both lipogenic and lipolytic depending upon the dose. Thus, GLP-1 could be acting through two different types of receptors in fat tissue where, in fact, this peptide not only stimulates inositolphosphoglycan (IPG) generation (17) but, distinct to its action in the liver and muscle (8,17-19), also increases cellular cAMP content (14). Concerning the effects of GLP-1 in these three extrapancreatic tissues, several protein kinases have been implicated (5,20,21).

GLP-1 inhibits gastric emptying and controls food intake by enhancing satiety (1). Moreover, a lower GLP-1 secretion, both basal and oral glucose- or meal-stimulated, has been reported in morbidly obese patients (22-24), which is reversed after bariatric surgery (24).

Exendin(1-39)amide (Ex-4), a peptide of non-mammalian nature, is 53% structurally homologous with GLP-1. It is also insulinotropic (25), and has GLP-1-like effects upon parameters related to the glucose metabolism in rat liver and skeletal muscle (26). Its truncated form exendin(9-39)amide (Ex-9), has been proven to be an antagonist of the GLP-1 receptor in various cell systems (27) and also of its effects in rat pancreas (28), liver cells and muscle tissue (26). Similar to Ex-4, it is agonist of its action in human muscle (8,29), and also of its receptor in adipocyte (30) and myocyte (12) cell lines.

In a recent study, we reported the effect of GLP-1 and exendins upon glucose transport and lipid metabolism in adipocytes from a type 2 diabetic rat model compared to

Correspondence to: Dr Maria L. Villanueva-Peñacarrillo, Metabolismo, Nutrición y Hormonas, Fundación Jiménez Díaz, Avda. Reyes Católicos 2, 28040 Madrid, Spain E-mail: mlvillanueva@fjd.es

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normal rat, which has further elucidated the mechanism of the beneficial action of these peptides in the diabetic state (5). Also recently, the reduced lipogenic effect and enhanced lipolytic action of GLP-1 has been observed in the fat tissue of morbidly obese patients, an effect adequate for its therapeutic use in obesity (31).

In this study we have explored the effect of GLP-1 and exendins, compared to that of insulin, on glucose transport in normal human adipocytes, the possible kinases involved and the differences in their action in cells from morbidly obese subjects.

Materials and methods

Reagents. The following reagents were used: human GLP-1 (7-36)amide (GLP-1, Bachem AG, Bubendorf, Switzerland); porcine insulin (Novo Biolabs, Bagsvaerd, Denmark); exendin(1-39)amide (Ex-4) and exendin(9-39)amide (Ex-9) (gifts from Dr John Eng, VAMC, NY, USA); PD98059 (PD) (Calbiochem®, La Jolla, CA, USA); collagenase P (Roche Diagnostics GmbH, Mannheim, Germany); ethylene glycol bis(ß-aminoethyl ether)-N,N,N'N'-tetraacetic acid (EGTA), ethylenedinitrilo-tetraacetic acid (EDTA), bovine serum albumin (BSA), wortmannin (W), cytochalasin B, phenylmethylsulfonyl fluoride (PMSF), phosphatidylinositol and phosphatidyl-serine (Sigma Chemical Co., St. Louis, MO, USA); aprotinin (Trasylol®, Bayer Leverkusen, Germany); 2-deoxy-D-[1,2-³H(N)]glucose (2-DOG, Moravek Biochemicals, Brea, CA, USA); Ultima Gold scintillation liquid (Packard, Gröninger, The Netherlands); [y³²P]ATP (30 Ci/nmol), horseradish peroxidase-conjugated donkey anti-rabbit immunoglobulin, Rainbow markers, ECL-Western blotting kit, Hyperfilm ECL (Amersham Biosciences, Buckinghamshire, UK); rabbit anti-total and antiphosphorylated form of p44/42MAP kinases, p70s6k and PKB (Cell Signalling Technology, New England Biolabs, Beverly, MA, USA); and rabbit anti-PI3-kinase p85 (Upstate Biotechnology, Lake Placid, NY, USA). All other commonly used chemicals were from Sigma or Merck (Merck Pharma Quimica, S.A., Barcelona, Spain).

Biological material. Residual samples of subcutaneous abdominal fat tissue were collected after informed consent was obtained from 31 normal subjects (14 female/17 male; 58 ± 3 years old; fasting plasma D-glucose, 5.00 ± 0.13 mM) during an inguinal hernia or plastic surgery, and from 9 morbidly obese patients (7 female/2 male; 43 ± 3 years old; BMI, 49.6 ±2.7 kg/m²; fasting plasma D-glucose, 111.2 ± 6.3 mM; cholesterol, 187.3 ± 13.0 mg/dl; triglycerides, 128.1 ± 11.7 mg/ml; HDL, 53.1 ± 4.9 mg/ml; and LDL, 106.8 ± 27.5 mg/ml) undergoing bariatric surgery. The patients were randomly selected. In all cases, comparisons between normal subjects and obese patients were restricted to results obtained under similar experimental conditions.

The study was approved by the Ethics Committee of the Fundación Jiménez Díaz, Madrid, in accordance with the guidelines proposed in The Declaration of Helsinki.

Cells. Adipocytes were isolated at 37°C by enzymatic digestion with collagenase P from subcutaneous fat tissue

(32). Then, cells were re-suspended in KRB supplemented with 10.9 mM HEPES, 500 KIU/ml Trasylol, BSA and without or with D-glucose, pH 7.4, at a density of 10⁶ cells/ml.

Kinase activity. Cells (106) were first incubated for 15 min in 1 ml KRB containing 30 mM HEPES, 500 KIU/ml Trasylol, 1% BSA, and 3.3 mM D-glucose, pH 7.4, followed by a 3-min incubation in the absence or additional presence of GLP-1, Ex-4, Ex-9 or insulin. For the measurement of PI3K activity and that of phosphorylated PKB, p70s6k and p44/42 MAPKs, the cells were homogenized and maintained at 4°C in 1.25% Triton containing 250 mM sucrose, 20 mM TrisHCl, pH 7.6, 2.5 mM MgCl₂, 50 mM 2-mercaptoethanol, 1.2 mM EGTA, 5 mM Na₄P₂O₇, 50 mM NaF, 1 mM Na₃VO₄, 30 U/ml bacitracin, 2 µM leupeptin, 2 µM pepstatin and 2 mM PMSF. After centrifugation at 10,000 x g, the infranatant containing cytosol and solubilized membranes was kept at -70°C until assay. An aliquot volume was taken from all membrane preparation samples for protein content determination (33).

PI3K activity was estimated as PIP₂ phosphorylation to PIP₃ (phosphatidylinositol bi- or tri-phosphate, respectively), in p85 immunoprecipitates obtained by treating each adipocyte membrane preparation with anti-PI3-kinase p85 and subsequent coupling to protein A agarose. The immunoprecipitates were incubated for 20 min at room temperature with 20 μ M [γ^{32} ATP] (5 μ Ci/nmol) in 6.25 mM HEPES, 5 mM MgCl₂ and 0.25 mM EGTA, and in the presence of 0.25 mg/ ml phosphatidylinositol/phosphatidylserine as substrate. The reaction was interrupted by the addition of 400 μ l chloroform/ methanol/HCl (1:2:1, v/v), 150 µl chloroform and 150 µl HCl. After centrifugation (10,000 x g), the organic phase was treated with an equal volume of methanol/100 mM HCl/ 2.5 mM EDTA (1:1:1, v/v), and the new organic phase was separated by centrifugation and then speed-vac dried. The lipidic extract, redissolved in chloroform, was spotted, together with PIP₃ standard, on a silicagel TLC plate, and developed in n-propanol/acetic acid/H₂O (66:2:33, v/v). Plates were dried, and radioactive PIP₃ was subsequently visualized by autoradiography and analyzed by densitometric scanning. In all experiments, the densitometric measurement of the band corresponding to cells incubated in the absence of peptide, was used as the control value (20).

For the measurements of the phosphorylation degree of the respective protein kinases by immunoblotting, equal amounts of each solubilized membrane preparation sample were subjected to SDS-PAGE (34) on an 8% resolving gel in parallel with molecular weight markers. The separated proteins were then transferred to a nitrocellulose membrane in a semidry system (Trans-blot SD semidry transfer cell, Bio-Rad). For immunodetection, a Western blotting kit was used following the manufacturer's instructions, using total and phosphorylated respective antibody for each protein kinase, and a horseradish peroxidase-conjugated donkey anti-rabbit immunoglobulin second antibody, with detection by the enhanced chemiluminescence method, and quantitation by densitometric scanning of the autoradiography (20). The densitometric measurement of the phosphorylated protein kinase was normalized with respect to the total (percent of

	PI3K	РКВ	p42 MAPK	p44 MAPK	p70s6k
Normal subjects					
Basal value	100	100	100	100	100
GLP-1	155±21 ^b	90±11	145±17 ^b	123±4 ^b	110±19
Ex-4	145±6 ^b	74±20	158±16 ^b	150±7 ^b	109±14
Ex-9	173±22 ^b	75±14	107±13	111±11	109±11
Insulin	151±15 ^b	231±33 ^b	141±7 ^b	135±18 ^b	94±13
Obese patients					
Basal value	100	100	100	100	100
GLP-1	158±15 ^b	99±6	135±9 ^b	124±7 ^b	101±8
Ex-4	144±5 ^b	107±11	172±10 ^b	157±9 ^b	106±11
Ex-9	162±18 ^b	100±11	154±14 ^b	120±17	105±7
Insulin	147±11 ^b	233±11 ^b	131±6 ^b	137±9 ^b	95±10

Table I. Effect of 10⁻⁹ M GLP-1, Ex-4, Ex-9 or insulin upon PI3K, PKB, p42/44 MAPK and p70s6k activity in adipocytes from 4-7 normal subjects and 4-6 morbidly obese patients.^a

^aData are means ± SEM; ^bp<0.05 vs basal value.

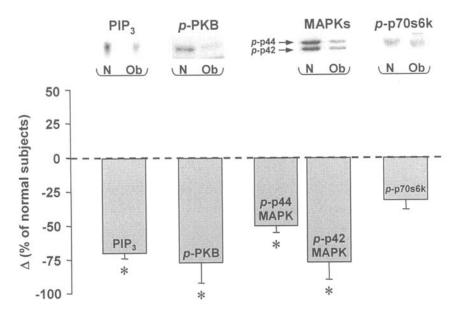


Figure 1. Basal PI3K, PKB, p44/42 MAPK and p70s6k activity in adipocytes from morbidly obese patients (Ob) compared to that in normal subjects (N). Representative immunoblot and percent increment (mean \pm SEM) relative to normal subjects, *p<0.05 vs normal basal value.

phosphorylated/total kinase), and the value obtained in the adipocytes incubated in the absence of peptide was used as the control value.

Glucose transport. Cells (10⁵) were incubated for 15 min at 37°C in 400 μ l KRB, 10.9 mM HEPES, 500 KIU/ml Trasylol and 2% BSA, pH 7.4, and either without (basal) or with GLP-1, Ex-4, Ex-9 or insulin, alone or combined with 10⁻⁶ M W -PI3K inhibitor- or 2.5x10⁻⁵ M PD -p44/42 MAPK inhibitor. This was followed by a 3-min incubation in the additional presence of 0.2 μ Ci (6.5 pmol) of 2-deoxy-D-[1,2-³H(N)] glucose (final concentration, 16.3 nM 2-DOG). Adipocytes, after being separated at 10,900 x g in 100 μ l dioctyl phthalate, were added to a 3-ml scintillation liquid for β -counting. The

total D-glucose content was corrected for the unspecific Dglucose uptake value, obtained in cell samples from each experiment treated in parallel with 0.175 mM cytochalasin B (5).

Statistical study. All data were presented as mean values (\pm SEM) together with the number of individual determinations. The statistical significance of differences was tested using either analysis of variance or the Student's t-test.

Results

Effect on kinase activity. In adipocytes from a total of 7 normal subjects (Table I), 10⁻⁹ M GLP-1, and its two

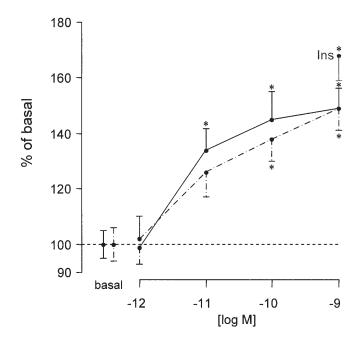


Figure 2. Effect of GLP-1 (—), Ex-4 (-·-) and insulin upon 2-deoxy-D-glucose uptake in adipocytes from normal subjects. Data (mean \pm SEM) are expressed in percent of the paired basal value found within the same experiment, and refer to 11 subjects. *p<0.02 vs basal value.

structurally related peptides, Ex-4 and Ex-9, all induced a clear increase (p<0.03 or lower) in the basal PI3K activity obtained in cells incubated in the absence of the hormones/ peptides. These effects were not significantly different from one another (p<0.9), with an overall mean value of $157\pm10\%$ of basal (n=13), indistinct (p=0.752) from that exerted by 10⁻⁹ M insulin (p<0.001). Except insulin, by which a clear increment in PKB phosphorylation was detected (p<0.001), GLP-1, Ex-4 and Ex-9 all failed to affect this enzyme. Ex-9 did not modify p42 or p44 MAP kinase activity, while both GLP-1 and Ex-4 induced a clear stimulation in p42 MAPK (overall mean value: 152±11% of basal, n=8, p<0.01), similar in magnitude (p=0.504) to that exerted by insulin (p<0.01); and the same was measured in the case of p44 MAPK (GLP-1, Ex-4 and insulin overall mean value: $134\pm7\%$ of basal, n=14, p<0.001). None of the hormones/peptides tested modified the p70s6 kinase phosphorylation degree (overall mean value: 106±5% of basal, n=21).

In adipocytes from a total of 5 morbidly obese patients (Table I), the pattern of the respective incremental response of the kinases tested to each of these four hormones/peptides, was basically the same as that observed in normal adipocytes, in contrast to the response to Ex-9, by which a clear increase in p42 MAP kinase activity (p<0.001) was measured.

The respective basal activity of PI3K, PKB and p42/44 MAP kinases (Fig. 1) in adipocytes from these obese patients was, in all cases, reduced (overall mean value: $58\pm5\%$ of normal, n=20, p<0.001) with respect to the corresponding value obtained in normal subjects (overall mean: $100\pm7\%$, n=21), unlike that of p70s6 kinase, whose activity (obese: $84\pm9\%$ of normal, n=5) showed no apparent difference (p=0.601) with that in normal cells ($100\pm12\%$, n=4).

Uptake of 2-deoxy-D-[1,2-³H]glucose. In adipose cells from normal subjects (Fig. 2), GLP-1 stimulated the basal glucose uptake (15.3±1.7 fmol glucose/10⁵ cells, n=37) in a doserelated manner. The increment induced was statistically significant at 10⁻¹¹ M up to 10⁻⁹ M of the peptide, with a maximum effect from 10⁻¹⁰ M GLP-1. At 10⁻⁹ M, the enhancing action gave a value indistinct (p=0.367) of that induced by the equimolar amount of insulin $(168\pm9\% \text{ of basal}, p<0.001)$. The GLP-1 structurally related peptide Ex-4 also exerted an increase in the corresponding basal glucose uptake value $(17.5\pm2.2 \text{ fmol glucose}/10^5 \text{ cells}, n=30)$; statistically significant at 10⁻¹⁰ and 10⁻⁹ M, concentrations at which the increments induced were indistinct of those exerted by the corresponding equimolar amounts of GLP-1. Ex-9 failed to affect the glucose transport at any concentration tested; the value obtained at 10-9 M representing only 114±6% of that of the basal value (14.1 \pm 2.1 fmol glucose/10⁵ cells, n=29).

The respective stimulatory action of 10^{-9} M GLP-1, Ex-4 or insulin (p<0.001 in all cases) upon glucose transport in normal subjects [overall mean: $140\pm7\%$ of basal (11.5±1.3 fmol glucose/10⁵ cells), n=52] was completely abolished (p<0.001) by wortmannin (overall mean: $91\pm5\%$ of basal, n=62), a PI3K inhibitor (Table II), while wortmannin alone did not apparently affect the basal value ($87\pm5\%$ of basal, n=29, p=0.092). The same blocking effect was detected (p<0.001) when PD 98059, MAP kinase inhibitory agent, was present. The overall mean value for the effect of these three hormones/peptides in the additional presence of PD 98059 represented $89\pm5\%$ of

Table II. Effect of wortmannin and PD 98059, inhibitors of PI3K and MAPK respective activities, on basal and 10⁻⁹ M hormone-stimulated 2-deoxy-D-glucose uptake in adipocytes from 4-6 normal subjects.^a

	Nil	Insulin	GLP-1	Ex-4
Wortmannin				
Nil	100±5	143±6 ^b	134±7 ^b	141±8 ^b
Inhibitor	87±5	100±6	87±10	84±7
PD 98059				
Nil	100±9	139±7 ^b	140±8 ^b	137±8 ^b
Inhibitor	86±6	89±9	85±9	95±12

^aData are means ± SEM; ^bp<0.001 vs basal value.

the basal value (14.1±2.3 fmol glucose/10⁵ cells, n=35) compared to that in its absence (overall mean: $139\pm4\%$, n=32, p<0.001). The MAP kinase inhibitor *per se* did not significantly alter the basal value (86±6\%, n=15).

The stimulatory action of GLP-1 and Ex-4 upon glucose transport in normal human adipocytes differed from our previous observations (31) in cells from morbidly obese patients, in which no effect of either of these two peptides could be detected, while cells maintained their normal response to insulin, accompanied also by a normal basal value.

Discussion

We have previously provided information on the role of GLP-1 in the control of glucotransporter expression, glucose and lipid metabolism in human and/or rat liver, muscle and fat tissue (2), and on the characteristics of its effects on the diabetic state (2,5,35). In recent studies of morbidly obese patients, we have also documented the possible participation of GLP-1 in the normalization of their metabolic conditions after bariatric surgery (24), and its action and that of exendins upon glucose transport and lipid metabolism of adipocytes (31).

The present study has explored the effect of GLP-1 and exendins upon the activity of kinases (known to be actively involved in other extrapancreatic tissues) (20,21,29) in the adipose tissue of morbidly obese patients, compared to normal subjects. All of the hormones/peptides tested (GLP-1, Ex-4, Ex-9 and insulin) induced a clear and similar increase in the PI3K activity in adipocytes from the obese group, equivalent in magnitude to that exerted in normal cells. No modification in PKB, though, was observed in any of the two groups with GLP-1 or either exendin, while the obese cells maintained their normal response capability to insulin. No effect on p70s6k was measured under any condition, indicating the absolute lack of participation of this enzyme. Our data also revealed that both GLP-1 and Ex-4 stimulated, like insulin, p42 and p44 MAP kinase activity in normal and obese cells. However, Ex-9, by which no effect could be detected in normal adipocytes, induced a clear increment in the p42 MAPK activity in the obese patients.

Despite the fact that the incremental responses of PI3K, PKB and MAPKs to GLP-1, Ex-4 and insulin in the obese cells were, as a rule, the same as those in normal adipocytes, this was not the case regarding the corresponding basal level of each enzyme which, in fact, proved to be significantly reduced. Apart from further considerations, the latter suggests that the net corresponding activity reached after stimulation with GLP-1, Ex-4 or insulin in the obese group was, in all cases, much lower than that in normal subjects. An alteration in cellular enzyme activity has been previously reported in other disorders associated with impaired glucose metabolism. Relative to this, a higher basal PI3K activity in muscle tissue or cells was detected in type 2 diabetic patients (35,36), accompanied by both a lower than normal glucose transport and glycogen synthase α activity and a higher glycogen synthesis (35). In fact, in obese patients (31), no significant effect of GLP-1 upon glucose transport, and a reduction of the adipocyte basal value by both exendins, were reported, while in the fat cells of the present group of normal subjects, GLP-1 and Ex-4 stimulated the uptake of glucose to

the same level as that reached by equimolar concentrations of insulin. In the normal group, no modification of the basal value by Ex-9 was measured, in contrast to its observed GLP-1-like effects upon parameters of the glucose or lipid metabolism in other human extrapancreatic tissues (8,29,31), or to its GLP-1-agonistic character in adipocyte (30) and myocyte (12) cell lines. Nevertheless, the present results do not allow us to conclude whether GLP-1 and Ex-4 exert their effects through the same or a different signalling pathway, but indicate a relevant role of PI3K and MAPKs in their action upstream in glucose metabolism. In fact, the GLP-1or Ex-4-induced increase in glucose uptake in normal adipocytes was highly reduced by respective inhibitors of these cellular enzymes.

The present results on PKB, which is stimulated by insulin but not by GLP-1 or either exendin in human adipocytes, were also observed in normal rats (5), whereas this negative effect was reversed. However, in the adipose cells of a STZ-induced type 2 diabetic rat model, where there was also evidence of other anomalies such as an increased basal PI3K activity. However, apart from possible species differences in hormonal action, the obese patients in this study, as far as we are aware, did not present a defect in insulin secretion or action. As a matter of fact, a section of the total participating population of obese patients here, all having the same characteristics, GLUT-4 expression, protein and mRNA, showed only a slight trend toward lower values as compared to normal subjects (31), contrary to other reports (37-39) in which patients suffering obesity associated with insulin resistance or type 2 diabetes presented significantly reduced adipocyte glucotransporter expression.

The increasing effect of Ex-9 upon p42 MAPK activity, solely detected in the obese group, associated with no modification of the adipocyte glucose uptake basal value as it occurs in normal conditions, indicates a possible correlation of this enzyme with an action of the peptide in fat tissue other than that upon glucose transport, such as that reported upon lipogenesis in morbidly obese patients (31).

To conclude, GLP-1 and Ex-4, like insulin, but not Ex-9, increase glucose transport in normal human adipocytes, and this effect is mediated by an activation of protein kinases. The fact that this stimulatory action does not seem to be maintained in morbidly obese subjects, suggests that the proposed participation of GLP-1 in the normalization of the metabolic conditions of these patients may occur through its effects upon the lipid metabolism; but perhaps also through its known action upon glucose transport and/or metabolism in other extrapancreatic tissues such as liver and muscle, which in human obesity remains to be investigated.

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