

Anti-adipogenic effects of 1,25-dihydroxyvitamin D3 are mediated by the maintenance of the wntless-type MMTV integration site/ β -catenin pathway

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Abstract. 1,25-dihydroxyvitamin D3 (1,25(OH)₂D₃), the active metabolite of vitamin D, was found to have anti-adipogenic activity, however, its mechanism of action has not been fully elucidated. In this study, 3T3-L1 preadipocytes were differentiated in the presence and absence of 1,25(OH)₂D₃, and the expression of the genes and proteins of the wntless-type MMTV integration site (WNT)/ β -catenin pathway were analyzed. While the expression of the members of the WNT/ β -catenin pathway were significantly downregulated during the adipogenesis of untreated 3T3-L1 cells, 1,25(OH)₂D₃ treatment was found to maintain the WNT/ β -catenin pathway. Among the members of the WNT/ β -catenin pathway, the levels of WNT10B and disheveled (DVL)2 as well as the phosphorylation of glycogen synthase kinase (GSK)3 β were maintained by 1,25(OH)₂D₃ treatment. The levels of nuclear β -catenin, which were downregulated during adipogenesis, were also maintained by 1,25(OH)₂D₃ treatment. The results of this study suggested that the anti-adipogenic effect of 1,25(OH)₂D₃ was mediated by the maintenance of the WNT/ β -catenin pathway, which was normally downregulated during adipogenesis.

Introduction

Obesity is a serious health problem worldwide, because it is a major risk factor for type 2 diabetes, hypertension, cardiovascular disease, and cancer (1). Overaccumulation of adipose tissue is the fundamental phenomenon of obesity (2,3). Therefore, the understanding of the molecular mechanisms underlying adipose tissue formation, i.e., adipogenesis, is required for the effective prevention and treatment of obesity. Adipogenesis is a complex process accompanying the differentiation of preadipocytes into adipocytes followed by the intracellular fat accumulation (4-6). 3T3-L1 preadipocyte has been used as an *in vitro* model of adipogenesis for decades (7), and its adipogenic differentiation was mediated by the upregulations of peroxisome proliferator-activated receptor (PPAR) γ and CCAAT/enhancer binding protein (C/EBP) α followed by the inductions of lipid metabolizing enzymes such as fatty acid binding protein (FABP)4, lipoprotein lipase (LPL) and fatty acid synthase (4-6).

Vitamin D is produced from 7-dehydrocholesterol in the skin by ultraviolet radiation and converted in the liver to 25-hydroxyvitamin D, followed by its conversion to 1,25-dihydroxyvitamin D3 (1,25(OH)₂D₃) in kidney and other tissues. 1,25(OH)₂D₃, the bioactive form of vitamin D, plays multiple physiological roles including the maintenance of calcium and phosphorus levels for bone homeostasis (8). Accumulating evidence suggests that a potential link exists between vitamin D deficiency and obesity in human population (9), and vitamin D deficiency was proposed as one of the causes of common obesity (10). Anti-obesity effect of vitamin D was suggested by the finding that 1,25(OH)₂D₃ efficiently inhibit adipogenesis at nanomolar concentration (11). 1,25(OH)₂D₃, the bioactive form of vitamin D, was found to inhibit the adipogenesis of 3T3-L1 cells and primary cultures of mouse epididymal fat (12). It is necessary to understand the anti-adipogenic mechanism of vitamin D for the effective prevention and treatment of obesity, however, the molecular mechanism for the anti-adipogenic effect of 1,25(OH)₂D₃ was not fully elucidated until now.

As a possible candidate of the mediator for the anti-adipogenic effect of vitamin D, we focused on the wntless-type MMTV integration site (WNT)/ β -catenin pathway. The WNT/ β -catenin pathway is known to be involved in various cellular processes such as differentiation, development, proliferation and tumorigenesis (13-15). Recently, an additional function of the WNT/ β -catenin

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Abbreviations: APC, adenomatous polyposis coli; C/EBP, CCAAT/enhancer binding protein; DVL, disheveled; 1,25(OH)₂D₃, 1,25-dihydroxyvitamin D3; FABP, fatty acid binding protein; GSK, glycogen synthase kinase; LPL, lipoprotein lipase; LRP, low density lipoprotein receptor-related protein; PPAR, peroxisome proliferator-activated receptor; WNT, wntless-type MMTV integration site

Key words: 1,25-dihydroxyvitamin D3, adipogenesis, β -catenin, vitamin D, wntless-type MMTV integration site

pathway was identified: the negative regulator of adipogenesis (16,17). When the WNT/ β -catenin pathway is inactive, β -catenin is proteasomally degraded by the destruction complex composed of adenomatous polyposis coli (APC), glycogen synthase kinase (GSK)3 β and AXIN. Activation of the WNT/ β -catenin pathway enhances the interaction between disheveled (DVL)s and AXIN. Consequently, the APC-GSK3 β -AXIN destruction complex is disrupted, thereby increasing stabilization and nuclear translocation of β -catenin (18,19). In the nucleus, β -catenin induces the expression of cyclin D1, c-Myc and PPAR δ , which have been reported to inhibit PPAR γ , a major adipogenic transcription factor (20-23). Until now, no study reported the association between the anti-obesity effects of vitamin D and the WNT/ β -catenin pathway. In this study, we found that the anti-adipogenic effect of 1,25(OH) $_2$ D $_3$ was accompanied by the maintenance of the WNT/ β -catenin pathway signaling, which was normally downregulated during adipogenesis.

Materials and methods

Chemicals and reagents. Cell culture reagents were obtained from Life Technologies Inc. (Grand Island, NY, USA). Anti-PPAR γ , anti-FABP4, anti-phospho-GSK3 β and anti-GSK3 β antibodies and secondary antibody were obtained from Cell Signaling (Beverly, MA, USA). Anti-C/EBP α , anti-DVL2, anti- β actin and anti-TBP antibody were purchased from Santa Cruz Biotechnology, Inc. (Santa Cruz, CA, USA). Anti- β -catenin antibody was purchased from BD Transduction Laboratories (Lexington, KY, USA). All other chemicals including 1,25(OH) $_2$ D $_3$ were purchased from Sigma-Aldrich (St. Louis, MO, USA).

Cell culture and differentiation. 3T3-L1 preadipocytes were purchased from the American Type Culture Collection (Manassas, VA, USA) and were subcultured every 2 or 3 days. The cells were seeded in 6-well plates at a density of 1.5×10^5 cells/well. Two days after reaching confluence (day 0), the 3T3-L1 cells were differentiated in Dulbecco's modified Eagle's medium (DMEM) containing 1 μ g/ml insulin, 0.25 μ M dexamethasone, 0.5 mM 3-isobutyl-1-methylxanthine and 10% fetal bovine serum (differentiation-induction medium) for 2 days. The cells were then maintained in DMEM containing 1 μ g/ml insulin and 10% fetal bovine serum (differentiation maintenance medium). The differentiation maintenance medium was changed every 2 days until the cells were harvested. To test its effects on adipogenesis, 1,25(OH) $_2$ D $_3$ was added to the differentiation induction medium and differentiation maintenance medium until the cells were harvested. Lipid droplets in the cells were stained with Oil Red O as previously described (24).

Real-time polymerase chain reaction (PCR). Cells were harvested, and total RNA was extracted using an RNeasy kit (Qiagen, Hilden, Germany). One microgram of total RNA was reverse-transcribed at 37°C using the cDNA Reverse Transcription kit (Applied Biosystems, Inc., Foster City, CA, USA). Briefly, the reaction was performed in a final volume of 20 μ l, which included reverse transcriptase reaction buffer, 100 mM dNTP mix, random primers, MultiScribe™ Reverse Transcriptase, RNase inhibitor and total RNA. The reaction mixtures were heated at 25°C for 10 min, 37°C for 120 min

and 85°C for 5 min. Real-time PCR was performed using the 7000 Real-Time PCR system (Applied Biosystems) in a final volume of 20 μ l, which included TaqMan gene expression master mix, 250 nM of TaqMan probe, an optimized concentration of each primer, and 2 μ l of the reverse transcription product containing cDNA. The reaction mixtures were preheated at 95°C for 10 min to activate the enzyme and then subjected to 40 cycles of melting at 95°C for 15 sec and annealing/extension at 60°C for 1 min. The assay-on-demand gene expression products (Applied Biosystems, Inc.) were used to evaluate the mRNA levels of PPAR γ (Mm00440945_m1), C/EBP α (Mm01265914_s1), FABP4 (Mm00445880_m1), LPL (Mm00434764_m1), WNT10B (Mm00442104_m1), LRP6 (Mm00999795_m1), DVL2 (Mm00432899_m1), GSK3 β (Mm00444911_m1) as well as the level of 18S rRNA (Hs99999901_s1). The 18S rRNA was used as an internal control as previously described (25). For each sample, the mRNA level was normalized against the level of 18S rRNA, and the ratio of normalized mRNA to the preadipocytes (day 0) was determined using the comparative Ct method (26).

Protein extraction and western blotting. Cells were harvested using a cell scraper and lysed with ice-cold RIPA buffer containing 25 mM Tris-HCl (pH 7.6), 150 mM NaCl, 1% Nonidet P-40, 1% sodium deoxycholate, 0.1% SDS and a protease inhibitor cocktail (Sigma-Aldrich) to obtain total cell lysates. The total cell lysates were then centrifuged at 20,000 \times g for 20 min at 4°C to remove the insoluble materials. The protein concentrations were determined using a BCA protein assay kit (Pierce, Rockford, IL, USA). Fifty micrograms of each protein extract was separated using 12% polyacrylamide gel electrophoresis and electrotransferred to nitrocellulose membranes at 150 mA for 1 h. The membranes were then blocked for 2 h at room temperature with phosphate buffered saline containing 5% skim milk and 0.1% Tween-20, and incubated with 1:1,000-dilutions of primary antibody overnight at 4°C followed by a 1:1,000-dilution of horseradish peroxidase-conjugated secondary antibody for 1 h at room temperature. Peroxidase activity was visualized using an ECL kit (Pierce). β -actin was used as a loading control for cytosolic proteins.

Analysis of nuclear β -catenin level. Cells were harvested using a cell scraper and then nuclear extracts were prepared using a nuclear extract kit (Active Motif, Carlsbad, CA, USA). Protein concentrations in the nuclear extracts were determined using a BCA protein assay kit (Pierce). Ten micrograms of nuclear protein were separated using 12% polyacrylamide gel electrophoresis and analyzed by western blotting using an anti- β -catenin antibody followed by a secondary antibody. TATA-box binding protein (TBP) was used as a loading control for nuclear proteins.

Statistical analysis. All data are expressed as means \pm standard deviations from at least three replicate experiments. Statistically significant differences between treated and untreated samples were detected using an unpaired t-test. All analyses were performed using SPSS ver. 19 (SPSS, Chicago, IL, USA).

Results

Inhibitory effects of 1,25(OH) $_2$ D $_3$ on the adipogenesis of 3T3-L1 cells. As shown in Fig. 1, treatments of nanomolar concentra-

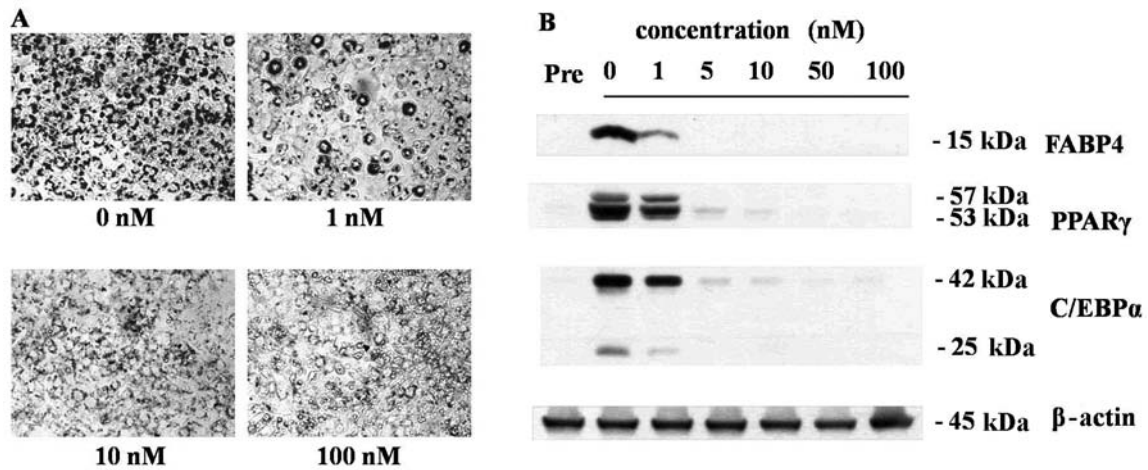


Figure 1. Dose response of the anti-adipogenic effects of 1,25(OH)2D3. (A) The inhibitory effects of the various concentrations of 1,25(OH)2D3 on the intracellular lipid droplet formation. Lipid droplets were stained with Oil Red O on day 6 of adipogenic differentiation and examined under light microscopy. (B) Effects of the various concentrations of 1,25(OH)2D3 on the expressions of adipocyte-specific marker proteins on day 6 of adipogenic differentiation. β -actin was used as a loading control for total cellular proteins. Each experiment was repeated independently at least twice, and the representative results are shown. Pre, undifferentiated preadipocytes.

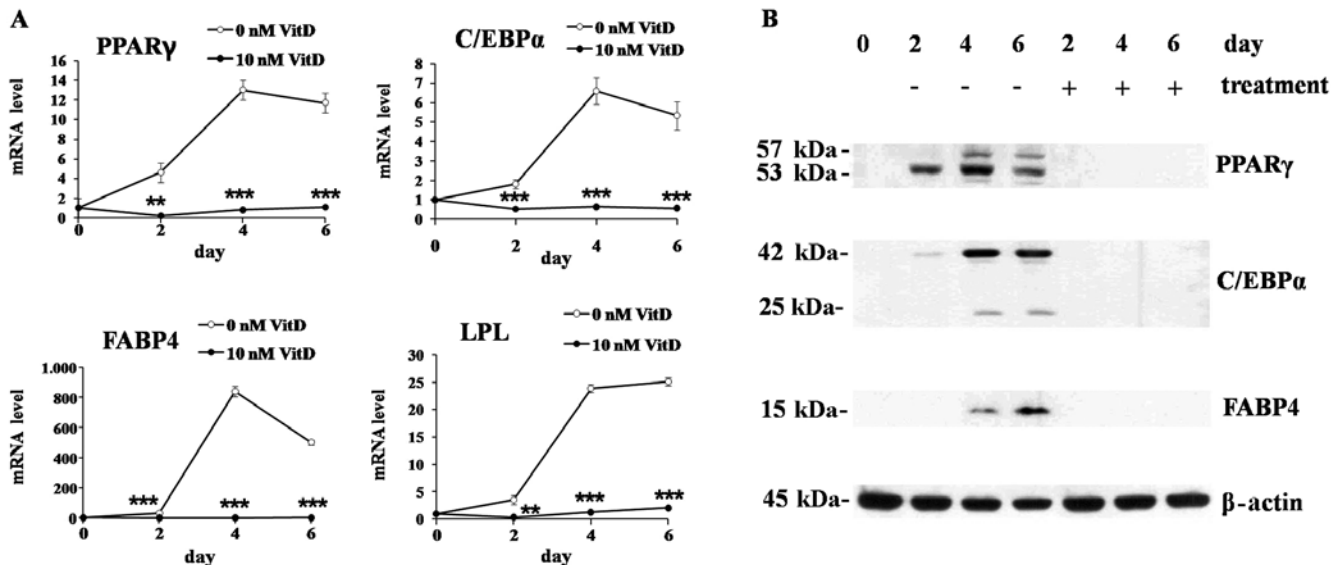


Figure 2. Time-course response of the anti-adipogenic effects of 1,25(OH)2D3. (A) Time-course response for the mRNA expression levels of the adipocyte-specific marker genes in the absence (open circle) or presence of 10 nM 1,25(OH)2D3 (closed circle). mRNA levels were expressed as the ratio to that of 0 day. (B) Time-course response for the levels of the adipocyte-specific marker proteins in the absence (-) or presence (+) of 10 nM 1,25(OH)2D3. The data represent the mean \pm standard deviation (SD) of triplicate assays. Each experiment was repeated independently at least twice, and the representative results are shown. ** $p < 0.01$, *** $p < 0.001$ compared with untreated cells on the same differentiation day.

tions of 1,25(OH)2D3 during the adipogenesis of 3T3-L1 cells effectively inhibited the adipogenesis of 3T3-L1 cells in a dose-dependent manner. At the concentration of 10 nM, 1,25(OH)2D3 completely inhibited lipid droplet formation (Fig. 1A) as well as the expressions of adipocyte maker protein, FABP4, PPAR γ and C/EBP α (Fig. 1B). Because 10 nM is the optimal anti-adipogenic concentration of 1,25(OH)2D3, all further experiments were conducted at this concentration. In time-course experiments, 1,25(OH)2D3 treatment significantly suppressed the mRNA expression of major transcription factors of adipogenesis, PPAR γ and C/EBP α , as well as the proteins involved in lipid metabolism, FABP4 and LPL (Fig. 2A). The protein expression was

also suppressed by 1,25(OH)2D3 showing similar patterns with mRNA expression (Fig. 2B).

Maintenance of the WNT signaling by 1,25(OH)2D3. To investigate the effects of 1,25(OH)2D3 on the WNT signaling, we measured the expressions of the genes involved in the WNT signaling in 3T3-L1 cells differentiated for 0, 2, 4 and 6 days in the presence or absence of 1,25(OH)2D3. The mRNA level of WNT10B was markedly downregulated during adipogenesis of untreated cells, but the treatment of 1,25(OH)2D3 maintained its expression levels (Fig. 3A). The mRNA level of LRP6 was mildly decreased during adipogenesis of untreated cells, but

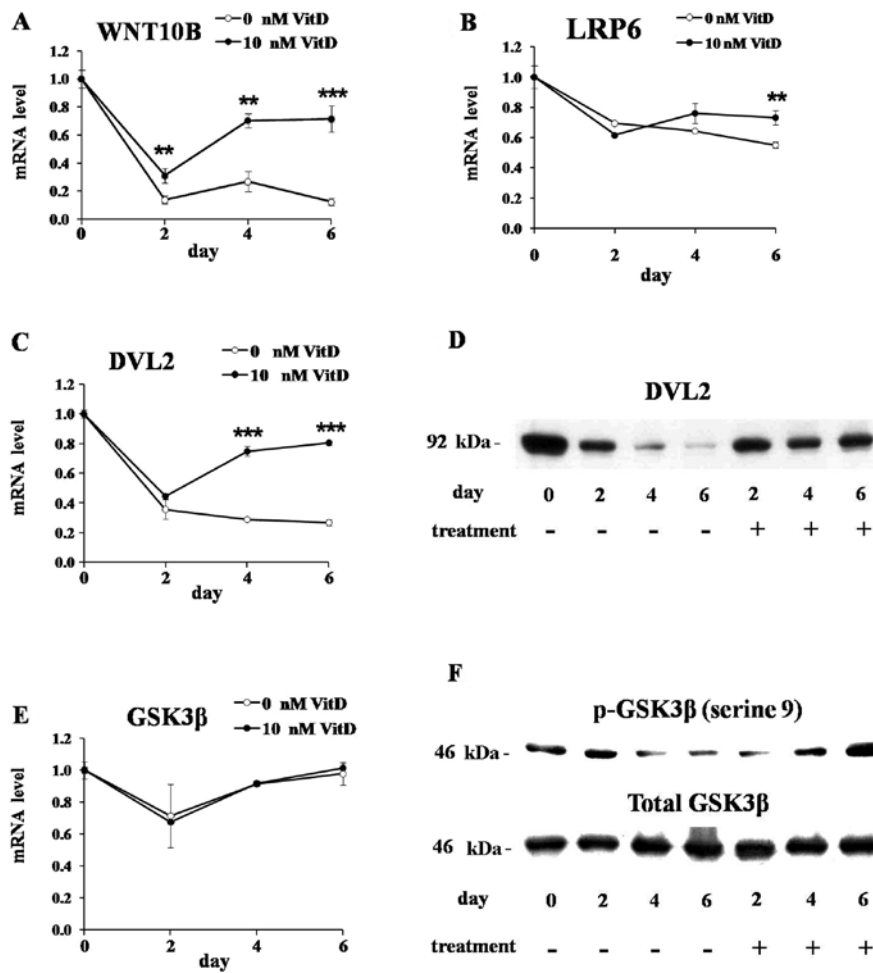


Figure 3. Effects of 1,25(OH)2D3 on WNT signaling during adipogenesis. (A-C and E) Time-course response for the mRNA expression levels of genes involved in WNT signaling in the absence (open circle) or presence of 10 nM 1,25(OH)2D3 (closed circle). mRNA levels were expressed as the ratio to that of 0 day. (D) Time-course response for the protein level of DVL2 in the absence (-) or presence (+) of 10 nM 1,25(OH)2D3. (F) Time-course response for the protein level of GSK3 β , for total and phosphorylated forms, in the absence (-) or presence (+) of 10 nM 1,25(OH)2D3. The data represent the mean \pm standard deviation (SD) of triplicate assays. Each experiment was repeated independently at least twice, and the representative results are shown. ** $p < 0.01$, *** $p < 0.001$ compared with untreated cells on the same differentiation day.

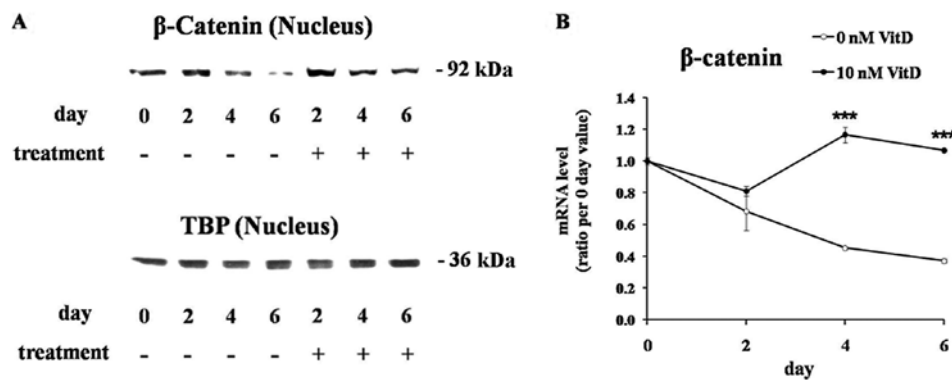


Figure 4. Effects of 1,25(OH)2D3 on β -catenin levels during adipogenesis. (A) Time-course response for the nuclear levels of β -catenin protein in the absence (-) or presence (+) of 10 nM 1,25(OH)2D3. TBP was used as a loading control for nuclear proteins. (B) Time-course response for the mRNA expression levels of β -catenin in the absence (open circle) or presence of 10 nM 1,25(OH)2D3 (closed circle). The data represent the mean \pm standard deviation (SD) of triplicate assays. Each experiment was repeated independently at least twice, and the representative results are shown. *** $p < 0.001$ compared with untreated adipocytes on the same differentiation day.

1,25(OH)2D3 maintained its level (Fig. 3B). DVL2 is reported to be activated by WNT signaling and prevent the formation of the APC-GSK3 β -AXIN destruction complex which degrade

β -catenin (27). The mRNA and protein levels of DVL2 were markedly downregulated during adipogenesis of untreated cells, but 1,25(OH)2D3 maintained their mRNA and protein levels

during adipogenesis (Fig. 3C and D). During adipogenesis of 3T3-L1 cells, treated or untreated, the mRNA and protein levels of GSK3 β were not significantly changed (Fig. 3E and F, lower panel), but its phosphorylation on serine 9 residue was significantly decreased in untreated cells (Fig. 3F, upper panel). 1,25(OH)2D3 efficiently elevated GSK3 β phosphorylation even though it has no effects on total GSK3 β protein levels as well as its mRNA levels. GSK3 β is a component of APC-GSK3 β -AXIN destruction complex which is involved in the proteolytic degradation of β -catenin, and it is also well-known that GSK3 β is inactivated by its phosphorylation (15,19). We found that GSK3 β was activated by its dephosphorylation during adipogenesis of untreated 3T3-L1 cells, but the treatment of 1,25(OH)2D3 efficiently elevated its phosphorylation levels resulting in its inactivation.

Maintenance of nuclear β -catenin levels by 1,25(OH)2D3. β -catenin is a transcriptional cofactor which plays a central role in the WNT/ β -catenin pathway (20). The nuclear level of β -catenin protein was significantly reduced during adipogenesis of untreated cells, but 1,25(OH)2D3 treatment maintained the nuclear β -catenin levels (Fig. 4A). TBP protein was used as an endogenous control for the loading of equal amounts of the nuclear protein. The levels of the β -catenin mRNA, which were significantly downregulated during the adipogenesis of untreated cells, were also maintained by 1,25(OH)2D3 treatment (Fig. 4B).

Discussion

The effects of vitamin D on WNT signaling in relation to adipogenesis were not reported until now. The relation of vitamin D and WNT signaling has only been reported in relation to cancer: vitamin D was reported to have a protective effect against colorectal cancer by its inhibitory action on the WNT/ β -catenin pathway (28). In the present study on the anti-adipogenic mechanisms of vitamin D, our interest in the WNT/ β -catenin pathway was prompted by recent findings of its role as a negative regulator of adipogenesis (16,17). WNT signaling was reported to inhibit adipogenesis (29), and was found to mediate the anti-adipogenic effects of some cytokines and chemical compounds: anti-adipogenic cytokines, tumor necrosis factor α and interleukin-6 inhibit adipogenesis by the activation of WNT signaling (25). Bisindolylmaleimide I, a specific inhibitor of protein kinase C, inhibits adipocyte differentiation through the activation of the WNT signaling (30), and curcumin, a component of curry, inhibits adipogenesis via the WNT/ β -catenin pathway (31).

In this study, the induction of PPAR γ , a major transcription factor of adipogenesis, was effectively suppressed by 1,25(OH)2D3 (Figs. 1 and 2), while the nuclear level of β -catenin was maintained by 1,25(OH)2D3 during the adipogenesis of 3T3-L1 cells (Fig. 4A). It is well-known that the WNT signaling upregulates β -catenin at post-translation levels by protein stabilization (18,19). Few reports exist on the transcriptional regulation of β -catenin during adipogenesis, and it is interesting that 1,25(OH)2D3 also maintained mRNA levels of β -catenin which were decreased during adipogenesis of untreated cells (Fig. 4B). Even though further study is necessary to elucidate the transcriptional modulation of β -catenin by 1,25(OH)2D3, these experimental data can

provide an explanation for the anti-adipogenic mechanism of 1,25(OH)2D3, considering that β -catenin inhibits PPAR γ directly (32), and indirectly by inducing its target genes (21-23). Our data suggest that the anti-adipogenic effect of 1,25(OH)2D3 is mediated by the maintenance of the nuclear β -catenin levels, which results in the suppression of PPAR γ , the major adipogenic transcription factor playing a central role in the expressions of lipid-metabolizing enzymes (4-6,33).

Among members of WNT family, WNT10B is most closely related to adipogenesis and obesity, and previous studies have established the role of WNT10B as an endogenous regulator of adipogenesis (34). WNT10B transgenic mice which express WNT10B from the adipocyte-specific promoter, showed 50% reduction of the total body fat and resisted the accumulation of adipose tissue when fed a high fat diet (35). Human obesity phenotype in various populations were reported to be associated with the genetic polymorphisms of WNT10B (36,37). In the present study, it was found that 1,25(OH)2D3 maintained the expression level of WNT10B, a WNT family member most closely related to adipogenesis and obesity (Fig. 3A).

In conclusion, 1,25(OH)2D3 maintained WNT10B expression levels resulting in the maintenance of the nuclear levels of β -catenin which is a suppressor of adipogenesis, suggesting that the WNT/ β -catenin pathway plays a role as a mediator for the anti-adipogenic and anti-obesity effects of vitamin D.

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References

1. Must A, Spadano J, Coakley EH, Field AE, Colditz G and Dietz WH: The disease burden associated with overweight and obesity. *JAMA* 282: 1523-1529, 1999.
2. Pi-Sunyer FX: The obesity epidemic: pathophysiology and consequences of obesity. *Obes Res* 10 (Suppl 2): S97-S104, 2002.
3. Spiegelman BM and Flier JS: Obesity and the regulation of energy balance. *Cell* 104: 531-543, 2001.
4. Farmer SR: Transcriptional control of adipocyte formation. *Cell Metab* 4: 263-273, 2006.
5. Rosen ED and MacDougald OA: Adipocyte differentiation from the inside out. *Nat Rev Mol Cell Biol* 7: 885-896, 2006.
6. Rosen ED, Walkey CJ, Puigserver P and Spiegelman BM: Transcriptional regulation of adipogenesis. *Genes Dev* 14: 1293-1307, 2000.
7. Green H and Meuth M: An established pre-adipose cell line and its differentiation in culture. *Cell* 3: 127-133, 1974.
8. Sutton AL and MacDonald PN: Vitamin D: more than a 'bone-a-fide' hormone. *Mol Endocrinol* 17: 777-791, 2003.
9. Martini LA and Wood RJ: Vitamin D status and the metabolic syndrome. *Nutr Rev* 64: 479-486, 2006.
10. Foss YJ: Vitamin D deficiency is the cause of common obesity. *Med Hypotheses* 72: 314-321, 2009.
11. Ishida Y, Taniguchi H and Baba S: Possible involvement of 1 α ,25-dihydroxyvitamin D3 in proliferation and differentiation of 3T3-L1 cells. *Biochem Biophys Res Commun* 151: 1122-1127, 1988.
12. Kong J and Li YC: Molecular mechanism of 1,25-dihydroxyvitamin D3 inhibition of adipogenesis in 3T3-L1 cells. *Am J Physiol Endocrinol Metab* 290: E916-E924, 2006.
13. Moon RT, Bowerman B, Boutros M and Perrimon N: The promise and perils of Wnt signaling through beta-catenin. *Science* 296: 1644-1646, 2002.

14. Veeman MT, Axelrod JD and Moon RT: A second canon. Functions and mechanisms of beta-catenin-independent Wnt signaling. *Dev Cell* 5: 367-377, 2003.
15. Wodarz A and Nusse R: Mechanisms of Wnt signaling in development. *Annu Rev Cell Dev Biol* 14: 59-88, 1998.
16. Bennett CN, Ross SE, Longo KA, Bajnok L, Hemati N, Johnson KW, Harrison SD and MacDougald OA: Regulation of Wnt signaling during adipogenesis. *J Biol Chem* 277: 30998-31004, 2002.
17. Prestwich TC and MacDougald OA: Wnt/beta-catenin signaling in adipogenesis and metabolism. *Curr Opin Cell Biol* 19: 612-617, 2007.
18. Cadigan KM and Liu YI: Wnt signaling: complexity at the surface. *J Cell Sci* 119: 395-402, 2006.
19. Huelsken J and Behrens J: The Wnt signalling pathway. *J Cell Sci* 115: 3977-3978, 2002.
20. Behrens J, von Kries JP, Kuhl M, Bruhn L, Wedlich D, Grosschedl R and Birchmeier W: Functional interaction of beta-catenin with the transcription factor LEF-1. *Nature* 382: 638-642, 1996.
21. Freytag SO and Geddes TJ: Reciprocal regulation of adipogenesis by Myc and C/EBP alpha. *Science* 256: 379-382, 1992.
22. Fu M, Rao M, Bouras T, Wang C, Wu K, Zhang X, Li Z, Yao TP and Pestell RG: Cyclin D1 inhibits peroxisome proliferator-activated receptor gamma-mediated adipogenesis through histone deacetylase recruitment. *J Biol Chem* 280: 16934-16941, 2005.
23. Shi Y, Hon M and Evans RM: The peroxisome proliferator-activated receptor delta, an integrator of transcriptional repression and nuclear receptor signaling. *Proc Natl Acad Sci USA* 99: 2613-2618, 2002.
24. Kasturi R and Joshi VC: Hormonal regulation of stearoyl coenzyme A desaturase activity and lipogenesis during adipose conversion of 3T3-L1 cells. *J Biol Chem* 257: 12224-12230, 1982.
25. Gustafson B and Smith U: Cytokines promote Wnt signaling and inflammation and impair the normal differentiation and lipid accumulation in 3T3-L1 preadipocytes. *J Biol Chem* 281: 9507-9516, 2006.
26. Livak KJ and Schmittgen TD: Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta C(T)) method. *Methods* 25: 402-408, 2001.
27. Gao C and Chen YG: Dishevelled: the hub of Wnt signaling. *Cell Signal* 22: 717-727, 2010.
28. Pendas-Franco N, Aguilera O, Pereira F, Gonzalez-Sancho JM and Munoz A: Vitamin D and Wnt/beta-catenin pathway in colon cancer: role and regulation of DICKKOPF genes. *Anticancer Res* 28: 2613-2623, 2008.
29. Ross SE, Hemati N, Longo KA, Bennett CN, Lucas PC, Erickson RL and MacDougald OA: Inhibition of adipogenesis by Wnt signaling. *Science* 289: 950-953, 2000.
30. Cho M, Park S, Gwak J, Kim DE, Yea SS, Shin JG and Oh S: Bisindolylmaleimide I suppresses adipocyte differentiation through stabilization of intracellular beta-catenin protein. *Biochem Biophys Res Commun* 367: 195-200, 2008.
31. Ahn J, Lee H, Kim S and Ha T: Curcumin-induced suppression of adipogenic differentiation is accompanied by activation of Wnt/beta-catenin signaling. *Am J Physiol Cell Physiol* 298: C1510-1516, 2010.
32. Liu J, Wang H, Zuo Y and Farmer SR: Functional interaction between peroxisome proliferator-activated receptor gamma and beta-catenin. *Mol Cell Biol* 26: 5827-5837, 2006.
33. Farmer SR: Regulation of PPARgamma activity during adipogenesis. *Int J Obes* 29 (Suppl 1): S13-S16, 2005.
34. Wend P, Wend K, Krum SA and Miranda-Carboni GA: The role of WNT10B in physiology and disease. *Acta Physiol* 204: 34-51, 2012.
35. Longo KA, Wright WS, Kang S, Gerin I, Chiang SH, Lucas PC, Opp MR and MacDougald OA: Wnt10b inhibits development of white and brown adipose tissues. *J Biol Chem* 279: 35503-35509, 2004.
36. Van Camp JK, Beckers S, Zegers D, Verrijken A, Van Gaal LF and Van Hul W: Genetic association between WNT10B polymorphisms and obesity in a Belgian case-control population is restricted to males. *Mol Genet Metab* 105: 489-493, 2012.
37. Kim IC, Cha MH, Kim DM, Lee H, Moon JS, Choi SM, Kim KS and Yoon Y: A functional promoter polymorphism -607G>C of WNT10B is associated with abdominal fat in Korean female subjects. *J Nutr Biochem* 22: 252-258, 2012.