

Cyanidin-3-glucoside isolated from mulberry fruit protects pancreatic β -cells against oxidative stress-induced apoptosis

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Abstract. The extract obtained from berries contains high amounts of anthocyanins, and this extract is used as a phytotherapeutic agent for different types of diseases. In this study, we examined the cytoprotective effects of cyanidin-3-glucoside (C3G) isolated from mulberry fruit against pancreatic β -cell apoptosis caused by hydrogen peroxide (H_2O_2)-induced oxidative stress. The MIN6 pancreatic β -cells were used to investigate the cytoprotective effects of C3G on the oxidative stress-induced apoptosis of cells. Cell viability was examined by MTT assay and lipid peroxidation was assayed by thiobarbituric acid (TBA) reaction. Immunofluorescence staining, flow cytometry and western blot analysis were also used to determine apoptosis and the expression of proteins associated with apoptosis. Our results revealed that H_2O_2 increased the rate of apoptosis by stimulating various pro-apoptotic processes, such as the generation of intracellular reactive oxygen species (ROS), lipid peroxidation, DNA fragmentation and caspase-3 activation. However, C3G reduced the H_2O_2 -induced cell death in the MIN6N pancreatic β -cells. In addition, we confirmed that H_2O_2 activated mitogen-activated protein kinases (MAPKs), such as extracellular signal-regulated kinase (ERK), c-Jun NH₂-terminal kinase (JNK) and p38 MAPK. C3G inhibited the phosphorylation of ERK and p38 without inducing the phosphorylation of JNK. Furthermore, C3G regulated the intrinsic apoptotic pathway-associated proteins, such as proteins belonging to the Bcl-2

family, cytochrome *c* and caspase-3. Taken together, our results suggest that C3G isolated from mulberry fruit has potential for use as a phytotherapeutic agent for the prevention of diabetes by preventing oxidative stress-induced β -cell apoptosis.

Introduction

Diabetes is one of the major metabolic diseases among adults worldwide. Type 1 diabetes, classified as insulin-dependent diabetes mellitus, is caused by the loss of insulin secretion due to the destruction of pancreatic β -cells (1). This leads to an increase in blood sugar levels and a decrease in the energy transfer to other organs, which results in various secondary complications (2). Therefore, the protection of pancreatic β -cells is crucial for preventing the onset of type 1 diabetes. Under homeostatic conditions, potentially toxic reactive oxygen species (ROS) are primarily generated by mitochondrial respiratory metabolism, and are subsequently neutralized effectively by cellular antioxidant defense mechanisms. However, the excessive generation of ROS, such as super oxide anions (O_2^-) and hydrogen peroxide (H_2O_2) due to environmental stress results in significant oxidative damage to cell structures, such as DNA, the mitochondria and cell membranes (3,4).

Recent studies have demonstrated that antioxidants present in natural products protect cells by reducing H_2O_2 -induced oxidative stress *in vitro* and *in vivo* (5-7). This indicates the importance of isolating compounds with antioxidant activity and the characterization of their antioxidant mechanisms. Mulberry fruit is a well known medicinal plant and has long been used for the treatment of different types of diseases (8-10). Recent studies have suggested that anthocyanins isolated from mulberry fruit exert anti-aging, anti-hyperlipidemic and anti-cancer effects (11-13). In particular, a representative of the anthocyanins isolated from mulberry fruit, cyanidin-3-glucoside (C3G), is present in high quantities in mulberry fruit and is considered a phytotherapeutic agent due to its anti-oxidant properties (11,14).

In the present study, we investigated the protective effects of C3G isolated from mulberry fruit against the H_2O_2 -induced apoptosis of MIN6N pancreatic β -cells.

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Materials and methods

Preparation of sample. The mulberry fruits (15 kg) were extracted in 70% ethanol (36 liters) for 24 h at room temperature 3 times. The solvent was filtered, concentrated using an evaporator, redissolved in distilled water and lyophilized (yield, 14.6%). The powder obtained from the mulberry extract was soaked in *n*-hexane for 24 h to remove fats and oils. C3G in the powder was extracted with acidified methanol (methanol/1.0 N HCl, 85:15, v/v), centrifuged at 12,000 × g for 15 min to remove the precipitate and then filtered through a 0.45-μm filter. The purified anthocyanin extract was evaporated at 46°C to dryness and stored 4°C. The sample was identified by diluting the purified anthocyanin powder with the solvent (methanol/H₂O/formic acid, 75:20:5) and performing high-performance liquid chromatography (HPLC). The pure C3G standard (kuromanin chloride) and purified anthocyanin isolated from the mulberry fruit were identified using the HPLC retention time (Fig. 1), and their purity was >99%, as shown by HPLC. Reversed-phase HPLC was performed using a Waters 486 detector (Water, Milford, MA, USA) under the following conditions: column, μBondapak C18 (Waters, 3.9x300 mm); flow rate, 0.5 ml/min; injection volume, 10 μl; column temperature, 46°C. The absorbance was recorded at 520 nm using a UV/Vis spectrophotometer (Spectronic Instruments; Thermo Fisher Scientific, Waltham, MA, USA). The identified C3G powder was redissolved in medium and filtered using a 0.22-μm filter before using it in cell culture.

Chemicals. Dulbecco's modified Eagle's medium (DMEM), fetal bovine serum (FBS), penicillin/streptomycin (PS) and trypsin-EDTA were purchased from Gibco (Grand Island, NY, USA). Dichlorodihydrofluorescein-diacetate (H₂DCF-DA) and the apoptosis assay kit were obtained from Molecular Probes (Carlsbad, CA, USA). The pure C3G standard (kuromanin chloride), H₂O₂, Hoechst 33342 and the mitochondria isolation kit were purchased from Sigma-Aldrich (St. Louis, MO, USA). The rat/mouse insulin enzyme-linked immunosorbent assay (ELISA) kit was obtained from Linco Research Inc. (St. Charles, MO, USA). Antibodies against c-Jun NH₂-terminal kinase (JNK; #9252), phosphorylated (p-)JNK (#9255S), extracellular signal-regulated kinase (ERK; #4695), p-ERK (#9101S), p38 (#9212), p-p38 (#4631S) and horseradish peroxidase (HRP)-linked anti-rabbit IgG (#7074) were purchased from Cell Signaling Technology (Beverly, MA, USA). Antibodies against β-actin (sc-47778), Bax (sc-493), Bcl-2 (sc-7382), caspase-3 (sc-7272) and HRP-linked goat anti-mouse IgG (sc-2005) were purchased from Santa Cruz Biotechnology, Inc. (Santa Cruz, CA, USA). The mitogen-activated protein kinase (MAPK) inhibitors, PD98059 and SB203580, were obtained from Calbiochem (La Jolla, CA, USA). All other chemicals were of analytical grade.

Cell culture and treatment. The MIN6N pancreatic β-cells were derived from a mouse pancreatic islet cell line. The cells were provided by Professor H.Y. Kwon (College of Medicine, Hallym University, Chuncheon, Korea). The MIN6N β-cells were cultured in DMEM (11 mM glucose; Gibco) supplemented with 10% inactivated FBS and 1% penicillin-streptomycin (PS) at 37°C in a humidified 5% CO₂ incubator. The cells were cultured to ~90% confluence and were harvested with 0.25%

trypsin-EDTA. The cells were harvested and subcultured for an additional 48 h in DMEM. The cells were maintained under these culture conditions for all the experiments. The cells were seeded in 6- or 12-well plates, pre-incubated with the indicated concentrations of C3G for 20 h at 37°C, and were then treated with H₂O₂.

Cell viability assay. The MIN6N β-cells were seeded into 12-well plates and were incubated for 20 h. The cells were exposed to the drug treatments (C3G, H₂O₂) at the indicated concentrations for the indicated periods of time. Cell viability was evaluated using the 3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyltetrazolium bromide (MTT) assay, which is based on the reduction of a tetrazolium salt by mitochondrial dehydrogenase in viable cells. After the treatments, 500 μg/ml MTT solution were added to each well followed by incubation for 3 h at 37°C. The formazan crystals in each well were dissolved in isopropyl alcohol and the absorbance of each well was measured at 595 nm using an ELISA microplate reader (model 550; Bio-Rad, Hercules, CA, USA).

Measurement of lipid peroxidation. Lipid peroxidation was assayed using the thiobarbituric acid (TBA) reaction, as previously described (15). Briefly, the treated cells were washed with cold phosphate-buffered saline (PBS), harvested with 0.25% trypsin-EDTA and homogenized in cold 1.15% KCl. Subsequently, a 100 μl aliquot of the homogenized cells was mixed with 0.2 ml of 8.1% sodium dodecyl sulfate (SDS), 1.5 ml of 20% acetic acid (pH 3.6) and 1.5 ml of 0.8% TBA. The mixture was then heated to 95°C for 2 h. After cooling to room temperature, an *n*-butanol/pyridine mixture (15:1, v/v) was added and the mixture was then shaken for 5 min prior to centrifugation at 1,000 × g for 10 min. The supernatant was isolated, and the absorbance was measured at 535 nm.

Measurement of intracellular ROS scavenging activity and image analysis. To determine the effects of C3G on oxidative stress-induced ROS generation, we added 5 μM of H₂DCF-DA in PBS (pH 7.38) to the treated cells, and measured the fluorescence at excitation and emission wavelengths of 485 and 535 nm, respectively, using a microplate spectrofluorometer (Molecular Devices Corp., Sunnyvale, CA, USA). Image analysis to determine the production of intracellular ROS was performed by seeding the MIN6N β-cells in coverslip-loaded 12-well plates. The cells were treated with the indicated concentrations of C3G for 20 h followed by the addition of 0.7 mM H₂O₂. Following incubation for 4 h, H₂DCF-DA solution was added to each well of the plate, and the cells were incubated further for 2 h at 37°C. Images of the stained cells were acquired using a fluorescence microscope (Nikon, Melville, NY, USA).

Determination of apoptotic cells. The determination of apoptotic cells was carried out as previously described (16). Cells undergoing apoptosis were examined using a fluorescein isothiocyanate (FITC)-labeled Annexin V/propidium iodide (PI) apoptosis detection kit (Molecular Probes, Eugene, OR, USA) according to the manufacturer's instructions. In brief, the cells were harvested, washed with PBS and centrifuged to collect the cell pellet. The number of cells was adjusted to 1x10⁶ cells/ml. Subsequently, the cells were resuspended in

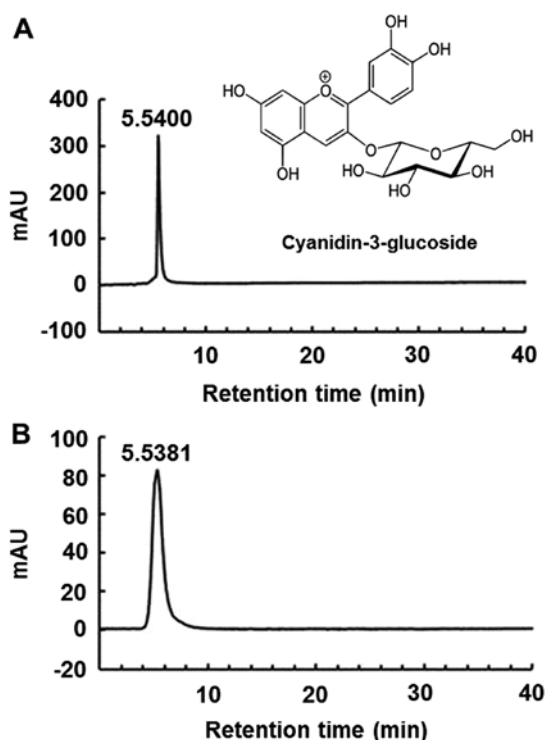


Figure 1. High-performance liquid chromatography (HPLC) chromatogram of cyanidin-3-glucoside (C3G) isolated from mulberry fruit. (A) HPLC chromatogram of the pure C3G standard (1 mg/ml). (B) C3G isolated from mulberry fruit (250 µg/ml). Detection was performed at 520 nm.

binding buffer [10 mM HEPES, 140 mM NaCl and 2.5 mM CaCl_2 (pH 7.4)] and stained with FITC-labeled Annexin V/PI at room temperature for 15 min under light-protected conditions. Flow cytometric analysis was performed using a FACSCalibur flow cytometer (Becton-Dickinson, Mountain View, CA, USA). The percentages of apoptotic cells were calculated using CellQuest software (Becton-Dickinson). Cells in the early phase of apoptosis were Annexin V-positive and PI-negative; however, cells in the late phase of apoptosis were positive for both Annexin V and PI. The apoptotic index (%) was calculated as the sum of cells in the early and late phases of apoptosis divided by the total number of events.

To determine the effects of C3G on H_2O_2 -induced DNA fragmentation, the MIN6N β -cells were labeled using the cell-permeable DNA-specific fluorescent dye, Hoechst 33342. The cells that showed homogeneously stained nuclei were considered viable, whereas the presence of chromatin fragmentation indicated apoptosis. The MIN6N β -cells treated with various concentrations of C3G were seeded in 12-well plates; Hoechst 33342 (stock 10 mg/ml) was added to each well followed by 15 min of incubation at room temperature. Images of the stained cells were acquired using a fluorescence microscope (Nikon) to examine the degree of DNA condensation and/or fragmentation.

Western blot analysis. The treated cells were washed in 1X PBS and lysed in lysis buffer for 30 min on ice. The lysates were centrifuged at 12,000 \times g for 30 min at 4°C. The supernatant was collected, and the protein content in the supernatant was measured using a Bio-Rad protein assay kit prior to analysis.

The total or fractionated protein samples were loaded and separated using SDS-polyacrylamide gel electrophoresis (PAGE) and transferred onto nitrocellulose membranes. The membranes were blocked with 5% non-fat powdered milk in 1X Tris-buffered saline containing 0.1% Tween-20 (TBS-T) for 1 h, and were then incubated with primary antibodies at 4°C overnight. Finally, the membranes were treated with HRP-linked secondary antibodies for 1 h at room temperature. The membranes were washed with TBS-T after each antibody binding reaction. The detection of each protein was performed using an enhanced chemiluminescence kit (Millipore Co., Billerica, MA, USA).

Measurement of caspase-3 activity. The treated MIN6N β -cells were lysed in 500 µl lysis buffer consisting of 10 mM Tris-HCl (pH 7.5), 10 mM $\text{NaH}_2\text{PO}_4/\text{NaHPO}_4$ (pH 7.5), 130 mM NaCl, 1% Triton X-100 and 10 mM NaPPi. Caspase-3 activity in the lysates was evaluated using a caspase assay kit (BD Biosciences, San Diego, CA, USA) according to the manufacturer's instructions.

Measurement of insulin secretion. The cell culture medium was collected from the treated cells, and the level of insulin released in the medium was measured using a rat/mouse insulin ELISA kit according to the manufacturer's instructions.

Statistical analysis. All measurements were from at least 3 independent experiments, and the values are expressed as the means \pm standard error (SE). Statistical analysis was performed using a Student's t-test. A value of $P < 0.05$ was considered to indicate a statistically significant difference.

Results

Optimal concentration of C3G and H_2O_2 in MIN6N β -cells. To determine the concentrations of C3G that could be used without affecting the viability of the MIN6N β -cells, we performed an MTT assay. The cells were seeded in 12-well plates and incubated for 24 h; subsequently, various concentrations of C3G were added to the cells followed by incubation for 20 h. C3G was well tolerated (cell viability, 86.6%) up to a concentration of 200 µg/ml; subsequently, cell viability decreased in a dose-dependent manner. At 300 and 500 µg/ml, C3G decreased cell viability by 83.9 and 80.3%, respectively. Thus, we concluded that the optimal concentration of C3G for use in further experiments was up to 200 µg/ml (Fig. 2).

In this study, H_2O_2 was used to induce oxidative stress in the MIN6N β -cells. After 4 h of treatment with H_2O_2 , the cell viability decreased in a dose- and time-dependent manner. Compared to the controls (untreated cells), the cells treated with 0.7 mM H_2O_2 for 4 h showed a 40% decrease in viability (Fig. 3).

Effects of C3G on lipid peroxidation and ROS generation. The inhibitory effects of C3G on lipid peroxidation in the H_2O_2 -treated MIN6N β -cells were determined by measuring the levels of thiobarbituric acid reactive substance (TBARS), a lipid peroxidation product. Lipid peroxidation was increased in the cells exposed to H_2O_2 , whereas C3G inhibited lipid peroxidation. Compared to the untreated cells, the cells treated

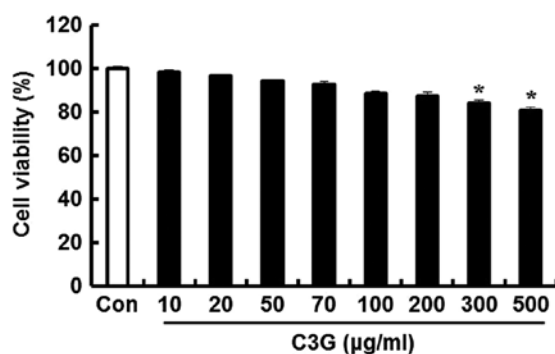


Figure 2. Viability of pancreatic MIN6N β -cells treated with various concentrations of cyanidin-3-glucoside (C3G). Dose-response experiments were performed to determine the maximum non-toxic concentration of C3G. Confluent cells were treated with various concentrations of C3G (10–500 μ g/ml) for 24 h. Data represent the means \pm SE. * P <0.05 vs. untreated control.

with 50 and 70 μ g/ml of C3G showed a 14.1 and 23.9% inhibition of lipid peroxidation, respectively (Fig. 4A). Intracellular ROS levels in the H_2O_2 -treated MIN6N β -cells were determined using the ROS-sensitive fluorescent probe, H_2DCF -DA, a cell-permeable dye that is cleaved by intracellular esterases into its non-fluorescent form, 2,7'-dichlorofluorescein (DCFH). This form, which is no longer membrane permeable, can be further oxidized by H_2O_2 to form the fluorescent compound. Compared to the untreated controls, the cells treated with C3G showed a significant dose-dependent increase in intracellular ROS scavenging activity (19.4 and 33.8% at 50 and 70 μ g/ml C3G, respectively; Fig. 4B). The H_2O_2 -dependent increase in the green fluorescence of DCF markedly decreased following treatment with 70 μ g/ml C3G and remained similar to the levels observed in the control cells (Fig. 4C). These data suggest that C3G inhibits lipid peroxidation and has ROS scavenging activity. On the basis of these results, we hypothesized that C3G inhibits oxidative stress-induced apoptosis in MIN6N β -cells.

Determination of apoptotic cells. To determine whether the inhibitory effects of H_2O_2 on MIN6N β -cells are associated with apoptosis, we performed double staining using FITC-labeled Annexin V and PI. Treatment with 0.7 mM H_2O_2 induced apoptosis in 46.6% of the cells (Fig. 5A). However,

pre-treatment with 70 μ g/ml C3G markedly inhibited the H_2O_2 -induced apoptotic cell death (32.2%; Fig. 5B). Furthermore, DNA fragmentation upon apoptosis induced by H_2O_2 was confirmed by staining the chromatin of the MIN6N β -cells using Hoechst 33342. Only the H_2O_2 -treated MIN6N β -cells underwent DNA fragmentation. The changes in chromatin condensation diminished following pre-treatment with 70 μ g/ml of C3G (Fig. 5C). These results indicate that C3G exerts protective effects against oxidative stress-induced apoptosis in MIN6N β -cells by inhibiting DNA fragmentation.

Effects of C3G on the phosphorylation of ERK and p38 MAPK.

In order to further examine the effects of C3G on H_2O_2 -induced MIN6N β -cell apoptosis, we examined the effects of C3G on the phosphorylation of MAPKs (ERK, JNK and p38) by western blot analysis. The levels of the phosphorylated forms of ERK, JNK and p38 significantly increased following treatment with H_2O_2 for 4 h. By contrast, treatment with 70 μ g/ml of C3G inhibited the H_2O_2 -induced phosphorylation of ERK and p38, but C3G did not inhibit the phosphorylation of JNK (Fig. 6A).

C3G regulates the activation of apoptosis-related proteins.

Recent studies have demonstrated that the phosphorylation of MAPKs is involved in the regulation of the mitochondrial permeability-mediated activation of apoptotic proteins, such as proteins of the Bcl-2 family and cytochrome *c* (Cyt *c*) (17–19). In this study, we confirmed the release of Cyt *c* and the induction of the expression of proteins of the Bcl-2 family, including Bcl-2 and Bax, in the MIN6N β -cells. The expression of Bcl-2 decreased and that of Bax increased in the H_2O_2 -treated cells (Fig. 6B). By contrast, the expression levels of proteins of the Bcl-2 family in the group pre-treated with 70 μ g/ml of C3G were similar to those of the controls. Subsequently, we investigated whether H_2O_2 induces the release of Cyt *c* from the mitochondria to the cytosol. Western blot analysis of the cytosolic fraction revealed the significant release of Cyt *c* from the mitochondria of the cells cultured with H_2O_2 . However, treatment with C3G (50 and 70 μ g/ml) induced a dose-dependent inhibition of the release of Cyt *c*.

Effect of C3G on H_2O_2 -induced caspase-3 activity and cell death.

We examined the effects of C3G on the H_2O_2 -induced

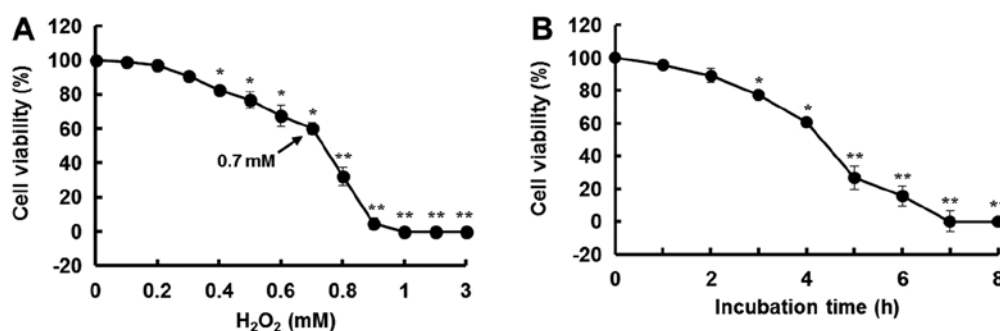


Figure 3. Viability of pancreatic MIN6N β -cells exposed to H_2O_2 . (A) The cytotoxicity of H_2O_2 was examined following exposure to various concentrations of H_2O_2 for 4 h. (B) The cytotoxicity of H_2O_2 was examined following exposure for different periods of time to 0.7 mM H_2O_2 . Data represent the means \pm SE. * P <0.05 and ** P <0.01 vs. untreated control.

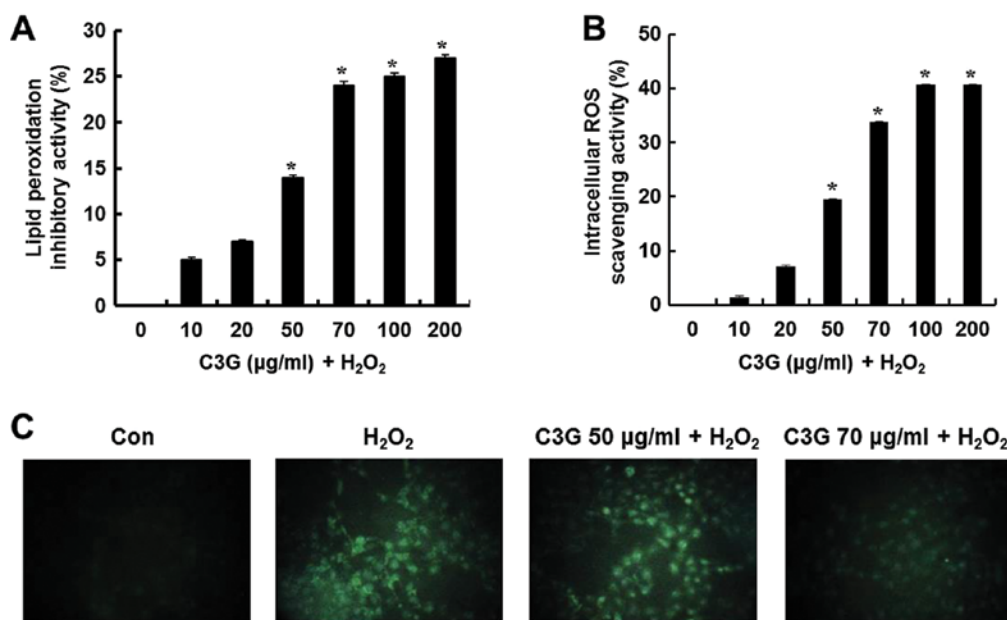


Figure 4. Inhibitory effects of cyanidin-3-glucoside (C3G) on H₂O₂-induced lipid peroxidation and reactive oxygen species (ROS) generation in MIN6N β-cells. (A) Lipid peroxidation in the treated cells was assayed by measuring the amount of thiobarbituric acid reactive substance (TBARS) formation. (B) The intracellular ROS scavenging activity was measured using the DCF-DA method. Data represent the means ± SE. *P<0.05 vs. untreated control. (C) The ROS levels in MIN6N β-cells were determined by fluorescence microscopy.

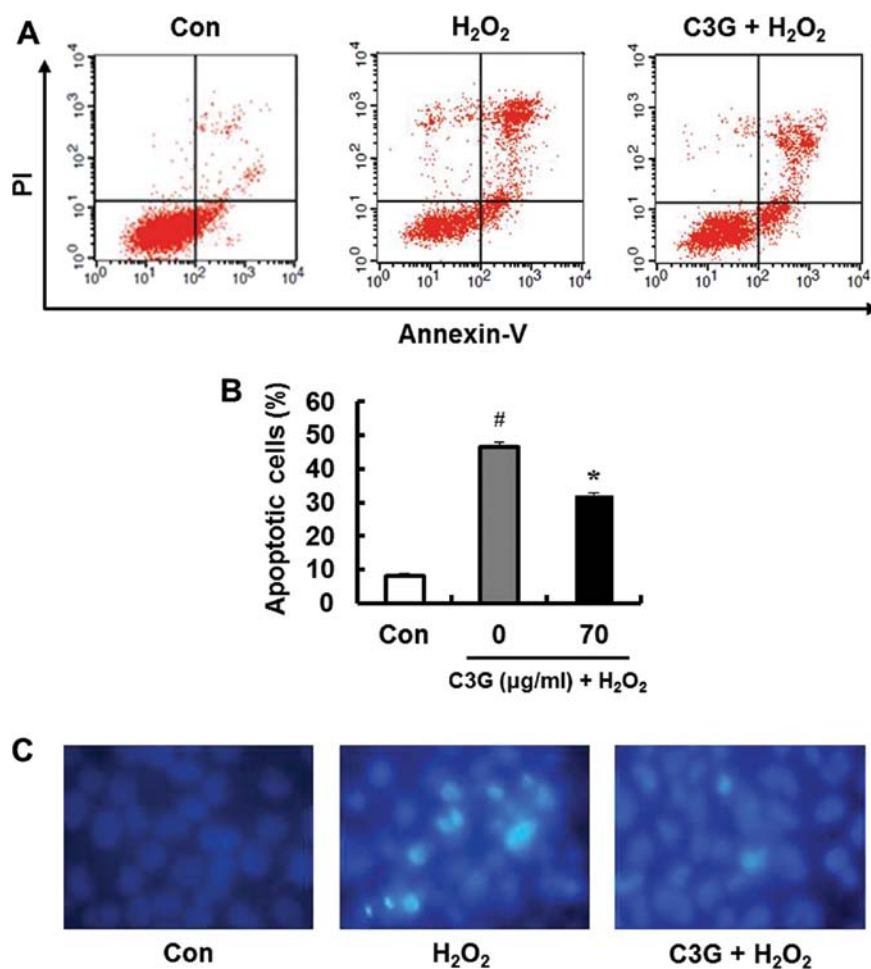


Figure 5. Inhibitory effects of cyanidin-3-glucoside (C3G) on apoptotic cell death of H₂O₂-treated MIN6N β-cells. (A) Apoptotic cells were detected Annexin V and PI double staining and analyzed by flow cytometry. Cells were either untreated [controls (Con)], treated with 0.7 mM H₂O₂, or treated with 70 μg/ml C3G + 0.7 mM H₂O₂. (B) The graph represents the percentage of apoptotic cells from the upper right plus lower right quadrants. Data represent the means ± SE. *P<0.05 vs. control; [#]P<0.05 vs. H₂O₂ alone. (C) The DNA fragmentation in MIN6N β-cells was determined by staining with the fluorescent dye, Hoechst 33342.

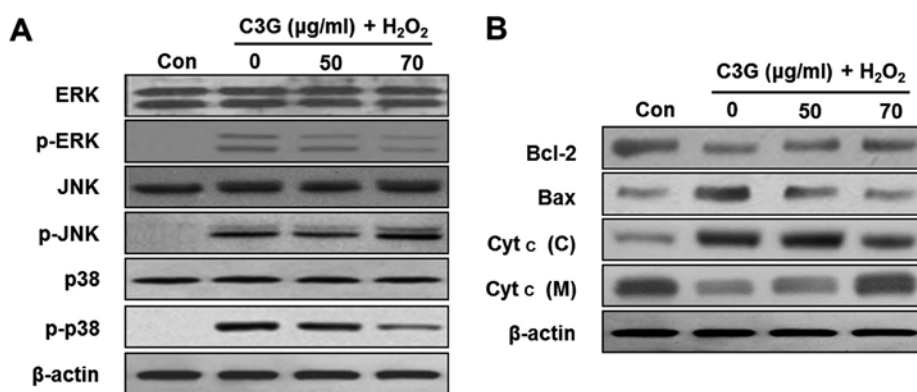


Figure 6. Effects of cyanidin-3-glucoside (C3G) on apoptosis-related proteins in H_2O_2 -treated MIN6N β -cells. The treated cells were harvested and lysates were prepared. (A) The expression levels of extracellular signal-regulated kinase (ERK), p-ERK, c-Jun NH₂-terminal kinase (JNK), phosphorylated (p-)JNK, p38 and p-p38 were assessed by western blot analysis. (B) The expression levels of Bcl-2 family proteins and fractional cytochrome *c* (Cyt *c*; C, cytosolic fraction; M, mitochondrial fraction) were determined by western blot analysis. Equal loading of total proteins in each sample was verified by β -actin expression.

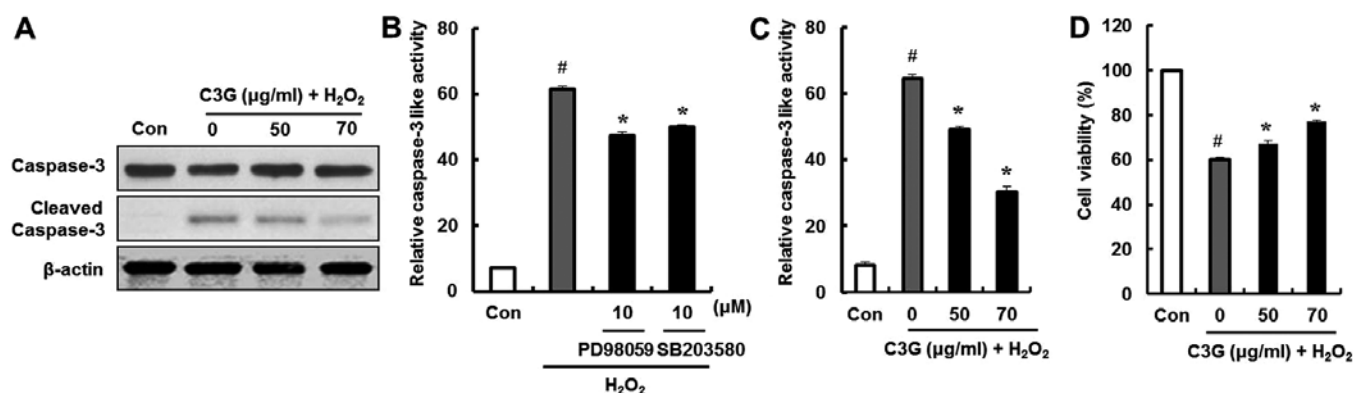


Figure 7. Effects of cyanidin-3-glucoside (C3G) on caspase-3 activation in H_2O_2 -treated MIN6N β -cells. The treated cells were harvested and lysates were prepared. (A) The expression levels of caspase-3 and cleaved caspase-3 were assessed by western blot analysis. Equal loading of total proteins in each sample was verified by β -actin expression. (B and C) Effect of the MAPK inhibitors (PD98059 and SB203580) and C3G on caspase-3 activation in H_2O_2 -treated MIN6N β -cells. (B) The cells were pre-treated with different types of inhibitors (10 μ M PD98059 and 10 μ M SB203580) or (C) indicated concentrations of C3G; after the treatments, the cells were exposed to 0.7 mM H_2O_2 for 4 h. Preparation of cell lysates and measurement of caspase-3 like activity was carried out using a caspase assay kit according to the manufacturer's instructions (BD Biosciences). (D) Effects of C3G on cell viability in H_2O_2 -treated MIN6N β -cells. Data represent the means \pm SE. # P <0.05 vs. control; * P <0.05 vs. H_2O_2 alone.

expression of cleaved caspase-3, which is an essential trigger of apoptosis, by western blot analysis. The cleaved form of caspase-3 was detected in the H_2O_2 -treated group. However, compared to the group treated with H_2O_2 , the group treated with C3G (50 and 70 μ g/ml) showed a decrease in the expression of cleaved caspase-3 (Fig. 7A). In addition, the ERK and p38 inhibitors, PD98059 and SB203580, were used to confirm that C3G inhibited the H_2O_2 -induced caspase-3 activation by regulating the phosphorylation of ERK and p38. The H_2O_2 -induced increase in caspase-3 activity was significantly decreased by the ERK and p38 inhibitors, PD98059 and SB203580 (Fig. 7B); by contrast, pre-treatment with C3G significantly decreased the H_2O_2 -induced caspase-3 activity (Fig. 7C). Additionally, the cytoprotective effects of C3G against the H_2O_2 -induced death of MIN6N β -cells were examined by MTT assay. The cells were pre-treated with C3G (50 and 70 μ g/ml) for 20 h followed by treatment with H_2O_2 . The cell viability decreased to 60% following treatment with H_2O_2 ; however, pre-treatment

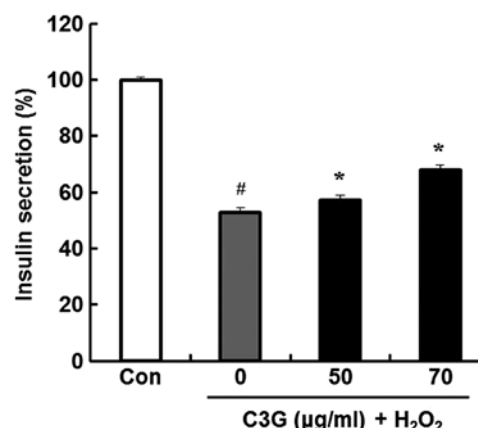


Figure 8. Effects of cyanidin-3-glucoside (C3G) on insulin secretion in H_2O_2 -treated MIN6N β -cells. After the treatments, the supernatants were collected, and insulin release was measured using a rat/mouse insulin enzyme-linked immunosorbent assay kit. Data represent the means \pm SE. # P <0.05 vs. control; * P <0.05 vs. H_2O_2 alone.

with 50 and 70 $\mu\text{g/ml}$ of C3G restored the cell viability to 67.8 and 76.1% of the controls, respectively (Fig. 7D). These results suggest that C3G protects pancreatic β -cells against death by inactivating caspase-3 and regulating the phosphorylation of ERK and p38.

Effects of C3G on H_2O_2 -induced insulin secretion. The anti-diabetic efficacy of C3G was determined by examining the effects of C3G on insulin release in MIN6N β -cells; we measured insulin secretion using a rat/mouse insulin enzyme-linked immunosorbent assay kit. Compared to the untreated group, the group pre-treated with 70 $\mu\text{g/ml}$ of C3G showed an increase in insulin secretion (52.6 vs. 68%) (Fig. 8).

Discussion

Deficiency of insulin caused by the destruction of pancreatic β -cells induces hyperglycemia, which leads to diabetes and serious pathological effects in humans (20,21). ROS are heavily implicated in the process of pancreatic β -cell destruction (22). ROS are routinely generated in the human body, and their production can be exacerbated by lifestyle choices, including eating, drinking and smoking habits (23). These factors can predispose individuals to diseases associated with ROS production, including cancer, stroke, cardiac disorders and diabetes (24-26). Thus, the inhibition of ROS generation is an important therapeutic goal. In apoptosis-inducing processes, H_2O_2 generation is attributed to the uncoupling of the electron transport chain in the mitochondria and to a decrease in mitochondrial membrane potential, leading the mitochondrial translocation of Bax and the cytosolic mobilization of Cyt c. Moreover, H_2O_2 has also been shown as a strong signal for the activation of the MAPK family of signaling proteins comprising of JNK, p38 and ERK (27). In this study, we used the H_2O_2 -induced toxicity of pancreatic β -cells as a model to evaluate the antidiabetic efficacy of C3G isolated from mulberry fruit. Pre-treatment with C3G reduced the levels of intracellular ROS and lipid peroxidation in the MIN6N cells treated with H_2O_2 (Fig. 4). In addition, the cells exposed to H_2O_2 showed distinct characteristics of apoptosis, such as DNA fragmentation and an increase in Annexin V staining. However, the cells pre-treated with C3G showed a significantly decreased percentage of apoptotic cells (Fig. 5). Studies have suggested that oxidative stress causes pancreatic β -cell apoptosis through the MAPK signaling pathway (28,29). MAPK proteins play a substantial role in the regulation of cell viability, proliferation and differentiation (30). The excessive accumulation of intracellular ROS activates MAPKs, which leads to apoptosis through the negative regulation of downstream proteins, such as proteins belonging to the Bcl-2 family, Cyt c and caspase-3 (31). Therefore, we confirmed the phosphorylation of MAPKs, the activation of Bcl-2 family proteins and the release of Cyt c from the mitochondria to the cytosol in the MIN6N β -cells damaged by H_2O_2 . All MAPKs were phosphorylated by treatment with H_2O_2 for 4 h. The cells treated with H_2O_2 showed an increase in the expression of the pro-apoptotic protein, Bax, and a decrease in the anti-apoptotic protein, Bcl-2. We confirmed the release of Cyt c from the mitochondria to the cytosol in the cells treated with H_2O_2 ; subsequently, cytosolic Cyt c activates caspase-3, which plays an important role in the

apoptotic pathway, and leads to cell apoptosis. However, we demonstrated that pre-treatment with C3G exerted protective effects against H_2O_2 -induced apoptosis by regulating the phosphorylation of ERK and p38. In addition, C3G regulated the proteins downstream of the MAPK signaling pathway, which resulted in the inactivation of caspase-3 (Figs. 6 and 7). Insulin secretion was significantly inhibited in the MIN6N β -cells exposed to H_2O_2 . By contrast, the cells treated with 70 $\mu\text{g/ml}$ of C3G prior to exposure to H_2O_2 showed an increase in insulin secretion (Fig. 8).

In conclusion, our results demonstrate that C3G isolated from mulberry fruit not only scavenges intracellular ROS and inhibits lipid peroxidation, but also regulates the apoptotic signaling pathways in pancreatic MIN6N β -cells following exposure to H_2O_2 , and thus results in a decrease in the apoptotic cell rate. Thus, our findings indicate that the anthocyanin isolated from mulberry fruit may prove to be a potential therapeutic agent for the treatment of diabetes.

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