

IL-1 β impedes the chondrogenic differentiation of synovial fluid mesenchymal stem cells in the human temporomandibular joint

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Abstract. Mesenchymal stem cell-based therapy has great therapeutic potential for temporomandibular joint (TMJ) cartilage repair. However, the behavior of mesenchymal stem cells in the inflammatory milieu following their delivery remains poorly understood. Synovial fluid-derived mesenchymal stem cells (SFMSCs) are a promising resource for TMJ cartilage repair, as they are easily obtained from patients with TMJ disorders (TMD). In this study, we obtained SFMSCs from patients with TMD and expanded them *in vitro*; we then stimulated the cells with interleukin (IL)-8, IL-1 β , IL-6, IL-10, tumor necrosis factor (TNF)- α and IL-12p. The cells expressed CD90, CD44, CD105 and CD73, and were negative for CD45, CD34, CD11b, CD19 and HLA-DR. They could be induced to differentiate into osteogenic, chondrogenic, adipogenic and neurogenic lineages *in vitro*. Only the levels of IL-6 and IL-8 were upregulated significantly following stimulation with IL-8, IL-1 β , IL-6, IL-10, TNF- α and IL-12p. Furthermore, IL-6 and IL-8 expression was driven mainly by IL-1 β -dependent nuclear factor- κ B (NF- κ B) pathway activation, and was independent of IL-8, IL-6, IL-10, TNF- α and IL-12p. IL-6 and IL-8 expression was inhibited completely by treatment with the NF- κ B inhibitor,

BAY11-7082. SRY-box 9 (*SOX9*) was downregulated and matrix metalloproteinase (*MMP*)13 was upregulated upon chondrogenic differentiation induced in the cells also exposed to IL-1 β . Sulfated glycosaminoglycan production was also reduced upon chondrogenic differentiation in the presence of IL-6, but not IL-8. Thus, IL-1 β in the inflammatory milieu is crucial in regulating SFMSCs. In doing so, IL-1 β impedes the chondrogenic differentiation of SFMSCs. The upregulation of IL-6 and NF- κ B pathway activation also contribute to this biological behavior. The findings of our study indicate the potential adverse effects of IL-1 β on the chondrogenic differentiation of SFMSCs, and may thus provide new insight into the pathogenesis of TMD.

Introduction

Temporomandibular joint disorder (TMD) is a common disease of the stomatognathic system. It is a degenerative disease, accompanied by internal derangement or osteoarthritis (OA), which is characterized by progressive articular cartilage destruction and decreased physical capabilities. Underlying these symptoms is the limited self-regenerating capacity of cartilage. Certain drugs, such as etanercept, infliximab and adalimumab, have been used clinically to treat TMD following the failure of traditional injection with corticosteroids or hyaluronic acid. However, a substantial proportion of patients still fail to respond to these biological therapies (1).

Mesenchymal stem cells (MSCs) are increasingly being considered attractive biological therapeutic agents for the treatment of inflammatory diseases and for the stimulation of tissue regeneration. They possess multi-differentiation potential and can be isolated from various human tissues, such as the bone marrow, adipose tissue, the synovium and periosteum. Among these sources, synovial fluid plays an important role in regulating temporomandibular joint (TMJ) physiology. Jones *et al* (2) were the first to identify MSCs in the synovial fluid of swollen osteoarthritic knee joints. These cells may originate from the MSCs in synovial tissue (3), as both populations express similar cell surface markers (4). Whereas the majority of subsequent studies have focused on large joints, information regarding synovial fluid-derived MSCs (SFMSCs) in the TMJ is limited. Koyama *et al* demonstrated that human pluripotent cells can be isolated from the synovial fluid of patients with TMD (5). Analogous to MSCs that are isolated from the fragments of syno-

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Abbreviations: TMJ, temporomandibular joint; MSCs, mesenchymal stem cells; TMD, temporomandibular joint disorders; SFMSCs, synovial fluid-derived mesenchymal stem cells; MPC, mesenchymal progenitor cell; OA, osteoarthritis; RA, rheumatoid arthritis; NF- κ B, nuclear factor- κ B; DI, dysfunction index

Key words: mesenchymal stem cells, cytokines, arthritis, temporomandibular, synovial fluid

vial tissue and synovial fluid, we hypothesized that the SFMSCs most likely originate from the disrupted intimal synovium (6). SFMSCs of the human TMJ can be obtained by arthrocentesis, which is a commonly used and less invasive method compared with arthroscopy or open operation to obtain sections of the synovium. Therefore, these cells constitute a promising resource for cell-directed articular cartilage repair.

Both the genetic background of MSCs and the local inflammatory milieu may affect the ability of MSCs to modulate disease activity *in vitro* (7,8). The number of SFMSCs reportedly increases in early human OA, and this increase correlates with cartilage degeneration and the severity of OA in the knee (9-11). Thus, these cells may play a potential role in joint homeostasis. Neidhart *et al* (12) found that adherent synovial fluid cells derived from patients with rheumatoid arthritis (RA) mediated cartilage destruction, independent of the accompanying synovial tissue hyperplasia. Heldens *et al* (13) demonstrated that human MSC chondrogenesis was inhibited in a diseased joint environment or upon treatment with pro-inflammatory cytokines. In another study, the amount of chondrogenic differentiation of mesenchymal progenitor cell (MPC) pellets in operated knees was significantly lower than that observed in control subjects. Additional treatment with interleukin (IL)-1 β also affected the degree of chondrogenic differentiation in cell pellets derived from bone marrow-derived MSCs (14). To the best of our knowledge, however, the effect of the inflammatory milieu on SFMSCs delivered to human TMJs remains poorly understood. To address this issue, in this study, we characterized the inflammatory factors that are secreted by SFMSCs following stimulation with IL-8, IL-1 β , IL-6, IL-10, tumor necrosis factor (TNF)- α and IL-12p *in vitro*. We found that the expression of only IL-6 and IL-8 increased significantly upon IL-1 β stimulation, and that this was independent of IL-8, IL-6, IL-10, TNF- α and IL-12p stimulation.

IL-1 β is a pivotal pro-inflammatory cytokine that mediates a variety of host defense processes, including inflammation and cellular response to injury, which can also lead to joint destruction if left unchecked (15). The level of IL-1 β is significantly higher in the synovial fluid from patients with TMD and chondral damage than in the synovial fluid from asymptomatic volunteers (16-18). IL-1 β reduces the expression of type II collagen and aggrecan by the increased secretion of matrix metalloproteinases (MMPs) by chondrocytes (19). This requires the activation of the nuclear factor- κ B (NF- κ B) pathway, which has been described as the link between inflammation and joint cartilage degeneration in OA (20-22). In this study, we further evaluated the chondrogenic differentiation potency and intracellular signaling pathways that are mediated by IL-1 β in SFMSCs.

Materials and methods

Ethics statement. This study was approved by the Institutional Ethics Board of the Hospital of Stomatology, Sun Yat-sen University, Guangzhou, China. Written informed consent was obtained from the sample providers prior to participation in this study.

Assay of inflammatory cytokines in the TMJ. Synovial fluid samples were obtained from 19 patients with TMD (19 laterals)

during arthrocentesis of the TMJ. These patients were aged between 16 and 57 years; 16 were female and 3 were male, and all were free of other systemic diseases. The patients with TMD were evaluated using the dysfunction index (DI) described in the study by Fricton JR and Schiffman (23). The synovial fluid samples were withdrawn with 2 ml of lidocaine (containing 25% vitamin B₁₂). We measured the levels of IL-8, IL-1 β , IL-6, IL-10, TNF- α and IL-12p in the synovial fluid samples with a BD Cytometric Bead Array (CBA) Human Soluble Protein Master Buffer kit (Human Inflammatory Cytokine kit; Cat. no. 551811; BD Biosciences, Bedford, MA, USA). The cytokine concentrations in the synovial fluid were calculated using the following formula: $C = \text{Casp}/[(\text{OD}_{\text{inj}} - \text{OD}_{\text{asp}})/\text{OD}_{\text{inj}}]$, C_{asp} refers to the levels of IL-8, IL-1 β , IL-6, IL-10, TNF- α and IL-12p in the synovial fluid samples; OD_{inj} refers to the optical density of lidocaine containing 25% vitamin B₁₂ before injection; OD_{asp} refers to the optical density of lidocaine containing 25% vitamin B₁₂ after injection, as previously described (16).

Human SFMSC isolation and expansion. The collected synovial fluid samples were centrifuged at 300 x g for 5 min at room temperature, and the supernatant was used in the inflammatory cytokine assay. The sediment was then cultured with complete culture medium [α minimum essential medium supplemented with 10% fetal bovine serum and 1X GlutaMAX (both from Gibco, Grand Island, NY, USA) at 37°C in a 5% CO₂ incubator. Portions of the cells from each sample were mixed and expanded in a monolayer culture for further experimentation. The cells were passaged at a density of 500 cells/cm².

Surface antigen identification. Surface markers, including CD90 (1:20; Cat. no. 562245; BD Biosciences), CD105 (1:11; Cat. no. 130-094-941; Miltenyi Biotec, Begisch Gladbach, Germany), CD73 (1:20), CD44 (1:20), and CD45/CD34/CD11b/CD19/HLA-DR (1:20) (Cat. no. 562245; all from BD Biosciences), were detected using an FC500 flow cytometer and MXP software, and the data were analyzed using CXP software (both from Beckman Coulter, Brea, CA, USA).

Osteogenic differentiation. The SFMSCs were seeded in 12-well plates at a density of 5,000 cells/cm². Following adherence, the complete culture medium was replaced with osteogenic induction medium consisting of H-DMEM, 10% fetal bovine serum (FBS) (both from Gibco), 10 mM sodium β -glycerophosphate (Santa Cruz Biotechnology, Inc., Santa Cruz, CA, USA), 10 nM 1,25-dihydroxyvitamin D3 (Sigma-Aldrich, St. Louis, MO, USA) and 50 μ g/l ascorbic acid-2-phosphate (Wako, Tokyo, Japan). The culture medium was replaced every 3 days. Following 28 days of induction, osteogenic differentiation was assessed by staining with a fresh 0.1% Alizarin Red solution (Cat. no. 130-22-3; Santa Cruz Biotechnology, Inc.) for 30 min at 37°C.

Adipogenic differentiation. The SFMSCs were seeded in 12-well plates at a density of 5,000 cells/cm². Following adherence, the complete culture medium was replaced with adipogenic induction medium consisting of H-DMEM, 10% FBS (both from Gibco), 200 mM indomethacin (Sigma-Aldrich), 0.5 mM isobutylmethylxanthine, 1 mM dexamethasone and 10 mg/ml insulin (all from MP Biomedicals, Irvine, CA, USA).

The medium was replaced every 3 days. After 28 days, cell staining was performed with a fresh 0.3% Oil Red O solution (Cat. no. 1320-06-5, Santa Cruz Biotechnology, Inc.) for 3 min.

Chondrogenic differentiation. The SFMSCs were harvested using 0.25% trypsin and approximately 3×10^5 cells were transferred to a 15 ml centrifuge tube. They were then centrifuged at $450 \times g$ for 8 min, and 500 μ l of chondrocyte differentiation induction medium [H-DMEM, 1X ITS-A (both from Gibco), 100 nM dexamethasone (MP Biomedicals), 50 mM ascorbic acid, 40 mg/ml proline (both from Sigma-Aldrich), and 10 ng/ml transforming growth factor- β 1 (PeproTech, Rocky Hill, NJ, USA)] were added. The medium was refreshed every 3 days. Following culture for 28 days, the chondrogenic pellets were fixed with 4% formalin and paraffin-embedded. Chondrogenic differentiation was assessed by 0.1% Safranin O staining (Cat. no. S2255; Seebio, Shanghai, China) and immunohistochemical staining for collagen II [rabbit anti-human collagen type II antibodies (Sigma-Aldrich) 1:80]. The control group was then incubated with complete culture medium.

Neurogenic differentiation. The cells were plated in 12-well plates and cultured until they reached 70% confluence, after which the complete culture medium was replaced with a neuronal induction medium that consisted of Neurobasal-A media containing 1X B-27 Supplement (both from Gibco), 20 ng/ml recombinant human epidermal growth factor (rhEGF), 40 ng/ml basic fibroblast growth factor (bFGF) (both from PeproTech), 10 ng/ml brain-derived neurotrophic factor (BDNF; ProSpec, East Brunswick, NJ, USA), 1 mM N6,2'-O-dibutyryl adenosine 3':5' cyclic monophosphate (dbCAMP; Sigma-Aldrich), 0.5 mM isobutylmethylxanthine (MP Biomedicals), and 10 ng/ml fibroblast growth factor (FGF)-8 (PeproTech) for 24 h to induce differentiation. The control group was incubated in complete culture medium. Neuronal differentiation was assessed by immunofluorescence staining for glial fibrillary acidic protein (GFAP; mouse anti-human antibody to GFAP, 1:200; Cat. no. MAB360; Millipore, Billerica, MA, USA).

RT-qPCR. Total RNA from the chondrogenic differentiation pellets and other cell samples was extracted using RNeasy mini kit reagents (Qiagen, Valencia, CA, USA) and TRIzol reagent (Roche, Indianapolis, IN, USA), respectively. cDNA was synthesized using the Transcriptor First Strand cDNA Synthesis kit (Roche). The relative expression levels of Runt-related transcription factor 2 (*RUNX-2*), osteocalcin (*OCN*), peroxisome proliferator activated receptor gamma 2 (*PPARG2*), lipoprotein lipase (*LPL*), SRY-box 9 (*SOX9*) and *MMP13* were quantified using a SYBR-Green qPCR kit (Roche). The primer nucleotide sequences are listed in Table I. The $2^{-\Delta\Delta Ct}$ method was used to analyze the relative gene expression levels by normalizing against the levels of glyceraldehyde 3-phosphate dehydrogenase (*GAPDH*) as an endogenous control and calibrating for amplification efficiency. $\Delta\Delta Ct$ was calculated using the following formula: $(Ct_{\text{target gene}} - Ct_{\text{gapdh}})_{\text{sample}} - (Ct_{\text{target gene}} - Ct_{\text{gapdh}})_{\text{control}}$.

Detection of lactate dehydrogenase (LDH) cytotoxicity. The human SFMSCs were seeded into 96-well plates at 10,000 cells/well and then treated with 10 ng/ml IL-1 β , 10 ng/ml TNF- α , 10 ng/ml

Table I. Oligonucleotide primers used in RT-qPCR.

Gene	Primer sequence	Product size (bp)
<i>GAPDH</i>	F: GACAGTCAGCCGCATCTTCT	178
	R: TTAAAAGCAGCCCTGGTGAC	
<i>RUNX2</i>	F: TCAACGATCTGAGATTTGTGGG	81
	R: GGGGAGGATTTGTGAAGACGG	
<i>OCN</i>	F: CCACCGAGACACCATGAGAG	267
	R: TCAGCCAACCTCGTCACAGTC	
<i>PPARG2</i>	F: GCAAACCCCTATTCCATGCTG	167
	R: CACGGAGCTGATCCCAAAGT	
<i>LPL</i>	F: CAAGAGTGAGTGAACAAC	189
	R: AATTATGCTGAAGGACAAC	
<i>SOX9</i>	F: ACACACAGCTCACTCGACCTTG	104
	R: AGGGAATTCTGGTTGGTCTCT	
<i>MMP13</i>	F: GACTGGTAATGGCATCAAGGGA	149
	R: CACCGGCAAAGCCACTTTA	

F, forward; R, reverse.

IL-8/IL-1 β /IL-6/IL-10/TNF- α /IL-12p compound, or 10 μ M NF- κ B inhibitor (BAY11-7082; Cat. no. 19542-67-7; Santa Cruz Biotechnology, Inc.) for 12 h. We then measured the activity of LDH released from the cells using the cytotoxicity detection kit (LDH; Roche) following the manufacturer's instructions.

Detection of cytokines secreted by SFMSCs. We measured the concentrations of IL-8, IL-1 β , IL-6, IL-10, TNF- α and IL-12p in the culture medium of SFMSCs following treatment with cytokines *in vitro* using a BD CBA Human Soluble Protein Master Buffer kit (Human Inflammatory Cytokine kit; Cat. no. 551811; BD Biosciences).

Immunofluorescence staining for NF- κ B. The human SFMSCs were plated in 24-well plates and allowed to grow onto coverslips placed in each well. At approximately 50% confluence, the wells received either the same volume of diluting agent as the control group or 10 ng/ml IL-1 β for 2 h. The medium was then aspirated, and the cells were washed twice with phosphate-buffered saline (PBS). The cells were then fixed in 4% paraformaldehyde for 30 min prior to treatment with 3% Triton X-100 for 15 min at approximately 26°C. The cells were blocked for 2 h with 5% bovine serum albumin. After washing with PBS, the cells were stained with rabbit anti-human NF- κ B p65 antibody (1:400; Cell Signaling Technology, Boston, MA, USA) for 18 h at 4°C. After washing with 1X PBS, the cells were incubated with the secondary antibody, DyLight 549 goat anti-rabbit IgG antibody (1:100; Cat. no. E032320-01; EarthOx LLC, San Francisco, CA, USA) for 60 min at 37°C. After washing with PBS, the cells were incubated with 4',6-diamidino-2-phenylindole (DAPI; Cell Signaling Technology) for 5 min. The cells were viewed under a fluorescent microscope (Olympus, Tokyo, Japan). As a specificity control, the cells were treated using the same process, but without incubation with a primary antibody.

Dual-luciferase reporter assay. The human SFMSCs were seeded into 96-well plates at 5,000 cells/well. pGL3-basic-NF- κ B-REVector and pRL-SV40-*Renilla* were transfected into the SFMSCs using ViaFect transfection reagent (both from Promega, Madison, WI, USA) according to the manufacturer's instructions. Following incubation for 48 h, cells were treated with the same volume of diluting agent as the control group or 10 ng/ml IL-1 β for 2 h. Luciferase activity in the SFMSCs was detected using the GloMax-MultiDetection system and the Dual-Luciferase Reporter assay kit (Cat. no. E4030) (both from Promega).

CBA assay. The human SFMSCs were seeded into 10-mm plates and cultured until they reached 80% confluence. The cells were treated with cytokines and/or inhibitors. Inhibitors [BAY11-7082 (Cat. no. 19542-67-7; Santa Cruz Biotechnology, Inc.), anti-TNF- α (167348; Ebioscience, San Diego, CA, USA)] were added 1 h prior to cytokine treatment. The empty plate using the same culture medium, but without cells as the negative control. IL-6 and IL-8 expression in the culture medium was determined using a CBA kit (Human Inflammatory Cytokine kit; Cat. no. 551811; BD Biosciences). Double-stranded DNA was detected using Quant-iT PicoGreen dsDNA reagent (Invitrogen, Carlsbad, CA, USA) as an endogenous control.

Western blot analysis. Cellular proteins were extracted using RIPA buffer (Cell Signaling Technology) and separated by sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE). The tissue samples were electrophoresed in a stacking gel (5% w/v) for 30 min and run on a running gel (10% w/v) for 120 min. Proteins were transferred electrophoretically onto PVDF membranes by applying 200 mA for 75 min. The membranes were first reacted with antibodies against NF- κ B p65 (1:1,000; Cat. no. 3033; Cell Signaling Technology) and GAPDH (1:2,000; Cat. no. Mab5465; MultiSciences Biotech Co., Hangzhou, China) overnight at 4°C, and then with IgG conjugated to horseradish peroxidase (Cat. no. 7076; Cell Signaling Technology) for 1 h at room temperature. The target bands were visualized by an enhanced chemiluminescence method. The corrected optical density of the protein bands was quantified using a Bio-Rad image analyzer (Bio-Rad, Hercules, CA, USA).

Effect of IL-1 β on the chondrogenic differentiation of SFMSCs. The chondrogenic differentiation procedure was performed as described above, with the addition of the corresponding cytokines. In order to evaluate the chondrogenic differentiation potential upon stimulation with rhIL-1 β , rhIL-6 or rhIL-8 (PeproTech), we measured the production of sulfated glycosaminoglycans (sGAG) in the chondrogenic pellets stimulated with rhIL-6 or rhIL-8, and the mRNA expression of *SOX9* and *MMP13* in the chondrogenic pellets, mediated by rhIL-1 β .

sGAG assay. The samples were digested overnight at 56°C using 100 μ l of a 50 μ g/ml proteinase K solution in 100 mM Na₂HPO₄ (pH 8.0). Proteinase K was then inactivated by heating the preparation for 10 min at 90°C. Following centrifugation, 500 μ l of working DMMB solution were added to 50 μ l of proteinase K-treated or untreated sample, and the mixture was vigorously vortexed for 30 min to promote complete

GAG/DMMB complexation. The samples were then centrifuged at 12,000 x g for 10 min. The supernatant was discarded, and the pellet was dissolved with DMMB decomplexation solution and vortexed for 30 min. The absorbance at 656 nm was used to quantify the sGAG levels.

Statistical analysis. In this study, the numerical data are expressed as the means \pm standard deviation (SD). Statistical significance was evaluated using the Student's t-test, and a value of $p < 0.05$ was considered to indicate a statistically significant difference.

Results

Cytokine expression in the synovial fluid of patients with TMD. In the present study, we randomly selected 19 patients with TMD (19 laterals) to evaluate the IL-8, IL-1 β , IL-6, IL-10, TNF- α and IL-12p expression levels in the synovial fluid of the TMJ. The levels of these cytokines were 18.23-282.24 pg/ml (average 88.25 pg/ml), 0.57-424.01 pg/ml (average 120.21 pg/ml), 2.80-498.44 pg/ml (average 120.73 pg/ml), 7.63-215.54 pg/ml (average 58.61 pg/ml), 0.00-321.55 pg/ml (average 85.48 pg/ml), and 36.39-482.77 pg/ml (average 157.32 pg/ml), respectively. The DI of these patients with TMD ranged from 0.05 to 0.57 (average 0.27). A more detailed summary of the cytokine expression profiles in the synovial fluid of the patients with TMD is depicted in Fig. 1.

Characteristics of SFMSCs in vitro. After the synovial fluid samples were plated and cultured for approximately 2 weeks, cell clones were observed in culture (Fig. 2A), and the cells exhibited a typical fibroblastic spindle shape (Fig. 2B). The SFMSCs expressed CD90, CD105, CD73 and CD44, whereas CD45, CD34, CD11b, CD19 and HLA-DR were not detected by FACS analysis (Fig. 3).

Differentiation of SFMSCs. Following culture in osteogenic induction medium for 4 weeks, calcium deposits were observed in the SFMSCs (Fig. 4B), while none were observed in the control group (Fig. 4A), as shown by Alizarin Red staining (Fig. 4A and B). The expression of the osteogenic transcription factor, *RUNX2*, was also found to be higher in the group subjected to osteogenic differentiation (induced group) compared to the controls (Fig. 4C). The expression of *OCN*, which is a marker for late-stage-matrix-producing osteoblasts, was significantly increased (Fig. 4D).

Following 28 days of culture in adipogenic induction medium, the SFMSCs had developed into lipid-laden fat cells that stained Oil Red O-positive (Fig. 4F), while this was not observed in the control group (Fig. 4E). In addition, *PPARG2* expression was significantly increased (Fig. 4G). The gene encoding *LPL*, a lipid exchange enzyme that is upregulated during adipogenesis, was significantly upregulated at day 28 of induction (Fig. 4H).

Following 28 days of culture in chondrogenic induction medium, the formation of cartilage pellets was evident (Fig. 4I), and this was not observed in the controls. The histological sections of the cartilage pellets were positively stained with Safranin O (Fig. 4J), as also shown in our previous study (6). In addition, there was extensive immunoreactivity for collagen type II, a characteristic

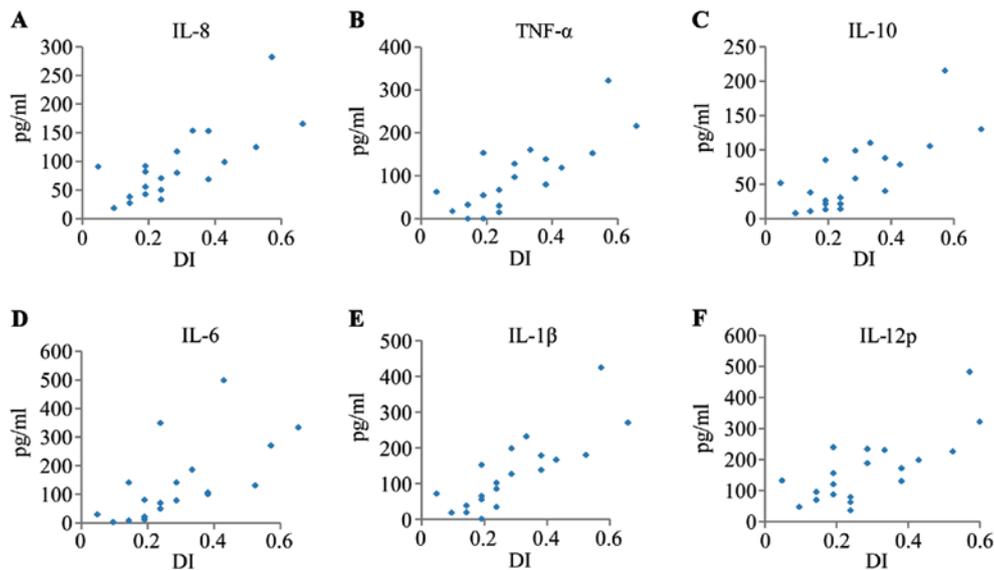


Figure 1. (A-F) Cytokine expression profiles of synovial fluid from patients with temporomandibular joint disorders (TMD). Subjects were evaluated using the dysfunction index (DI) described in the study by Friction JR and Schiffman (23).

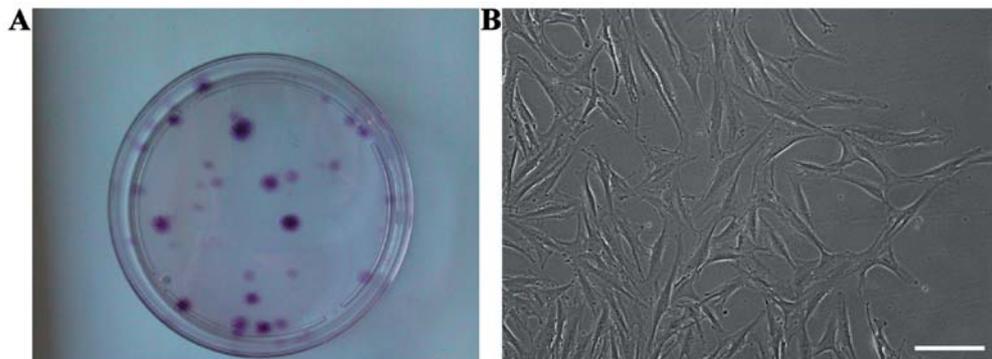


Figure 2. Characteristics of synovial fluid-derived mesenchymal stem cells (SFMSCs) *in vitro*. (A) Clones of SFMSCs were observed after *in vitro* culture of synovial fluid samples for 2 weeks. (B) Shape of SFMSCs in monolayer culture. Scale bars, 100 μ m.

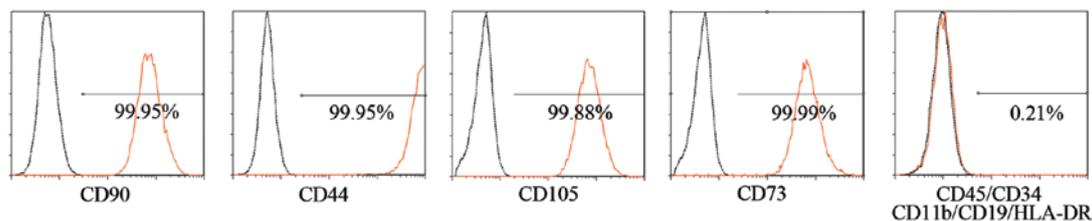


Figure 3. Mesenchymal stem cell (MSC) surface marker assay in synovial fluid-derived mesenchymal stem cells (SFMSCs).

collagen found in cartilage (Fig. 4K), whereas no immunostaining was detected in the respective controls (Fig. 4L).

Following 24 h of culture in neurogenic induction medium, the cells exhibited a bipolar and stellate morphology (Fig. 4N), while the cells in the control group exhibited a typical fibroblastic spindle shape (Fig. 4M). The expression of the glia-associated marker, glial fibrillary acidic protein (GFAP, a specific marker for the astrocyte lineage) in SFMSCs was demonstrated by immunofluorescence staining (Fig. 4O). No immunostaining was detected in the respective control cells (Fig. 4P).

Cytotoxicity detection. None of the treatments induced a significant increase in LDH activity compared to the controls,

indicating that they did not exert cytotoxic effects on the SFMSCs (data not shown).

Cytokine secretion from SFMSCs stimulated with the inflammatory cytokines, IL-8, IL-1 β , IL-6, IL-10, TNF- α and IL-12p. We then determined which cytokines were secreted by SFMSCs upon exposure to the inflammatory cytokines, IL-8, IL-1 β , IL-6, IL-10, TNF- α and IL-12p. We found that only the IL-8 and IL-6 levels increased significantly following stimulation with IL-1 β , independent of IL-8/IL-1 β /IL-6/IL-10/TNF- α /IL-12p compound stimulation (Fig. 5E and F). This secretion profile was also observed following stimulation with, or the blockade of, TNF- α signaling (Fig. 6). IL-8 and IL-6 expres-

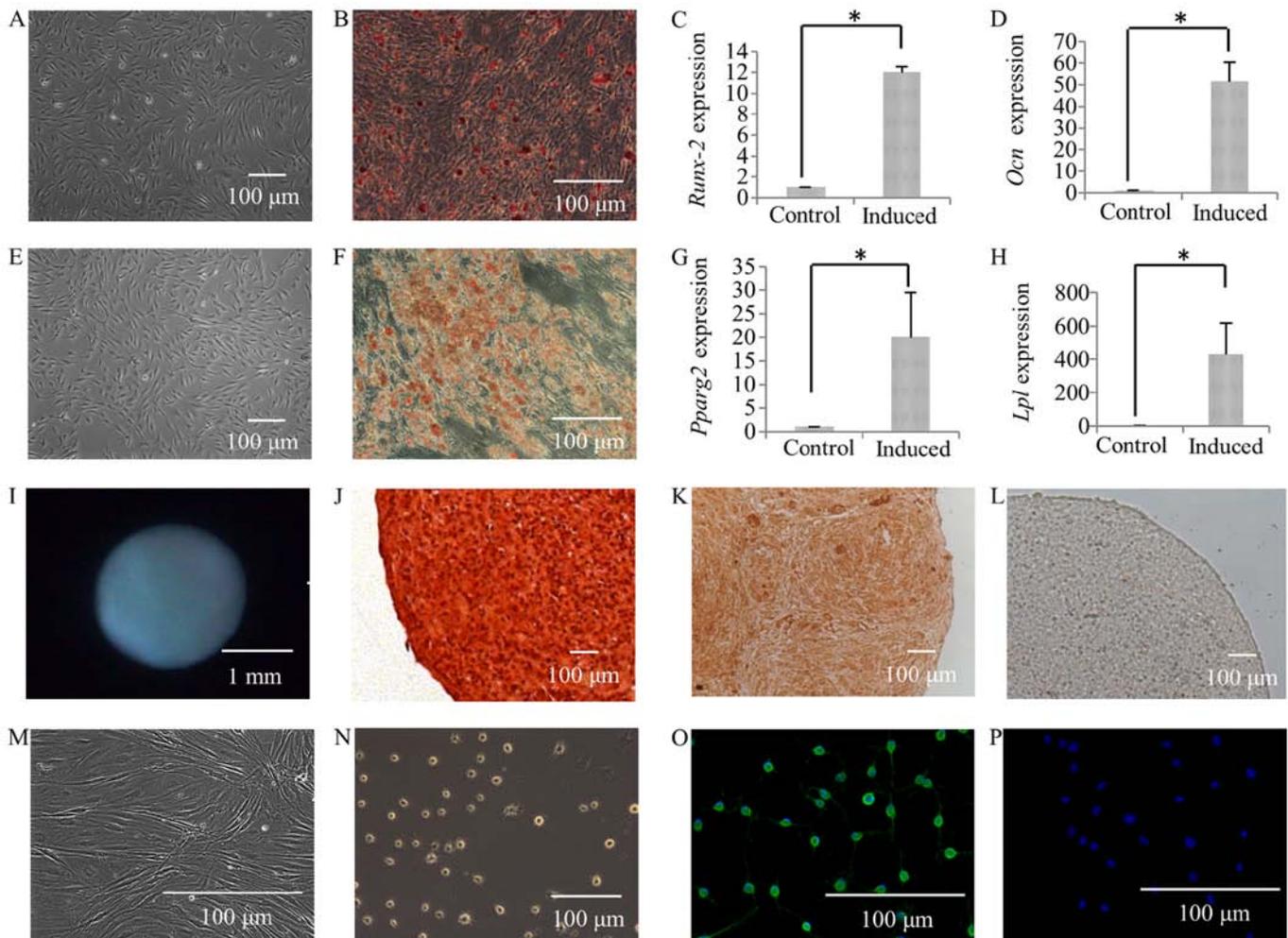


Figure 4. Alizarin Red staining of SFMSCs prior to culture in osteogenic induction medium (A), and following culture in osteogenic induction medium for 4 weeks (B), and the expression of *RUNX2* (C) and *OCN* (D) was compared between the osteogenic induction group and the control group. Oil Red O staining of SFMSCs prior to culture in adipogenic induction medium (E) and following culture in adipogenic induction medium for 4 weeks (F), and the expression of *PPARG2* (G) and *LPL* (H) in the adipogenic induction group was compared to that in controls. Following culture in chondrogenic induction medium for 4 weeks, cartilage pellets formed in the tube (I), and the pellets were subjected to Safranin O staining (J) and immunohistochemical staining for type II collagen (K). The control group sections, which were treated using the same immunohistochemical staining process but without incubation with type II collagen antibody, exhibited negative staining (L). The morphology of SFMSCs prior to culture in neurogenic induction medium (M) and following culturing in neurogenic induction medium for 24 h (N); Cells after induction (O) were also stained for GFAP, while the control group (P) was not. Significant differences between the 2 groups are indicated by single asterisks (* $p < 0.05$).

sion was markedly dependent on the concentration of IL-1 β used (Fig. 5A-E).

To determine whether NF- κ B was responsible for the effects of IL-1 β , we performed immunofluorescence staining using an antibody to the NF- κ B p65 subunit (Fig. 7A and B). Compared to the control group, the nuclei of the SFMSCs stimulated with IL-1 β were strongly stained, indicating the induction of NF- κ B activity. The results of western blot analysis revealed that NF- κ B p65 expression increased in response to IL-1 β (Fig. 7D). The increased NF- κ B activity was also confirmed by dual-luciferase reporter assays (Fig. 7C). Compared with the untreated control group, the levels of IL-8 and IL-6 were much higher in the SFMSCs stimulated with 10 ng/ml IL-1 β for 2 h (Fig. 8A-C). Pre-treatment with NF- κ B inhibitor for 1 h significantly attenuated the IL-1 β -dependent induction of IL-8 and IL-6 expression (Fig. 8C and D). Taken together, these data indicate that the secretion of IL-6 and IL-8 by SFMSCs upon stimulation with IL-1 β *in vitro* requires the activation of the NF- κ B pathway.

Influence of cytokines on the chondrogenesis of SFMSCs in vitro. Compared to the untreated cells, the induction of chondrogenic differentiation in the cells stimulated with IL-1 β was associated with the downregulation of *SOX9* and the upregulation of *MMP13* (Fig. 9).

To determine the relative contributions of IL-6 and IL-8 to the inhibition of chondrogenesis, we exposed the SFMSCs to IL-6 and IL-8 and quantitatively analyzed the sGAG content. The sGAG levels of the cartilage pellets exposed to 0, 0.1, 1 and 10 ng/ml IL-6 decreased from 13.17 ± 0.48 to 9.20 ± 0.39 μ g/pellet (300,000 cells/pellet) (Fig. 10A). By contrast, there were no significant changes observed in sGAG synthesis following exposure to IL-8 (Fig. 10B).

Discussion

MSCs are adult stem cells with notable self-renewal and multi-lineage differentiation capacities (24). MSC-based technology

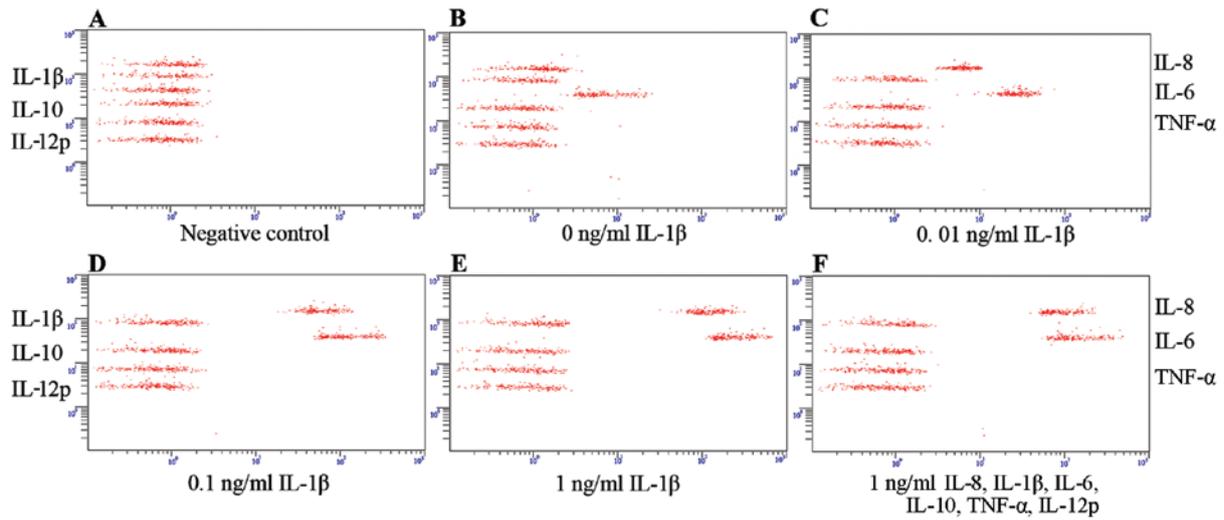


Figure 5. Secreted cytokine profiles of synovial fluid-derived mesenchymal stem cells (SFMSCs). (A) In culture medium without cells, (B) without interleukin (IL)-1 β stimulation, (C-E) following stimulation with IL-1 β at the indicated doses, and (F) with 1 ng/ml IL-8/IL-1 β /IL-6/IL-10/tumor necrosis factor (TNF)- α /IL-12p for 12 h.

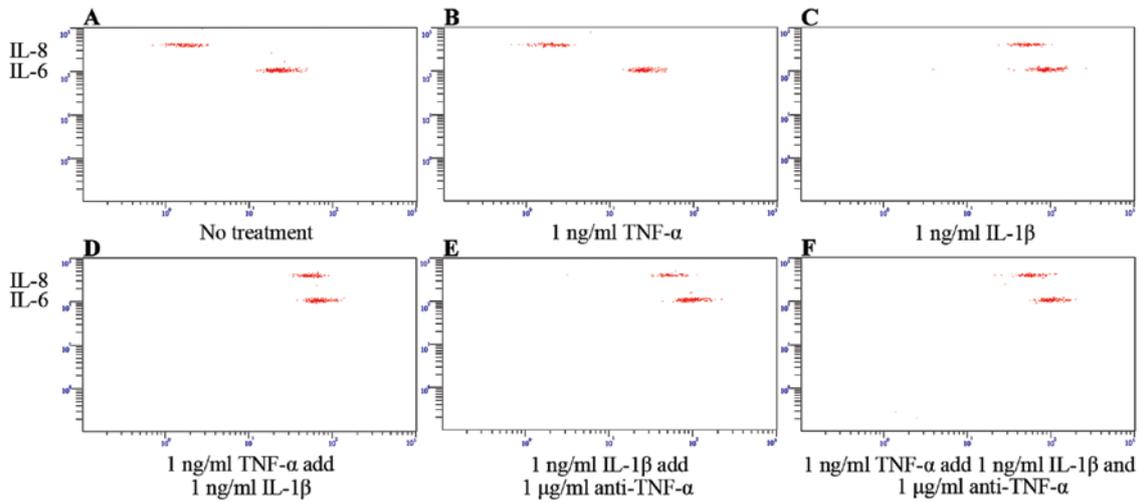


Figure 6. Tumor necrosis factor- α (TNF- α)- and interleukin (IL)-1 β -dependent cytokine secretion from synovial fluid-derived mesenchymal stem cells (SFMSCs). (A) In culture medium without cytokine stimulation, and (B) upon stimulation with 1 ng/ml TNF- α , (C) 1 ng/ml IL-1 β , (D) 1 ng/ml TNF- α and 1 ng/ml IL-1 β , (E) 1 ng/ml IL-1 β and 1 μ g/ml anti-TNF- α , and (F) 1 ng/ml TNF- α and 1 ng/ml IL-1 β with the addition of 1 μ g/ml anti-TNF- α for 12 h.

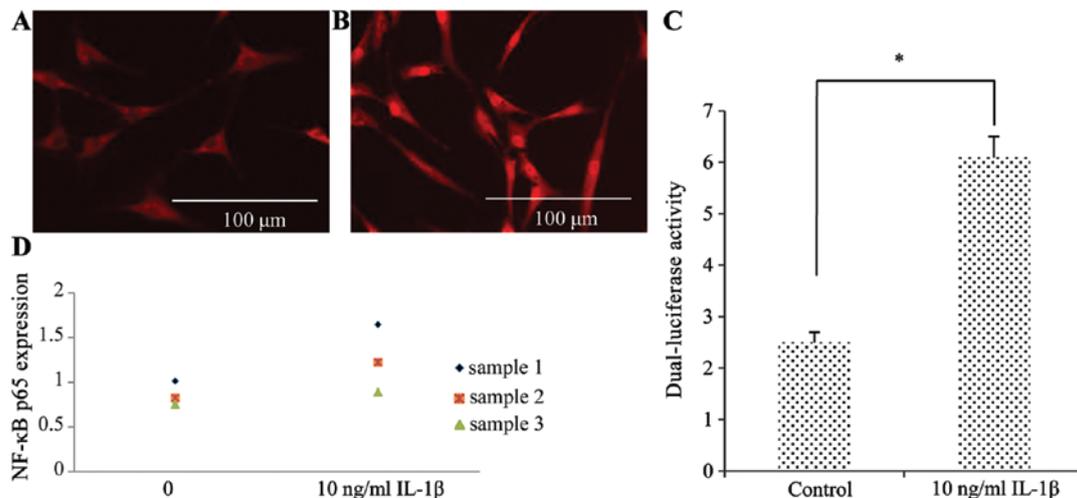


Figure 7. Interleukin-1 β (IL-1 β)-dependent activation of the nuclear factor- κ B (NF- κ B) pathway in synovial fluid-derived mesenchymal stem cells (SFMSCs). Microscopic image of NF- κ B immunofluorescence staining in SFMSCs (A) without and (B) with IL-1 β stimulation. (C) Quantitative analysis of NF- κ B activity in response to IL-1 β as determined by a dual-luciferase assay. (D) Western blot analysis of phospho-NF- κ B in IL-1 β -treated and control groups. Significant differences between the 2 groups are indicated by a single asterisk (* p <0.05).

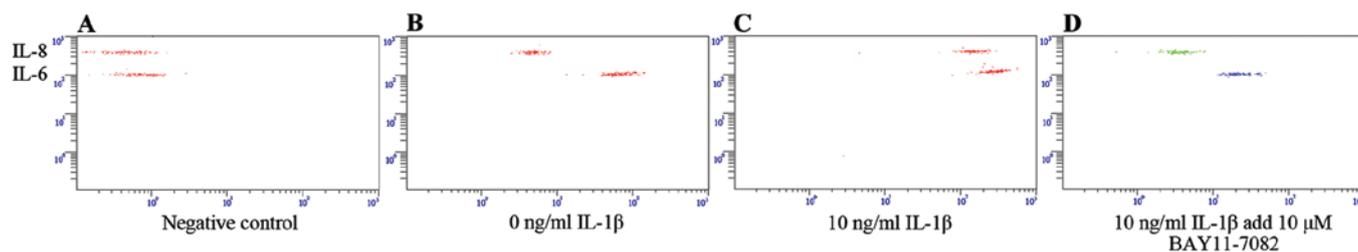


Figure 8. Interleukin (IL)-1 β -induced cytokine secretion from synovial fluid-derived mesenchymal stem cells (SFMSCs) requires nuclear factor- κ B (NF- κ B). (A) In culture medium without cell growth, (B) without IL-1 β stimulation, (C) following stimulation with 10 ng/ml IL-1 β , (D) following a 1 h pre-incubation with 10 μ M BAY11-7082 (NF- κ B inhibitor) prior to the addition of 10 ng/ml IL-1 β .

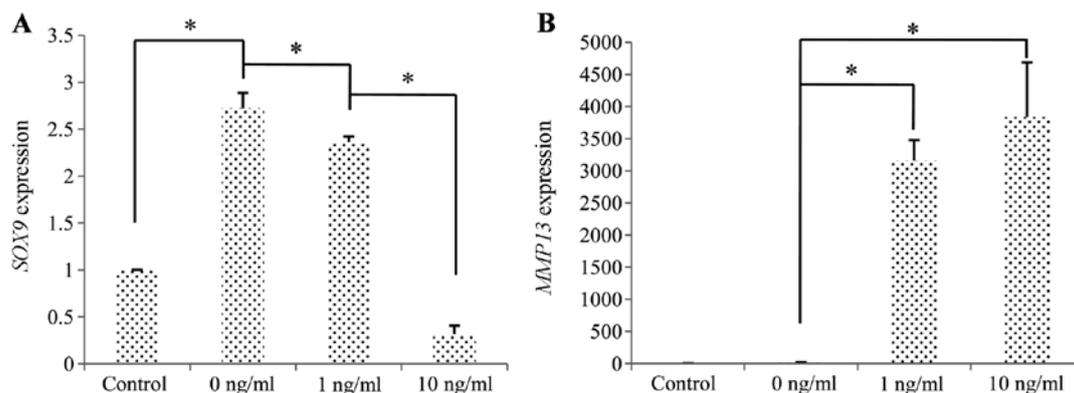


Figure 9. Influence of interleukin (IL)-1 β on chondrogenesis of synovial fluid-derived mesenchymal stem cells (SFMSCs) *in vitro*. RT-qPCR of chondrogenic markers. Statistically significant suppression of (A) *SOX9* and the upregulation of (B) *MMP13* are shown. Significant differences between the 2 groups are indicated by a single asterisk (* p <0.05).

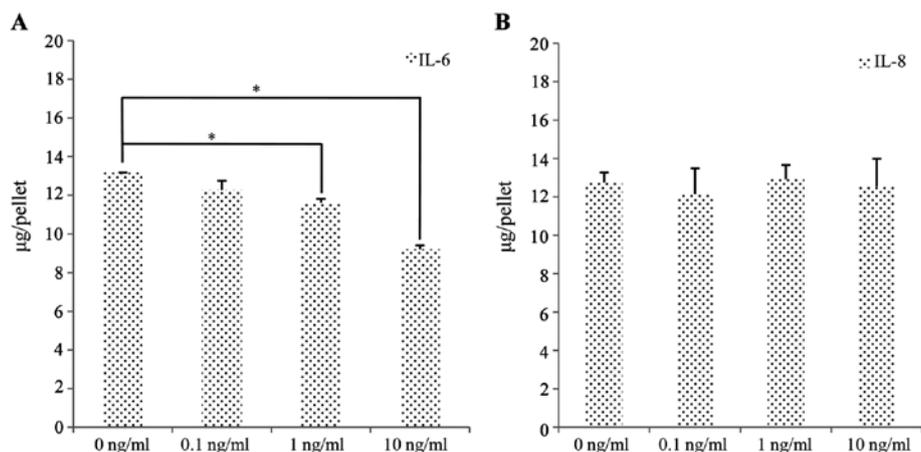


Figure 10. Effect of interleukin (IL)-6 and IL-8 on the levels of sulfated glycosaminoglycans (sGAG), a marker of chondrogenesis. The levels of sGAG was significantly decreased in (A) IL-6-treated chondrogenesis pellets in a concentration-dependent manner. The levels of sGAG were not significantly decreased in (B) the IL-8-treated chondrogenic pellets. Significant differences between the 2 groups are indicated by a single asterisk (* p <0.05).

is considered one of the most promising cell-based treatment options for several diseases, including those associated with tissue injury (25). Various stem cell sources have been investigated for the repair of damaged tissue. Synovium-derived MSCs exist in the same microenvironment as articular cartilage and display the optimal potential for both proliferation and chondrogenesis when compared with other MSC sources (26-28).

In this study, cells isolated from synovial fluid samples exhibited a fibroblastic spindle-like shape and expressed CD90, CD105, CD73 and CD44; they did not express CD45,

CD34, CD11b, CD19 or HLA-DR. These cells also exhibited multilineage potential and thus match the criteria for human MSCs proposed by the Mesenchymal and Tissue Stem Cell Committee of the International Society for Cellular Therapy (ISCT) (24). SFMSCs are a promising cellular resource for autologous articular cartilage repair, as they are simple to isolate and culture, and their introduction into the host does not cause much trauma. Although the existence of SFMSCs has been documented in some studies (2,4,5), research into the biological characteristics of SFMSCs in

TMD is limited. However, particular attention should be paid to the way in which SFMSCs are used in the clinic, as the safety and efficacy of the procedure are paramount.

Pro-inflammatory cytokines are key contributors to TMD pathology. Indeed, cytokine regulatory networks are well-documented features of arthritis. The concentrations of the pro-inflammatory cytokines, IL-1 β , IL-6, IL-8, IL-10, IL-12p and TNF- α , in patients with TMD are much higher than those in non-affected individuals; this leads to synovitis and the production of MMPs. The effects of individual cytokines are target-tissue-dependent. Fibroblast-like synoviocytes in the joint are considered major effectors of cartilage destruction, and they perpetuate inflammation by producing cytokines and degradative enzymes (29). In this study, we simulated a pro-inflammatory environment by treating SFMSCs with IL-1 β , IL-6, IL-8, IL-10, IL-12p and TNF- α *in vitro*.

We observed a robust secretion of IL-6 and IL-8 by SFMSCs following stimulation with IL-1 β in a concentration-dependent manner, and this was independent of IL-8, IL-6, IL-10, TNF- α and IL-12p stimulation. IL-8 is one of the most potent chemoattractants for neutrophils and T lymphocytes (30). IL-6 is important not only in the orchestration of the overall cytokine network, but also as it plays a direct role in destruction of the joint (30). Therefore, SFMSCs appear to be an alternative source of inflammatory cytokines.

Pro-inflammatory cytokines in the inflammatory milieu can modulate some of the fundamental characteristics of MSCs (31-33). There are studies on the beneficial effects of BMSCs on cartilage function in OA (34,35). However, Djouad *et al* observed a switch in the behavior of MSCs depending on the inflammatory environment. Specifically, the injection of MSCs into the inflamed tissue in a collagen-induced arthritis model had no beneficial effects on arthritis severity, and in some cases it even aggravated the collagen-induced arthritis symptoms (36). Augello *et al* (34) attributed these effects to the allogeneic C3H10T1/2 cells used by Djouad *et al*, as it is an immortalized cell line that may have lost the expression of immunosuppressors. Liu and Hwang reported that the continuous exposure of MSCs from cord blood to IL-1 β facilitated adipogenic maturation (37). However, whether or not SFMSCs in the TMD microenvironment have beneficial effects on cartilage repair remains unclear.

We further evaluated the chondrogenic differentiation potential of SFMSCs following stimulation with IL-1 β , a critical cytokine that mediates the activation of inflammatory and degradative pathways that are associated with TMD. We found that the expression of *SOX9*, which is a chondrogenic marker, was significantly decreased following exposure to IL-1 β . By contrast, we observed an increased expression of the gene encoding *MMP13*, the major protease involved in degradation of the cartilage extracellular matrix, and a factor in OA progression. These results indicate that IL-1 β impedes the chondrogenic differentiation of SFMSCs in the human TMJ. However, further studies are required to elucidate the molecular mechanisms that underlie this process.

Djouad *et al* reported that MSC-like cells from the synovial membrane expressed higher levels of transcripts for macrophage inflammatory protein (MIP2)- α , IL-6 and IL-8 (38). IL-6 and IL-8, which are produced mainly in response to IL-1 β , are significant contributors to TMJ disorders, such as synovitis,

arthralgia and OA (30,39,40). We further demonstrated that IL-6 significantly inhibited the SFMSC-dependent sGAG production that accompanies the induction of chondrogenesis. By contrast, IL-8 did not have this effect, indicating that IL-6 is a key mediator of the IL-1 β -dependent inhibition of chondrogenic differentiation.

The NF- κ B pathway plays a major role in transducing inflammatory and catabolic signals in joint degeneration (41). In this study, to further determine whether NF- κ B plays a pivotal role in IL-1 β -mediated IL-6 upregulation, we first examined the association between NF- κ B activity and the presence of IL-6 and IL-8. We found that NF- κ B activity in the SFMSCs increased significantly upon exposure to IL-1 β . Furthermore, the IL-1 β -dependent expression of IL-6 and IL-8 was reduced to the basal levels when the cells were pre-treated with an NF- κ B inhibitor, confirming that the NF- κ B pathway indeed contributes to the IL-1 β -mediated IL-6 upregulation. Thus, the detailed regulatory mechanisms involved in the NF- κ B pathway and the biological effects of SFMSCs *in vivo* require further investigation.

In conclusion, in this study, we found that IL-1 β plays an important role in regulation of the biological behavior of SFMSCs in an inflammatory milieu. In doing so, IL-1 β impedes the chondrogenic differentiation of SFMSCs. The upregulation of IL-6 and the activation of the NF- κ B pathway contribute to this biological behavior. Our results indicate the potential adverse effects of IL-1 β on the chondrogenic differentiation of SFMSCs, which may provide new insight into the pathogenesis of TMD. Our findings also provide an experimental basis for the existence of MSCs that initiate self-renewal in the synovial fluid.

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