

Isopsoralen-mediated suppression of bone marrow adiposity and attenuation of the adipogenic commitment of bone marrow-derived mesenchymal stem cells

JIAN WANG^{1*}, SHENG-FA LI^{2*}, TING WANG^{3*}, CHUN-HAN SUN^{2*}, LIANG WANG⁴, MIN-JUN HUANG⁴, JIAN CHEN⁵, SHAO-WEI ZHENG², NAN WANG⁶, YING-JUN ZHANG⁷ and TIAN-YU CHEN⁴

¹Department of Orthopaedics, People's Hospital of Inner Mongolia Autonomous Region, Hohhot 010050;

²Department of Orthopaedics, The First People's Hospital of Huizhou, Huizhou, Guangdong 516003;

³Department of Cell Biology, School of Basic Medical Science, Southern Medical University, Guangzhou, Guangdong 510515;

⁴Department of Orthopedics, Guangdong Orthopedics Academy, The Third Affiliated Hospital of Southern Medical University, Guangzhou, Guangdong 510665; ⁵Department of Orthopaedics, Three Gorges Central Hospital of Chongqing, Chongqing 404100;

⁶Research Center of Clinical Medicine, Nanfang Hospital, Southern Medical University, Guangzhou, Guangdong 510515;

⁷Department of Medical Imaging, Hunan University of Medicine, Huaihua, Hunan 418000, P.R. China

Received October 4, 2015; Accepted October 4, 2016

DOI: 10.3892/ijmm.2017.2880

Abstract. Osteoporosis (OP) increases the risk of bone fractures and other complications, and is thus a major clinical problem. In this study, we examined the effect of isopsoralen on the differentiation of bone-derived marrow mesenchymal stem cells (BMSCs) into osteoblasts and adipocytes, as well as bone formation under osteoporotic conditions. Primary femoral BMSCs isolated from C57BL/6 mice were used to evaluate the isopsoralen-mediated regulation of the expression of alkaline phosphatase (ALP), osteocalcin (OCN) and runt-related transcription factor 2 (RUNX2) during osteogenesis 2 weeks. We also examined the expression of peroxisome proliferator-activated receptor γ (PPAR γ) and CCAAT/enhancer binding protein β (C/EBP β) under adipogenic conditions for 1 and 2 weeks. In addition, ovariectomized (OVX) mice were used to examine the effects of isopsoralen on bone formation for 2 months. Finally, mammalian target of rapamycin complex 1 (mTORC1) signaling was examined under osteogenic and adipogenic conditions. We found that following treatment with isopsoralen, the expression levels of ALP, OCN and RUNX2 were upregulated, whereas those of PPAR γ and C/EBP β were downregulated. mTORC1 signaling was also inhibited *in vitro* and *in vivo*. In the OVX

mice that were intragastrically administered isopsoralen, bone parameters (trabecular thickness, bone volume/total volume and trabecular number) in the distal femoral metaphysis were significantly increased and the adipocyte number was decreased. On the whole, our findings demonstrate that isopsoralen promoted BMSC differentiation into osteoblasts and suppressed differentiation into adipocytes.

Introduction

Osteoporosis is a bone disease which affects bone strength. It is known to increase the risk of bone fractures and other complications, and has become a major clinical problem that affects over 200 million individuals worldwide (1). Current strategies for the treatment of osteoporosis, include the administration of teriparatide, strontium and bisphosphonates, mainly aimed at reducing bone resorption (which is regulated by osteoclasts) and increasing bone formation (which is regulated by osteoblasts). However, these synthetic medicines have several side-effects that reduce their efficacy, such as jaw bone osteonecrosis induced by large doses of bisphosphonates (which act as inhibitors of bone resorption) or an increased risk of osteosarcoma induced by prolonged treatment with teriparatide (2,3). Thus, healthier and safer agents that can be used in the prevention or treatment of bone-related disorders are highly desired.

Previous studies have indicated that the decreased bone volume in osteoporosis is associated with increased adipose tissue in bone marrow (4-6). Osteoblasts and marrow adipocytes differentiate from a common precursor, namely bone marrow-derived mesenchymal stem cells (BMSCs) (7). It has been previously demonstrated that compared with the osteogenic capability of mesenchymal stem cells (MSCs) derived from healthy women, those derived from post-menopausal women with osteoporosis have a much lower osteogenic potential (8,9).

Correspondence to: Dr Tian-Yu Chen, Department of Orthopedics, Guangdong Orthopedics Academy, The Third Affiliated Hospital of Southern Medical University, 183 West Zhongshan Avenue, Guangzhou, Guangdong 510665, P.R. China
E-mail: 402970117@qq.com

*Contributed equally

Key words: biochemistry, bone, chemicals, immunobiology, endocrinology/metabolism, molecular

Previous studies have demonstrated a large degree of plasticity of osteoblasts and adipocytes. Specifically, fully-differentiated osteoblasts derived from human MSCs (hMSCs) are capable of dedifferentiation and transdifferentiation into adipocytes and vice versa (10,11). There is a reciprocal association and balance between the differentiation of adipocytes and osteoblasts (12).

The majority of previous data suggest that bone marrow adipose tissue simply plays the role of filling the marrow cavity and lacks hematopoietic function (13,14). However, Elbaz *et al* reported a lipotoxic effect from marrow adipocytes on osteoblast differentiation and function (15). Additionally, free-fatty acids released by adipocytes can inhibit osteoblast proliferation and induce osteoblast apoptosis (16).

Existing data demonstrate that an excessive amount of bone marrow adipocytes may be a significant negative risk factor for skeletal health (17). Accordingly, inhibiting adipocyte differentiation in bone marrow, while simultaneously accelerating osteogenesis may be a therapeutic approach for age-related osteoporosis.

The fruit of *Psoralea corylifolia* L. (*P. corylifolia*) is a widely used Chinese herbal medicine and is specifically used in the treatment of fractures, and bone and joint diseases. It has also been used in the treatment of other disorders, such as skin diseases, cardiovascular diseases, tumors and asthma (18,19). It has been found that psoralen and isopsoralen (ISO) have growth inhibitory effects on transplanted tumors in nude rats with osteosarcoma; however, following the administration of high doses of psoralen and ISO, toxic reactions such as writhing, lassitude and hypoactivity were observed (19). Thus, the efficacy and safety of any agent in the treatment of osteoporosis should also be considered.

ISO is the main active ingredient extracted from the seeds of *P. corylifolia* (psoralen is its isomer). Previous studies have proposed a positive role for psoralen in promoting osteoblast differentiation (20), as well as its stimulatory effects on bone formation (21,22). However, only a few studies have evaluated the beneficial effects of ISO on bone development (23,24), and to the best of our knowledge, none have examined its underlying mechanisms of action.

In addition, to date, at least to the best of our knowledge, there are no studies available on the role and mechanisms of action of ISO in bone marrow adipogenesis. Thus, in the present study, by employing a mouse model of osteoporosis induced by ovariectomy along with ISO treatment, we aimed to examine the effects of ISO on bone marrow adipogenesis *in vitro*. Furthermore, we evaluated the effects of ISO on the differentiation of osteoblasts and adipocytes derived from BMSCs isolated from C57BL/6 mice in an effort to clarify the probable underlying cellular and molecular mechanisms that occur during this period. Our results may contribute to the development of novel therapeutic approaches for the treatment of bone-related diseases.

Materials and methods

Ethics approval. Ethics approval was provided by the Medical Ethics Committee of Southern Medical University, Guangzhou, China.

Materials and reagents. ISO (chemical structure shown in Fig. 1A) was obtained from Sigma (St. Louis, MO, USA;

purity, 99%; molecular weight, 186.1635). Stock solutions of ISO were prepared in dimethyl sulfoxide (DMSO; Sigma) and stored at -20°C .

Animal model and animal feeding. Female C57BL/6 mice (aged 2 months; $n=18$) were provided by the Experimental Animal Center of Southern Medical University. The mice were randomly divided into the sham-operated, ovariectomized (OVX) or OVX plus ISO ($n=6/\text{group}$) groups. The mice in the OVX plus ISO group were intragastrically administered ISO at a dose of 20 mg/kg/day for 5 days prior to being subjected to ovariectomy and this was maintained for 2 months after the mice were subjected to ovariectomy. A previous study found that a significant amount of bone loss in vertebrae and femurs was observed in C57BL/6 mice at only 4 weeks following ovariectomy (25). Thus, ISO treatment was maintained for 2 months in this study. The mice were anesthetized with 1% pentobarbital sodium. The operation area was disinfected by iodophor. The back skin was then longitudinally cut step by step at the second lumbar level by one incision (1.5 cm in length) and 2 sides of the ovaries were removed. Ovary peripheral vascular flow was blocked by ligation using sutures (4-0#). The incision was then closed and the mice were allowed to recover from the anesthesia at room temperature. The mice in the sham-operated group only had some fat tissue around the ovaries removed. The mice were sacrificed by cervical dislocation. The femurs were then removed using ophthalmic scissors and fixed in 10% paraformaldehyde solution.

Bone marrow adiposity analyses. For bone marrow adiposity analyses, the distal portion of the femurs was fixed, decalcified and sectioned into 2- μm -thick sections, and subjected to hematoxylin and eosin staining according to standard histological protocols. To quantify proximal metaphyseal adipocyte parameters, we calculated the adipocyte number (AD#, per mm^2). A uniform number of fields was screened in all sections by 3 authors (Jian Wang, Sheng-Fa Li and Ting Wang), starting 3 fields from the left end and 3 fields from the top endocortical surface, excluding adipocytes with disruption in the fields. To avoid any bias in the final analyses, all sections were interpreted while blinded, without knowledge of the groups (sham-operated, CON or CON + ISO). Images were obtained at $\times 20$ magnification using an Olympus BX51RF stereomicroscope (Olympus).

Microcomputed tomography (μCT) analyses. Micro-CT (μCT 80; Scanco Medical, Bassersdorf, Switzerland) analyses were performed on the distal portion of the femurs. Distal femurs were selected for scanning and corrected for the CT value, a 70 kV scanning voltage, 30 W power, 429 μA current, and 20 μm scan thickness. Our analyses included various bone parameters: trabecular thickness (Tb.Th), bone volume (BV)/total volume (TV) and trabecular number (Tb.N). The assessment of bone microstructure was carried out according to the guidelines provided in the study by Buxsein *et al* (26).

Isolation of BMSCs and cell culture. BMSCs were isolated from C57BL/6 mice ($n=10$; aged 4 weeks; also from the Southern Medical University). The mice were sacrificed using CO_2 , and the mouse femurs were dissected free of surrounding soft tissue. The bone marrow was flushed with α -MEM (Invitrogen, Carlsbad, CA, USA). The marrow content from 4 bones was

plated in culture flasks containing BMSC growth medium [α -MEM containing 10% fetal bovine serum (FBS), 100 U/ml penicillin, 100 mg/ml streptomycin sulfate (Gibco, Auckland, New Zealand). We used a centrifuge (1,000 rpm for 5 min) to isolate the cells from the extra soft tissue. The cells were re-suspended and non-adherent cells were removed, and adherent BMSCs were cultured and expanded for further experiments. The cell culture medium were replaced every 3 days. The cells were seeded in 96-well plates and 6-well plates at densities of 1×10^4 and 1×10^6 cells/well, respectively, and cultured in a humidified atmosphere of 5% CO₂ and 95% air at 37°C. After examining the levels of different proteins characteristic of BMSCs by western blot analysis (see below), we found that the cells were CD29⁺ and Sca-1⁺, and CD45⁻ and CD11b⁻; this confirmed these cells were actually BMSCs.

For osteogenic differentiation, the BMSCs were grown to 90% confluence in 6-well-plates. The culture medium was replaced with osteogenic medium (α -MEM supplemented with 15% fetal calf serum (FCS) plus 1% penicillin/streptomycin, 100 nM dexamethasone, 50 μ g/ml ascorbate-2-phosphate and 10 mM β -glycerol phosphate). The medium was changed every 3 days.

For adipocyte-induced culture, the putative BMSCs were harvested and seeded into 6-well cell culture plates. Upon attaining 90% confluency, the cells were treated with adipogenic differentiation medium: BMSC maintenance medium supplemented with 1 μ M dexamethasone, 100 μ M indomethacin and 500 μ M 1-methyl-3-isobutylxanthine (all from Sigma). For the initial induction, 5 μ g/ml bovine insulin (Sigma) were added together with adipogenic differentiation medium. The medium was changed every fourth day.

Cell proliferation assays. Primary BMSCs were seeded in 96-well plates at a density of 1×10^4 cells/well. Following culture for 2 days, the cells were treated for 48 h with ISO at concentrations of 0, 0.1, 1, 10, 100 and 1,000 μ M. Cell proliferation assays were performed using Caspase-8 Colorimetric Assay kits (KeyGEN Biotech, Nanjing, Jiangsu, China) according to the manufacturer's instructions. The absorbance in the wells was measured at 450 nm.

Alkaline phosphatase (ALP) staining assay. The BMSCs were cultured in the presence of osteogenic inducers and ISO (0, 5, 10 and 20 μ M) for 2 weeks. ALP activity was also evaluated in the cells stained using an ALP staining kit (Beyotime, Nanjing, Jiangsu, China) according to the manufacturer's instructions. For ALP staining, the cells were washed in phosphate-buffered saline (PBS), fixed in 4% paraformaldehyde for 20 min at room temperature, and rinsed in distilled water. The ALP staining mixture was added for 30 min at room temperature in the dark. The cells were rinsed in distilled water and PBS to reduce non-specific staining.

Oil Red O staining. The BMSCs were cultured in the presence of adipogenic inducers and ISO (0, 5, 10 and 20 μ M) for 7 and 14 days. Fat droplets that formed in differentiated adipocytes from the BMSCs were observed using Oil Red O staining. The cells were fixed in 4% formaldehyde for 15 min at room temperature, washed in PBS, and stained with 0.6% (w/v) Oil Red O solution (60% isopropanol, 40% water) for 1 h at

37°C. The cells were washed with PBS to remove unbound dye and isopropyl alcohol (1 ml) was added to the culture plates.

Immunocytochemical staining. The murine BMSCs (10^4 cells/well) were inoculated into 12-well assay plates with collagen-coated coverslips. Various concentrations of ISO (0, 5, 10 or 20 μ M) were added at room temperature. The cells were fixed in 4% paraformaldehyde for 15 min at room temperature, rinsed with PBS, and incubated for 1 h with blocking buffer (1X PBST, 0.5% Triton X-100) supplemented with 5% serum from the same species as the secondary antibody. The blocking solution was removed and primary mouse monoclonal antibodies against osteocalcin (OCN; Cat. no. sc-365797; 1:100) and proliferator-activated receptor γ (PPAR γ ; sc-7273; 1:200) (both from Santa Cruz Biotechnology, Inc., Santa Cruz, CA, USA) were added followed by incubation overnight at 4°C. Subsequently, the secondary antibody, goat anti-mouse IgG (SAB4600004; 1:200; Sigma) was added, followed by incubation for 1 h at room temperature. The secondary antibody was aspirated and the wells were washed 3 times with PBST. DAB (Sangon Biotech, Shanghai, China) was added followed by incubation for 5 min at room temperature in the dark. The slides were restained with hematoxylin (Sigma). Images were acquired using an Olympus BX51RF stereomicroscope (Olympus, Tokyo, Japan). The mean optical density (MOD) was examined using Image-Pro Plus software (IPP; Media Cybernetics, Rockville, MD, USA).

Immunofluorescence staining and microscopic analyses. The BMSCs were grown on collagen-coated coverslips, washed with PBS, fixed for 15 min at room temperature in 4% paraformaldehyde, washed, permeabilized with 0.5% Triton X-100 (5 min, room temperature) and blocked for 30 min in blocking buffer (5% FBS in PBST). The cells were sequentially probed with primary antibodies against runt-related transcription factor 2 (RUNX2; Cat. no. 12556; 1:100) and CCAAT/enhancer binding protein β (C/EBP β ; Cat. no. 3802; 1:200) (both from Cell Signaling Technology, Danvers, MA, USA), and washed 3 times with PBST. The FITC-conjugated sheep anti-rabbit IgG (Cat. no. F5137; 1:500) and TRITC-conjugated sheep anti-mouse IgG (Cat. no. T5393; 1:500) (both from Sigma) were added followed by incubation for 1 h at room temperature, followed by PBST washes. The cells were incubated with 4',6-diamidino-2-phenylindole (DAPI; 1 μ g/ml) for 15 min and then rinsed 3 times with PBST. Immunofluorescence staining was also performed on the femur histological sections using a standard protocol. Following ovariectomy and ISO treatment, the mice were then sacrificed. The femurs were fixed in 4% paraformaldehyde in 0.1 M phosphate buffer (pH 7.4). The fixed bones were decalcified by immersion in 10% EDTA (pH 7.0) for 14 days at room temperature and embedded in paraffin. The paraffin-embedded longitudinal bone sections (5- μ m-thick) were blocked in 5% goat serum for 1 h at room temperature, and incubated overnight with the primary antibodies. The sections were then incubated with secondary antibodies for 1 h at room temperature. Femur histological sections were imaged using a FV1000 confocal microscope and positive cells were evaluated using the IPP software program. The cells and femur histological sections were imaged using a laser-scanning confocal microscope (FV1000; Olympus). Finally, MOD was measured using IPP software.

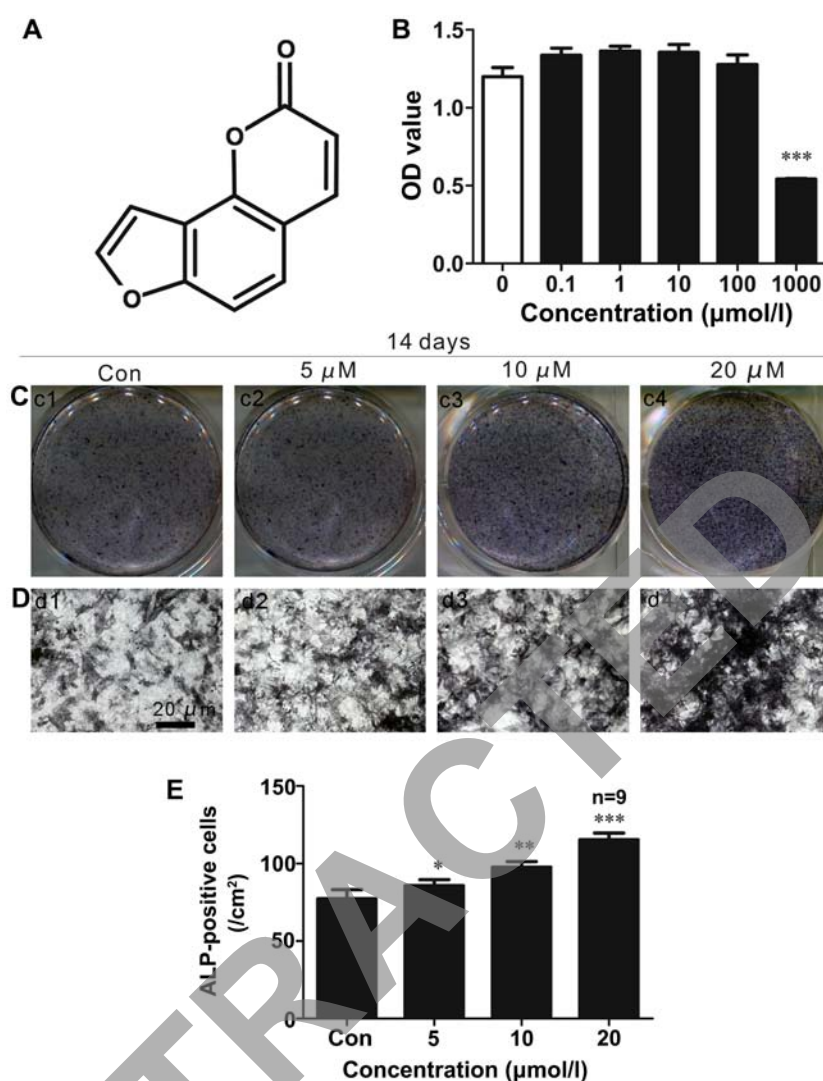


Figure 1. Isopsoralen (ISO) promotes bone-derived marrow mesenchymal stem cell (BMSC) differentiation into osteoblasts. (A) Molecular structure of ISO. (B) CCK8 assays showed that ISO did not significantly affect cell growth at the concentrations used (0.1–100 μM) following treatment for 2 days. (C–E) Alkaline phosphatase (ALP) staining showed that ISO treatment at concentrations of 5–20 μM increased ALP activity in BMSCs under osteogenic differentiation conditions for 2 weeks. * $P < 0.05$, ** $P < 0.01$ and *** $P < 0.001$ compared to the group without ISO (Con, control). Columns represent the means \pm SD from 9 wells/group (B and E).

Western blot analysis. Western blot analysis was performed using a sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) electrophoresis system. Protein samples (20 μg) were resuspended in reduced sample buffer, electrophoresed on a 7.5–10% Tris gel with Tris running buffer, blotted onto PVDF membranes, and sequentially probed with primary antibodies against RUNX2 (Cat. no. 12556; 1:1,000; Cell Signaling Technology), OCN (Cat. no. sc-365797; 1:1,000; Santa Cruz Biotechnology, Inc.), eukaryotic translation initiation factor 4E-binding protein 1 (4E/BP1; Thr37/46; Cat. no. 9644; 1:1,000; Cell Signaling Technology), phospho-S6 ribosomal protein (P-S6; S235/S236; Cat. no. sc-293143; 1:1,000), PPAR γ (Cat. no. sc-7273; 1:1,000) (both from Santa Cruz Biotechnology, Inc.), C/EBP β (Cat. no. 3802; 1:2,000), Sca-1 (Cat. no. 9664; 1:4,000), CD29 (Cat. no. 4706; 1:2,000), CD45 (Cat. no. 13917; 1:6,000), CD11b (Cat. no. 14271; 1:3,000) and β -actin (Cat. no. 3700; 1:2,500) (all from Cell Signaling Technology). Horseradish peroxidase-conjugated goat anti-rabbit (Cat. no. SAB4600223; 1:1,000) or anti-mouse (Cat. no. SAB4600004; 1:1,000) antibodies (both from Sigma)

were added, and secondary antibodies were detected using enhanced chemiluminescence (ECL Plus; General Electric Healthcare, Milwaukee, WI, USA).

Statistical analyses. We used one-way analyses of variance (ANOVA) to analyze the data. The homogeneity of the variance tests was used to evaluate data homogeneity (IBM SPSS Statistics 19.0 software). If the variance was confirmed to be equal, least-significant difference tests were used for data analyses. If the variance was determined to be unequal, Dunnett's T3 tests were used. The results are presented as the means \pm standard deviation (SD). A value of $P < 0.05$ was considered to indicate a statistically significant difference.

Results

Cell proliferation assays. We examined the effects of ISO on the proliferation of primary mouse BMSCs using CCK8 assays. As shown in Fig. 1B, ISO did not significantly affect cell growth at concentrations of 1–100 μM after 48 h. However,

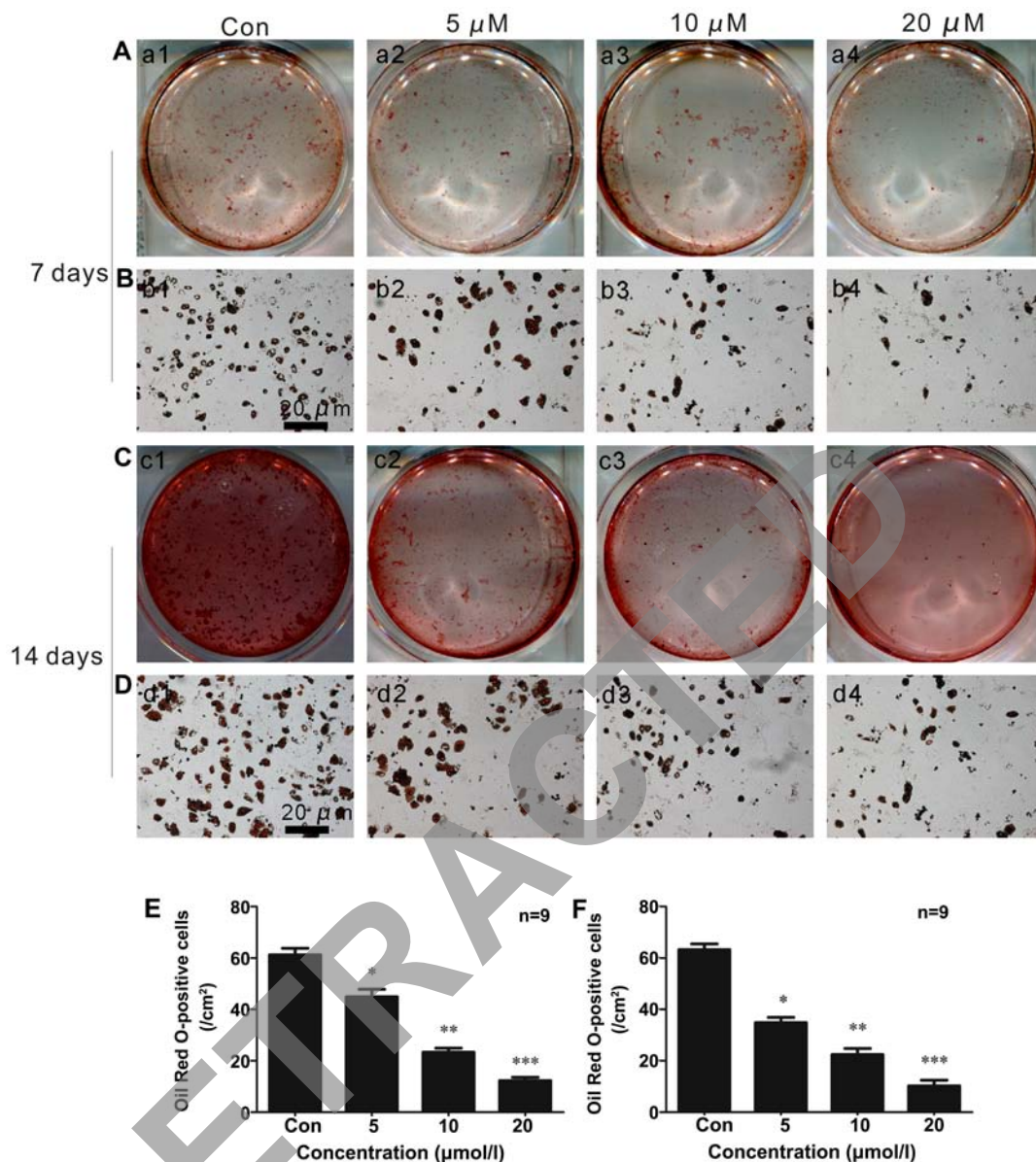


Figure 2. Isoprosalen (ISO) inhibits bone marrow stromal cell (BMSC) differentiation into adipocytes. ISO (0, 5, 10 or 20 μ M) was added to primary mouse BMSCs under adipogenic differentiation conditions. (A and B) Fewer lipid droplets appeared in the cytoplasm of adipocytes that were stained with Oil Red O in cells treated with ISO under adipogenic differentiation conditions for 7 days (E). (C and D) After 2 weeks of adipogenic differentiation, the adipogenesis rate was at its lowest level in the presence of 20 μ M ISO (F). * $P < 0.05$, ** $P < 0.01$ and *** $P < 0.001$ compared to the group without ISO (Con, control). Columns represent the means \pm SD from 9 wells/group (E and F).

BMSC proliferation was significantly inhibited following treatment with 1,000 μ M ISO.

ALP staining in vitro. To determine whether ISO affects BMSC differentiation into osteoblasts, ALP staining (Fig. 1C-E) was performed as increased ALP activity is an important indicator of osteoblast differentiation. We found that ISO enhanced ALP activity in the primary BMSCs in a dose-dependent manner (Fig. 1C-D). The most significant effects were observed at 2 weeks after osteogenic induction with a concentration of 20 μ M ISO (Fig. 1E). These results suggested that ISO significantly stimulated BMSC differentiation into osteoblasts.

Oil Red O staining in vitro. ISO inhibited the adipocytic differentiation of BMSCs induced by adipogenic inducers in

a concentration-dependent manner. As shown in Fig. 2A-F, fewer lipid droplets appeared in the cytoplasm of adipocytes that were stained with Oil Red O in the cells treated with ISO (Fig. 2E) under adipogenic differentiation conditions for 7 days (Fig. 2A, B and E). After 2 weeks of adipocytic differentiation, the adipogenic rate was at its lowest level in the presence of 20 μ M ISO (Fig. 2C and D and F) compared to the controls (untreated cells; Fig. 2F). These data suggested that ISO inhibited BMSC differentiation into adipocytes.

OCN and PPAR γ immunocytochemistry. Two weeks following osteoblast differentiation, OCN expression (brown) increased (Fig. 3A and C), with a maximum level observed in the cells treated with a concentration of 20 μ M ISO (Fig. 3C). Under adipogenic conditions for 2 weeks, the generation of

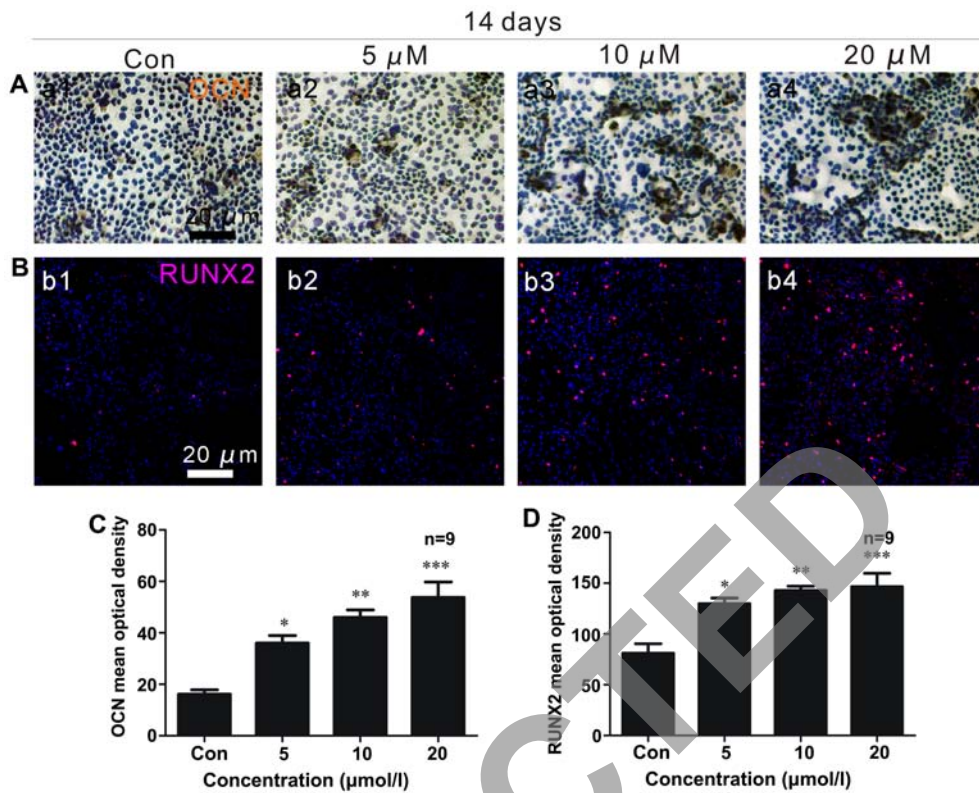


Figure 3. Isopsoralen (ISO) stimulates bone-derived marrow mesenchymal stem cell (BMSC) differentiation into osteoblasts *in vitro*. ISO (0, 5, 10 or 20 μ M) was added to primary mouse BMSCs under osteogenic differentiation conditions for 2 weeks. (A) Osteocalcin (OCN) immunocytochemistry and (B) runt-related transcription factor 2 (RUNX2) immunofluorescence were performed to examine the expression of osteoblast-specific proteins following treatment with ISO. (A) ISO increased OCN expression (brown) in BMSCs in a dose-dependent manner, with more pronounced effects at 20 μ M ISO (C). (B) ISO dose-dependently enhanced RUNX2 (red) activity in BMSCs, particularly at a concentration of 20 μ M (D). * P <0.05, ** P <0.01 and *** P <0.001 compared to the group without ISO (Con, control). Columns represent the means \pm SD from 9 wells/group (C and D).

PPAR γ (brown) was significantly reduced (Fig. 4A and D). The lowest expression of PPAR γ was noted in the presence of 20 μ M ISO (Fig. 4A and D).

RUNX2 and C/EBP β immunofluorescence. After 2 weeks of osteogenic differentiation, RUNX2 expression (red) in the osteoblasts was upregulated (Fig. 3B and D), and ISO promoted RUNX2 expression in a dose-dependent manner. In the presence of 20 μ M ISO, RUNX2 expression was at its maximum level (Fig. 3D). Additionally, after 2 weeks of adipogenic differentiation, C/EBP β (green) expression decreased (Fig. 4B and C). The minimum density of C/EBP β staining occurred in the presence of 20 μ M ISO (Fig. 3C).

RUNX2 immunofluorescence in vivo. RUNX2 is very important for bone remodeling, and functions by regulating the differentiation of osteoblasts (27,37). The intensity of positive RUNX2 staining (red) was reduced in the OVX group (Fig. 5A), whereas it was significantly elevated in the OVX + ISO group (Fig. 5A and C).

PPAR γ immunofluorescence in vivo. Increased PPAR γ (green) expression was observed in the OVX group compared with the OVX + ISO group (Fig. 5B and D). PPAR γ expression was markedly increased in mice in the OVX group (Fig. 5B, panel b2 and D) compared to the mice in the OVX + ISO

group (Fig. 5B, panel b3 and D). In addition, there were also statistically significant differences between the sham-operated group (Fig. 5B, panel b1 and D) and the OVX group. PPAR γ is considered indispensable for adipocyte differentiation.

Bone histomorphometries. Representative hematoxylin and eosin images of bone from the OVX and OVX + ISO groups (Fig. 6A) indicated that ISO reduced the number of adipocytes (AD#) in the bone marrow (Fig. 6A and G).

Micro-CT analyses. We performed μ CT scans on the mouse distal femurs (Fig. 6B-F). Two months after the surgery, bone turnover was significantly decreased in the OVX mice compared with the sham-operated group (Fig. 6D-F). Treatment with ISO treatment significantly rescued this bone loss. The values for Tb.Th (Fig. 6D), BV/TV (Fig. 6E) and Tb.N (Fig. 6F) in the OVX group were markedly decreased compared with the sham-operated group. Following treatment with ISO for 2 months, these indicators were significantly improved (Fig. 6D and E). However, compared to the sham-operated group, the levels of Tb.N, Tb.Th and BV/TV were still decreased in the OVX + ISO group (Fig. 6D-F). This result reminded us of the fact that ISO also cannot be a perfect substitute for estrogen.

Western blot analysis. To determine whether ISO affects BMSC differentiation, we performed western blot anal-

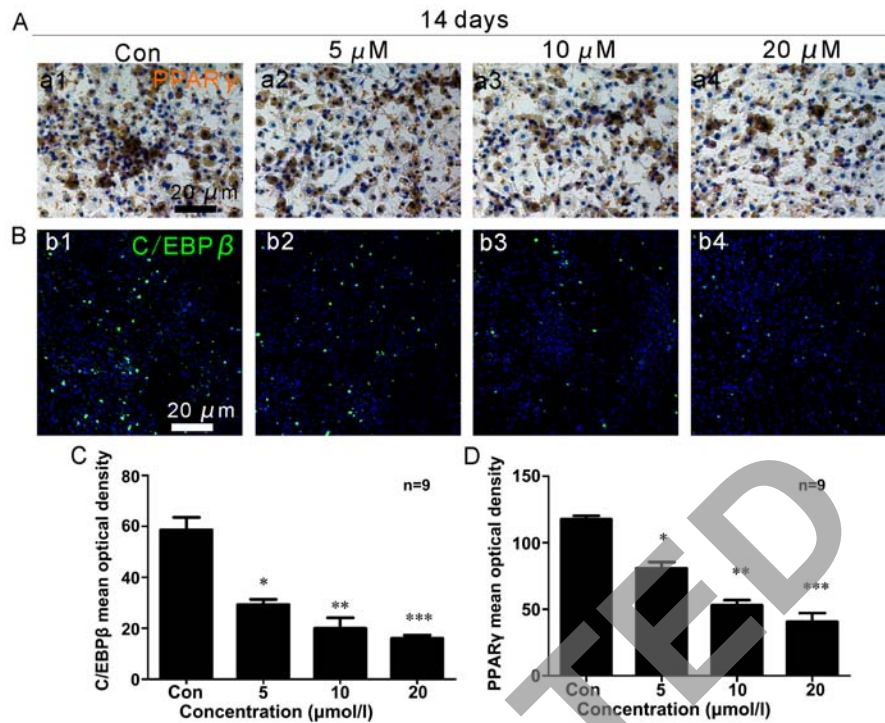


Figure 4. Isopsoralen (ISO) inhibits bone-derived marrow mesenchymal stem cell (BMSC) differentiation into adipocytes *in vitro*. ISO (5, 10 or 20 μ M) was added to primary mouse BMSCs following adipogenic differentiation. (A) Peroxisome proliferator-activated receptor γ (PPAR γ) immunocytochemistry and (B) CCAAT/enhancer binding protein β (C/EBP β) immunofluorescence assays were performed to examine the expression of adipocyte-specific proteins following treatment with ISO. (A) ISO increased PPAR γ expression (brown) in BMSCs in a dose-dependent manner, with the most pronounced effect at 20 μ M (C). (B) ISO dose-dependently enhanced C/EBP β activity (green) in BMSCs, particularly at 20 μ M (D). * P <0.05, ** P <0.01 and *** P <0.001 compared to the group without ISO (Con, control). Columns represent the means \pm SD from 9 wells/group (C and D).

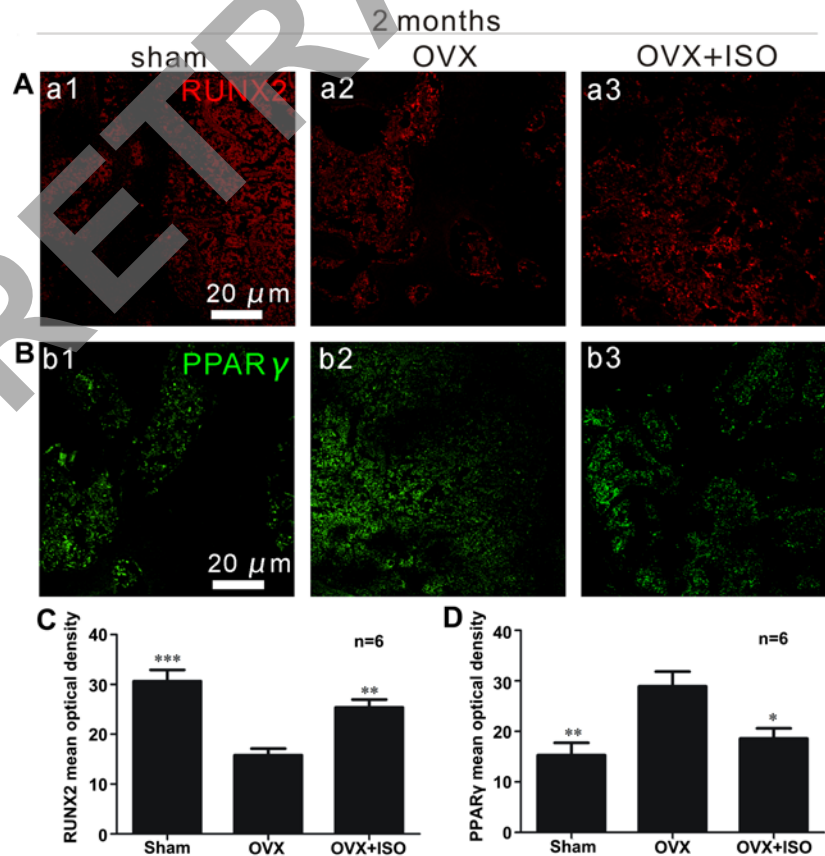


Figure 5. Isopsoralen (ISO) promotes bone-derived marrow mesenchymal stem cell (BMSC) differentiation into osteoblasts and inhibits differentiation into adipocytes 2 months following ovariectomy (ovariectomized mice, OVX group). (A) Runt-related transcription factor 2 (RUNX2) immunofluorescence in the distal femur was elevated by ISO treatment. (B) Peroxisome proliferator-activated receptor γ (PPAR γ) immunofluorescence in the distal femur was inhibited by ISO. * P <0.05, ** P <0.01 and *** P <0.001 vs. OVX. Columns represent the means \pm SD from 6 mice/group (C and D). Sham, sham-operated.

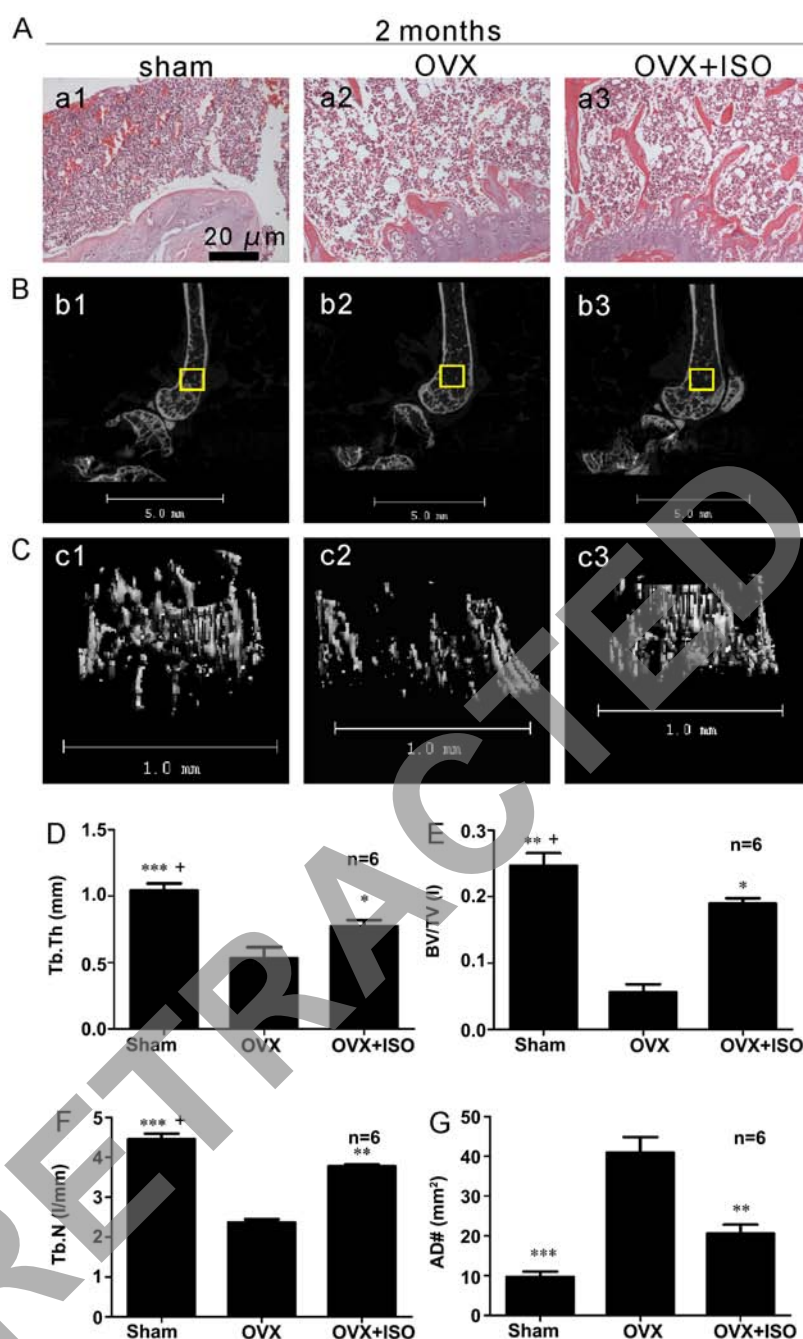


Figure 6. Isopsoralen (ISO) decreases the rate of bone loss following 2 months of treatment. (A) Hematoxylin and eosin staining indicated that ovariectomy (ovariectomized mice, OVX group) increased the number of adipocytes (AD#), whereas ISO decreased the number of adipocytes (G). (B and C) Micro-CT morphometry of murine distal femora shows that the (D) trabecular thickness (Tb.Th), (E) bone volume/total volume (BV/TV), and (F) trabecular number (Tb.N) values in the OVX group were decreased compared to the sham-operated and OVX plus ISO groups. These three indicators were significantly improved by ISO treatment compared to OVX mice. Compared to sham-operated (Sham) group the (D) Tb.Th, (E) BV/TV and (F) Tb.N levels in the OVX + ISO group were also decreased. * $P < 0.05$, ** $P < 0.01$ and *** $P < 0.001$ vs. OVX; * $P < 0.05$ vs. OVX + ISO. Columns represent the means \pm SD from 6 mice/group (D-G).

ysis (Fig. 7). ISO promoted BMSC osteoblast differentiation in a dose-dependent manner, as demonstrated by the upregulation of the osteoblast-specific markers, OCN (Fig. 7A and G) and RUNX2 (Fig. 7A and H). OCN activity was increased in the cells treated with ISO for 2 weeks (Fig. 7A and G). ISO had the most significant effect on OCN expression at a concentration of 20 μ M (Fig. 7G). In addition, the maximum expression of RUNX2 (Fig. 7A and H) was observed at this concentration of ISO, although ISO regulated RUNX2 expression in a dose-dependent manner (Fig. 7H).

PPAR γ and C/EBP β are adipocyte-specific markers. One week after adipogenic induction, PPAR γ (Fig. 7C and K) and C/EBP β (Fig. 7C and L) activity was decreased in the cells treated with ISO in a dose-dependent manner (Fig. 7K and L). We also found that the expression of PPAR γ (Fig. 7E and O) and C/EBP β (Fig. 7E and P) was markedly decreased in the presence of ISO at day 14 in a dose-dependent manner (Fig. 7O and P).

To explore the signaling pathways involved in the regulatory effects of ISO on BMSC adipogenesis, we assessed mammalian target of rapamycin complex 1 (mTORC1) signaling. It

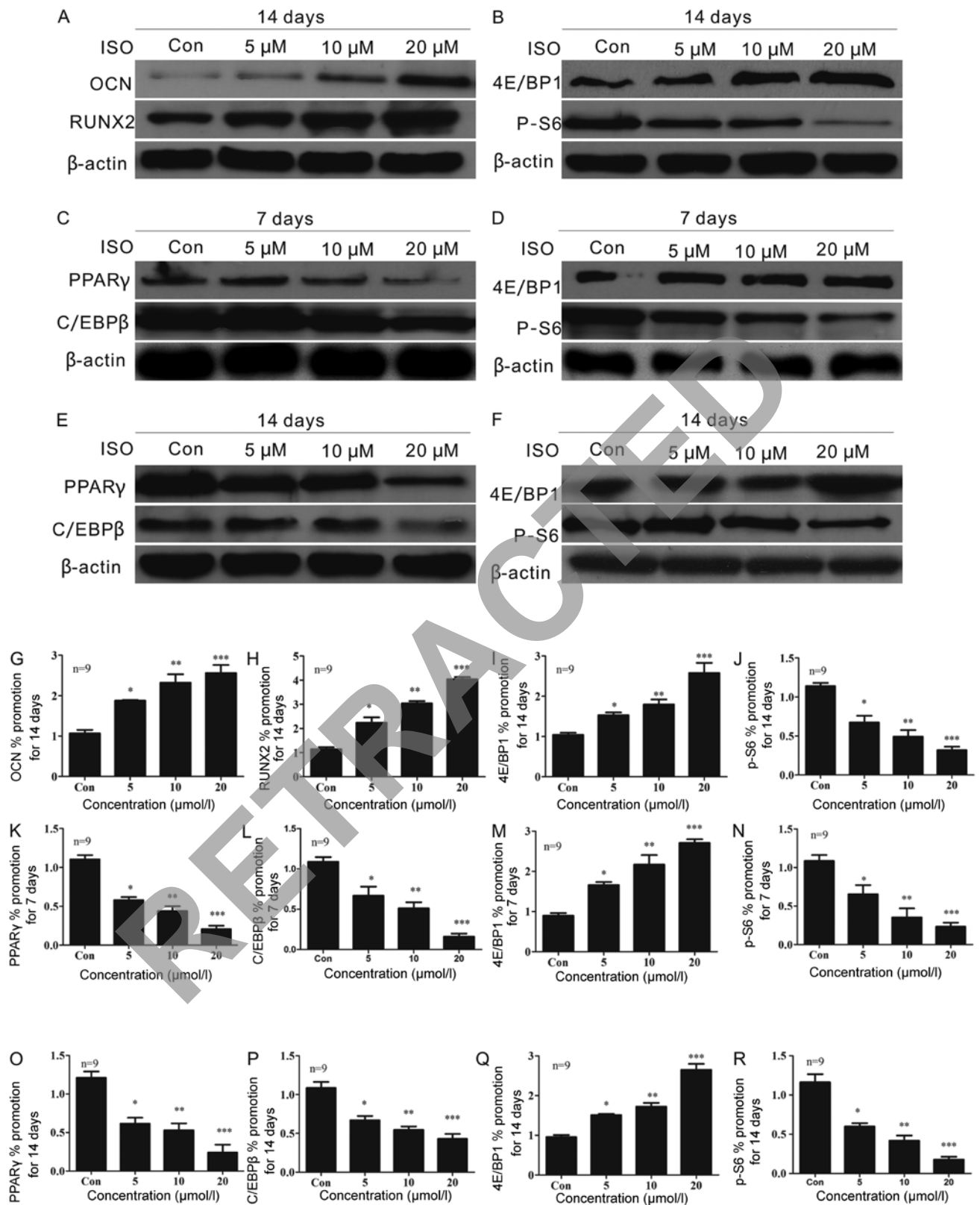


Figure 7. Isoprosalen (ISO) suppresses the adipogenic and promoted osteogenic differentiation of bone-derived marrow mesenchymal stem cells (BMSCs) by inhibiting the activation of the mTORC1 pathway *in vitro*. (A) ISO altered the expression of osteocalcin (OCN) (G) and runt-related transcription factor 2 (RUNX2) (H) during the osteoblastic differentiation of cultured BMSCs for 7 days. Under adipogenic differentiation conditions for 1 week, ISO inhibited peroxisome proliferator-activated receptor γ (PPAR γ) (C and K) and CCAAT/enhancer binding protein β (C/EBP β) expression (C and L). PPAR γ (E and O) and C/EBP β (E and P) expression was markedly decreased in the presence of ISO at day 14 in a dose-dependent manner (O and P). mTORC1 signaling was down-regulated in (B) the osteoblastic differentiation process and (D and F) under adipogenic conditions. (B and I) Eukaryotic translation initiation factor 4E-binding protein 1 (4E/BP1; Thr37/46) expression was upregulated and (B and J) phospho-S6 ribosomal protein (P-S6; S235/236) expression was decreased 2 weeks following osteogenic induction. The expression of (M and Q) 4E/BP1 (Thr37/46) and (N and R) P-S6 (S235/236) in adipogenic medium at (M and N) 7 days or (Q and R) 14 days was similar to that in BMSCs cultured under osteogenic conditions for 7 days. *P<0.05, **P<0.01 and ***P<0.001 compared to the group without ISO (Con, control). Columns represent the means \pm SD from 9 wells/group (G-R).

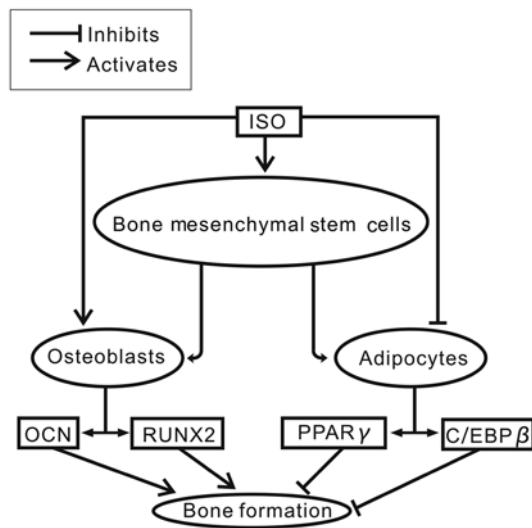


Figure 8. Isopsoralen (ISO) suppresses the adipogenic and promotes the osteogenic differentiation of bone-derived marrow mesenchymal stem cells (BMSCs) via the mTORC1 pathway.

is notable that mTORC1 signaling plays an important role in PPAR γ -mediated adipogenesis (28-30). We demonstrated that the phosphorylation of S6 (S235/236) was inhibited and the phosphorylation of 4E/BP1 (Thr37/46) was promoted by ISO. These proteins are direct downstream effectors of mTORC1, suggesting that ISO may prevent BMSC adipogenesis via the inhibition of mTORC1 signaling. To determine whether the ISO-induced BMSC differentiation was dependent on the inhibition of mTOR signaling (Fig. 7B, D and F), we measured the *in vitro* expression of 4E/BP1 (Thr37/46) (Fig. 7I, M and Q) and P-S6 (S235/236) (Fig. 7J, N and R). We observed an increased 4E/BP1 (Thr37/46) expression (Fig. 7B and I) and a decreased P-S6 (S235/236) expression (Fig. 7B and J) 2 weeks after osteogenic induction. Importantly, similar results were observed 7 days after the induction of adipocyte differentiation (Fig. 7D, M and N). In adipogenic differentiation conditions and in the presence of ISO for 2 weeks, we observed that the expression of 4E/BP1 (Thr37/46) (Fig. 7F and Q) was significantly upregulated and that of P-S6 (S235/236) remained suppressed (Fig. 7F and R). These results were observed in a dose-dependent manner (Fig. 7Q and R).

Discussion

The active components extracted from the seeds of *P. corylifolia* have been widely investigated, including psoralen, which has been shown to exert beneficial effects on skeletal health (20,31,32). Psoralen and ISO are the two main active ingredients of *P.corylifolia* fruit extracts. However, studies evaluating the protective effects of ISO against osteoporosis are limited, particularly studies that investigate its cellular and molecular mechanisms (19,23,33).

In this study, we demonstrated that 2 months of ISO treatment (20 mg/kg/day) increased the bone mass of the distal femoral metaphysis in OVX mice (Tb.Th, BV/TV and Tb.N). The above bone metabolism parameters (BV/TV, Tb.Th and Tb.N) are usually important indicators reflect the level of osteoporosis *in vivo*. Based on the close association between

bone marrow adipogenesis and bone loss during the pathogenesis of osteoporosis, the effects of ISO on bone marrow were evaluated using histological and immunofluorescence analyses. In ISO-treated OVX mice, we observed increased trabecular bone in parallel with reduced adipose tissue in bone marrow. Moreover, an enhanced RUNX2 secretion and decreased PPAR γ secretion were observed.

PPAR γ and C/EBP β are essential transcription factors for adipogenesis (34). Its expression and/or activity determines the commitment of BMSCs into the osteoblasts or adipocyte lineage (35). ALP, RUNX2 and OCN are key regulators of osteoblast differentiation and play an important role in bone formation (36). We used the above parameters as measurements of osteogenesis in the present study.

To the best of our knowledge, this is the first study to demonstrate a role for ISO in the promotion of osteogenesis and the attenuation of adipogenesis *in vivo*. This mechanism of action, which focuses on BMSCs, differs from the mechanisms of currently available agents for osteoporosis that target mature osteoblasts and osteoclasts (37).

We then found that the ISO-enhanced ALP activity is a dose-dependent osteogenic inducer of BMSCs that does not affect cell growth in bone marrow. This result is similar to that of previous studies on psoralen (20,38). As previously demonstrated, in patients with age-related osteoporosis, BMSCs in bone marrow preferentially differentiate into adipocytes rather than osteoblasts (8,39). Accordingly, in this study, BMSCs were cultured under adipogenic conditions, which mimics osteoporosis in humans. Our data indicated that ISO inhibited BMSC adipogenesis in a dose-dependent manner. The pro-osteogenic and anti-adipogenic effects of ISO on BMSCs were further confirmed by evaluating the expression levels of key osteogenic and adipogenic transcription factors. As we had hypothesized, our results indicated that treatment with ISO increased RUNX2 expression. By contrast, PPAR γ expression was reduced by treatment with ISO. These findings were consistent with the data from our *in vitro* experiments.

Based on the observation that decreased numbers of osteoblastic cells occur concomitantly with increased fat content in bone marrow during aging and osteoporosis, studies have suggested a reciprocal association between the adipocyte differentiation and osteoblast differentiation of BMSCs (40-42).

As osteoblasts and adipocytes are both derived from BMSCs (43), we hypothesized that ISO may exert its anti-osteoporotic effects under low estrogen conditions by modulating the RUNX2/PPAR γ balance, thereby inhibiting the differentiation of BMSCs towards adipocytes (7).

There are several limitations to the current study. First, we did not measure the serum lipid levels of mice in our *in vivo* experiments. However, previous studies have demonstrated that compared with other fat types, alterations in serum lipid levels do not significantly affect the role and function of fat in bone marrow (44,45). Additionally, although our results have indicated a protective effect of ISO on osteoporotic bone and bone marrow adipose tissue, we did not include a positive control group, such as strontium and estrogen, which prevent bone loss due to the inhibition of bone marrow adipogenesis (46,47).

In conclusion, the present study demonstrated that ISO attenuated bone marrow adipogenesis, which was indicated by

increased RUNX2 levels and decreased PPAR γ levels. This shifts BMSC lineage differentiation toward osteoblasts rather than adipocytes. Moreover, we suggest that mTORC1 signaling may be the underlying signaling pathway involved in this process (Fig. 8). Our data suggest that the naturally occurring agent, ISO, is safer and more cost-effective for use in the treatment of post-menopausal bone-related diseases.

Acknowledgements

We would like to thank Dr Xin Wang (Guangzhou Huayin Medical Technology Co., Guangdong, China) for providing excellent technical support with the micro-CT analysis. This study was supported by the Science and Technology Department of Hunan Province (grant no. 2014FJ4233) and the Education Department of Hunan Province (grant no. 14C0914).

References

- Schurman L, Bagur A, Claus-Hermberg H, Messina OD, Negri AL, Sánchez A, González C, Diehl M, Rey P, Gamba J, et al: Guías 2012 para el diagnóstico, la prevención y el tratamiento de la osteoporosis. *Medicina (B Aires)* 73: 55-74, 2013 (In Spanish).
- Wang ZQ, Li JL, Sun YL, Yao M, Gao J, Yang Z, Shi Q, Cui XJ and Wang YJ: Chinese herbal medicine for osteoporosis: A systematic review of randomized controlled trials. *Evid Based Complement Alternat Med*: 2013: 356260, 2013.
- Tastekin N and Zateri C: Probable osteosarcoma risk after prolonged teriparatide treatment: comment on the article by Saag et al. *Arthritis Rheum* 62: 1837-1838, 2010.
- Nuttall ME and Gimble JM: Is there a therapeutic opportunity to either prevent or treat osteopenic disorders by inhibiting marrow adipogenesis? *Bone* 27: 177-184, 2000.
- Burkhardt R, Kettner G, Böhm W, Schmidmeier M, Schlag R, Frisch B, Mallmann B, Eisenmenger W and Gilg T: Changes in trabecular bone, hematopoiesis and bone marrow vessels in aplastic anemia, primary osteoporosis, and old age: A comparative histomorphometric study. *Bone* 8: 157-164, 1987.
- Griffith JF, Yeung DK, Antonio GE, Wong SY, Kwok TC, Woo J and Leung PC: Vertebral marrow fat content and diffusion and perfusion indexes in women with varying bone density: MR evaluation. *Radiology* 241: 831-838, 2006.
- Majors AK, Boehm CA, Nitto H, Midura RJ and Muschler GF: Characterization of human bone marrow stromal cells with respect to osteoblastic differentiation. *J Orthop Res* 15: 546-557, 1997.
- Rodríguez JP, Montecinos L, Ríos S, Reyes P and Martínez J: Mesenchymal stem cells from osteoporotic patients produce a type I collagen-deficient extracellular matrix favoring adipogenic differentiation. *J Cell Biochem* 79: 557-565, 2000.
- Sekiya I, Larson BL, Vuoristo JT, Cui JG and Prockop DJ: Adipogenic differentiation of human adult stem cells from bone marrow stroma (MSCs). *J Bone Miner Res* 19: 256-264, 2004.
- Song L and Tuan RS: Transdifferentiation potential of human mesenchymal stem cells derived from bone marrow. *FASEB J* 18: 980-982, 2004.
- Park SR, Oreffo RO and Triffitt JT: Interconversion potential of cloned human marrow adipocytes in vitro. *Bone* 24: 549-554, 1999.
- Beresford JN, Bennett JH, Devlin C, Leboy PS and Owen ME: Evidence for an inverse relationship between the differentiation of adipocytic and osteogenic cells in rat marrow stromal cell cultures. *J Cell Sci* 102: 341-351, 1992.
- Gimble JM: The function of adipocytes in the bone marrow stroma. *New Biol* 2: 304-312, 1990.
- Tavassoli M: Marrow adipose cells and hemopoiesis: An interpretative review. *Exp Hematol* 12: 139-146, 1984.
- Elbaz A, Wu X, Rivas D, Gimble JM and Duque G: Inhibition of fatty acid biosynthesis prevents adipocyte lipotoxicity on human osteoblasts in vitro. *J Cell Mol Med* 14: 982-991, 2010.
- Dong X, Bi L, He S, Meng G, Wei B, Jia S and Liu J: FFAs-ROS-ERK/P38 pathway plays a key role in adipocyte lipotoxicity on osteoblasts in co-culture. *Biochimie* 101: 123-131, 2014.
- Greco EA, Lenzi A and Migliaccio S: The obesity of bone. *Ther Adv Endocrinol Metab* 6: 273-286, 2015.
- Pae HO, Cho H, Oh GS, Kim NY, Song EK, Kim YC, Yun YG, Kang CL, Kim JD, Kim JM and Chung HT: Bakuchiol from *Psoralea corylifolia* inhibits the expression of inducible nitric oxide synthase gene via the inactivation of nuclear transcription factor-kappaB in RAW 264.7 macrophages. *Int Immunopharmacol* 1: 1849-1855, 2001.
- Lu H, Zhang L, Liu D, Tang P and Song F: Isolation and purification of psoralen and isopsoralen and their efficacy and safety in the treatment of osteosarcoma in nude rats. *Afr Health Sci* 14: 641-647, 2014.
- Tang DZ, Yang F, Yang Z, Huang J, Shi Q, Chen D and Wang YJ: Psoralen stimulates osteoblast differentiation through activation of BMP signaling. *Biochem Biophys Res Commun* 405: 256-261, 2011.
- Wong RW and Rabie AB: Effect of psoralen on bone formation. *J Orthop Res* 29: 158-164, 2011.
- Tsai MH, Huang GS, Hung YC, Bin L, Liao LT and Lin LW: *Psoralea corylifolia* extract ameliorates experimental osteoporosis in ovariectomized rats. *Am J Chin Med* 35: 669-680, 2007.
- Ming L, Ge B, Chen K, Ma H and Zhai Y: Effects of isopsoralen on bone marrow stromal stem cells differentiate and proliferate in vitro. *Zhongguo Zhong Yao Za Zhi* 36: 2124-2128, 2011 (In Chinese).
- Ming LG, Cheng KM, Ge BF, Ma HP and Zai YK: Effect of isopsoralen on the proliferation and differentiation of osteoblasts in vitro. *Zhong Yao Cai* 34: 404-408, 2011 (In Chinese).
- Lee SK, Kalinowski JF, Jacquin C, Adams DJ, Gronowicz G and Lorenzo JA: Interleukin-7 influences osteoclast function in vivo but is not a critical factor in ovariectomy-induced bone loss. *J Bone Miner Res* 21: 695-702, 2006.
- Bouxsein ML, Boyd SK, Christiansen BA, Guldberg RE, Jepsen KJ and Muller R: Guidelines for assessment of bone microstructure in rodents using micro-computed tomography. *J Bone Miner Res* 25: 1468-1486, 2010.
- Rossini M, Gatti D and Adami S: Involvement of WNT/ β -catenin signaling in the treatment of osteoporosis. *Calcif Tissue Int* 93: 121-132, 2013.
- Cybulski N, Polak P, Auwerx J, Rüegg MA and Hall MN: mTOR complex 2 in adipose tissue negatively controls whole-body growth. *Proc Natl Acad Sci USA* 106: 9902-9907, 2009.
- Xiang X, Yuan F, Zhao J, Li Z, Wang X, Guan Y, Tang C, Sun G, Li Y and Zhang W: Deficiency in pulmonary surfactant proteins in mice with fatty acid binding protein 4-Cre-mediated knockout of the tuberous sclerosis complex 1 gene. *Exp Physiol* 98: 830-841, 2013.
- Polak P, Cybulski N, Feige JN, Auwerx J, Rüegg MA and Hall MN: Adipose-specific knockout of raptor results in lean mice with enhanced mitochondrial respiration. *Cell Metab* 8: 399-410, 2008.
- Lim SH, Ha TY, Kim SR, Ahn J, Park HJ and Kim S: Ethanol extract of *Psoralea corylifolia* L. and its main constituent, bakuchiol, reduce bone loss in ovariectomized Sprague-Dawley rats. *Br J Nutr* 101: 1031-1039, 2009.
- Wang D, Li F and Jiang Z: Osteoblastic proliferation stimulating activity of *Psoralea corylifolia* extracts and two of its flavonoids. *Planta Med* 67: 748-749, 2001.
- Yuan X, Bi Y, Yan Z, Pu W, Li Y and Zhou K: Psoralen and isopsoralen ameliorate sex hormone deficiency-induced osteoporosis in female and male mice. *Biomed Res Int* 2016: 6869452, 2016.
- Wood RJ: Vitamin D and adipogenesis: New molecular insights. *Nutr Rev* 66: 40-46, 2008.
- Moerman EJ, Teng K, Lipschitz DA and Lecka-Czernik B: Aging activates adipogenic and suppresses osteogenic programs in mesenchymal marrow stroma/stem cells: the role of PPAR-gamma2 transcription factor and TGF-beta/BMP signaling pathways. *Aging Cell* 3: 379-389, 2004.
- Yoshida CA, Furuichi T, Fujita T, Fukuyama R, Kanatani N, Kobayashi S, Satake M, Takada K and Komori T: Core-binding factor beta interacts with Runx2 and is required for skeletal development. *Nat Genet* 32: 633-638, 2002.
- Peng S, Zhang G, He Y, Wang X, Leung P, Leung K and Qin L: Epimedium-derived flavonoids promote osteoblastogenesis and suppress adipogenesis in bone marrow stromal cells while exerting an anabolic effect on osteoporotic bone. *Bone* 45: 534-544, 2009.

38. Yang Z, Huang JH, Liu SF, Zhao YJ, Shen ZY, Wang YJ and Bian Q: The osteoprotective effect of psoralen in ovariectomy-induced osteoporotic rats via stimulating the osteoblastic differentiation from bone mesenchymal stem cells. *Menopause* 19: 1156-1164, 2012.
39. Duque G: Bone and fat connection in aging bone. *Curr Opin Rheumatol* 20: 429-434, 2008.
40. Chan GK and Duque G: Age-related bone loss: Old bone, new facts. *Gerontology* 48: 62-71, 2002.
41. Verma S, Rajaratnam JH, Denton J, Hoyland JA and Byers RJ: Adipocytic proportion of bone marrow is inversely related to bone formation in osteoporosis. *J Clin Pathol* 55: 693-698, 2005.
42. Rodríguez JP, Garat S, Gajardo H, Pino AM and Seitz G: Abnormal osteogenesis in osteoporotic patients is reflected by altered mesenchymal stem cells dynamics. *J Cell Biochem* 75: 414-423, 1999.
43. Owen M: Marrow stromal stem cells. *J Cell Sci Suppl* 10: 63-76, 1988.
44. Gasparrini M, Rivas D, Elbaz A and Duque G: Differential expression of cytokines in subcutaneous and marrow fat of aging C57BL/6J mice. *Exp Gerontol* 44: 613-618, 2009.
45. Ecklund K, Vajapeyam S, Feldman HA, Buzney CD, Mulkern RV, Kleinman PK, Rosen CJ, Gordon CM: Bone marrow changes in adolescent girls with anorexia nervosa. *J Bone Miner Res* 25: 298-304, 2010.
46. Fournier C, Perrier A, Thomas M, Laroche N, Dumas V, Rattner A, Vico L and Guignandon A: Reduction by strontium of the bone marrow adiposity in mice and repression of the adipogenic commitment of multipotent C3H10T1/2 cells. *Bone* 50: 499-509, 2012.
47. Syed FA, Oursler MJ, Hefferanm TE, Peterson JM, Riggs BL and Khosla S: Effects of estrogen therapy on bone marrow adipocytes in postmenopausal osteoporotic women. *Osteoporos Int* 9: 1323-1330, 2008.

RETRACTED