

Geraniin protects bone marrow-derived mesenchymal stem cells against hydrogen peroxide-induced cellular oxidative stress *in vitro*

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Abstract. Administration of bone marrow-derived mesenchymal stem cells (MSCs) has emerged as a potential therapeutic approach for the treatment of myocardial infarction (MI). However, the increase in reactive oxygen species (ROS) in ischemic cardiac tissue compromises the survival of transplanted MSCs, thus resulting in limited therapeutic efficiency. Therefore, strategies that attenuate oxidative stress-induced damage and enhance MSC viability are required. Geraniin has been reported to possess potent antioxidative activity and exert protective effects on numerous cell types under certain conditions. Therefore, geraniin may be considered a potential drug used to modulate MSC-based therapy for MI. In the present study, MSCs were pretreated with geraniin for 24 h and were exposed to hydrogen peroxide (H₂O₂) for 4 h. Cell apoptosis, intracellular ROS levels and mitochondrial membrane potential were measured using Annexin V-fluorescein isothiocyanate/propidium iodide staining, the 2',7'-dichlorodihydrofluorescein diacetate fluorescent probe and the membrane permeable dye JC-1, respectively. Glutathione and malondialdehyde levels were also investigated. The expression levels of apoptosis-associated proteins and proteins of the phosphoinositide 3-kinase (PI3K)/protein kinase B (Akt) signaling pathway were analyzed by western blotting. The results demonstrated that geraniin could significantly attenuate H₂O₂-induced cell damage by promoting MSC survival, reducing cellular ROS production and maintaining mitochondrial function. Furthermore, geraniin modulated the expression levels of phosphorylated-Akt in a time- and dose-dependent manner. The cytoprotective

effects of geraniin were suppressed by LY294002, a specific PI3K inhibitor. In conclusion, the present study revealed that geraniin protects MSCs from H₂O₂-induced oxidative stress injury via the PI3K/Akt pathway. These findings indicated that cotreatment of MSCs with geraniin may optimize therapeutic efficacy for the clinical treatment of MI.

Introduction

Myocardial infarction (MI) is among the main causes of mortality worldwide (1). Irreversible loss of cardiac myocytes and concomitant cicatrization are induced by MI; therefore, patients exhibit poor cardiac pump function and congestive heart failure. Cell-based therapy for MI represents an emerging strategy in biological therapeutics (2,3). As one of the most frequently investigated cellular populations, bone marrow-derived mesenchymal stem cells (MSCs) are particularly attractive therapeutic candidates. MSC-based therapy relies on the self-renewal capability of MSCs, and the ability of MSCs to differentiate into cardiovascular cells and secrete multitudinous bioactive molecules. These actions subsequently activate endogenous neovascularization, immunomodulation and cardiac regeneration, thus resulting in restoration of cardiac function (4). However, current evidence indicates that poor viability of engrafted MSCs in the infarcted myocardium is a primary limitation of the therapeutic efficacy of MSCs (5,6). Reactive oxygen species (ROS), including hydrogen peroxide (H₂O₂), superoxide radicals and hydroxyl radicals, are produced during infarction (7) or reperfusion of ischemic hearts (8). ROS may lead to impaired cell metabolism and decreased cell viability, thus inhibiting transplanted MSCs from taking effect. Therefore, protecting MSCs from apoptosis, together with enhancing their ability to survive under oxidative stress, is crucial for optimizing MSC-based therapy.

Polyphenols, or polyphenolic compounds, are widely distributed in natural plants, and range from simple structures, such as flavonoids, to highly complex polymeric substances, including proanthocyanidins and ellagitannins (9). Due to their various biological activities, polyphenols exhibit potential as effective therapeutic drugs. In addition, they have

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been demonstrated to display numerous pharmacological activities, including anticarcinogenic (10), antibacterial (11) and antidiabetic (12) effects. Furthermore, polyphenols are strong antioxidants, due to their free radical-scavenging activities (13). Geraniin is a typical ellagitannin, which has been identified as the major active compound extracted from *Geranium sibiricum*. A previous study reported that geraniin possesses marked nitric oxide-scavenging, superoxide radical-scavenging and β -carotene-linoleic acid-bleaching properties due to its unique chemical structure (Fig. 1A) (14). Furthermore, geraniin has been confirmed to protect liver cells against ethanol-induced cytotoxicity (15) and inhibit apoptosis of pulmonary fibroblasts under γ -radiation conditions (16). Our previous study demonstrated that geraniin may exert strong ROS-scavenging activities when preventing THP-1 macrophages from switching to an M1 phenotype under lipopolysaccharide stimulation (17). Since H_2O_2 is often used *in vitro* to simulate the oxidative stress microenvironment detected in ischemic heart tissue, the present study hypothesized that geraniin may defend MSCs against H_2O_2 -induced damage. The present study aimed to investigate the cytoprotective effects of geraniin on MSCs against H_2O_2 -induced cellular injury, as well as the underlying mechanism.

Materials and methods

Materials. Geraniin (purity $\geq 98\%$) was purchased from Shanghai Yuanye Bio-Technology Co., Ltd. (Shanghai, China), and was dissolved in dimethyl sulfoxide (DMSO) and stored at -80°C . The stock concentration of geraniin was 10 mM. DMSO and H_2O_2 were purchased from Sigma-Aldrich (Merck KGaA, Darmstadt, Germany). Dulbecco's modified Eagle's medium (DMEM)/F12 and fetal bovine serum (FBS) were obtained from Gibco (Thermo Fisher Scientific, Inc., Waltham, MA, USA). The conjugated antibodies used to identify MSCs: Fluorescein isothiocyanate (FITC)-labeled anti-CD29 (555005) and anti-CD44 (561859), and phycoerythrin-labeled anti-CD45 (553091) and anti-CD90 (551401), as well as the Annexin V-FITC/propidium iodide (PI) apoptosis detection kit, were all purchased from BD Biosciences (Franklin Lakes, NJ, USA). Cell Counting kit-8 (CCK-8) was from Dojindo Molecular Technologies, Inc. (Kumamoto, Japan). Hoechst 33342, JC-1 dye, 2,7'-dichlorodihydrofluorescein diacetate (DCFH-DA) fluorescent probe, glutathione (GSH) kit (S0053), malondialdehyde (MDA) kit (S0131), ROS scavenger N-acetyl-L-cysteine (NAC), cell mitochondrial protein isolation kit (C3601), radioimmunoprecipitation assay (RIPA) lysis buffer, bicinchoninic acid (BCA) protein assay kit and mouse polyclonal anti- β -actin (AA128) were all purchased from Beyotime Institute of Biotechnology (Beijing, China). Rabbit antibodies against phosphorylated-protein kinase B [p-Akt (Ser473); 4060s], Akt (9272s), caspase-3 (9662s), B-cell lymphoma 2 (Bcl-2; 2876s), Bcl-2-associated X protein (Bax; 2772s) and cytochrome *c* (Cyt C; 4272s), and LY294002 [phosphoinositide 3-kinase (PI3K) specific inhibitor], were all purchased from Cell Signaling Technology, Inc. (Danvers, MA, USA). Horseradish peroxidase (HRP)-conjugated anti-rabbit (ZB-2301) and anti-mouse (ZB-2305) secondary antibodies were obtained from OriGene Technologies, Inc., (Beijing, China).

MSCs isolation and culture. MSCs were isolated and harvested from male Sprague-Dawley rats (age, 3 weeks old; weight, 60–80 g), as previously described with minor modifications (18). A total of 10 SPF Sprague-Dawley rats were purchased from the Laboratory Animal Science Department of the Second Affiliated Hospital of Harbin Medical University (Harbin, China). They were kept under standard animal housing conditions (temperature, $21\pm 1^\circ\text{C}$; humidity, $55\pm 5\%$), at a 12-h dark/light cycle and had access to unlimited food and water. The present study was approved by the Local Ethics Committee on Animal Care and Use of Harbin Medical University. Briefly, total bone marrow was flushed from the tibias and femurs of the rats with 10 ml DMEM/F12 using a sterile syringe. After centrifugation at $300 \times g$ for 5 min, the remaining pellets were resuspended in 5 ml DMEM/F12 supplemented with 10% FBS and 1% penicillin/streptomycin, and were then seeded into cell culture flasks at 37°C in an atmosphere containing 5% CO_2 . Following incubation for 48 h, the culture medium and non-adherent cells were discarded and fresh medium was added. The medium was replaced every 72 h thereafter. Cells were cultured until they reached 80% confluence, after which they were passaged. Cells were split using 0.25% trypsin and were expanded at a 1:2 or 1:3 dilution. Cells from passages 3–5 were used in the subsequent experiments. The MSC population was characterized according to positive (CD44, CD29 and CD90) and negative (CD45) cell surface markers by flow cytometry, as reported in our previous studies (18,19).

Cell treatments. All treatments were conducted at 37°C in the incubator. MSCs were seeded into six-well plates or a 25 cm^2 culture flask. Once cell density reached 60–70%, H_2O_2 (100, 200, 300, 400 and $500\text{ }\mu\text{M}$), mixed with serum-free DMEM/F12 for 4 h, was used to establish an *in vitro* oxidative stress model. Geraniin, at 1, 5, 10 and $20\text{ }\mu\text{M}$, was separately preincubated in DMEM/F12 for 24 h. The inhibitor of PI3K, LY294002 ($25\text{ }\mu\text{M}$), or the ROS scavenger, NAC ($500\text{ }\mu\text{M}$), was added 1 h prior to H_2O_2 treatment without geraniin co-treatment. Cells cultured in complete medium without any specific treatment comprised the control group.

Cell viability assay. MSC viability was assessed using the CCK-8 assay. Briefly, cells were plated into 96-well plates (3×10^3 cells/well). Following cell adhesion to the plates, appropriate treatments were administered. Subsequently, the medium was removed and replaced with $100\text{ }\mu\text{l}$ fresh DMEM/F12 and $10\text{ }\mu\text{l}$ CCK-8 solution in each well. The plates were maintained at 37°C for 1 h. Finally, absorbance was detected at 450 nm using a Tecan Infinite 200 PRO microplate reader (Tecan Austria GmbH, Grödig, Austria).

Measurement of cell apoptosis. Apoptosis of MSCs was determined using the Annexin V-FITC/PI staining method. Following treatment, the cells were harvested, washed with ice-cold PBS and resuspended in $400\text{ }\mu\text{l}$ binding buffer. The cell suspension was then incubated with $5\text{ }\mu\text{l}$ Annexin V solution for 15 min at room temperature in the dark, followed by incubation with $5\text{ }\mu\text{l}$ PI for an additional 5 min. The cells were immediately analyzed by flow cytometry using BD FACSCanto II (BD Biosciences). Approximately 1×10^5 cells were detected in each sample. According to the

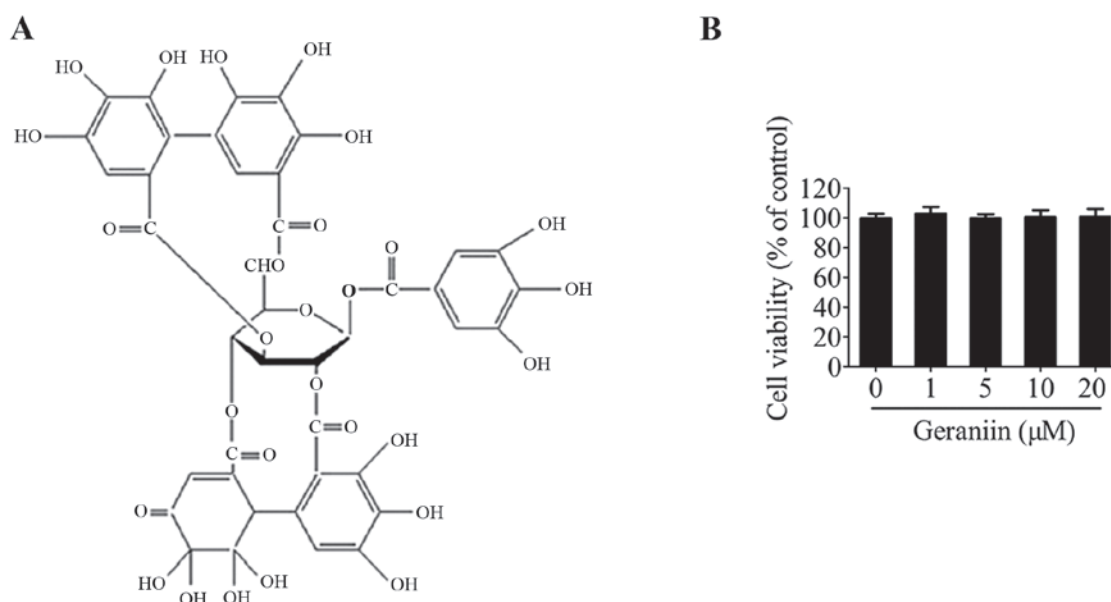


Figure 1. Chemical structure of geraniin and effects of geraniin on MSC viability. (A) Chemical structure of geraniin. (B) MSCs were treated with various concentrations of geraniin for 24 h, and viability was measured using the Cell Counting kit-8 assay. Data are presented as the mean \pm standard deviation from five replicate wells and are representative of three independent experiments. MSCs, mesenchymal stem cells.

reaction principles, Annexin V/PI⁻ staining signified viable cells, Annexin V⁺/PI⁻ indicated early apoptotic cells, and Annexin V⁺/PI⁺ represented late apoptotic or necrotic cells.

Assessment of morphological alterations. MSCs were treated with geraniin and H₂O₂ in six-well plates. Hoechst 33342 was used to detect cell nuclear condensation and fragmentation. After fixing in 4% paraformaldehyde for 15 min at room temperature, cells were washed twice with PBS, stained with 5 μg/ml Hoechst 33342 for 5 min and washed a further two times with PBS. Finally, cells were assessed by fluorescence microscopy. Apoptotic cells were identified by condensed or fragmented nuclei.

ROS, GSH and MDA assays. Intracellular ROS levels were determined using a ROS assay kit. Briefly, cells were incubated with the diluted fluorescent probe, DCFH-DA, for 20 min at 37°C. Cells were then washed three times with serum-free DMEM/F12, collected and analyzed using a flow cytometer. Due to the important roles of GSH and MDA in ROS-associated oxidative stress, their concentrations were also measured using commercial kits according to the manufacturer's protocols.

Detection of mitochondrial membrane potential (Ψ_m). JC-1 was used to measure alterations in Ψ_m . Briefly, cells were washed with PBS, stained with 5 μM JC-1 and maintained for 20 min at 37°C. Subsequently, cells were washed twice with ice-cold JC-1 staining buffer and were then directly observed under a fluorescence microscope, or were collected and analyzed by flow cytometry.

Protein extraction and western blot analysis. Cells were washed with ice-cold PBS, lysed with RIPA lysis buffer and centrifuged at 12,000 \times g for 15 min at 4°C. The extraction of cytoplasmic and mitochondrial proteins was conducted in accordance with the manufacturer's protocol. BCA protein assay was used to quantify

protein concentrations. Equal amounts of total protein (50 μg/lane) were separated by 8-12% SDS-PAGE and were transferred onto polyvinylidene fluoride membranes. The membranes were blocked with 5% skim milk diluted in Tris-buffered saline containing 0.05% Tween-20 (TBST) at 37°C for 1 h, and were then incubated with diluted primary antibodies against p-Akt (S473) (1:1,000), total-Akt (1:1,000), cleaved caspase-3 (1:1,000), Bax (1:1,000), Bcl-2 (1:1,000), Cyt C (1:1,000) and β -actin (1:800) overnight at 4°C. After washing three times with TBST, membranes were incubated with the corresponding HRP-conjugated secondary antibodies (1:5,000) for 1 h at room temperature. The images of the immune complexes were developed by ECL in the dark, and images were captured using a Tanon-5200 (Tanon Science and Technology Co., Ltd., Shanghai, China). Band density was determined using ImageJ (1.48u; National Institutes of Health, Bethesda, MD, USA).

Statistical analysis. All data are presented as the mean \pm standard deviation and were analyzed using SPSS 19.0 software (IBM Corp., Armonk, NY, USA). Differences between groups were analyzed by one-way ANOVA with a Tukey's post hoc test. $P < 0.05$ was considered to indicate a statistically significant difference.

Results

Effects of geraniin on MSC viability. Initially, the effects of geraniin on MSC viability were examined using the CCK-8 assay. Treatment with geraniin for 24 h had little influence on cell viability compared with the 0 μM geraniin group (Fig. 1B). This result indicated that geraniin did not exert toxic effects on MSCs.

Geraniin significantly inhibits H₂O₂-induced apoptosis of MSCs. H₂O₂ has been reported to induce apoptosis of MSCs at various concentrations and time-points (20,21). To establish a successful *in vitro* oxidative stress model, the present

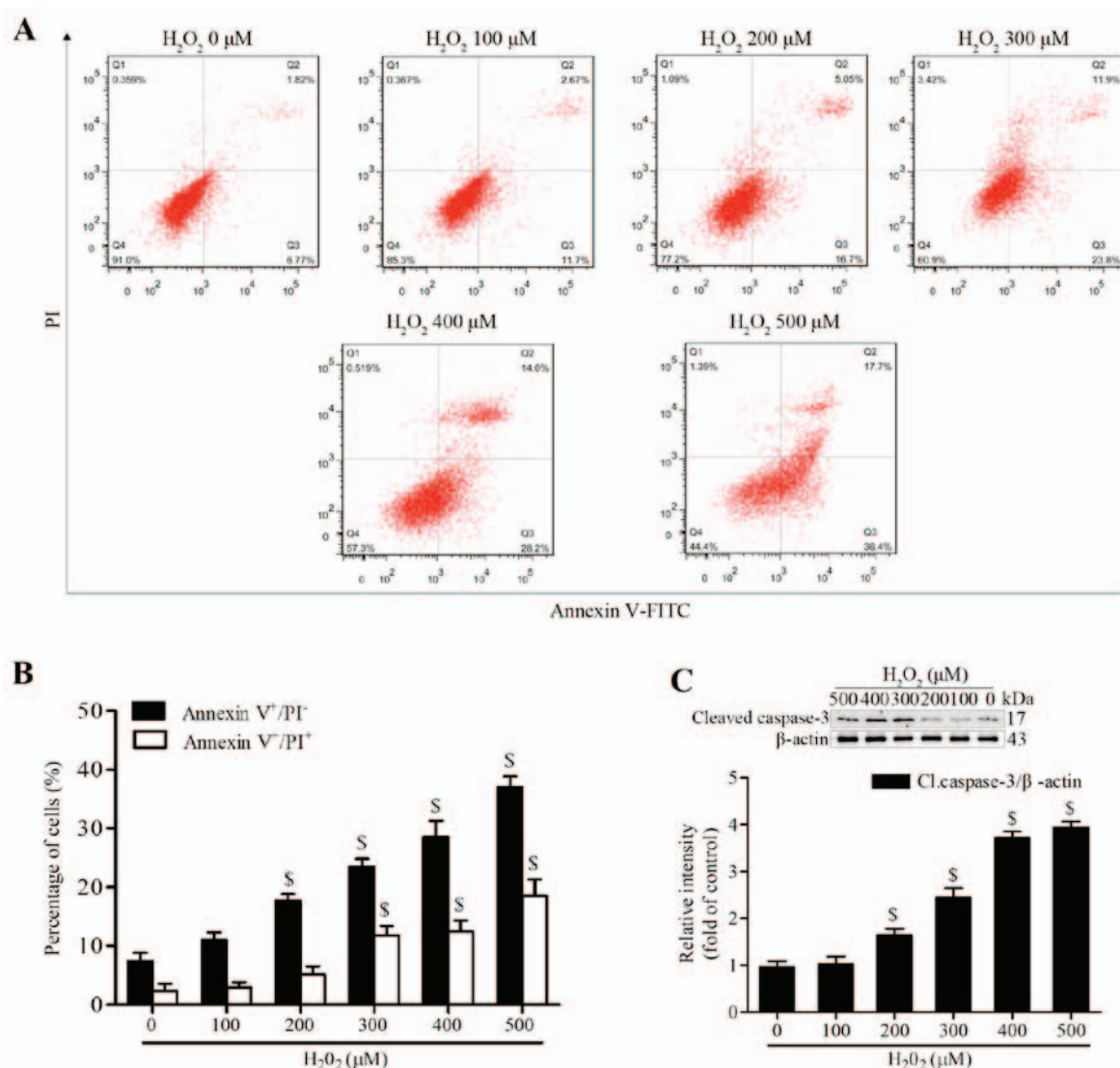


Figure 2. H₂O₂ induces apoptosis of MSCs. (A and B) Cells were treated with a range of H₂O₂ concentrations for 4 h, and apoptosis was analyzed by Annexin V-FITC/PI staining assay. (C) H₂O₂-induced cleaved caspase-3 expression was measured by western blotting, β -actin was used for normalization. Data are presented as the mean \pm standard deviation of three independent measurements. * $P < 0.05$ compared with the 0 μ M H₂O₂ group. Cl.caspase-3, cleaved caspase-3; FITC, fluorescein isothiocyanate; H₂O₂, hydrogen peroxide; MSCs, mesenchymal stem cells; PI, propidium iodide.

study investigated the proapoptotic effects of H₂O₂ on MSCs. Following treatment with various concentrations of H₂O₂ (100–500 μ M) in serum-free medium for 4 h, MSCs were stained with Annexin V-FITC/PI. The proportion of early apoptotic cells (Annexin V⁺/PI⁻) was significantly increased following treatment with ≥ 200 μ M H₂O₂, whereas the proportion of late apoptotic or necrotic MSCs (Annexin V⁺/PI⁺) was markedly increased following treatment with ≥ 300 μ M (Fig. 2A and B). In addition, the expression levels of cleaved caspase-3 were significantly increased at 200 μ M and continued to increase to 500 μ M (Fig. 2C). We then selected treatment with H₂O₂ at 300 μ M for 4 h as the condition to induce effective apoptosis, since this concentration generated moderate apoptotic cells.

To determine whether geraniin rescues MSCs from H₂O₂-induced apoptosis, MSCs were pretreated with increasing concentrations of geraniin (1, 5, 10 and 20 μ M) for 24 h. MSCs were then co-treated with geraniin and H₂O₂. A significant reversal in the percentage of H₂O₂-induced Annexin V⁺/PI⁺ cells, in response to geraniin, was observed by flow cytometry

(geraniin 1 μ M, 20.53 \pm 1.25%; 5 μ M, 18.67 \pm 1.89%; 10 μ M, 11.57 \pm 1.01%; 20 μ M, 7.23 \pm 0.31%; $P < 0.05$ vs. H₂O₂, 23.83 \pm 1.32%) (Fig. 3A and B). However, 1 μ M geraniin had no effect on the proportion of Annexin V⁺/PI⁺ cells, whereas the other concentrations significantly reduced the percentage of late apoptotic or necrotic cells (geraniin 5 μ M, 7.88 \pm 2.58%; 10 μ M, 7.05 \pm 1.72%; 20 μ M, 4.40 \pm 0.48%; $P < 0.05$ vs. H₂O₂, 12.97 \pm 1.65%) (Fig. 3A and B). Furthermore, in apoptotic cells, nuclei become condensed or fragmented, whereas in normal cells, nuclei are circular or oval. Hoechst 33342 staining was used to confirm the presence of nuclear morphological alterations. Cells treated with H₂O₂ appeared to possess shrunken and fragmented nuclei; however, those pretreated with geraniin exhibited marked amelioration of H₂O₂-induced nuclear impairment. These data indicated that geraniin may effectively attenuate H₂O₂-induced MSC apoptosis (Fig. 3C).

Geraniin exerts protective effects by regulating ROS generation. Since ROS serves a pivotal role in proapoptotic signaling

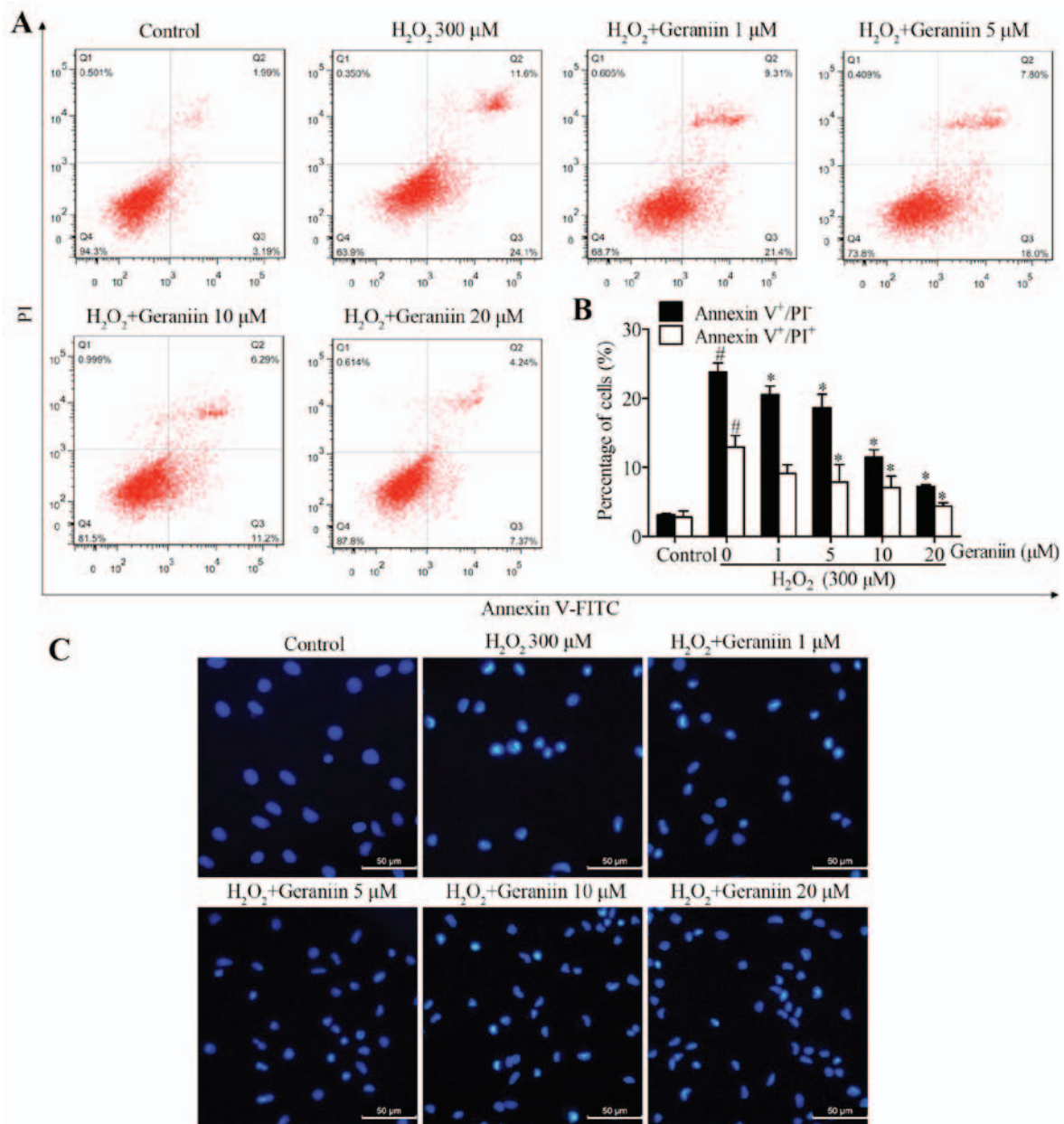


Figure 3. Geraniin reduces H₂O₂-induced MSC apoptosis. MSCs were treated with increasing concentrations of geraniin for 24 h and were then exposed to 300 μM H₂O₂ for 4 h. Apoptosis was reduced by geraniin in a dose-dependent manner, as identified by (A and B) flow cytometry and (C) Hoechst 33342 staining (original magnification, 200x; scale bar, 50 μm). *P<0.05 compared with the control group; #P<0.05 compared with the 300 μM H₂O₂ group. FITC, fluorescein isothiocyanate; H₂O₂, hydrogen peroxide; MSCs, mesenchymal stem cells PI, propidium iodide.

cascades (22), the present study examined the effects of geraniin on ROS generation using flow cytometry. The results demonstrated that H₂O₂ induced a 7.6-fold increase in ROS production compared with the control group. However, pretreatment with geraniin markedly suppressed ROS generation in a concentration-dependent manner (Fig. 4A and B). Following treatment with NAC, a general ROS scavenger, similar results were recorded compared with 20 μM geraniin (mean fluorescence intensity: Geraniin 20 μM, 476.33±46.65; NAC, 443.80±53.15, P>0.05) (Fig. 4B). Furthermore, alterations in intracellular GSH and MDA contents were investigated; cells pretreated with geraniin had significantly increased GSH levels, whereas MDA production was suppressed by geraniin (Fig. 4C and D). These findings indicated that geraniin is able to enhance the cellular antioxidant system and remove redundant ROS.

Geraniin protects MSCs against oxidative stress through stabilizing the Ψ_m. Mitochondria in eukaryotic cells are the primary components of respiration, and are critical in the defense against oxidative stress-induced damage (23). Maintaining the Ψ_m is essential to ensure the scavenging efficiency of ROS, and to prevent cell apoptosis or other stress-associated events induced by excessive ROS (24). JC-1 is a Ψ_m-sensitive dye, which aggregates in the mitochondrial matrix and exhibits red fluorescence in normal cells. However, when the Ψ_m is reduced, JC-1 is converted to its monomer state, which exhibits green fluorescence. H₂O₂ resulted in a marked reduction in Ψ_m within MSCs, whereas geraniin markedly upregulated the Ψ_m, as identified by fluorescence microscopy (Fig. 5A). In addition, the ratio of red/green fluorescence intensity was significantly downregulated under H₂O₂

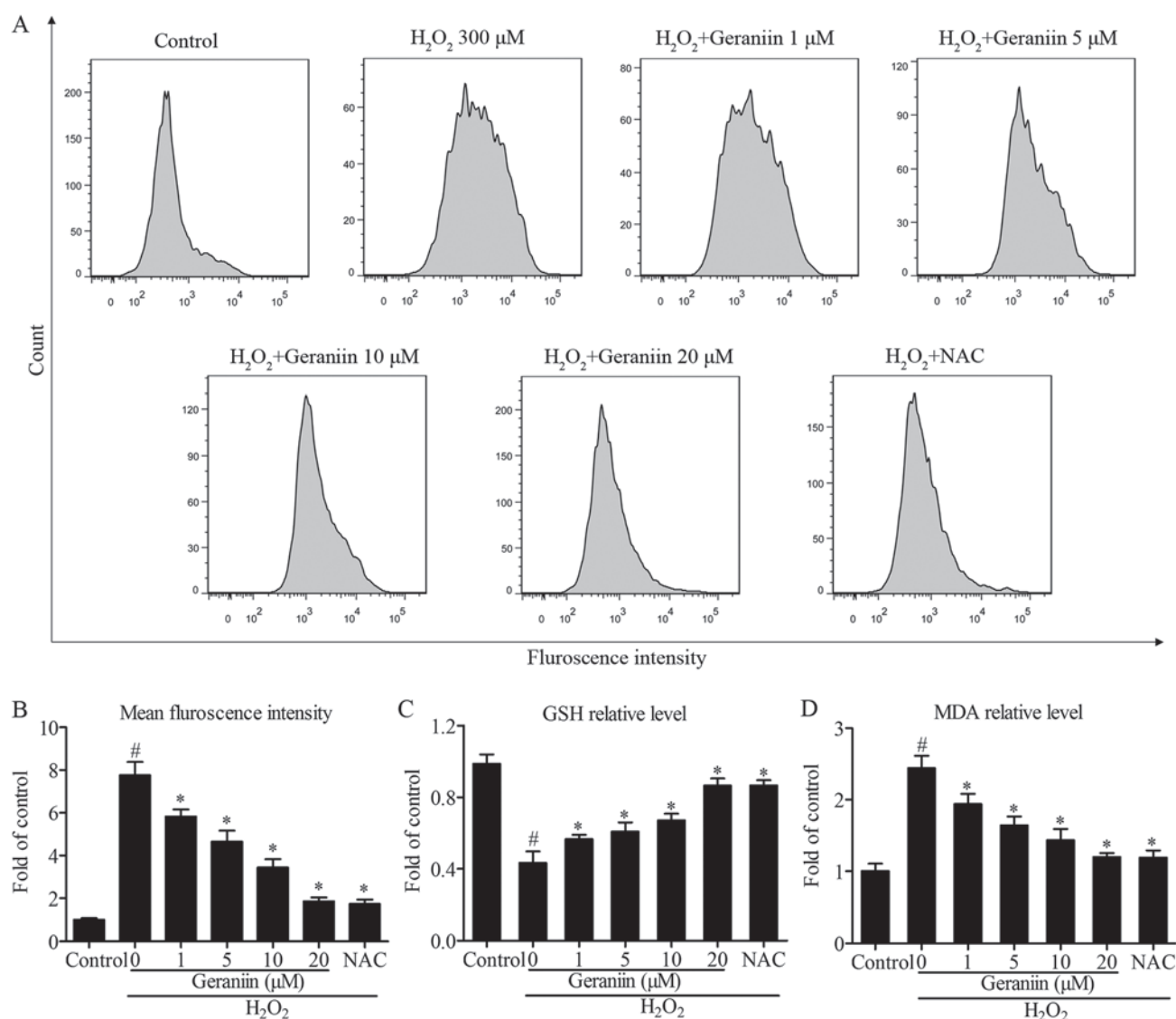


Figure 4. Geraniin exerts protective effects by regulating ROS generation. MSCs were pretreated with the indicated concentrations of geraniin for 24 h or NAC for 1 h, followed by exposure to H₂O₂ for 4 h. (A) Intracellular ROS levels were evaluated by flow cytometry and (B) data were quantified. Alterations in (C) GSH and (D) MDA levels were measured. Data are presented as the mean \pm standard deviation of three independent experiments. [#]P<0.05 compared with the control group; ^{*}P<0.05 compared with the H₂O₂-treated group. GSH, glutathione; H₂O₂, hydrogen peroxide; MDA, malondialdehyde; MSCs, mesenchymal stem cells; NAC, N-acetyl-L-cysteine; ROS, reactive oxygen species.

exposure compared with in the control group; however, this effect was reversed by geraniin in a concentration-dependent manner (Fig. 5B and C). These findings suggested that geraniin may exert beneficial effects on mitochondrial function.

PI3K/Akt signaling is required for geraniin to exert anti-apoptotic effects on MSCs. It has previously been reported that geraniin exerts cytoprotective effects on HepG2 cells via activation of the extracellular signal-regulated kinase 1/2 and PI3K/Akt pathways (25). Due to the importance of the PI3K/Akt pathway on classical survival signals (26), the present study aimed to determine the association between geraniin and the PI3K/Akt pathway in MSCs. Cells were treated with geraniin (20 μM) for the indicated periods of time; the protein expression levels of p-Akt (Ser473) were transiently upregulated at 15 min and peaked at 60 min, prior to subsequent downregulation (Fig. 6A). In addition, the effects of 1 h treatment with various concentrations of geraniin on p-Akt (Ser473)

expression in MSCs was investigated; p-Akt expression was upregulated in a dose-dependent manner. Conversely, p-Akt expression was markedly inhibited following pretreatment with LY294002, a PI3K-specific inhibitor (Fig. 6B). These findings indicated that geraniin may activate the PI3K/Akt signaling pathway in a time- and dose-dependent manner.

To gain further insight into the role of the PI3K/Akt pathway in the protective effects of geraniin on MSCs, cells were preconditioned with LY294002 and were then exposed to 20 μM geraniin and H₂O₂. PI3K inhibition significantly attenuated the anti-apoptotic effects of geraniin on MSCs under H₂O₂ treatment, as evidenced by a decrease in cell survival rate using CCK-8 assay (Fig. 6C). Since reductions in the Ψ_m and increased cleaved caspase-3 expression are initiating and amplifying factors of the mitochondrial apoptosis pathway, it may be suggested that H₂O₂ induces apoptosis of MSCs through regulating the mitochondrial apoptosis pathway. Therefore, the expression levels of Cyt C, cleaved caspase-3, Bax and Bcl-2

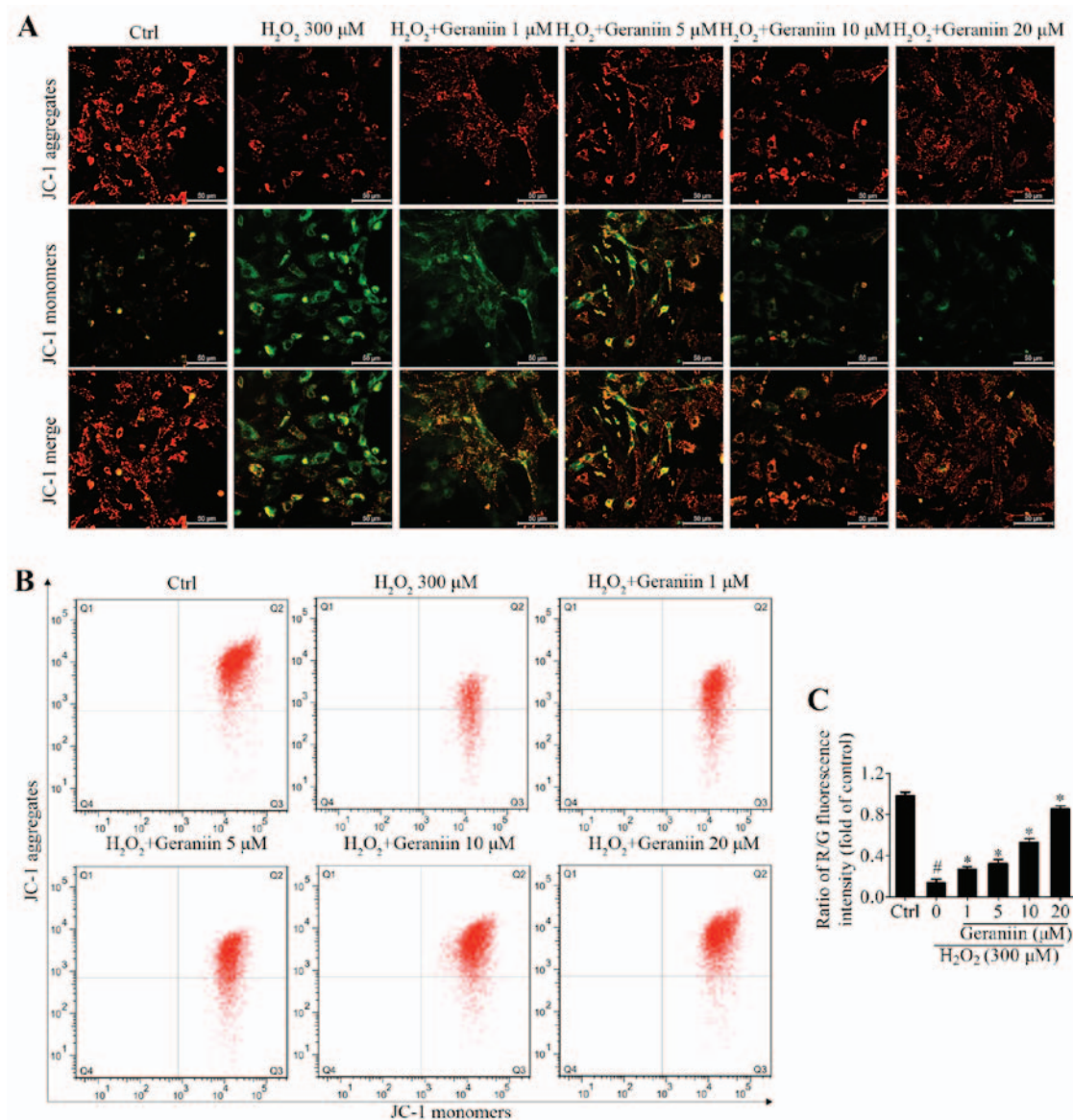


Figure 5. Geraniin protects MSCs against oxidative stress through stabilizing the Ψ_m . MSCs were treated with the indicated concentrations of geraniin for 24 h, followed by H_2O_2 administration for 4 h. Alterations in Ψ_m were analyzed by (A) fluorescence microscopy (original magnification, x200; scale bar, 50 μm) and (B) flow cytometry using the JC-1 staining method. (C) Ratio of R/G fluorescence intensity was quantified. Data are presented as the mean \pm standard deviation of three separate experiments. $^{\#}P < 0.05$ compared with the control group; $^*P < 0.05$ compared with the H_2O_2 -treated group. Ψ_m , mitochondrial membrane potential; Ctrl, control; H_2O_2 , hydrogen peroxide; MSCs, mesenchymal stem cells; R/G, red/green.

were investigated. Treatment with geraniin induced a marked increase in the expression levels of mitochondrial Cyt C and Bcl-2, and a decrease in the expression levels of cytoplasmic Cyt C, cleaved caspase-3 and Bax (Fig. 6D-F). These effects were reversed by LY294002. These data indicated that the PI3K/Akt signaling pathway may contribute to the prosurvival role of geraniin in MSCs under H_2O_2 treatment.

Discussion

The present study demonstrated that geraniin could attenuate H_2O_2 -induced cell damage through promoting MSC survival, reducing cellular ROS production and maintaining mitochondrial function. Furthermore, the effects of geraniin were mediated by activating the PI3K/Akt signaling pathway. Conversely, inhibition of the PI3K/Akt pathway weakened the

protective effects of geraniin. To the best of our knowledge, the present study is the first to report the cytoprotective effects of geraniin on MSCs and reveal the underlying mechanism.

MSCs are easily isolated and expanded, and can be transplanted in the heart; therefore, they are considered leading candidates for cellular therapy (27). Substantial data from preclinical and clinical studies support the cardioprotective effects of MSCs. Amado *et al* demonstrated that, following MSC implantation into swine, reappearance of myocardial tissue and restoration of cardiac contractility could be detected using serial computed tomography imaging (28). Furthermore, the POSEIDON randomized trial demonstrated that intravenous administration of allogeneic MSCs within 7 days of acute MI markedly attenuated cardiac hypertrophy, reduced ventricular arrhythmia, improved heart function and decreased rehospitalization for cardiac complications (27). However, the practical

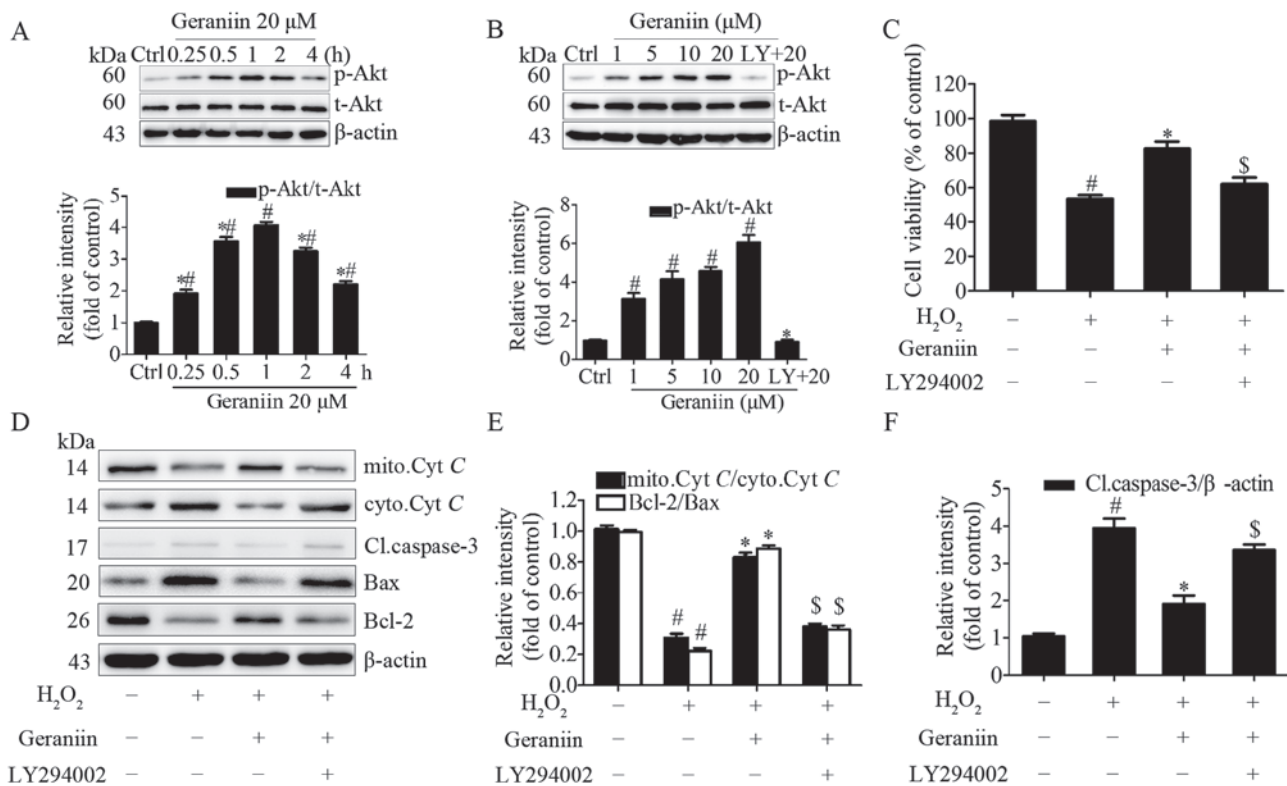


Figure 6. PI3K/Akt signaling is involved in the anti-apoptotic effects of geraniin on MSCs. (A) Cells were treated with geraniin (20 μ M) for the indicated time-periods or (B) with the indicated concentrations of geraniin for 1 h. The expression levels of p-Akt were evaluated by western blotting. (C) Cells were pretreated with LY294002, and were then incubated with geraniin and H₂O₂; cell viability was measured using the Cell Counting kit-8 assay. Expression levels of proteins associated with the mitochondrial apoptosis pathway were assessed (D) by western blotting and (E and F) data were semi-quantified. Data are presented as the mean \pm standard deviation of three independent experiments. [#]P<0.05 compared with the control group; ^{*}P<0.05 compared with the group treated with 20 μ M geraniin for 1 h (A and B); ^{*}P<0.05 compared with the H₂O₂ group (C-F); ^{\$}P<0.05 compared with the geraniin + H₂O₂ group. Akt, protein kinase B; Bax, Bcl-2-associated X protein; Bcl-2, B-cell lymphoma 2; Cl.caspase-3, cleaved caspase-3; ctrl, control; cyto.Cyt C, cytoplasmic cytochrome c; H₂O₂, hydrogen peroxide; LY, LY294002; mito.Cyt C, mitochondrial cytochrome c; p-, phosphorylated; t-, total.

applications of MSCs are restricted by their poor survival rate. A previous study indicated that after 4 days, only 0.44% of engrafted MSCs survived and resided in the myocardium (29). This low survival rate is mainly due to the hostile microenvironment in injured heart tissue, and ROS burst in the infarcted region is the major risk factor (30). As one of the most stable ROS, H₂O₂ is often used to establish an *in vitro* oxidative stress model for studies regarding apoptosis-associated mechanisms (31). In the present study, treatment with 300 μ M H₂O₂ for 4 h markedly increased apoptosis of MSCs, thus indicating that exogenous ROS burst results in the critical inducement of transplanted MSC apoptosis. In addition, this result reflects the necessity to identify novel methods to enhance MSC survival upon oxidative stress injury.

Pharmacological studies have confirmed that polyphenols are efficacious in treating ischemic heart diseases. Polyphenols extracted from grapes and wine have been reported to increase coronary flow *in vivo* (32). Epigallocatechin gallate, which is the major polyphenolic compound in green tea, has been revealed to prevent oxidative stress-induced cardiomyocyte apoptosis *in vitro* (33). In addition, emerging evidence demonstrated that the mechanism of action of polyphenols is predominantly ascribed to their strong free radical-scavenging activity. Polyphenols are rich in hydroxyl groups, which can capture free radicals and inactivate ROS (34). There are numerous hydroxyl groups within geraniin, indicating its strong antioxidant activity. Therefore,

the present study detected the antioxidative effects of geraniin on MSCs exposed to H₂O₂; the results confirmed that geraniin pretreatment may reduce excessive cellular ROS levels and preserve mitochondrial function, which may contribute to inhibiting H₂O₂-induced apoptosis. Furthermore, cells can defend themselves against oxidative stress through specific scavenging mechanisms, in which GSH and MDA are vital participants. GSH is an important intracellular antioxidative mediator that interacts with redundant ROS and balances cellular oxidation status. MDA is the end product of cellular lipid peroxidation in response to ROS damage. The severity of oxidative injury is, to some degree, reflected by cellular GSH and MDA levels. In the present study, geraniin was able to restore GSH levels, and decrease the amount of MDA, thus suggesting that geraniin functions through enhancing the intrinsic antioxidant repair system to inhibit ROS production and prevent accumulation of intracellular oxidative damage. Furthermore, reduced Ψ m indicates severe mitochondrial respiratory chain dysfunction and increased generation of ROS, which enhances oxidative stress and activates caspase family-mediated mitochondrial apoptosis (35). The present study indicated that geraniin treatment increases Bcl-2 expression, and decreases the expression levels of Bax and cleaved caspase-3, thus suggesting that geraniin may act on Ψ m stabilization. In addition, in normal cells, Cyt C is located in the mitochondrial inner membrane and works as an electron transmitter. Numerous proapoptotic stimuli, including ROS, increase permeability of

the outer membrane and promote mobilization of Cyt C from the mitochondria to the cytosol. In the cytosol, Cyt C mediates maturation of the caspase family, including caspase-3, which ultimately induce cell death (36). In the present study, geraniin treatment inhibited the release of Cyt C from the mitochondria to the cytosol, which indirectly indicated that geraniin preserved mitochondrial membrane integrity. Taken together, these results revealed that geraniin improved MSC viability under oxidative stress by enhancing the ability of the antioxidant defense system to remove excessive ROS, maintaining mitochondrial function and inhibiting the mitochondrial apoptosis pathway.

The PI3K/Akt pathway has been reported to serve an important role in the biological behavior of MSCs *in vitro* and in MSC engraftment for ischemic treatment *in vivo*. A previous study confirmed that cell survival, proliferation and migration were enhanced when the PI3K/Akt pathway was activated in MSCs (21). In addition, transplantation of MSCs overexpressing Akt preserved normal cardiac metabolism and pH post-MI (37). In the present study, the results verified that geraniin could activate the PI3K/Akt signaling pathway, whereas PI3K inhibition abolished the effects of geraniin on MSCs. However, inhibiting PI3K using LY294002 could not absolutely abrogate the protective effects of geraniin, thus suggesting that other pathways may be involved in the actions of geraniin. Numerous studies have indicated that geraniin activates the PI3K/Akt and mitogen-activated protein kinase (MAPK) pathways (25,38). Whether MAPK or other pathways also contribute to the beneficial effects of geraniin requires further study. Furthermore, the stromal cell-derived factor 1/C-X-C chemokine receptor type 4 (CXCR4) axis serves a vital role in mobilization and recruitment of MSCs to the infarcted area (39). PI3K/Akt activation has also been demonstrated to induce an increase in CXCR4 expression in MSCs (40). Whether geraniin may promote CXCR4 expression via the PI3K/Akt pathway remains unknown and requires further research.

In conclusion, the present study provided preliminary evidence to suggest that geraniin exerts prosurvival effects on MSCs against H₂O₂-induced cellular oxidative stress, predominantly by activating the PI3K/Akt signaling pathway. These findings indicated that geraniin may be considered a potential drug to enhance MSC-based therapeutic efficacy for the future treatment of MI.

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