

A 16-gene expression signature to distinguish stage I from stage II lung squamous carcinoma

RUI WANG^{1*}, YUXING CAI^{2*}, BAOPING ZHANG³ and ZHENGXIA WU²

¹Department of VIP and Geriatrics, Xi'an Gaoxin Hospital, Gaoxin Industrial Development District, Xi'an, Shanxi 710075; Departments of ²Respiratory Medicine and ³Thoracic Surgery, Baoji Central Hospital, Baoji, Shanxi 721008, P.R. China

Received July 17, 2017; Accepted December 8, 2017

DOI: 10.3892/ijmm.2017.3332

Abstract. The present study aimed to perform screening of a gene signature for the discrimination and prognostic prediction of stage I and II lung squamous carcinoma. A microarray meta-analysis was performed to identify differentially expressed genes (DEGs) between stage I and II lung squamous carcinoma samples in seven microarray datasets collected from the Gene Expression Omnibus database via the MetaQC and MetaDE package in R. The important DEGs were selected according to the betweenness centrality value of the protein-protein interaction (PPI) network. Support vector machine (SVM) analysis was performed to screen the feature genes for discrimination and prognosis. One independent dataset downloaded from The Cancer Genome Atlas was used to validate the reliability. Pathway enrichment analysis was also performed for the feature genes. A total of 924 DEGs were identified to construct a PPI network consisting of 392 nodes and 686 edges. The top 100 of the 392 nodes were selected as crucial genes to construct an SVM classifier, and a 16-gene signature (caveolin 1, eukaryotic translation elongation factor 1 γ , casein kinase 2 α 1, tyrosine 3-monooxygenase/tryptophan 5-monooxygenase activation η , tyrosine 3-monooxygenase/tryptophan 5-monooxygenase activation θ , pleiotrophin, insulin receptor, insulin receptor substrate 1, 3-phosphoinositide-dependent protein kinase-1, specificity protein 1, COP9 signalosome subunit 6, N-myc downstream regulated gene 1, retinoid X

receptor α , heat shock protein 90 α A1, karyopherin subunit β 1 and erythrocyte membrane protein band 4.1) with high discrimination accuracy was identified. This 16-gene signature had significant prognostic value, and patients with stage II lung squamous carcinoma exhibited shorter survival rates, compared with those with stage I disease. Seven DEGs of the 16-gene signature were significantly involved in the phosphoinositide 3-kinase-Akt signaling pathway. The 16-gene signature identified in the present study may be useful for stratifying the patients with stage I or II lung squamous carcinoma and predicting prognosis.

Introduction

Lung cancer is the leading cause of cancer-associated mortality worldwide, with an estimated 224,390 newly diagnosed cases and a mortality rate of 158,080 in the United States in 2016 (1). The prognosis of patients with lung cancer has been demonstrated to be associated with stage at diagnosis, and the survival rate for patients at early stages I and II is generally substantially higher, compared with those at advanced stages III and IV (2,3). Therefore, several studies have been performed to attempt to identify diagnostic biomarkers, which may assist in distinguishing early-stage from advanced-stage lung cancer in order to plan suitable therapeutic strategies for improving prognosis (4,5). The discrimination of stage I and II lung cancer is also clinically meaningful as patients with stage I or II have different susceptibilities to the development of distant metastases and chance of succumbing to mortality within 5 years (2,3). However, few studies have been performed on the further discrimination of stage I and II lung cancer.

Lung cancer consists of two major types, and non-small cell lung cancer (NSCLC) is the most common form, accounting for ~85% of lung cancer cases. NSCLC can be further divided into three subtypes: Adenocarcinoma (40%), squamous carcinoma (40%) and large-cell cancer (20%). Currently, the majority of studies on the identification of gene signatures for staging and prognosis have been focused on NSCLC (6-8) and its adenocarcinoma subtype (9,10). However, there have been fewer reports specific for lung squamous carcinoma (11,12).

The aim of the present study was to perform screening of a gene expression signature for the discrimination and predicting the prognosis of stage I and II lung squamous carcinoma by performing a meta-analysis of microarray datasets.

Correspondence to: Dr Baoping Zhang, Department of Thoracic Surgery, Baoji Central Hospital, 8 Jiang Tan Road, Weibin, Baoji, Shanxi 721008, P.R. China
E-mail: bjbaopzhang@163.com

Dr Zhengxia Wu, Department of Respiratory Medicine, Baoji Central Hospital, 8 Jiang Tan Road, Weibin, Baoji, Shanxi 721008, P.R. China
E-mail: wuzx135@tom.com

*Contributed equally

Key words: lung squamous carcinoma, staging, prognosis, gene signature

The results of the meta-analysis may increase the statistical power and improve the reliability of investigations, compared with single study analysis (13).

Materials and methods

Microarray data collection and preprocessing. The Gene Expression Omnibus (GEO) database at the National Center for Biotechnology Information (<http://www.ncbi.nlm.nih.gov/geo/>) was used to retrieve expression profile datasets of human lung squamous carcinoma. The inclusion criteria were as follows: i) mRNA expression profiles; ii) lung cancer tissues of patients with lung squamous carcinoma; iii) specific pathological stage I and II; and iv) number of samples >50. Finally, seven microarray datasets were obtained, including GSE43580, GSE50081, GSE42127, GSE41271, GSE17710, GSE68793 and GSE33532 (Table I).

The raw data (CEL files) of GSE43580, GSE50081, GSE68793 and GSE33532 were downloaded from the Affymetrix Human Genome U133 Plus 2.0 Array platform (<https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GPL570>). Background correction (MAS 5.0), normalization (quantile), log₂-transformation and median polish summarization were then performed by using the robust multichip average algorithm (14) implemented in the affy package from R/Bioconductor (<http://www.bioconductor.org/packages/release/bioc/html/affy.html>). The expression value data (TXT files) of the other three microarray datasets (GSE42127, GSE41271 and GSE17710) were directly downloaded from their corresponding platforms. The probe IDs were converted into gene symbols according to the annotation files. The probes without gene symbols were filtered and the average expression value of the probes was calculated as the expression value for the gene with multiple probes, followed by data normalization using the Linear Models for Microarray Analysis package (15) in R (<http://www.R-project.org>).

Meta-analysis of multiple microarray datasets for screening DEGs. In order to obtain more reliable genes associated with the stage of lung squamous carcinoma, a meta-analysis of the above seven microarray datasets was performed as previously described (16). The MetaQC package in R was used to eliminate the bias among datasets downloaded from different platforms by filtering out low-quality studies. This procedure provided six quantitative quality control measures, including the internal quality control (IQC, homogeneity of coexpression structure across studies), external quality control (EQC, consistency of coexpression information with pathway database), accuracy quality control (AQCg and AQCp, accuracy of differentially expressed gene and enriched pathway detection, respectively), and consistency quality control (CQCg and CQCp, consistency of differentially expressed gene and enriched pathway ranking, respectively) (17). In addition, the principal component analysis and standardized mean rank summary score were applied to assist in the identification of problematic studies (17). The datasets of high quality were screened for identification of DEGs using the MetaDEES algorithm of MetaDE package in R (18). The thresholds of homogeneity were set as $\tau^2=0$ and $Q_{pval} > 0.05$. A false discovery rate (FDR) of <0.05 was considered as the cut-off

criterion for identifying DEGs between stage I and II of lung squamous carcinoma samples.

Protein-protein interaction (PPI) network-based prioritization of DEGs. Cytoscape is software for biological network visualization and analysis (19), in which the PPIs are downloaded from acknowledged Human Protein Reference Database (<http://www.hprd.org>) (20). The PPI network for the identified DEGs was constructed using Cytoscape software 3.5.0 to investigate the DEGs crucial for the development of lung squamous carcinoma. In addition, the non-differentially expressed genes, which interacted with at least 10 DEGs were included in this PPI network. The signaling pathways enrichment analysis was performed for the genes in the constructed PPI network with the criterion of $P < 0.05$.

Betweenness centrality (BC) is one of the important topological characteristics of the PPI network for determining the highly connected nodes (hubs). BC is defined as the fraction of shortest paths, which pass through a node (v), and nodes with BC values closer to 1 have a higher hub degree. BC can be calculated using the following equation:

$$C_B(v) = \sum_{t \neq v \in V} \frac{\sigma_{st}(v)}{\sigma_{st}}$$

Where σ_{st} is the total number of shortest paths from node s to node t , and $\sigma_{st}(v)$ is the number of those paths that pass through v .

Support vector machine (SVM) modeling and classification accuracy evaluation. Using the top 100 genes ranked by the BC value, a supervised SVM classifier was constructed with the e1071 package for R under the default parameters (radial basis function kernel, $\gamma=0.5$; cost=4; cross=10-fold) (21). Following screening of the optimal combination of genes, which completely distinguished between stage I or II lung squamous carcinoma samples by utilizing the dataset with the maximum sample size as the training sets, its predictive accuracy was computed using the other datasets as the test sets. The effects of classification were evaluated based on five parameters, including the accuracy, sensitivity, specificity, positive predictive value, negative predictive value, and area under the receiver operating curve (AUC).

Prognostic validation analysis. To further demonstrate the classification reliability and prognostic potential of the optimal combination of genes, an independent dataset, comprising 67 stage I lung squamous carcinoma samples, 27 stage II lung squamous carcinoma samples and survival rates was downloaded from The Cancer Genome Atlas (TCGA; <http://tcga-data.nci.nih.gov/tcga/>; TCGA_LUSC_exp_u133a level 3). Cox regression analysis was then performed for this independent dataset using the R survival package (22).

Pathway enrichment analysis. Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway enrichment analysis (<http://www.kegg.jp/>) was performed for the gene signature with a two-tailed Fisher's exact test based on the hypergeometric distribution. The P-value was adjusted for multiple testing using the Benjamini and Hochberg approach (23), and adjusted $P < 0.05$ was set as the threshold.

Table I. General information of included expression profiles.

GEO accession	Chip	Probe number	Sample size	Stage I	Stage II
GSE43580	HG-U133_Plus_2	54,676	73	34	39
GSE50081	HG-U133_Plus_2	54,676	43	27	16
GSE42127	Illumina HumanWG-6	48,803	33	23	10
GSE41271	Illumina HumanWG-6	48,803	48	31	17
GSE17710	Agilent-UNC	33,421	53	34	19
GSE68793	HT_HG-U133A	22,277	101	70	31
GSE33532	HG-U133_Plus_2	25,906	16	8	8

GEO, Gene Expression Omnibus.

Table II. Results of QC measures and SMRs.

Study	IQC	EQC	CQCg	CQCp	AQCg	AQCp	SMR
GSE43580	4.52	2.00	307.65	133.86	32.71	82.01	2.17
GSE42127	4.96	1.70	307.65	101.51	30.42	29.94	2.58
GSE50081	5.42	2.00	307.65	60.02	29.49	29.18	3.33
GSE41271	4.58	2.00	19.10	90.31	8.24	58.61	3.25
GSE17710	0.23	0.37	0.09	1.89	0.05	0.36	6.83
GSE68793	5.92	1.10	276.14	35.54	19.03	10.66	3.92
GSE33532	1.79	5.00	0.59	1.69	0.52	3.21	5.67

QC, quality control; IQC, internal; EQC, external; CQCg, consistency QC of differentially expressed gene; CQCp, CQC of enriched pathway; AQCg, accuracy QC of differentially expressed gene; AQCp, AQC of enriched pathway; SMR, standardized mean rank.

Results

Identification of DEGs with microarray meta-analysis. Among the seven microarray datasets downloaded from the GEO, five datasets (GSE43580, GSE50081, GSE42127, GSE41271 and GSE68793) were included in the meta-analysis for DEGs following MetaQC analysis. The other two datasets (GSE17710 and GSE33532) were excluded due to small sample size and lower quantitative quality control scores (Table II and Fig. 1). A total of 964 DEGs between the stage I and II lung squamous carcinoma samples were identified using the MetaDE.ES algorithm under the cut-off criterion of FDR <0.05. The top 10 DEGs ranked by the FDR value are listed in the Table III.

Prioritization of DEGs by PPI network analysis. The PPI network was constructed by mapping the 964 DEGs into PPI data, consisting of 392 nodes (proteins) and 686 edges (interactions). A total of five pathways were significantly enriched for genes in the constructed PPI network, including proteasome, Wnt signaling pathway, ribosome, apoptosis and focal adhesion (Fig. 2). The top 100 genes ranked by BC value were then selected as important genes in order to construct an SVM classifier for distinguishing between stage I and II lung squamous carcinoma samples (Table IV). Using the GSE68793 dataset as a training set, the classification accuracy was found to gradually improve with the increase in the number of DEGs. When

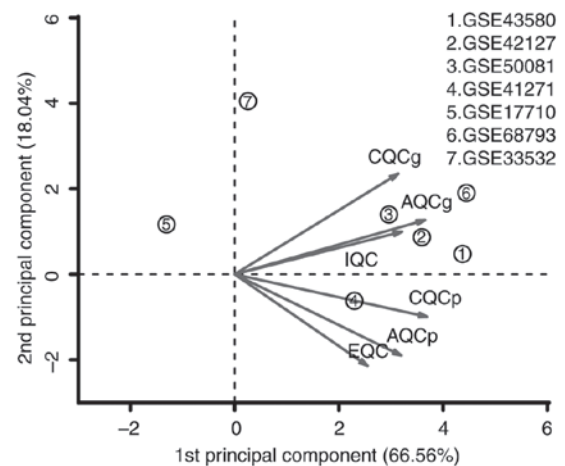


Figure 1. Principal component analysis biplots of QC measures in seven microarray datasets. The numbers in circles represent the overall rank of a study based on the standardized mean rank score. Smaller numbers correspond to higher quality studies. QC, quality control; CQCg, consistency QC of differentially expressed gene; CQCp, CQC of enriched pathway; AQCg, accuracy QC of differentially expressed gene; AQCp, AQC of enriched pathway; IQC, internal QC; EQC, external QC.

the 16 DEGs, comprising 15 upregulated DEGs in stage II lung squamous carcinoma, including caveolin 1, (*CAVI*), eukaryotic translation elongation factor 1 γ (*EEF1G*), casein kinase 2 α 1 (*CSNK2A1*), tyrosine 3-monooxygenase/tryptophan

Table III. Top 10 differentially expressed genes screened using microarray meta-analysis.

Symbol	P-value	FDR	tau ²	Q-value	QP-value
<i>PLP2</i>	1.30E-10	1.59E-09	0	3.67E+00	4.52E-01
<i>EEF1G</i>	1.33E-10	1.62E-09	0	1.33E+00	8.56E-01
<i>RPL19</i>	1.33E-10	1.62E-09	0	3.62E+00	4.60E-01
<i>C7</i>	1.35E-10	1.64E-09	0	2.90E+00	5.75E-01
<i>MDH2</i>	1.36E-10	1.66E-09	0	8.43E-01	9.33E-01
<i>IGFBP6</i>	1.37E-10	1.67E-09	0	2.57E+00	6.33E-01
<i>OR12D2</i>	1.39E-10	1.69E-09	0	2.12E+00	7.14E-01
<i>SERPINI1</i>	1.44E-10	1.74E-09	0	3.88E+00	4.23E-01
<i>TNS4</i>	1.51E-10	1.81E-09	0	2.30E+00	6.82E-01
<i>KDR</i>	1.54E-10	1.84E-09	0	1.34E+00	8.54E-01

FDR, false discovery rate.

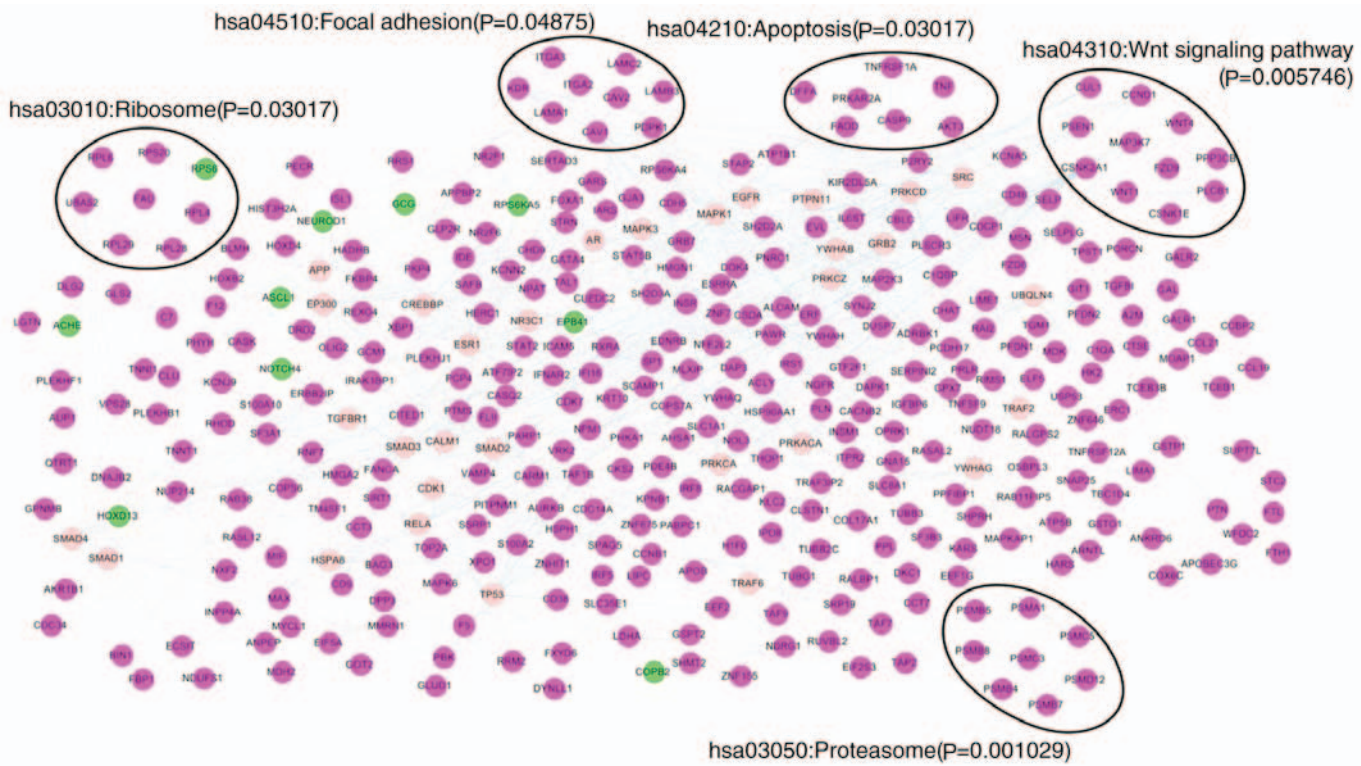


Figure 2. Protein-protein interaction network of critical genes. A total of 392 nodes and 686 edges were included. The nodes represented the differentially expressed genes. Purple represents upregulated genes in stage II; green represents downregulated genes in stage II; light pink represents non-differentially expressed genes.

5-monooxygenase activation η (*YWHAH*), tyrosine 3-monooxygenase/tryptophan 5-monooxygenase activation θ (*YWHAQ*), pleiotrophin (*PTN*), insulin receptor (*INSR*), insulin receptor substrate 1 (*IRS1*), 3-phosphoinositide-dependent protein kinase-1 (*PDPK1*), specificity protein 1 (*Sp1*), COP9 signalosome subunit 6 (*COPS6*), N-myc downstream regulated gene 1 (*NDRG1*), retinoid X receptor α (*RXR α*), heat shock protein 90 α A1 (*HSP90AA1*) and karyopherin subunit β 1 (*KPNB1*) and one downregulated DEG in stage II lung squamous carcinoma, namely erythrocyte membrane protein band 4.1 (*EPB41*) were included in the top 40 crucial genes,

100% accuracy was achieved (Fig. 3). It was also found that high accuracy was obtained using the other four datasets (GSE43580: 94.5%, 69/73; GSE50081: 97.7%, 42/43; GSE41271: 97.9%, 47/48; GSE42127: 100%, 33/33) as test sets (Table V).

The present study also examined the prognostic value of the 16 DEGs by performing survival analysis with the GSE42127, GSE50081 and GSE41271 (Fig. 4) datasets. The results indicated that patients with stage I lung squamous carcinoma exhibited significantly higher survival rates, compared with the patients with stage II lung squamous carcinoma in GSE50081 ($P=0.000138$) using these 16 genes (Fig. 4B).

Table IV. Top 10 genes with highest BC scores.

Gene	BC score	Expression	P-value	FDR	tau ²	Q-value	QP-value
<i>PORCN</i>	0.6667	1	2.45E-02	3.26E-01	0	5.66E-01	9.67E-01
<i>WNT1</i>	0.5000	1	1.44E-02	2.90E-01	0	2.25E+00	6.89E-01
<i>WNT4</i>	0.5000	1	1.50E-02	2.95E-01	0	3.03E+00	5.54E-01
<i>CAV1</i>	0.1587	1	2.00E-03	1.45E-01	0	7.05E-01	9.51E-01
<i>CSNK2A1</i>	0.1452	1	4.00E-02	3.74E-01	0	2.39E+00	6.64E-01
<i>HSP90AA1</i>	0.1436	1	1.14E-02	2.66E-01	0	2.09E+00	7.20E-01
<i>YWHAG</i>	0.1087	-	9.95E-02	5.04E-01	7.70E-02	7.86E+00	9.70E-02
<i>TP53</i>	0.0993	-	8.75E-01	9.68E-01	2.15E-01	1.47E+01	5.40E-03
<i>SP1</i>	0.0966	1	1.70E-03	1.40E-01	0	3.41E+00	4.92E-01
<i>YWHAQ</i>	0.0861	1	1.01E-02	2.54E-01	0	2.08E+00	7.21E-01

Expression: 1, expression higher in stage II than stage I; -, not differentially expressed; BC, betweenness centrality; FDR, false discovery rate.

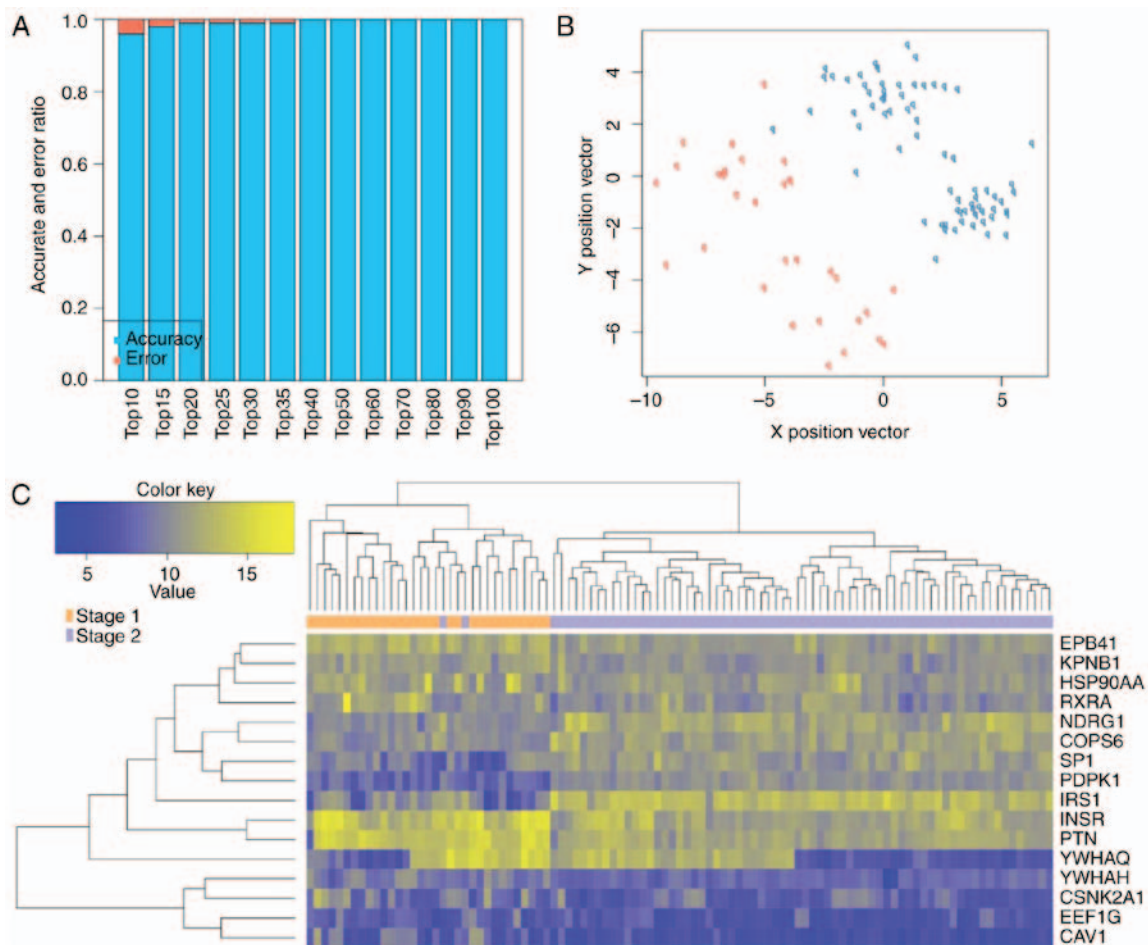


Figure 3. SVM modeling and classification accuracy evaluation. (A) Accurate rate using crucial genes (screened by the betweenness centrality score) as classifier; 100% accuracy was achieved in the top 40. (B) Visualization of classification result using the 16-gene signature as an SVM classifier and GSE68793 as a training dataset (orange, stage II sample; blue, stage I sample). (C) Hierarchical clustering analysis indicated samples of the same type, which were clustered using the 16-gene signature (yellow, higher expression; blue, lower expression). SVM, support vector machine.

Therefore, this classification model may also be effective for clinical prognosis and treatment guidance.

To further confirm the classification reliability of the above selected 16 genes, another independent dataset was downloaded from TCGA database. The results indicated

that these 16 genes provided a predictive accuracy of 100% (67/67) for stage I lung squamous carcinoma samples, a predictive accuracy of 92.59% (25/27) for stage II lung squamous carcinoma, an overall predictive accuracy of 97.87% (92/94) and an AUC of 0.996 (Fig. 5A). The survival analysis also

Table V. Support vector machine classifier confirmation in test datasets.

Dataset	Sample size (n)	Correct rate, %	Sensitivity	Specificity	PPV	NPV	AUROC
GSE43580	73	94.5	0.941	0.949	0.941	0.949	0.989
GSE50081	43	97.7	1.000	0.936	0.964	1.000	0.991
GSE41271	48	97.9	1.000	0.941	0.969	1.000	0.994
GSE42127	33	100.0	1.000	1.000	1.000	1.000	1.000

PPV, positive predictive value; NPV, negative predictive value; AUROC, area under receiver operating curve.

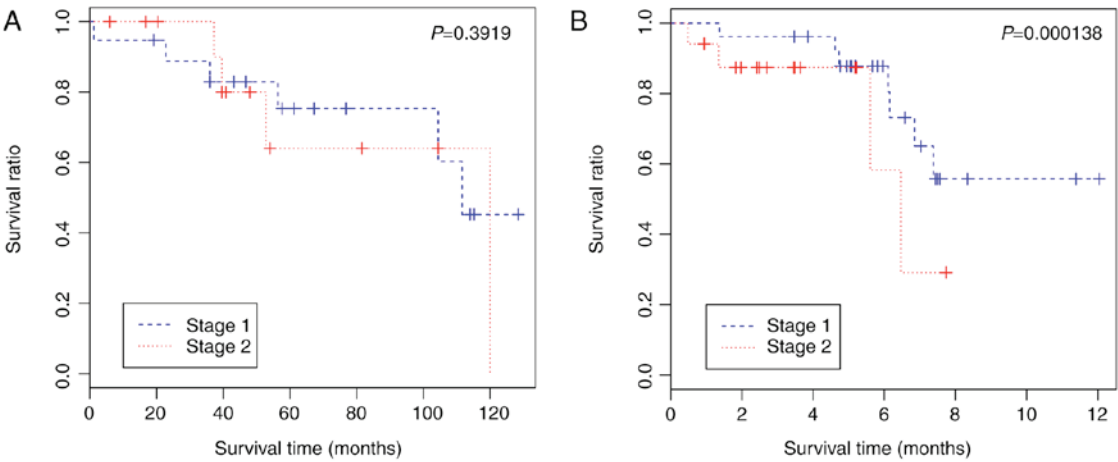


Figure 4. Survival analysis. Survival analysis for dataset (A) GSE42171 and (B) GSE50081.

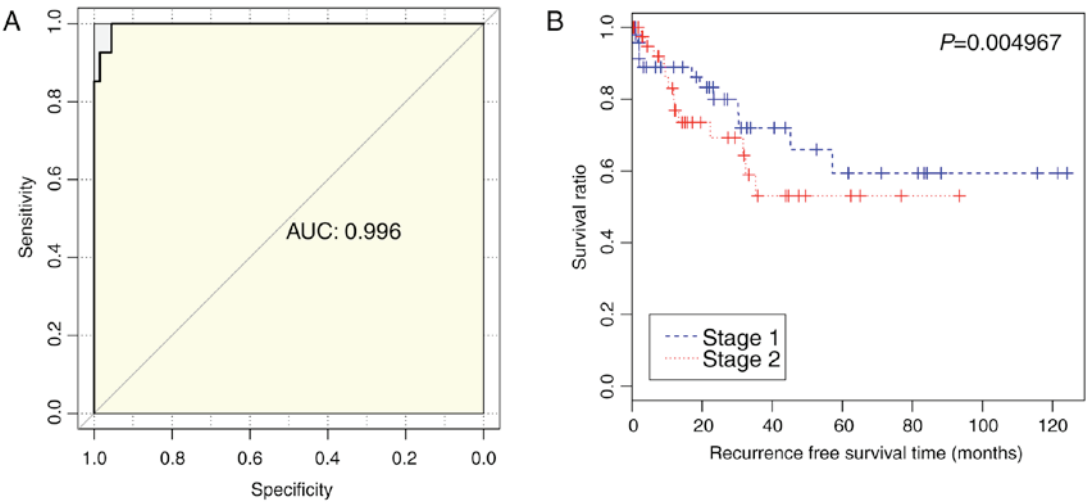


Figure 5. Support vector machine classifier confirmation and survival analysis using another independent dataset. (A) Receiver operating characteristic curve. (B) Kaplan-Meier curve. AUC, area under curve.

demonstrated that the survival rates of patients with stage II lung squamous carcinoma were significantly lower, compared with those of patients with stage I lung squamous carcinoma ($P=0.004967$) (Fig. 5B).

Pathway enrichment analysis. Only one significant KEGG pathway, the phosphoinositide 3-kinase (PI3K)-Akt signaling pathway (adjusted $P=0.020268797$), was enriched for seven DEGs (*YWHAQ*, *IRSI*, *PDPK1*, *HSP90AA1*, *INSR*, *RXRA* and

YWHAH). No pathway was significantly enriched for the other nine DEGs.

Discussion

The present study identified 16 DEGs, which can be used as a gene signature to distinguish stage I from stage II lung squamous carcinoma and are of value for predicting prognosis. There were 15 upregulated DEGs (*CAVI*, *EEFIG*, *CSNK2A1*,

YWHAH, *YWHAQ*, *PTN*, *INSR*, *IRS1*, *PDPK1*, *Sp1*, *COPS6*, *NDRG1*, *RXRA*, *HSP90AA1* and *KPNB1*) and one downregulated DEG (*EPB41*) in stage II lung squamous carcinoma.

CAVI, encoding a 22-24 kDa structural protein component of caveolae in the plasma membrane, has been demonstrated to be upregulated in NSCLC (24). In addition, high expression of *CAVI* has shown to be associated with poor prognosis in patients with NSCLC (25,26). The high expression level of *EEF1G* may promote the synthesis of several proteins to prevent oxidative stress and nutrient deprivation, resulting in the excessive proliferation of cells and development of cancer (27). However, direct investigations of *EEF1G* remain limited and further confirmation of *EEF1G* in lung squamous carcinoma remains essential. *CSNK2A1*, encoding an α catalytic subunit of protein kinase CK2, promotes tumor cell growth, adhesion and migration by activating the Janus kinase/signal transducer and activator of transcription, nuclear factor- κ B, PI3K/AKT, Hsp90, Wnt and Hedgehog pathways (28). A study by O-charoenrat *et al* indicated that *CSNK2A1* may serve as a potential predictor of poor prognosis for lung squamous carcinoma (29), which is consistent with the present study to a certain extent. *PTN* is a heparin-binding growth factor, with high secretion levels in NSCLC cells (30). *PTN* regulates cell proliferation and migration in NSCLC by activating its cell surface receptors and PI3K signaling (31,32), which may lead to a poor prognosis (33). *Sp1*, an oncogenic transcription factor for lung squamous carcinoma, may be involved in regulating the promoter activity of its target genes (34,35). Detecting the expression level of *Sp1* is considered to be an effective approach for the diagnosis and prognosis of lung cancer (36).

COPS6, encoding one of the eight subunits of the COP9 signalosome, mediates the upregulation of *E6AP* by reducing *E6AP* poly-ubiquitination and targeting p53 for proteasome-mediated degradation, which in turn decreases the transcriptional activity of p53 target genes to induce cervical cell growth and motility (37). However, the underlying mechanism of *COPS6* in lung squamous carcinoma remains to be fully elucidated. *NDRG1*, a member of the N-myc downstream regulated gene family, can be overexpressed in several types of lung cancer, including lung squamous cell carcinoma (38,39). The survival rate of patients negative for the nuclear expression of *NDRG1* has been reported to be significantly higher, compared with that in patients who positive for *NDRG1* (38). In addition, a high expression of *NDRG1* may promote the progression of lung squamous cell carcinoma by upregulating vascular endothelial growth factor-A and interleukin-8/C-X-C-motif chemokine ligand 8 (38). *EPB41*, encoding the erythrocyte membrane protein band 4.1, has been shown to function as a tumor suppressor gene in meningioma cell lines (40). However, the role and mechanism of *EPB41* in lung cancer remains to be fully elucidated and further experiments are essential.

According to the pathway enrichment analysis in the present study, seven DEGs (*YWHAQ*, *IRS1*, *PDPK1*, *HSP90AA1*, *INSR*, *RXRA* and *YWHAH*) among the 16 DEGs were significantly involved in the PI3K-Akt signaling pathway. Increasing evidence suggests that activation of the PI3K-Akt signaling pathway is correlated with poor prognosis in patients with NSCLC, as this pathway antagonizes apoptosis and contributes to the proliferation and metastasis

of NSCLC cells (41,42). It has been reported that upregulated expression levels of *HSP90AA1* and *PDPK1* are significantly associated with shorter overall survival rates in patients with NSCLC (43,44). *INSR* can be activated by the insulin ligand to trigger the PI3K/AKT signaling pathway and enhance tumor growth, migration and angiogenesis (45,46). Previous multivariate analysis revealed patients with high expression levels of *INSR* had shorter overall survival rates, compared with those with low expression levels (47). A truncated version of *RXR- α* has been demonstrated to significantly contribute to the anchorage-independent growth of cancer cells by activation of the PI3K/AKT pathway (48). *YWHAH* and *YWHAQ* mediate oncogenic transformation by stimulating PI3K signaling, and these two genes may be correlated with a more advanced pathologic stage and lower overall survival rate (49,50). Therefore, it may be that these seven upregulated DEGs involved in the PI3K-Akt signaling pathway may also be associated with decreased survival rate in patients with stage II lung squamous carcinoma.

In conclusion, the 16-gene expression signature identified in the present study may be of value in the discrimination of patients with stage I or II lung squamous carcinoma and prediction of prognosis. Further investigations aim to analyze the association between the gene expression levels of this 16-gene signature and prognosis based on the prognosis of clinical patients.

References

1. Siegel RL, Miller KD and Jemal A: Cancer statistics, 2017. *CA Cancer J Clin* 66: 7-30, 2017.
2. McPhail S, Johnson S, Greenberg D, Peake M and Rous B: Stage at diagnosis and early mortality from cancer in England. *Br J Cancer* 112 (Suppl 1): S108-S115, 2015.
3. Walters S, Maringe C, Coleman MP, Peake MD, Butler J, Young N, Bergström S, Hanna L, Jakobsen E, Kölbek K, *et al*: Lung cancer survival and stage at diagnosis in Australia, Canada, Denmark, Norway, Sweden and the UK: A population-based study, 2004-2007. *Thorax* 68: 551-564, 2013.
4. Klarod K, Hongprabhas P, Khampitak T, Wirasorn K, Kiertiburanakul S, Tangrassameeprasert R, Daduang J, Yongvanit P and Boonsiri P: Serum antioxidant levels and nutritional status in early and advanced stage lung cancer patients. *Nutrition* 27: 1156-1160, 2011.
5. Lim W, Zhang WH, Miller CR, Watters JW, Gao F, Viswanathan A, Govindan R and McLeod HL: PTEN and phosphorylated AKT expression and prognosis in early- and late-stage non-small cell lung cancer. *Oncol Rep* 17: 853-857, 2007.
6. Lu Y, Lemon W, Liu PY, Yi Y, Morrison C, Yang P, Sun Z, Szoke J, Gerald WL, Watson M, *et al*: A gene expression signature predicts survival of patients with stage I non-small cell lung cancer. *PLoS Med* 3: e467, 2006.
7. Der SD, Sykes J, Pintilie M, Zhu CQ, Strumpf D, Liu N, Jurisica I, Shepherd FA and Tsao MS: Validation of a histology-independent prognostic gene signature for early-stage, non-small-cell lung cancer including stage IA patients. *J Thorac Oncol* 9: 59-64, 2014.
8. Huang S, Reitze NJ, Ewing AL, McCreary S, Uihlein AH, Brower SL, Wang D, Wang T, Gabrin MJ, Keating KE, *et al*: Analytical performance of a 15-gene prognostic assay for early-stage non-small-cell lung carcinoma using RNA-stabilized tissue. *J Mol Diagn* 17: 438-445, 2015.
9. Skrzypski M, Jassem E, Taron M, Sanchez JJ, Mendez P, Rzyman W, Gulida G, Raz D, Jablons D, Provencio M, *et al*: Three-gene expression signature predicts survival in early-stage squamous cell carcinoma of the lung. *Clin Cancer Res* 14: 4794-4799, 2008.
10. Krzystanek M, Moldvay J, Szűts D, Szallasi Z and Eklund AC: A robust prognostic gene expression signature for early stage lung adenocarcinoma. *Biomarker Res* 4: 1, 2016.

11. Hwang JA, Song JS, Yu DY, Kim HR, Park HJ, Park YS, Kim WS and Choi CM: Peroxiredoxin 4 as an independent prognostic marker for survival in patients with early-stage lung squamous cell carcinoma. *Int J Clin Exp Pathol* 8: 6627-6635, 2015.
12. Zhu CQ, Strumpf D, Li CY, Li Q, Liu N, Der S, Shepherd FA, Tsao MS and Jurisica I: Prognostic gene expression signature for squamous cell carcinoma of lung. *Clin Cancer Res* 16: 5038-5047, 2010.
13. Kim TH, Choi SJ, Lee YH, Song GG and Ji JD: Gene expression profile predicting the response to anti-TNF treatment in patients with rheumatoid arthritis; analysis of GEO datasets. *Joint Bone Spine* 81: 325-330, 2014.
14. Bolstad BM, Irizarry RA, Åstrand M and Speed TP: A comparison of normalization methods for high density oligonucleotide array data based on variance and bias. *Bioinformatics* 19: 185-193, 2003.
15. Smyth GK: Limma: Linear models for microarray data. In: *Bioinformatics and computational biology solutions using R and Bioconductor*. Springer, pp397-420, 2005.
16. Yang ZH, Zheng R, Gao Y and Zhang Q: Identification of suitable genes contributes to lung adenocarcinoma clustering by multiple meta-analysis methods. *Clin Respir J* 10: 631-646, 2016.
17. Kang DD, Sibille E, Kaminski N and Tseng GC: MetaQC: Objective quality control and inclusion/exclusion criteria for genomic meta-analysis. *Nucleic Acids Res* 40: e15, 2012.
18. Chang LC, Lin HM, Sibille E and Tseng GC: Meta-analysis methods for combining multiple expression profiles: Comparisons, statistical characterization and an application guideline. *BMC Bioinformatics* 14: 368, 2013.
19. Kohl M, Wiese S and Warscheid B: Cytoscape: Software for visualization and analysis of biological networks. *Methods Mol Biol* 696: 291-303, 2011.
20. Peri S, Navarro JD, Kristiansen TZ, Amanchy R, Surendranath V, Muthusamy B, Gandhi TK, Chandrika KN, Deshpande N, Suresh S, *et al*: Human protein reference database as a discovery resource for proteomics. *Nucleic Acids Res* 32: D497-D501, 2004.
21. Meyer D and Wien FT: Support vector machines. The interface to libsvm in package, e1071, 2015.
22. Therneau T: A package for survival analysis in S. R package version 2.37-4. URL <http://CRAN.R-project.org/package=survival> Box 980032: 23298-20032, 2013.
23. Benjamini Y and Hochberg Y: Controlling the false discovery rate: A practical and powerful approach to multiple testing. *J Royal Statistical Soc Series B* 57: 289-300, 1995.
24. Sunaga N, Miyajima K, Suzuki M, Sato M, White MA, Ramirez RD, Shay JW, Gazdar AF and Minna JD: Different roles for caveolin-1 in the development of non-small cell lung cancer versus small cell lung cancer. *Cancer Res* 64: 4277-4285, 2004.
25. Chen D, Shen C, Du H, Zhou Y and Che G: Duplex value of caveolin-1 in non-small cell lung cancer: A meta analysis. *Familial Cancer* 13: 449-457, 2014.
26. Ho CC, Kuo SH, Huang PH, Huang HY, Yang CH and Yang PC: Caveolin-1 expression is significantly associated with drug resistance and poor prognosis in advanced non-small cell lung cancer patients treated with gemcitabine-based chemotherapy. *Lung Cancer* 59: 105-110, 2008.
27. Matassa D, Amoroso MR, Agliarulo I, Maddalena F, Sisinni L, Paladino S, Romano S, Romano MF, Sagar V, Loreni F, *et al*: Translational control in the stress adaptive response of cancer cells: A novel role for the heat shock protein TRAP1. *Cell Death Dis* 4: e851, 2013.
28. Zheng Y, McFarland BC, Drygin D, Yu H, Bellis SL, Kim H, Bredel M and Benveniste EN: Targeting protein kinase CK2 suppresses prosurvival signaling pathways and growth of glioblastoma. *Clin Cancer Res* 19: 6484-6494, 2013.
29. O-charoenrat P, Rusch V, Talbot SG, Sarkaria I, Viale A, Socci N, Ngai I, Rao P and Singh B: Casein kinase II alpha subunit and C1-inhibitor are independent predictors of outcome in patients with squamous cell carcinoma of the lung. *Clinical Cancer Res* 10: 5792-5803, 2004.
30. Jäger R, List B, Knabbe C, Souttou B, Raulais D, Zeiler T, Wellstein A, Aigner A, Neubauer A and Zugmaier G: Serum levels of the angiogenic factor pleiotrophin in relation to disease stage in lung cancer patients. *Br J Cancer* 86: 858-863, 2002.
31. Feng ZJ, Gao SB, Wu Y, Xu XF, Hua X and Jin GH: Lung cancer cell migration is regulated via repressing growth factor PTN/RPTP β/ζ signaling by menin. *Oncogene* 29: 5416-5426, 2010.
32. Gao S, Feng Z, Xu B, Wu Y, Yin P, Yang Y, Hua X and Jin GH: Suppression of lung adenocarcinoma through menin and polycomb gene-mediated repression of growth factor pleiotrophin. *Oncogene* 28: 4095-4104, 2009.
33. Du ZY, Shi MH, Ji CH and Yu Y: Serum pleiotrophin could be an early indicator for diagnosis and prognosis of non-small cell lung cancer. *Asian Pac J Cancer Prev* 16: 1421-1425, 2014.
34. Brown KC, Perry HE, Lau JK, Jones DV, Pulliam JF, Thornhill BA, Crabtree CM, Luo H, Chen YC and Dasgupta P: Nicotine induces the up-regulation of the $\alpha 7$ -nicotinic receptor ($\alpha 7$ -nAChR) in human squamous cell lung cancer cells via the Sp1/GATA protein pathway. *J Biol Chem* 288: 33049-33059, 2013.
35. Li M, Ling B, Xiao T, Tan J, An N, Han N, Guo S, Cheng S and Zhang K: Sp1 transcriptionally regulates BRK1 expression in non-small cell lung cancer cells. *Gene* 542: 134-140, 2014.
36. Cha N, Lv M, Zhao YJ, Yang D, Wang EH and Wu GP: Nicotine utility of VEGF mRNA and SP1 mRNA expression in bronchial cells of patients with lung cancer. *Respirology* 19: 544-548, 2014.
37. Gao S, Fang L, Phan LM, Qdaisat A, Yeung SC and Lee MH: COP9 signalosome subunit 6 (CSN6) regulates E6AP/UBE3A in cervical cancer. *Oncotarget* 6: 28026-28041, 2015.
38. Azuma K, Kawahara A, Hattori S, Taira T, Tsurutani J, Watari K, Shibata T, Murakami Y, Takamori S, Ono M, *et al*: NDRG1/Cap43/Drg-1 may predict tumor angiogenesis and poor outcome in patients with lung cancer. *J Thorac Oncol* 7: 779-789, 2012.
39. Wang D, Tian X and Jiang Y: NDRG1/Cap43 overexpression in tumor tissues and serum from lung cancer patients. *J Cancer Res Clin Oncol* 138: 1813-1820, 2012.
40. Robb VA, Li W, Gascard P, Perry A, Mohandas N and Gutmann DH: Identification of a third Protein 4.1 tumor suppressor, Protein 4.1 R, in meningioma pathogenesis. *Neurobiol Dis* 13: 191-202, 2003.
41. Zhou L, Luan H, Liu Q, Jiang T, Liang H, Dong X and Shang H: Activation of PI3K/Akt and ERK signaling pathways antagonized sinomenine-induced lung cancer cell apoptosis. *Mol Med Rep* 5: 1256-1260, 2012.
42. Zhang C, Lan T, Hou J, Li J, Fang R, Yang Z, Zhang M, Liu J and Liu B: NOX4 promotes non-small cell lung cancer cell proliferation and metastasis through positive feedback regulation of PI3K/Akt signaling. *Oncotarget* 5: 4392-4405, 2014.
43. Gallegos Ruiz MI, Floor K, Roepman P, Rodriguez JA, Meijer GA, Mooi WJ, Jassem E, Niklinski J, Muley T, van Zandwijk N, *et al*: Integration of gene dosage and gene expression in non-small cell lung cancer, identification of HSP90 as potential target. *PLoS One* 3: e0001722, 2008.
44. Han L, Zhang G, Zhang N, Li H, Liu Y, Fu A and Zheng Y: Prognostic potential of microRNA-138 and its target mRNA PDK1 in sera for patients with non-small cell lung cancer. *Med Oncol* 31: 129, 2014.
45. Taniguchi CM, Emanuelli B and Kahn CR: Critical nodes in signalling pathways: Insights into insulin action. *Nat Rev Mol Cell Biol* 7: 85-96, 2006.
46. Heidegger I, Kern J, Ofer P, Klocker H and Massoner P: Oncogenic functions of IGF1R and INSR in prostate cancer include enhanced tumor growth, cell migration and angiogenesis. *Oncotarget* 5: 2723-2735, 2014.
47. Kim JS, Kim ES, Liu D, Lee JJ, Solis L, Behrens C, Lippman SM, Hong WK, Wistuba II and Lee HY: Prognostic impact of insulin receptor expression on survival of patients with non-small cell lung cancer. *Cancer* 118: 2454-2465, 2012.
48. Wang GH, Jiang FQ, Duan YH, Zeng ZP, Chen F, Dai Y, Chen JB, Liu JX, Liu J, Zhou H, *et al*: Targeting truncated retinoid X receptor- α by CF31 Induces TNF- α -dependent apoptosis. *Cancer Res* 73: 307-318, 2013.
49. Radhakrishnan VM and Martinez JD: 14-3-3gamma induces oncogenic transformation by stimulating MAP kinase and PI3K signaling. *PLoS One* 5: e11433, 2010.
50. Fan T, Li R, Todd NW, Qiu Q, Fang HB, Wang H, Shen J, Zhao RY, Caraway NP, Katz RL, *et al*: Up-regulation of 14-3-3zeta in lung cancer and its implication as prognostic and therapeutic target. *Cancer Res* 67: 7901-7906, 2007.



This work is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International (CC BY-NC-ND 4.0) License.