

Reversal of the Warburg effect with DCA in PDGF-treated human PASMCM is potentiated by pyruvate dehydrogenase kinase-1 inhibition mediated through blocking Akt/GSK-3 β signalling

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Abstract. There is accumulating evidence indicating that the growth inhibitory effect of dichloroacetate (DCA) on pulmonary arterial smooth muscle cells (PASCs) may be associated with the reversal of the Warburg effect and initiation of the mitochondria-dependent apoptotic pathway. Previous studies indicated that platelet-derived growth factor (PDGF) promoted the Warburg effect and resulted in apoptotic resistance of PASCs, which was attributed to

activation of the phosphatidylinositol 3-kinase (PI3K)/protein kinase B (Akt) signalling pathway. However, the mechanism underlying the pro-apoptotic effect of DCA on PDGF-treated PASCs has not been thoroughly elucidated, and the effect of the Akt/glycogen synthase kinase-3 β (GSK-3 β) pathway inhibition concomitant with the effect of DCA on PASC proliferation remains unclear. The growth of human PASCs and the lactate concentration in extracellular medium of PASCs were detected by Cell Counting Kit-8 assays and a Lactate Colorimetric Assay kit, respectively. Cell apoptosis was evaluated by fluorescence activated cell sorting. The mitochondrial membrane potential ($\Delta\Psi_m$) was assessed with 5,5',6,6'-tetrachloro-1,1',3,3'-tetraethylbenzimidazol-carbocyanine iodide assays. The expression levels of phosphorylated Akt and GSK-3 β , pyruvate dehydrogenase, cleaved caspase-3, pyruvate dehydrogenase kinase-1 (PDK-1), hypoxia inducible factor-1 α (HIF-1 α) and hexokinase-2 (HK-2) were measured with western blot analysis. Confocal analyses were employed to determine HK-2 co-localisation with the mitochondria. The results indicated that DCA inhibited human PASC proliferation in a dose-dependent manner. DCA at 10 mM promoted apoptosis and the upregulation of activated caspase-3 in PASCs pre-treated with 20 ng/ml PDGF-homeodimer BB (BB). Treatment with 5 μ M LY294002 produced minimal anti-proliferative effects on human PASCs and barely induced cellular apoptosis and caspase-3 activation. However, co-administration of 10 mM DCA with LY294002 significantly decreased the cell proliferation index and induced cell apoptosis and caspase-3 activation. The combined administration of LY294002 with DCA significantly decreased lactate concentration, promoted the depolarisation of the $\Delta\Psi_m$ and repressed HIF-1 α upregulation and HK-2 activation in PASCs treated with PDGF, which was attributed to the potentiation of DCA-induced PDK-1 inhibition by LY294002 via blockade of the Akt/GSK-3 β /HIF-1 α signalling pathway. In conclusion, inhibition of the Akt/GSK-3 β pathway improved the pro-apoptotic effect of DCA on human PASCs, which may be attributed to a reversal of the Warburg effect by

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Abbreviations: DCA, dichloroacetate; PASCs, pulmonary arterial smooth muscle cells; HK-2, hexokinase-2; GSK-3 β , glycogen synthase kinase-3 β ; $\Delta\Psi_m$, mitochondria membrane potential; HIF-1 α , hypoxia inducible factor-1 α ; PAH, pulmonary arterial hypertension; PDH, pyruvate dehydrogenase; PDK, pyruvate dehydrogenase kinase; VDAC, voltage dependent anion channel; PI3K, phosphatidylinositol 3-kinase; FBS, foetal bovine serum; JC-1, 5,5',6,6'-tetrachloro-1,1',3,3'-tetraethylbenzimidazol-carbocyanine iodide

Key words: apoptosis, dichloroacetate, pyruvate dehydrogenase kinase-1, phosphatidylinositol 3-kinase/protein kinase B/glycogen synthase kinase-3 β , hypoxia inducible factor-1, hexokinase-2

blocking the mutual interaction between HIF-1 α and PDK-1, consequently downregulating HK-2. Therefore, combinatory treatment with DCA and PI3K inhibitors may represent a novel therapeutic strategy for the reversal of apoptosis resistance exhibited by PASMCs as a result of mitochondrial bioenergetic abnormalities, as well as the treatment of pulmonary vascular remodelling in pulmonary arterial hypertension.

Introduction

Hyperplasia and phenotypic remodelling of pulmonary arterial smooth muscle cells (PASMCs) serve key roles in the progressive narrowing and occlusion of the distal pulmonary arterioles in pulmonary arterial hypertension (PAH) (1,2). As in cancer cells, mitochondria from proliferating PASMCs may undergo diverse molecular changes and respond by switching their cellular metabolism towards cytoplasm-based glycolysis, a condition associated with apoptosis resistance (3,4). Previous studies indicated that platelet-derived growth factor (PDGF) expression was increased in neointimal lesions of rats treated with hypoxia or monocrotaline; the application of a tyrosine receptor inhibitor, imatinib, exhibited a beneficial effect in abrogating these pulmonary vasculature lesions (5,6). Previous results from Xiao *et al* (7) suggested that PDGF promoted the Warburg effect, which refers to enhanced glycolysis effects under normoxic conditions, and induced excessive proliferation and apoptosis resistance in PASMCs (8). The activation of the phosphatidylinositol 3-kinase (PI3K)/protein kinase B (Akt) signalling pathway accounted for this transition in metabolic phenotype (glucose oxygenation to glycolysis), as pharmacological ablation of PI3K with LY294002 facilitated the reversal of the Warburg effect (7).

Dichloroacetate (DCA), a selective inhibitor of pyruvate dehydrogenase kinase-1 (PDK-1), activates pyruvate dehydrogenase (PDH), which catalyses the decarboxylation of pyruvate into acetyl-CoA, promoting substrate entry into the Krebs cycle. DCA-mediated inhibition of PDKs triggers a switch in pyruvate metabolism towards glucose oxidation to CO₂ in mitochondria, along with a destabilisation of hypoxia-inducible factor-1 α (HIF-1 α), which reverses the hyper-polarised mitochondrial membrane potential ($\Delta\Psi_m$) and induces apoptosis of cancer cells (3). DCA also has beneficial effects on PAH due to its 'lung selective' characteristic and ameliorates pulmonary vascular remodelling in several rodent models of PAH (9-11). The reversal of HIF-1 α activation and the Warburg effect is considered to serve a pivotal role in mitigating the vascular lesions in rats subjected to carotid intimal balloon injury (12). Furthermore, the hypoxia response element is ubiquitously present in the promoter regions of genes encoding glycolytic enzymes, including PDK-1, glucose transporter-1 and hexokinase (13). Hexokinases (HKs) catalyse the first essential step in glucose metabolism by phosphorylation of glucose to glucose-6-phosphate. There are 4 major isoforms (HK-1, HK-2, HK-3, and HK-4) characterised in mammalian tissue (14). Among these, HK-2 has been extensively investigated and identified to be associated with a blocking of cytochrome *c* release through the induction of the pro-apoptotic proteins B-cell lymphoma 2 (Bcl-2)-associated X protein and BH3 interacting-domain death agonist, and protection of cancer cells from apoptosis (15). The interaction

between HIF-1 α and HK-2 in cancer cells is well-documented yet poorly exploited in vascular cells (16,17). A study by Lambert *et al* (12) indicated that the hyper-proliferating smooth muscle cells (SMCs) from injured carotid arteries exhibited activation of HIF-1 α and HK-2 and increased glycolysis. Functional suppression using small interfering RNA (siRNA) targeting HIF-1 α promoted SMC apoptosis through the inhibition of HK-2 expression and translocation of HK-2 with voltage dependent anion channel (VDAC) on the outer membrane of the mitochondria (12). Additionally, HK-2 was identified to be upregulated in neointimal lesion regions in pre-capillary pulmonary arterioles in a rat model of severe PAH, accompanied by a high expression of HIF-1 α (18). In addition to the transcriptional regulation of HK-2 by HIF-1 α , HK-2 expression is precisely controlled by multiple signalling pathways, particularly the PI3K/Akt/glycogen synthase kinase-3 β (GSK-3 β) axis (19). A study by Bonnet *et al* (20) indicated that dehydroepiandrosterone, a steroid hormone, decreased proliferation and induced apoptosis of vessel SMCs *in vivo* and *in vitro*, which was mediated by inhibition of Akt/GSK-3 β , interfering with the HK-2/VDAC interaction.

In the present study, whether PDK-1 and HK-2 inactivation was involved in DCA-induced growth retardation and whether the pro-apoptotic effect of DCA in PASMCs exposure to PDGF was potentiated by blocking the PI3K/Akt signalling pathway were investigated. The results of the present study revealed that DCA induced a pro-apoptotic effect on human PASMCs associated with the downregulation of PDK-1 and HIF-1 α and subsequent HK-2 inactivation via blocking of the mitochondrial interaction. Furthermore, this effect may be potentiated via the suppression of PDK-1 by blocking the Akt/GSK-3 β /HIF-1 α signalling pathway.

Materials and methods

Ethics approval. The present study was approved by the Ethics Committee of Nanjing University Drum Tower Hospital for Institutional Animal Care and Use.

Cell culture. Human PASMCs were purchased from Wuxi BioHermes Biomedical Technology Co., Ltd (Wuxi, China). They were trypsinised and subcultured subsequent to reaching 90% confluence. A monoclonal antibody against smooth muscle α -actin (1:1,000; cat. no. M085129-2; Agilent Technologies, Inc., Santa Clara, CA, USA) was used to assess the purity (>99%) of the SMC culture according to the manufacturer's protocol. Unless otherwise indicated, primary cultures of PASMCs were maintained in RPMI-1640 containing 10% foetal bovine serum (FBS) and 1% antibiotics (penicillin and streptomycin mixture), all purchased from Thermo Fisher Scientific Inc. (Waltham, MA, USA).

All experiments examining HK-2, HIF-1 α activation, Akt, GSK-3 β , PDH phosphorylation and PDH kinase-1 (PDK-1) expression, and cell viability analysis, were performed on cells that were cultured for 3 days at passages 3-5. Following this, cells were trypsinised and seeded at a distinct cell density into 96-well plates (5,000 cells/well) or culture flasks (5 \times 10⁵ cells/flask) and subcultured in RPMI-1640 containing 10% FBS and 1% antibiotics in a humidified 5% CO₂ atmosphere at 37°C overnight. Cells were then exposed to

PDGF-BB (Cytolab Ltd., Rehovot, Israel), SB216763 (S1075; Selleck Chemicals, Houston, TX, USA), DCA (5, 10, 20 and 50 mM) and LY294002 (Sigma-Aldrich; Merck KGaA, Darmstadt, Germany) at numerous concentrations (5, 10 and 20 μ M) or a combination of two drugs (10 mM DCA and 5 μ M LY294002) as indicated. In the control groups, an equal volume of PBS was substituted for the reagents.

Determination of cell viability [Cell-Counting-Kit 8 (CCK-8) colorimetric assay]. Cells were seeded at 5,000 cells/well into 96-well plates and grown in RPMI-1640 supplemented with 10% FBS for 72 h. After 48 h of serum starving with 0.5% FBS, the cells were then incubated with 20 ng/ml PDGF, followed by increasing concentrations of DCA (5, 10, 20 and 50 μ M), LY294002 or a combination of the two drugs for 72 h. Following drug exposure, cells were incubated in a humidified 5% CO₂ atmosphere at 37°C with 10 μ l CCK-8 (Dojindo Molecular Technologies, Inc., Kumamoto, Japan) for 1 h. The optical densities in the 96-well plates were determined using a microplate reader at a wavelength of 450 nm. The cell proliferation percentage was determined in the treated (AT), PDGF (Ap), blank (Ab) groups using the following formula: Proliferation Index = (AT-Ab/Ap-Ab) x 100; optical density was measured at a 450 nm wavelength.

Cell apoptosis and 5,5',6,6'-tetrachloro-1,1',3,3'-tetraethylbenzimidazol-carbocyanine iodide (JC-1) assays. Cells were seeded in 25 cm² tissue culture flasks at 5x10⁵ cells/flask and incubated in standard conditions. Cells were maintained in complete medium for 16 h, followed by serum starvation for 24 h prior to treatment with 20 ng/ml PDGF with 5 μ M LY294002, DCA at 10 mM, a combination of LY294002 and 10 mM DCA or alone for 48 h. Following drug treatment, PSMCs were harvested and centrifuged at 4°C at 600 x g for 5 min; the pellets were washed twice with PBS and then resuspended in 100 μ l Annexin V binding buffer (0.14 M NaCl, 2.5 mM CaCl₂, 0.01 M HEPES pH 7.4). Annexin V (1 μ l; Invitrogen; Thermo Fisher Scientific, Inc.) and 5 μ l propidium iodide (50 μ g/ml) were added to the samples and incubated at room temperature in the dark for 15 min. Samples were kept on ice following incubation until fluorescence activated cell sorting (FACS) analysis was performed. For the assays measuring mitochondrial membrane potential ($\Delta\Psi$ m), the cells were treated as indicated above, and then 1 μ l JC-1 (Beyotime Institute of Biotechnology, Haimen, China) was added in the medium at a final concentration of 2 μ M, and the cells were stained at room temperature for 30 min in the dark. Subsequent to JC-1 staining, cells were trypsinised, washed twice with PBS and resuspended in 100 μ l PBS. JC-1 fluorescence intensity was examined using FACS and data was analysed with FlowJo 7.61 software (Tree Star Inc., Ashland, OR, USA).

Lactate measurement. The PSMCs were seeded into 25 cm² tissue culture flasks at a cell density of 5x10⁵ cell/flask and cultured with RPMI-1640 complete culture medium for 16 h followed by serum starvation with 0.5% FBS for 24 h. The cells were then incubated at 37°C with 20 ng/ml PDGF with 5 μ M LY294002, DCA at 10 or 20 mM, a combination of LY294002 and 10 mM DCA, or alone for 48 h, subsequent to

which the medium was removed from cells and lactate levels in the extracellular medium were measured at room temperature using the Lactate Colorimetric Assay kit (Abcam, Cambridge, MA, USA) according to the manufacturer's protocol. Lactate concentration was normalised to sample cell number.

Immunoblotting and densitometric analysis. The PSMCs were seeded into T25 culture flasks at a cell density of 1x10⁶ and cultured with RPMI-1640 complete culture medium for 16 h followed by serum starvation with 0.5% FBS for 24 h. Thereafter, the cells were incubated under different interventions for 24 h, as aforementioned, while PBS was substituted for reagents in control cells cultured in medium with 0.5% FBS. Immunoblotting was performed as described previously (18). Briefly, PSMCs were harvested and resuspended in ice-cold cell lysis solution (Thermo Fisher Scientific, Inc.) and the homogenate was centrifuged at 400 x g for 15 min at 4°C. Protein concentration was determined using a Bicinchoninic Acid Protein Assay kit (cat. no. PICPI23223; Thermo Fisher Scientific, Inc.). The supernatant was subsequently transferred to a fresh tube and 75 μ g protein from each sample was then separated via 12% SDS-PAGE. Following this, the proteins were then transferred to polyvinylidene difluoride membranes. Membranes were then blocked using 5% fat-free milk for 1 h at room temperature and subsequently incubated at 4°C overnight with the following primary antibodies specific to Akt, phosphorylated (p)-Akt, GSK3 β , p-GSK3 β (Cell Signaling Technology, Inc., Danvers, MA, USA; cat. nos., 4685, 4060, 9315 and 9313, respectively), HK-2, PDK-1 (1:1,000; Cell Signaling Technology, Inc.; cat. nos., 2867 and 3820), phospho(S293)-PDH E1 α subunit (1:1,000; Abcam, Cambridge, UK; cat. no., ab177461), HIF-1 α (1:500; Novus Biologicals, LLC, Littleton, CO, USA; cat. no., NB100-449) and β -actin (1:2,000; Santa Cruz Biotechnology, Inc., Dallas, TX, USA; cat. no., sc-47778). Following this, membranes were incubated for 2 h at room temperature with the following secondary antibodies: Horseradish peroxidase-conjugated goat anti-rabbit IgG (cat. no. ab6721; 1:5,000; Abcam) or HRP-conjugated anti-mouse IgG (cat. no. 5450-0011; 1:5,000; KPL, Gaithersburg, MD, USA). β -actin protein expression served as an internal control and was used to normalise the protein band intensity. To determine the phosphorylation levels of AKT and GSK-3 β , membranes were washed with stripping buffer (cat. no. P0025S; Beyotime Institute of Biotechnology) at 50°C for 30 min, followed by blocking the membrane with 5% bovine serum albumin (cat. no. 37525; Thermo Fisher Scientific Inc.) in PBST at room temperature for 4 h. The membrane was then re-probed at 4°C overnight with p-AKT (1:1,000) and p-GSK-3 β antibodies (1:1,000; Cell Signalling Technology, Inc., Danvers, MA, USA). Western blots were detected using the Western chemiluminescent detection system (Pierce; Thermo Fisher Scientific, Inc.) and exposure to X-ray film. Images were acquired using a CanoScan 8600F flatbed scanner, quantified using ImageJ software (version 1.4; National Institutes of Health, Bethesda, MD, USA) and standardised to β -actin in each lane. Experiments were repeated in triplicate.

Immunofluorescence confocal microscopy. Human PSMCs were fixed with 1% paraformaldehyde and permeabilised

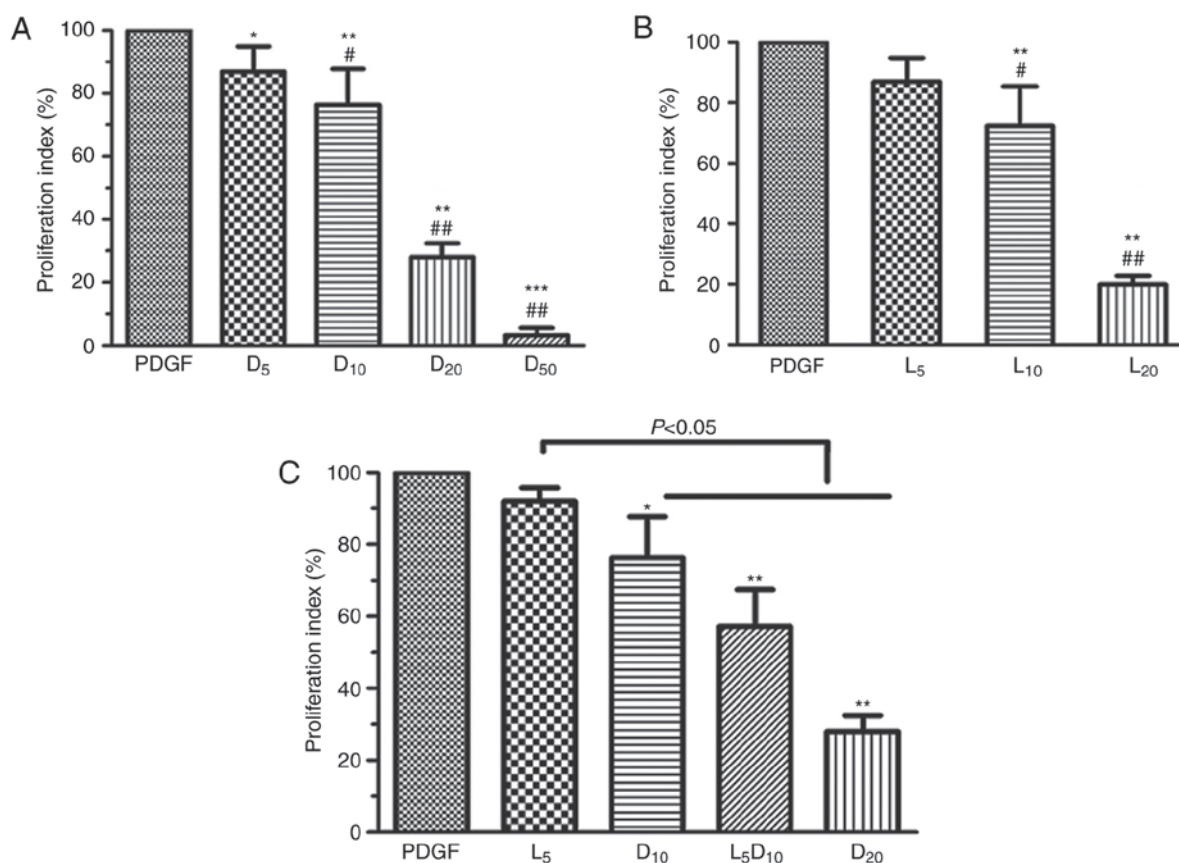


Figure 1. Effect of DCA, LY294002 or combination of DCA and LY294002 on the growth of human PSMCs. The human PSMCs were seeded in 96-well plates in RPMI-1640 medium supplemented with 10% FBS followed by 48 h serum starving prior to exposure to 20 ng/ml PDGF or with increased concentrations of (A) DCA or (B) LY294002 in fresh culture medium with 0.5% foetal bovine serum for 72 h. * $P < 0.05$, ** $P < 0.01$ and *** $P < 0.001$ vs. PDGF-treated cells; # $P < 0.05$ and ## $P < 0.01$ vs. cells treated with 5 mM DCA or 5 μ M LY294002. (C) Cells were exposed to PDGF or 5 μ M LY294002, 10 or 20 mM DCA and combination of 5 μ M LY294002 and DCA (10 mM) for 24 h. * $P < 0.05$, ** $P < 0.01$ as compared with PDGF cells. The data are presented as the mean \pm standard deviation of 6 duplicated wells in three separate experiments. PSMCs, pulmonary arterial smooth muscle cells; DCA, dichloroacetate; PDGF, platelet-derived growth factor; D₅, D₁₀, D₂₀ and D₅₀, cells treated with dichloroacetate at 5, 10, 20, 50 mM, respectively, following PDGF exposure. L₅, 10 and 20, cells treated with LY294002 at 5, 10, 20 μ M, respectively, following PDGF exposure. L₅D₁₀, cells treated in a combination of 5 μ M LY294002 and 10 mM dichloroacetate following PDGF exposure.

with 0.2% Triton X-100. The $\Delta\Psi_m$ was determined using MitoTracker® Red CMXRos (Invitrogen; Thermo Fisher Scientific, Inc.). The PSMCs were then incubated at 37°C with M7512- Mito tracker Red (50 nM) for 30 min and maintained in a dark room at 4°C overnight for confocal detection. HK-2 detection was performed using Alexa Fluor 488 (10 μ g/ml; Invitrogen; Thermo Fisher Scientific, Inc.) for green fluorescent protein staining at 37°C for 1.5 h. Cell imaging was performed using an FV1000 confocal microscope (magnification, x100) equipped with a live cell apparatus (Olympus Corporation, Tokyo, Japan).

Statistical analysis. Data are presented as the mean \pm standard deviation. Calculations were performed using the GraphPad Prism software package (version 4.0.1; GraphPad Software, Inc., La Jolla, CA, USA), and one-way analysis of variance with Tukey's post-hoc test was applied. $P < 0.05$ was considered to indicate a statistically significant difference.

Results

DCA inhibits the proliferation of human PSMCs stimulated with PDGF-BB in a dose-dependent manner, which was

improved by co-incubation with LY294002. As demonstrated in Fig. 1A, DCA suppressed human PSMC proliferation in a dose-dependent manner (0–50 mM). DCA at 5 mM exhibited minimal inhibitory effects on PSMC proliferation after 72 h incubation. The cell proliferation index was significantly decreased in the 10, 20 and 50 mM groups compared with the 5 mM DCA group ($P < 0.01$). The inhibitory effect of DCA on the growth of human PSMCs was mimicked by LY294002. LY294002 (10 μ M) caused a 27.7% decrease in cell proliferation rate, whereas only a 13.0% decrease was observed after 72 h of treatment with 5 μ M LY294002 ($P > 0.05$; Fig. 1B). Notably, the cell proliferation index was significantly decreased to 57.2% after 72 h incubation with a combination of DCA (10 mM) and LY294002 (5 μ M) compared with the cells treated by DCA alone (76.4%; $P < 0.05$; Fig. 1C).

Growth inhibitory effect of DCA on PSMCs potentiated by LY294002 may be attributed to increased mitochondria-associated apoptosis. As indicated in Fig. 2A, the percentage of apoptotic cells was ~18.8% following treatment with 10 mM DCA, which was significantly increased compared with cells treated with 5 μ M LY294002 alone (9.2%; $P < 0.05$). However, the combined treatment with LY294002 and 10 mM DCA

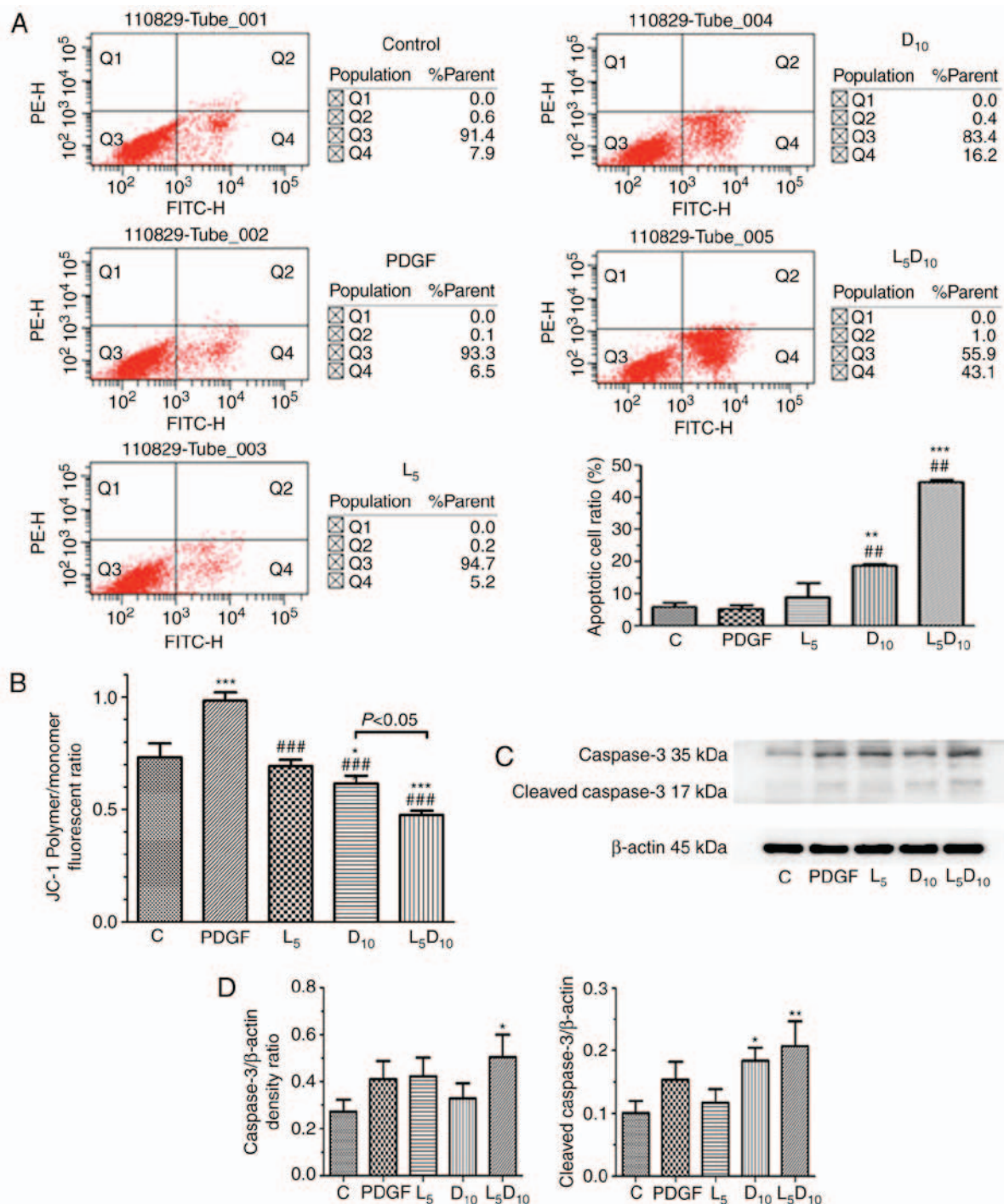


Figure 2. Effect of DCA, LY294002 or combination of DCA and LY294002 on the apoptosis and mitochondria membrane potential of human PASMCs. The PASMCs were seeded into 25 cm² tissue culture flask at a density of 5×10⁵ cell/flask and cultured in RPMI-1640 complete culture medium for 16 h followed by serum starvation for 24 h. The cells were then exposed to PDGF alone or 5 μM LY294002, DCA at 10 mM or a combination of 5 μM LY294002 and 10 mM DCA for 48 h prior to (A) apoptosis or (B) JC-1 assay. (C) The expression levels of caspase-3 and cleaved caspase-3 were analyzed with western blot analysis. The representative change of one of the three experiments is presented, as all assays exhibited identical results. (D) Results were pooled from three separate experiments and are presented as mean ± standard deviation. *P<0.05, **P<0.01 and ***P<0.001 vs. control cells. ##P<0.01 and ###P<0.001 vs. cells treated with PDGF. C, control; PASMCs, pulmonary arterial smooth muscle cells; DCA, dichloroacetate; PDGF, platelet-derived growth factor; D5 and D10, cells treated with DCA at 5 and 10 mM, respectively, following PDGF exposure; L5, cells treated with LY294002 at 5 μM following PDGF exposure; L5D10, cells treated with a combination of 5 μM LY294002 and 10 mM DCA following PDGF exposure.

markedly increased the apoptotic cell percentage to 44.9% following 48 h incubation. As demonstrated in Fig. 2C and D, although 5 μM LY294002 only marginally affected the expression of cleaved caspase-3, it significantly increased cleaved caspase-3 expression levels when added in combination with

10 mM DCA. The human PASMCs exhibited an increased ΔΨ_m, following PDGF incubation for 48 h compared with the control cells (P<0.05), as expressed by the ratio of red and blue fluorescence intensity following JC-1 staining in Fig. 2B. DCA at 10 mM or LY294002 significantly decreased the ΔΨ_m

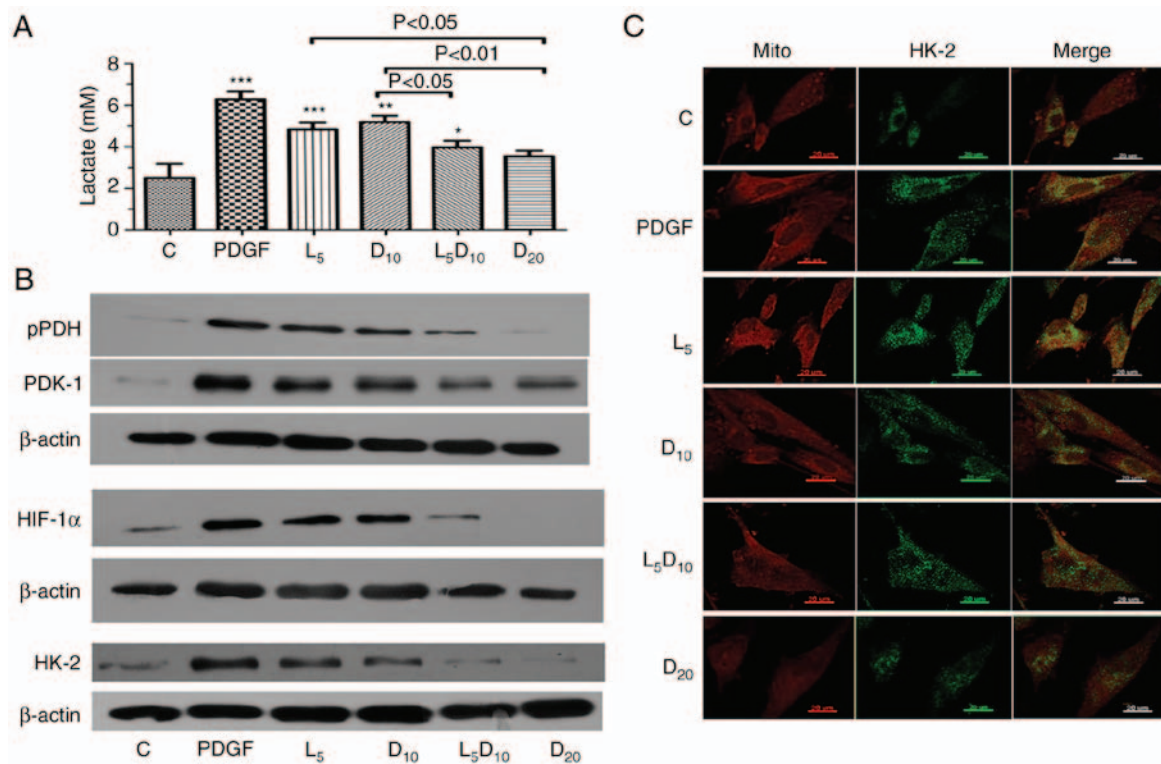


Figure 3. Effect of DCA, LY294002 or combination of DCA and LY294002 on extracellular lactate concentration, key glycolysis-associated enzymes expression and HK-2 activation. The PSMCs were seeded into 25 cm² tissue culture flask at a density of 5x10⁵ cell/flask and cultured with RPMI-1640 complete culture medium for 16 h followed by serum starvation for 24 h. (A) The cells were then exposed to PDGF or 5 μ M LY294002, DCA at 10 or 20 mM, and a combination of LY294002 and 10 mM DCA for 48 h prior to measurement of lactate concentration. *P<0.05, **P<0.01 and ***P<0.001 vs. control cells. (B) Cell lysates were extracted for PDK-1, PDH, HIF-1 α and HK-2 expression analysis using western blot analysis. The representative change of one of the three experiments is presented, as all assays exhibited identical results. (C) HK-2 activation was analyzed with immune-fluorescent confocal microscopy. PDGF-stimulated PSMCs exhibited significant co-localization between HK-2 (green) and the mitochondria (red), giving a yellow pattern in the merged images. The cells treated with 20 mM DCA and the combination of LY294002 and 10 mM DCA demonstrated diffuse cytoplasmic staining of HK-2 (no colocalization of HK-2 to the mitochondria). C, control; DCA, dichloroacetate; PDGF, platelet-derived growth factor; L₅, D₁₀ and D₂₀, cells treated with DCA at 5, 10 and 20 mM, respectively, following PDGF exposure; L₅, cells treated with LY294002 at 5 μ M following PDGF exposure; L₅D₁₀, cells treated in combination of 5 μ M LY294002 and 10 mM dichloroacetate following PDGF exposure; PSMCs, pulmonary arterial smooth muscle cells; HK-2, hexokinase-2; PDK-1, pyruvate dehydrogenase kinase-1; PDH, pyruvate dehydrogenase; HIF-1 α , hypoxia-inducible factor-1 α ; mito, mitochondria.

in PSMCs. Furthermore, the $\Delta\Psi_m$ was markedly decreased after 48 h of treatment with a combination of 10 mM DCA and LY294002 compared with DCA or LY294002 alone (P<0.05).

Co-treatment with LY294002 and DCA decreases lactate concentration in extracellular culture medium, PDK-1, p-PDH, HIF-1 α and HK-2 expression and potentially represses HK-2 activation, which are all associated with a reversal of the Warburg effect. As suggested in Fig. 3A, DCA at 10 mM or 5 μ M LY294002 significantly decreased the lactate concentration in PSMCs stimulated with PDGF-BB. Additionally, the increased concentration of DCA (20 mM) markedly decreased PDGF-induced lactate production (3.55 vs. 6.28 mM, respectively; P<0.01). The lactate concentration markedly decreased after 48 h of treatment with the combination of 10 mM DCA and LY294002 compared to 10 mM DCA alone (3.97 vs. 5.18 mM; P<0.05). Furthermore, the upregulation of PDK-1, p-PDH and HIF-1 α induced by PDGF was abrogated by treatment with 10 mM DCA in PSMCs. The inhibitory effect on PDK-1, p-PDH and HIF-1 α expression in PSMCs was improved by a combination of 10 mM DCA with LY294002. The protein expression of HK-2 increased significantly following PDGF treatment compared with the

control cells. Treatment with 5 μ M LY294002 or 10 mM DCA significantly decreased HK-2 expression. The combination of LY294002 and DCA or 20 mM of DCA elicited an additional decrease in HK-2 expression compared with the cells treated with 10 mM DCA (Fig. 3B). In addition, the inhibitory effect of DCA or LY294002 resulted in an uncoupling of HK-2 from the mitochondria. The increased co-localisation of HK-2 with mitochondria in the cytoplasm following PDGF-BB stimulation was significantly attenuated by 10 and 20 mM DCA treatment. Notably, LY294002 increased the dislocation of HK-2 from the mitochondria induced by 10 mM DCA, which is demonstrated clearly in Fig. 3C as a visible decrease in the yellow fluorescence intensity in the cytoplasm.

Downregulation of PDK-1 through Akt/GSK-3 β signal blocking may be involved in the potentiation of DCA-induced human PSMCs apoptosis by LY294002. As indicated in Fig. 4A-D, LY294002 inhibited Akt/GSK-3 β axis activation in PSMCs exposed to PDGF-BB in a concentration-dependent manner (1–20 μ M). The PDK-1 upregulation induced by PDGF in PSMCs was apparently inhibited by LY294002 in a concentration-dependent manner (1–20 μ M), as demonstrated in Fig. 4E and F (P<0.05). To investigate the role of the Akt/GSK-3 β axis

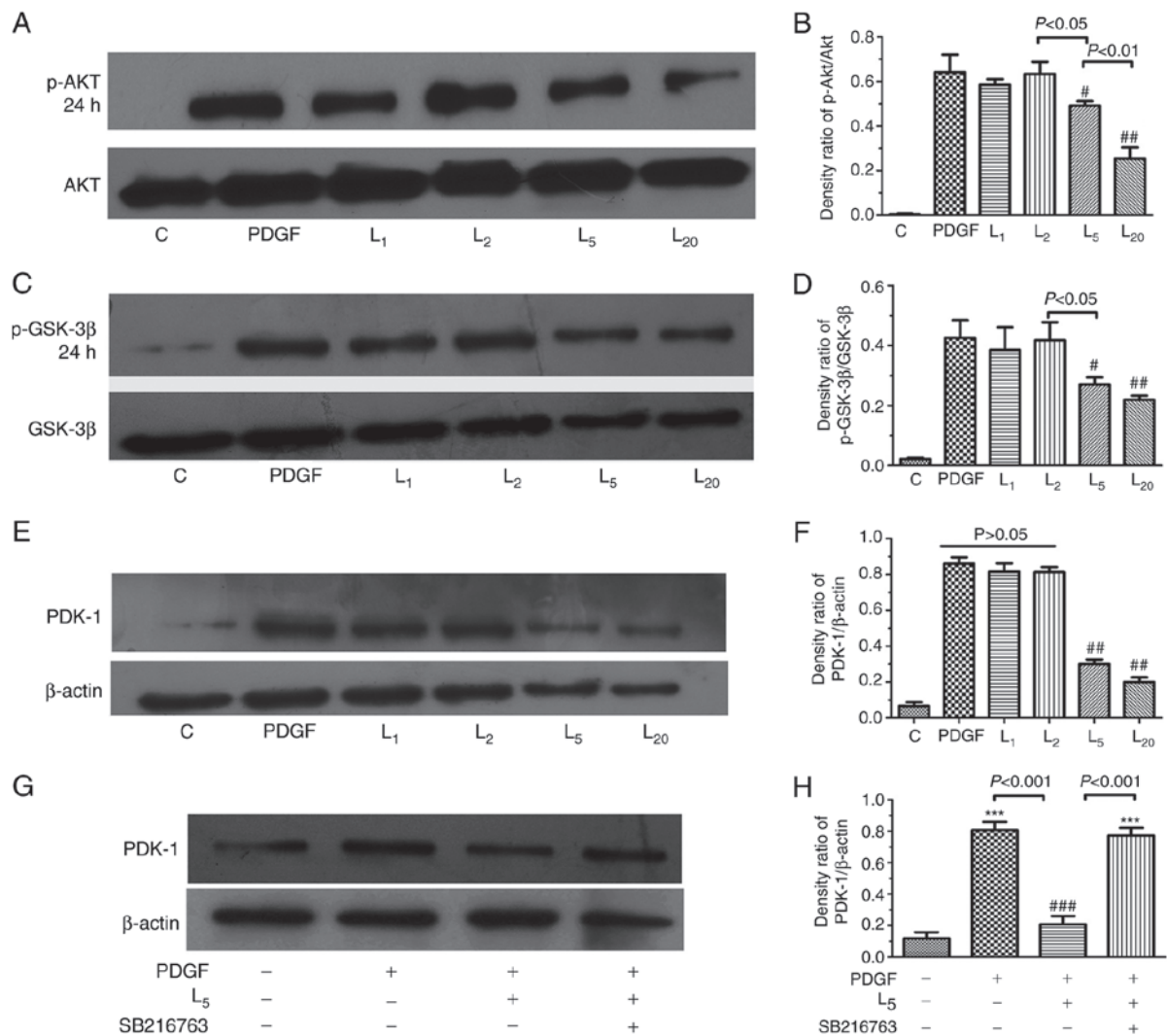


Figure 4. Effect of LY294002 on Akt/ GSK-3 β /PDK-1 signaling and the inhibition of GSK-3 β activity on PDK-1 expression. The cells were exposed to PDGF, or with increasing LY294002 concentrations of 1, 2, 5 and 20 μ M for 24 h, prior to detection of the expression of (A and B) p-Akt, (C and D) GSK-3 β or (E and F) PDK-1. (G and H) The cells were treated with PDGF, or with 5 μ M LY294002 with/without 30 min pre-exposure to 5 μ M SB216763 for 24 h, prior to the detection of PDK-1 expression. [#] $P < 0.05$ and ^{##} $P < 0.01$ vs. cells treated with PDGF. ^{***} $P < 0.001$ vs. control cells. Results were pooled from three separate experiments and are presented as mean \pm standard deviation. A, C and E are the representative change of p-Akt or p-GSK-3 β and PDK-1 expression, respectively. The representative change of one of the three experiments is presented, as all assays exhibited identical results. Semi-quantitative analyses of p-Akt, p-GSK-3 β and PDK-1 expression levels are presented in B, D, F and H, respectively. C, control; PDGF, platelet-derived growth factor; L₁, L₂, L₅ and L₁₀, cells treated with LY294002 at 1, 2, 5 and 20 μ M following PDGF exposure; Akt, protein kinase B; GSK-3 β , glycogen synthase kinase-3 β ; PDK-1, pyruvate dehydrogenase kinase-1; p-, phosphorylated.

in the downregulation of PDK-1 by LY294002, PASMCs were treated with a GSK-3 β inhibitor (5 μ M SB216763) for 30 min prior to PDGF-BB and 5 μ M LY294002 exposure for 24 h. The inhibitory effect of LY294002 on PDK-1 was completely abrogated by SB216763 (Fig. 4G-H).

Discussion

The hyperplasia of PASMCs within the vascular wall of distal pulmonary arterioles, causing medial hypertrophy and neointimal lesion formation, is a hallmark of pulmonary vascular remodelling in PAH (2). Abnormality in energy metabolism is one of the contributing factors that results in aberrant growth of PASMCs (21,22). In accordance with previous studies in cancer cell lines (23,24), DCA in the present study inhibited human PASMC proliferation stimulated by

PDGF in a dose-dependent manner (0-50 mM). Reversal of the Warburg effect by downregulating PDK-1 and HIF-1 α and inactivation of HK-2 may account for the repression of the aberrant growth of human PASMCs treated by DCA. Notably, the pro-apoptotic effect of DCA may also be potentiated by the inhibition of PDK-1, PDH phosphorylation and HK-2 inactivity through blocking Akt/GSK-3 β /HIF-1 α signalling with a PI3K inhibitor.

Accumulating evidence has suggested that PDGF was highly expressed in the neointimal lesions of pulmonary arterioles from rats subjected to monocrotaline or hypoxia insult (5,25). Targeting of the PDGF receptor using imatinib exhibited beneficial effects on PASMC proliferation and mitigated the neointimal formation of the affected pulmonary arterioles. Repression of PDGF receptor phosphorylation and subsequent mitogen activated protein kinase activation underlies the alleviation of PAH

by imatinib (5), but the precise mechanism remains unclear. A previous study from Xiao *et al* (7) provided novel insights into the current understanding of PDGF-induced PASMOC proliferation. Consistent with the mitochondrial metabolic shifting paradigm in cancer (26), PDGF induced increased expression of a glycolysis-associated gate-keeper enzyme PDK-1, glucose transporter-1 and HIF-1 α , preferentially converting pyruvate to lactate in PASMOCs (7). DCA selectively inhibits PDK activity, thereby indirectly inactivating PDH and shifting metabolism to mitochondrial glucose oxygenation, thereby reversing apoptotic resistance; this change has been demonstrated to be important in attenuating PASMOC hyperplasia in several rodent models of PAH (9,27). Consistent with these studies, the results of the present study suggested that a high dose of DCA facilitated the reversal of PASMOC glycolytic status, which was confirmed by decreased lactate content in culture medium. Along with rectifying this abnormal glycolysis metabolism phenotype, DCA treatment promoted apoptotic sensitivity and inhibited proliferation of PASMOCs.

One notable result from the present study was that the inhibitory effect of DCA on PASMOC growth was potentiated by blocking the PI3K/Akt signalling pathway. Although 5 μ M LY294002 exhibited a negligible effect on cell apoptosis, it increased caspase-3 levels and promoted apoptosis in PASMOCs treated with 10 mM DCA. Previous studies demonstrated that the median lethal dose of DCA causing growth retardation in human cancer cell lines was 5-20 mM (28-30). DCA has a relatively wide therapeutic margin, and few adverse reactions have been described, even following long-term ingestion, with the exception of a few case reports of peripheral nerve lesions (31). However, a previous study suggested that the incubation of rat sciatic nerves with 20 mM DCA caused a 15% decrease in the amplitude of the compound action potential, while 10 mM DCA had no effect on the nerve transduction (32). Therefore, combined pharmaceutical intervention is recommended to avoid a potential adverse reaction due to overdose by single drug use, as multiple studies have indicated that the anti-tumour effect of DCA may be improved by sensitising the pro-apoptotic signalling pathway in cancer cell lines (33,34).

The results of the present study indicated that the reversal of the Warburg effect mediated through the downregulation of HIF-1 α and inactivation of HK-2 may account for the increased PASMOC apoptosis following DCA intervention. DCA is a well-characterised inhibitor of the protein kinase PDH. DCA-mediated inhibition of PDKs triggers a switch in pyruvate metabolism towards glucose oxidation to CO₂ in the mitochondria; during the process of electron transport in the Krebs cycle, the intermediate substrates (mitochondrial reactive oxygen species or α -ketoglutarate) serve key roles in the destabilisation of HIF-1 α , which also reverses hyper-polarised $\Delta\Psi_m$ and induces apoptosis of cancer cells (3). HIF-1 α upregulation was also demonstrated by Lambert *et al* (12), in a rodent model with carotid arterial neointimal formation induced by balloon injury. Targeting HIF-1 α using siRNA effectively reversed the aerobic glycolysis of SMCs and mitigated the neointimal lesions in diseased carotid arteries. In Fawn-Hooded rats, DCA inhibited proliferation of PASMOCs by reversing the mitochondrial metabolic phenotype and inactivation of HIF-1 α and downstream voltage-gated potassium channel subtype 1.5 (27). HK-2 has been recognised

as one of the key molecules in cell proliferation, differentiation and metastasis during carcinogenesis: Inhibition of HK-2 using 2-DG or siRNA exhibited beneficial effects on tumour growth in a xenograft model (35,36). However, the role of HK-2 in DCA-induced PASMOC apoptosis remains unclear. Previous studies indicated that the HK-2 gene promoter comprising a 5'-RCGTG-3' box known as the hypoxia response element was under precise transcriptional regulation by HIF-1 α (37). HIF-1 α activation was demonstrated to serve a critical role in the subsequent upregulation of HK-2 in diverse cancer cells, but has rarely been described in vascular cells, particularly in PASMOCs. The present study indicated that with the downregulation of HIF-1 α following DCA treatment, the expression and activity of HK-2 decreased. Therefore, the present results suggested that HIF-1 α induced HK-2 activation in the apoptotic-resistant PASMOCs. Consistent with previous studies (12,20), these data indicated that in addition to HK-2 downregulation, the dislocation of HK-2 from VDAC on the mitochondrial outer membrane was involved in cell apoptosis following DCA treatment, which was associated with an efflux of cytochrome *c* into the cytoplasm and caspase-3 activation. However, although there was no significant difference in the expression levels of cleaved caspase-3 in cells treated with 10 mM DCA alone or a combination of LY294002 and DCA, the number apoptotic cells was markedly increased in the combined treatment group. We hypothesize that the activation of PI3K/Akt signalling is associated with the cytosolic sequestration of the apoptotic protein B-cell lymphoma 2 (Bcl-2)-associated death protein (BAD); therefore, pharmacologic ablation of PI3K facilitates BAD translocation with B-cell lymphoma-extra large on the outer membrane of the mitochondria, which activates caspase-9 and -3 and causes the opening of the mitochondrial membrane transit pore (MTP). In addition to the release of cytochrome *c* following the MTP opening, the apoptotic inducing factor may also be released into the cytoplasm. The latter may initiate the non-caspase-3-dependent apoptosis pathway.

PDK-1 has been proposed as a key regulator of the Warburg effect and inhibits acetyl-CoA entry into the Krebs cycle, accompanied with decreased ketoglutarate synthesis, leading to decreased HIF-1 α proteasomal degradation (38). DCA is a pyruvate mimetic that targets PDKs, inhibiting the phosphorylation of PDH (31). Therefore, we hypothesised that inhibiting the activity of PDK-1 promoted the degradation of HIF-1 α and rectified the pseudo-hypoxic status in mitochondria of PASMOCs exposed to PDGF followed by DCA treatment. In addition, the results of the present study suggested that DCA inhibited PDK-1 expression in addition to downregulating PDK-1 activity. The mechanism of the DCA-induced decrease in PDK-1 is poorly delineated, and we postulate that PDK-1 activation induces a pseudo-hypoxic state in mitochondria and HIF-1 α activation, while HIF-1 α inhibits the tricarboxylic acid cycle by directly activating the gene encoding PDK-1, which initiates a positive regulatory loop. The results of the present study suggested that DCA inhibits PDK-1 activity and disrupts the vicious cycle among aerobic glycolysis-associated molecules, accounting for the decreased expression of PDK-1 in PASMOCs. A previous study indicated that the activation of PDGF receptor increased PDK-1 activity, mitochondrial respiration and cell proliferation (7); however, the effect of

PI3K suppression on PDK-1 and its target molecule (PDH) in growth factor receptor tyrosine kinase-induced glycolytic flux remains unclear (39). Several previous data indicate that the PI3K/Akt/HIF-1 α axis serves an essential role in PDGF-induced vascular SMC proliferation mediated through enhanced glycolytic flux (7,12). Inhibition of PI3K decreases extracellular acidification rates through HIF-1 α downregulation in vascular SMCs stimulated with growth factors (40). PDK-1 downregulation with LY294002 was abrogated by the GSK-3 β inhibitor in the present study; therefore, the downregulation of HIF-1 α by GSK-3 β signal blockade is the key factor responsible for disrupting the mutual positive feedback regulation between HIF-1 α and PDK-1 when co-incubated with DCA. In contrast to the results of the present study, PI3K blockade with LY294002 facilitated the phosphorylation of PDH at the E1 α -subunit (inactivation of its function) and decreased the oxygen consumption rate in SQ20B cells (41). Therefore, the role of PI3K/Akt activation in regulation of downstream signals of glycolysis-associated enzymes varies among diverse cells.

There are certain limitations in the present study to be addressed: Firstly, with the exception of GSK-3 β , other downstream molecules of Akt, including mechanistic target of rapamycin, may be affected following PI3K/Akt signal blocking (42), which is closely associated with cell survival and autophagy, and may have potential effects on PSMCs apoptosis. Therefore, the possibility that other signal molecules may be involved in the DCA-induced decrease in cell growth cannot not be excluded. Secondly, LY-294002 may also affect cell cycle proteins, including cyclin-dependent kinases, and DCA was indicated to inhibit cell cycle progression, had no association with cell apoptosis and inhibited cell hyperplasia in certain cell lines (43). Therefore, the synergistic effect of DCA and PI3K blockade on PSMC growth may be mediated by cell cycle suppression, and additional studies are warranted.

Collectively, the data in the present study indicated that the pro-apoptotic effect of DCA on human PSMCs was associated with PDK-1 and HIF-1 α downregulation, which led to HK-2 inactivation by blocking the interaction with mitochondria. Furthermore, this effect may be potentiated by the suppression of PDK-1 via blockade of Akt/GSK-3 β /HIF-1 α signalling. These results suggest that the inhibition of Akt/GSK-3 β signalling may improve the pro-apoptotic effect of DCA on human PSMCs, which suggests that combined therapy may be used in the treatment of PAH and may decrease the probability of peripheral neuropathic lesions or liver damage following long-term ingestion of high doses of DCA.

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Availability of data and materials

All data generated or analyzed during this study are included in this published article.

Authors' contributions

BL and JY contributed to experimental design, performed experiments, data analysis and wrote the final manuscript. YZ, QS, LC, YT and CY performed experiments and contributed to preparation of the manuscript. All authors read and approved the final manuscript.

Ethics approval and consent to participate

The approval of this study was obtained from the ethic committee of Drum tower Hospital of Nanjing University Medical School for Laboratory Animal Use and Care.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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