Abstract. Telomeres are nucleotide tandem repeats located at the tip of eukaryotic chromosomes that maintain genomic integrity. The gradual shortening of telomeres leads to cellular senescence and apoptosis, a key mechanism of aging and age-related chronic diseases. Epigenetic factors, such as nutrition, exercise and tobacco can affect the rate at which telomeres shorten and can modify the risk of developing chronic diseases. In this study, we evaluated the effects of a combination of nutraceutical supplements (NS) on telomere length (TL) in healthy volunteers with no medical history of any disease. Participants (n=47) were selected from healthy outpatients visiting a private clinic and were divided into the experimental group (n=16), that received the NS and the control group (n=31). We estimated the length of single telomeres in metaphase spread leukocytes, isolated from peripheral blood, using quantitative-fluorescent in situ hybridization (Q-FISH) analysis. The length of the whole telomere genome was significantly increased (P<0.05) for the mean, 1st quartile and median measurements in the experimental group. Similar findings were observed for short TL (20th percentile) (P<0.05) for the median and 3rd quartile measurements in the NS group, compared to the control group. The beneficial effects of the supplements on the length of short telomeres remained significant (P<0.05) following adjustment for age and sex. Telomeres were moderately longer in female patients compared to the male patients. On the whole, the findings of this study suggest that the administration of NS may be linked to sustaining the TL.

Introduction

Telomeres are DNA regions positioned at the tips of chromosomes. They are composed of repetitive nucleotide sequences (5'-TTAGGG-3'). These functional non-coding repeats protect the ends of eukaryotic chromosomes from attrition and damage. Telomeres shorten during mitotic cell division due to the inability of DNA polymerase to fully replicate the end part of the lagging strand of DNA. This is commonly referred to as the 'end-replication problem'. Telomere length (TL) gradually decreases, by approximately 20-40 base pairs per year (1). The shortening of telomeres can be reversed by the enzyme telomerase, which is active in high-proliferating cells, such as male germ cells, activated lymphocytes, stem cells and cancer cells (2,3). In most adult human somatic cells though, the expression of telomerase is deficient, and TL decreases with age (1). Telomerase activity has been associated with pregnancy complications (4,5). Furthermore, attention has been paid to telomere shortening...
in various diseases and age-related diseases, such as cancer, cardiovascular diseases (CAD), diabetes, rheumatoid arthritis and psychiatric disorders (6-10). In addition, TL is a biomarker of the replicative past and the replicative potential of somatic cells. Previous studies have focused on the estimation of mean TL (MTL) in age-related diseases (11,12); however, current concepts highlight the pivotal role of critically short telomeres in cell homeostasis (13). Vakonaki et al previously demonstrated an association between TL and drug abuse, which leads to premature biological aging (14). Recently, the creation of a database named ‘BIOTEL version 2.4’, Telomere Length Database Project (TLDP), was reported (15). This database is a functional semi-automated worksheet that calculates a wide range of TL statistics and is thus a useful tool with applications in research on telomere biology, as well as in biological age estimation (15). It is important to mention that when the length of the telomeres shortens below a threshold limit, cell growth is restricted, and cells undergo cellular senescence or apoptosis (16). Continuous telomere shortening in cells that fail to undergo senescence, leads to critically short telomeres (<3 kb), prone to chromosome-end fusion. Fused chromosomes break unequally in mitosis, leading to genomic instability and chromosomal rearrangements during cell divisions (17). Indeed, a short telomere load has emerged as a more potent biomarker than MTL or the expression of telomerase (18,19). Actually, recent studies have indicated that a short TL is associated with several age-associated diseases, including CAD, stroke, cancer, arthritis, osteoporosis, cataracts, diabetes type2, hypertension, mental diseases, chronic obstructive pulmonary disease and dementia (9,20-22). Goglin et al suggested that individuals with shorter telomeres have a higher mortality rate, as compared to those with longer telomeres (23). Human studies have shown that TL can be inherited through the direct inheritance of the telomeres and through the specific genetic factors found in the parental gametes (24). Furthermore, telomere shortening can be affected by environmental factors, including physical activity, body mass index (BMI), hormone replacement therapy, smoking, chronic inflammation, oxidative stress, dietary antioxidants and importantly vitamin intake (25-28).

Several studies have examined the correlation of individual nutrients with TL. Indeed, B12 vitamin deficiency has been associated with TL, and an altered DNA methylation (26). In addition, a lack of vitamin B12 and folate can lead to inadequate DNA methylation, genomic instability and telomere shortening (29-31). In a previous study, the beneficial effects of omega-3 (n-3) fatty acids on TL observed in patients with chronic kidney disease, was attributed to reduced oxidative stress (32). Furthermore, it has been shown that omega-3 polyunsaturated fatty acids (PUFAs) can reduce inflammation and decrease oxidative stress (33-35) and thus, protect telomeres from attrition (36). Another study suggested that a shorter TL is associated with a significantly increased risk for breast cancer in pre-menopausal women and women with a lower dietary and supplemental β-carotene vitamin C or E intake (37). In vitro experiments have demonstrated that telomere shortening is delayed by ascorbic acid or its derivatives (38-40) and the lifespan of certain somatic cells has been shown to be increased by α-tocopherol (41). To date, TL has been associated with single vitamin or micronutrient intake, while little is known about the effects of their combination on telomere status. A recent study by Fragkiadaki et al indicated that the synergistic effects of nutrients and natural compounds can activate telomerase enzyme in vitro, in human peripheral blood mononuclear cells; human clinical studies with these formulations are required in order to reveal the health benefits of telomerase activation through natural molecules (42). Epidemiological studies have demonstrated that the deficiency of multiple concurrent micronutrients is very common globally and is associated with chronic diseases; these studies stress the need to restore these deficiencies through diet and nutraceutical supplements (NS) (43,44).

The aim of the present study was to analyze the TL profile of a population using a combination of NS and to evaluate the putative effects of supplements on TL, in comparison to a healthy population not receiving any supplements.

Materials and methods

Materials and reagents. One capsule of My Health (Lumis Research SA) contains the following: Mix Vitamin (ascorbic acid, vitamin E acetate 50% natural, niacin, vitamin B1, vitamin K2 Mena Q7 0.2%, vitamin B6, β-carotene, vitamin B12 1%), anti-caking agents (microcrystalline cellulose, mono- and diglycerides of fatty acids, magnesium stearate and silica dioxide). One drop of vitamin D (Pure D3, Natural Doctor) contains 2,000 IU vitamin D. One capsule of Complete D3&K2 Cofactors (Natural Doctor) contains 2,000 IU vitamin D3, 100 µg K2 and 56 mg magnesium. Two capsules of Omega 3-6-9 (Lumis Research SA) contain the following: 740 mg fish oil, 740 mg linseed oil, 740 mg borage oil (Borrego officinalis L.), 200 mg Krill oil, 370 mg alpha linolenic acid, 312.6 mg eicosapentaenoic acid (EPA), 154.2 mg docosahexaenoic acid (DHA), 162.8 mg gamma linolenic acid (GLA), 400 mg linoleic acid and 273.8 mg oleic acid. Four capsules of My Antioxidant (Lumis Research SA) contain the following: Mix Vitamin (ascorbic acid, calcium ascorbate, ascorbyl palmitate, natural vitamin E acetate 50%, vitamin B3, vitamin B12 1%, β-carotene, vitamin B5, vitamin B6, vitamin B2, vitamin B1, folic acid, vitamin D3 biotin), One capsule of My Probiotics (Lumis Research SA) contains the following: Lyophilized kefir grains, Lactobacillus casei (LMG-S27763), Lactobacillus reuteri (LMG-S27759), Bifidobacterium bifidum (B. bifidum; LMG-S27761), Lactobacillus acidophilus (LMG-S27762), inulin, acacia fiber and hydroxypropyl cellulose. One scoop of My Gastro (Lumis Research SA) contains the following: 2,500 mg L-glutamine (Kyowa®) and 500 mg oligo fructose (FOS). One capsule of My Ubiquinol (Q10) (Lumis Research SA) contains 50 mg of ubiquinol.

Study design. The process of TL estimation is depicted in Fig. 1. The participants were selected from individuals visiting a private clinic in Athens for routine checkup, as well as from personnel of the University Hospital of Heraklion. Basic demographic data and a medical history of chronic diseases were obtained. Demographic data included the name, surname, date of birth and sex. Medical history included all the self-reported statements for chronic diseases, such as diabetes, cancer, rheumatoid arthritis, etc., possible exposure to pesticides (occupational) and use of antidepressant medication. Individuals that reported having a medical history of
any chronic disease were excluded. Based on these criteria, 47 individuals were selected for the study. They had never taken any supplements prior to the study and they were in the 40-55 age group. The participants were healthy and were not on a restricted diet. They were divided into 2 groups as follows: An experimental group that consisted of 16 individuals supplemented with NS and the control group with 31 individuals that did not receive any supplements. The daily dose of all supplements was assigned according to the suggestions of the manufacturers. Blood samples were collected from all individuals and citrate was used as an anticoagulant. The supplementation period for all participants lasted for 6 to 12 months. The protocol of this study was approved by the Ethics Committee for Patients and Biological Material of the University of Crete with reference no. 63/22.03.2019. Biological Material and information of patients were obtained with written informed consent according to the EU General Data Protection Regulation (GDPR). All procedures performed in studies involving human participants were under the ethical standards with the 1964 Helsinki declaration and its later amendments, or comparable ethical standards.

Quantitative-fluorescent in situ hybridization (Q-FISH) analysis. Peripheral blood samples (2.5 ml) were collected from all participants. Heparinized blood was cultured in RPMI-1640 culture medium supplemented with 10% fetal bovine serum, 1% L-glutamine, 1% penicillin and streptomycin (all from Sigma-Aldrich) and stimulated for 72 h in a CO₂ incubator with phytohemagglutinin. For metaphase chromosome preparations, 10 µg/ml colcemid was added to each culture 2 h before harvesting, followed by KCl hypotonic shock and methanol/acetic (3:1) fixation. Q-FISH analysis on metaphases was performed as previously described (14,15).

Images were captured for each slide, using a 63X objective and a charge-coupled device camera at 1,024x1,024 pixels resolution and 8-bit depth. For each slide, >10 different scanned images were obtained from 3 independent biological replicates. Maximum projections and deconvolution of the images were performed using Leica Q-FISH software [Leica Application Suite-Advanced Fluorescence (LAS-AF) version 3.1.3 for Leica TCS SP8] and telomere fluorescence intensity was analysed using ImageJ (https://imagej.nih.gov/ij/) by two independent investigators. Telomere fluorescence values were converted into kb by using L5178Y-S cells (cat. no. 93050408; Culture Collections, Public Health England) with a stable and known TL, which was estimated to be approximately 7 kb (45).

Statistical analysis. Percentiles of TL (of the whole telomere genome) and the TL of short telomeres (TLS) (TL <20th percentile) were estimated. The examination of TL and TLS normality was assessed using Kolmogorov-Smirnov’s test with Liliefors correction. Differences in proportions were examined using Pearson’s Chi-square test or Fisher’s exact test when assumptions were not reached. Age comparison was tested using an independent samples t-test. Multiple linear regressions were applied for examining the effect of NS on TL and TLS length following adjustment for age and sex. Box and Whisker plot and bar-charts were used for data graphical representation. IBM SPSS Statistics 23.0 software was used for data analysis and a level of acceptance of null hypothesis was set at P=0.05.
**Results**

The demographic characteristics of the participants in the control group (CG) and NS group (NSG) are presented in Table I. The sex distribution between the 2 groups did not differ significantly (P=0.609), with a similar proportion of males in both the NSG (n=9, 37.5%) and the CG (n=15, 62.5%). Age distribution, after dividing the population to 5-year interval groups, revealed that the 2 groups were age-matched (P=0.505). To elucidate whether demographics had any effect on TL, we performed statistical analysis in both groups with respect to sex. A separate analysis was performed in the respective age groups. As shown in Table II, females had a longer TL compared to males with a difference of 248 to 474 bases in TL and a difference of 195 to 200 bases in TLS. These differences however, did not reach statistical significance, as none of the quartiles in whole or in short telomeres exhibited a statistically significant difference between the 2 sexes (P>0.300).

TL (of the whole telomere genome) and TLS (<20th percentile) was estimated in the participants of both groups. Differences in TL and TLS between the NSG and CG are shown in Figs. 2 and 3. Quartiles of TL and TLS were estimated from raw data for each patient separately. There was a statistically significant difference (P<0.05) in the TL values of the NSG group, compared to the CG, with increased mean quartiles values for the 1st and 2nd quartile (median). TL measurements were 844 and 953 bases higher in the NSG group compared to the CG. Accordingly, there were statistically significant increased values of TLS in the 2nd (median) and 3rd quartile with values of P=0.044 and P=0.033, respectively. A tendency for difference was found for the 1st quartile comparisons (P=0.051).

The position of NSG and CG in the nomograms produced from TLDP data (13) is shown in Fig. 4. Lines of 1st, 2nd (median) and 3rd quartile of TL vs. age were presented, showing the age decrement. Mean age with 2SD of age and TL with 2SD of both groups were placed onto the diagram. It is clear that the mean TL levels of the NSG are over the line of the median of a healthy population, while the mean TL levels of the CG are under the line of the median of a healthy population. A difference of approximately 1,000 bases was observed.

Furthermore, as demonstrated in Fig. 5, and in more details in Table SI, there were no significant differences in TL among the age groups examined (P>0.500). P-values of TL vs. age were 0.964 for the 1st quartile, 0.908 for the median and 0.537 for the 3rd quartile, while P-values for TLS vs. age were 0.741 for the 1st quartile, 0.826 for the median and 0.912 for the 3rd quartile.

Multiple linear regression models were applied for mean TL (or TLS) and their quartiles were set as dependent variables. Explanatory variables were NSG, age and sex of participants. The results of each regression model are presented in Fig. 6. There was a positive effect for mean TL (beta: 918; 95% CI: 38-1799), for the 1st quartile TL (beta: 889; 95% CI: 65-1714), for the median TL (beta: 1004; 95% CI: 125-1883), but not for the 3rd quartile TL (beta: 953; 95% CI: -128-2035) in the NSG. There was also a positive effect on the median TLS (beta: 0.728; 95% CI: 1-1521) and 3rd quartile TLS (beta: 842; 95% CI: 45-1639), and tendencies for an effect were observed in the mean TLS (P=0.051) and in 1st quartile TLS (P=0.060). Furthermore, Fig. 6 shows the effect (beta coefficients) as derived from multiple linear regression, following adjustment for age and sex (beta and 95% CI).

**Table I. Demographic characteristics of the participants in the nutraceutical supplements group (NSG) and control group (CG).**

<table>
<thead>
<tr>
<th>Sex (n, %)</th>
<th>NSG (n=16)</th>
<th>CG (n=31)</th>
<th>Total</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Male</td>
<td>9</td>
<td>15</td>
<td>24</td>
<td>0.609</td>
</tr>
<tr>
<td>Female</td>
<td>7</td>
<td>16</td>
<td>23</td>
<td></td>
</tr>
<tr>
<td>Age, years (means ± SD)</td>
<td>46.4</td>
<td>3.5</td>
<td>47.6</td>
<td>4.0</td>
</tr>
<tr>
<td>Age groups, (n, %)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>40-45</td>
<td>6</td>
<td>8</td>
<td>14</td>
<td>0.505</td>
</tr>
<tr>
<td>45-50</td>
<td>8</td>
<td>15</td>
<td>23</td>
<td></td>
</tr>
<tr>
<td>50-55</td>
<td>2</td>
<td>8</td>
<td>10</td>
<td></td>
</tr>
</tbody>
</table>

Data were analyzed using *Pearson's Chi-square test; †an independent samples t-test; or ‡Fisher's exact test.*

**Table II. Comparison of whole telomere genome (TL) and short telomere length (TLS) (<20th percentile) (in bases) between the sexes.**

<table>
<thead>
<tr>
<th>Sex</th>
<th>Male (n=24)</th>
<th>Female (n=23)</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Mean ± SD</td>
<td>Mean ± SD</td>
<td></td>
</tr>
<tr>
<td>TL</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>1st Quartile</td>
<td>8,444 ± 1,296</td>
<td>8,692 ± 1,374</td>
<td>0.528</td>
</tr>
<tr>
<td>Median</td>
<td>10,142 ± 1,315</td>
<td>10,490 ± 1,537</td>
<td>0.407</td>
</tr>
<tr>
<td>3rd Quartile</td>
<td>12,107 ± 1,505</td>
<td>12,582 ± 1,910</td>
<td>0.348</td>
</tr>
<tr>
<td>TLS</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>1st Quartile</td>
<td>6,321 ± 1,288</td>
<td>6,516 ± 1,079</td>
<td>0.577</td>
</tr>
<tr>
<td>Median</td>
<td>7,052 ± 1,270</td>
<td>7,247 ± 1,173</td>
<td>0.587</td>
</tr>
<tr>
<td>3rd Quartile</td>
<td>7,592 ± 1,283</td>
<td>7,792 ± 1,293</td>
<td>0.598</td>
</tr>
</tbody>
</table>

*Data were analyzed using an independent samples t-test.*
Discussion

The results of the present study suggest that the daily administration of NS favorably affects the length of whole and short telomeres, independent of sex and age. Although the underlying mechanisms of the effects of NS remain unclear, their effects on several key mechanisms, including oxidative stress and chronic inflammation, could be linked to TL maintenance (46). Indeed, studies have demonstrated that increased oxidative stress and inflammation are associated with a reduced
leukocyte TL in patients with periodontitis (47), as well as in patients with Parkinson's disease (48). Furthermore, Ma \textit{et al} reported that older individuals with central obesity, hyperglycemia, insulin resistance and lower antioxidant levels had a shorter leukocyte TL; specifically, the authors demonstrated that a decreased TL correlated with increased oxidative stress in patients with type 1 and type 2 diabetes (49). Furthermore, oxidative stress has been linked to TL shortening during aging, in a comparative study that concluded that elderly males living in Greece have lower indices of oxidative stress and higher antioxidant levels, compared to a corresponding population of elderly Dutch males (50). The authors suggested that the length of telomeres in leukocytes was significantly greater in the Greek compared to the Dutch population, as
the latter live in a more stressful urban setting and follow a diet poor in antioxidant nutrients. Indeed, antioxidants and anti-inflammatory agents have been shown to reduce the pace of TL shortening (51). In a previous study, Paul associated the availability of vitamin B and D with leukocyte (LTL) (52). Furthermore, in that study, serum folate and its metabolites were positively associated with LTL. It has been suggested that the mechanisms through which these nutritional factors attenuate telomere attrition are antioxidant activity, DNA methylation and the prevention of DNA damage (53). In addition, it has been indicated that carotenoids, omega-3-fatty acid, coenzyme Q10, selenium and vitamin D can reduce the rate of telomeric shortening (51). In 2013, Kiecolt-Glaser et al demonstrated that men and women aged 40-85 years who had been taking supplementation, including omega-3 fatty acids, had reduced oxidative stress and an increased TL (54).

In this study, we present findings from the evaluation of TL in participants receiving a combination of vitamin supplements. An advantage of the present study is the methodology of TL measurement. We measured the TL of peripheral blood leukocytes using a method of high sensitivity (Q-FISH) to produce data on TL and TLS (<20th percentile) percentages using BIOTEL (15). We determined a statistically significant increase in TLS and TL by NS. This is a very interesting finding, as there is evidence indicating that a short telomere load is more critical to cell homeostasis than MTL, or the expression of telomerase (18,17). The increase of short telomeres, measured as a percentage, predicts longevity in mammals and is reversely associated with the majority of age-related diseases. Although the majority of measurements of TL in leukocyte subpopulations have been performed by flow-FISH or real-time PCR, which gives the average length of short telomeres, Q-FISH provides results on median length of single telomeres, the percentage of short telomeres and provides an overview of the rate of telomere shortening and longevity (13).

In this study, we performed an analysis of the TL and TLS between the 2 sexes and did not find a statistically significant difference between males and females. This finding may be attributed to the relatively small size of the group. A previous study by Herrmann et al suggested that telomeres are longer in women due to higher estrogen levels, which increase telomerase activity and present antioxidant effects (53).

The limitations of this study include the small number of participants, as well as the fact that not all lifestyle parameters that can affect TL and telomerase activity were determined. Taking this limitation into account, the effects of NS on TL presented herein highlight the need for the further evaluation of the underlying mechanisms.

In conclusion, in this study, we demonstrate that NS may affect the length of short telomeres. Short telomeres are critical to cell homeostasis and the only reliable method for the evaluation of short telomere distribution, to date, is Q-FISH. Therefore, this study provides evidence that the antioxidant effects of NS is implicated in TL maintenance. Future research is required to focus on the mechanisms through which NS affect telomere shortening.

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Availability of data and materials
The datasets presented in this study are available from the corresponding author upon reasonable request.

Authors’ contributions
DT, PF, AT, DAS and DC conceived and designed the study and wrote the manuscript. PF, EV and AOD performed the data processing and quality control assessment. EvS, AKA and DN performed the statistical analysis and data interpretation. LK interpreted the data and critically revised the article. EK and ErS performed the imaging analysis. All authors reviewed and approved the manuscript prior to submission and have approved the final manuscript.

Ethics approval and consent to participate
The protocol of this study was approved by the Ethics Committee for Patients and Biological Material of the University of Crete with reference no. 63/22.03.2019. Biological Material and information of patients were obtained with written informed consent according to the EU General Data Protection Regulation (GDPR). All procedures performed in studies involving human participants were under the ethical standards with the 1964 Helsinki declaration and its later amendments, or comparable ethical standards.

Patient consent for publication
Not applicable.

Competing interests
DAS is the Editor-in-Chief for the journal, but had no personal involvement in the reviewing process, or any influence in terms of adjudicating on the final decision, for this article. DT is a scientific advisor for Lumis Research SA and Natural Doctor SA. The other authors declare no competing interests. To avoid any bias in the collection of the experimental data, the experiments were conducted by the Laboratory of Toxicology of the Medical School of the University of Crete, Lumis Research SA and Natural Doctor SA had no involvement in the preparation of the manuscript, the results and the supervision of the study.

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