

# miR-135b-5p enhances the sensitivity of HER-2 positive breast cancer to trastuzumab via binding to cyclin D2

ZHILAN LI<sup>1\*</sup>, YIYU QIN<sup>2\*</sup>, PEIHONG CHEN<sup>1</sup>, QIONG LUO<sup>2</sup>, HAIYAN SHI<sup>2</sup> and XIUDI JIANG<sup>1\*</sup>

<sup>1</sup>Department of Clinical Laboratory, Seventh People's Hospital of Shanghai University of Traditional Chinese Medicine, Shanghai 200137; <sup>2</sup>School of Clinical Medicine, Jiangsu Medical Vocational College, Yancheng, Jiangsu 224005, P.R. China

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**Abstract.** Trastuzumab has led to a marked improvement in the outcomes of patients with human epidermal growth factor receptor 2 (HER-2)-positive breast cancer. However, the effects of trastuzumab on HER-2-positive breast cancer are limited by the emergence of its cardiotoxic side effects. MicroRNA (miR)-135b-5p has been shown to inhibit tumor metastasis in breast cancer. The present study aimed to explore the effects of miR-135b-5p overexpression on the efficacy of trastuzumab in HER-2-positive breast cancer. Reverse transcription-quantitative PCR was performed to detect the levels of miR-135b-5p. Cell viability was evaluated with a Cell Counting Kit-8 assay. Annexin V/propidium iodide staining was employed to detect the number of apoptotic cells. Flow cytometry assay was performed to investigate the cell cycle. Western blotting was used to detect the expression levels of Bax, cleaved caspase-3, Bcl-2, cyclin D2, p27<sup>Kip1</sup> and cyclin E1. Cell migration and invasion were detected by Transwell assay. Luciferase assays were conducted to identify the target gene of miR-135b-5p. In addition, an *in vivo* tumor xenograft model was established. miR-135b-5p agomir significantly enhanced the anti-proliferative effect of trastuzumab on HER-2-positive breast cancer cells via the induction of apoptosis, whereas the anti-metastatic effect of trastuzumab was enhanced by miR-135b-5p agomir treatment. Subsequently, luciferase assays indicated that cyclin D2 was the direct target of miR-135b-5p, whereas overexpression of the latter arrested cell cycle during the G<sub>0</sub>/G<sub>1</sub> phase. Moreover, miR-135b-5p agomir notably increased the antitumor effect of trastuzumab *in vivo*. The data demonstrated that miR-135b-5p sensitized HER-2-positive breast cancer cells to trastuzumab *in vitro* and *in vivo* by

directly binding to cyclin D2. These results suggested that the combination of miR-135b-5p with trastuzumab may be a therapeutic strategy for patients with HER-2-positive breast cancer.

## Introduction

Breast cancer is a complex and heterogeneous disease that is considered the leading cause of cancer-related death among females worldwide (1). In the past decades, advances in molecular biology have contributed to significant improvements in the diagnosis and classification of breast cancer (2,3). Surgery, radiotherapy and chemotherapy (hormone and targeted) have shown remarkable survival benefits in patients with breast cancer (4). Approximately 30% of breast tumors overexpress human epidermal growth factor receptor 2 (HER-2) (5). These cells are characterized by high rates of cell proliferation, metastasis and low overall survival (5). The humanized HER-2 monoclonal antibody trastuzumab, also known as Herceptin, targets HER-2 and has been approved by the United States Food and Drug Administration (6). Trastuzumab drug has become the standard treatment for patients with HER-2-positive cancer in early or advanced breast cancer (7). Trastuzumab treatment has led to a marked improvement in the outcomes of patients with HER-2-positive breast cancer (8). It was reported that trastuzumab markedly inhibits tumor proliferation, angiogenesis and metastasis via downregulation of HER-2 (9-12). Although this antibody has led to optimal stratification of breast cancer, while the prognosis of patients with HER-2 breast cancer remains poor (13,14).

MicroRNAs (miRs/miRNAs) are small regulatory RNAs that modulate the expression of their target genes (15). Several miRNAs have shown preferentially conserved interactions with the majority of human mRNAs; they can influence specific developmental processes and the progression of several diseases (16). Notably, miRNAs are well known for their roles in regulating cell growth, which is critical to cancer development (17). Several studies have investigated the combination of miRNAs with trastuzumab for the treatment of patients with HER-2-positive breast cancer. It was reported that miR-16 (18), miR-26a, miR-30b (1), miR-141-3p (19), miR-770-5p (20) and miR-205-5p (21) are involved in mediating trastuzumab response in HER-2-positive breast cancer. Moreover, repression of miR-135b-5p was reported to promote

**Correspondence to:** Dr Xiudi Jiang, Department of Clinical Laboratory, Seventh People's Hospital of Shanghai University of Traditional Chinese Medicine, 358 Datong Road, Shanghai 200137, P.R. China  
E-mail: jiangxiudi123@126.com

\*Contributed equally

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metastasis of early-stage breast cancer (22), whereas it was also shown to enhance the antitumor effect of doxorubicin in breast cancer cells by targeting anterior gradient 2 (23). The present study hypothesized that miR-135b-5p may be associated with regulating the sensitivity of breast cancer cells to trastuzumab. Therefore, the focus of the current study was to investigate of the expression of miR-135b-5p, which was markedly down-regulated in HER-2-positive breast cancer cells. In addition, the mechanism by which miR-135b-5p improved trastuzumab sensitivity was investigated.

## Materials and methods

**Cell culture and transfection.** HER-2-positive breast cancer cells BT-474 and SK-BR-3 and the normal breast epithelial cell line MCF-10A were obtained from the American Type Culture Collection. Both cell lines were grown in Roswell Park Memorial Institute medium (Thermo Fisher Scientific, Inc.), which was supplemented with 10% fetal bovine serum (Thermo Fisher Scientific, Inc.) and 1% penicillin/streptomycin. The cells were maintained at 37°C in a humidified atmosphere at 5% CO<sub>2</sub>.

miR-135b-5p agomir (5'-UAUGGCUUUUUAUCCUGUGUGA-3') and its negative control (NC; 5'-UUUGUACUACACAAAAGUACUG-3') were synthesized and purchased from Guangzhou RiboBio Co., Ltd. The cells were transfected with 50 nM miR-135b-5p agomir and its NC (50 nM) using Lipofectamine 3000 (Invitrogen; Thermo Fisher Scientific, Inc.) for 24 h according to the manufacturer's protocol. Cells in the blank group were not given any treatment.

pcDNA3.1-cyclin D2 was purchased from Shanghai GenePharma Co., Ltd. BT-474 cells were transiently transfected with 2 µg/ml pcDNA3.1-NC or 2 µg/ml pcDNA3.1-cyclin D2 using Lipofectamine<sup>TM</sup> 3000 (Invitrogen; Thermo Fisher Scientific, Inc.) for 48 h according to the manufacturer's instructions.

**Cell viability.** The cell viability of BT-474 and SK-BR-3 cells was evaluated using Cell Counting Kit-8 (CCK-8; Dojindo Molecular Technologies, Inc.) according to the manufacturer's protocol. The cells were seeded into 96-well plates at a density of 5×10<sup>4</sup> cells/ml overnight. Subsequently, the cells were treated with 10 nM miR-135b-5p agomir or agomir NC for 48 h. Finally, 10 µl CCK-8 (5 mg/ml) solution was added to the culture medium in each well and the cells were incubated for 2 h at 37°C. The absorbance was measured at a wavelength of 450 nm using a multifunctional microplate reader (Bio-Rad Laboratories, Inc.).

**Reverse transcription-quantitative PCR (RT-qPCR).** miR-135b-5p levels were quantified using RT-qPCR. Total RNA was extracted from BT-474, SK-BR-4 or MCF-10A cells using TRIzol<sup>®</sup> reagent (Invitrogen; Thermo Fisher Scientific, Inc.) according to the manufacturer's protocol. cDNA synthesis was performed using a High Capacity cDNA Synthesis kit (Thermo Fisher Scientific, Inc.) with miRNA-specific primers. The following specific primers were used for RT: U6, 5'-AACGCTTCACGAATTTGCGT-3' and miR-135b-5p, 5'-CTCACTGGTGTCTGAGTCGGCAATTCAGTTGAGTCA

CATAG-3'. qPCR was performed on an Applied Biosystems 7500 Real-Time PCR machine (Thermo Fisher Scientific, Inc.) with the EnTurbo<sup>TM</sup> SYBR Green PCR SuperMix (ELK Biotechnology Co., Ltd.). The following primer pairs were used for the qPCR: miR-135b-5p forward, 5'-AGCTATGGC TTTTCATTCCTATG-3' and reverse, 5'-CTCAACTGG TGTCGTGGAGTC-3'; U6 forward, 5'-CTCGCTTTCGGCA GCACAT-3' and reverse, 5'-AACGCTTCACGAATTTGC GT-3'; GAPDH forward, 5'-CGGACCAATACGACCAAA TCCG-3' and reverse, 5'-AGCCACATCGCTCAGACACC-3'; and cyclin D2 forward, 5'-GAACTCGAGGAGAGCCAT CT-3' and reverse, 5'-AGTTCGAATCTGCACCGTAG-3'. The relative level of miR-135b-5p was normalized to U6, while the relative level of cyclin D2 normalized to GAPDH according to the 2<sup>-ΔΔC<sub>q</sub></sup> method (24). miRNA RT-qPCR primer sets (one RT primer and one pair of qPCR primers) specific for miR-135b-5p, and mRNA RT-qPCR primers specific for cyclin D2 were obtained from Genecreate. Cyclin D2 levels were analyzed by RT-qPCR in BT-474 breast cancer cells transfected with miR-135b-5p agomir for 24 and 48 h.

**Detection of apoptosis and cell cycle analysis.** Apoptosis induction was analyzed using the Annexin V-FITC apoptosis detection kit (BD Biosciences) according to the manufacturer's protocol. The cells were harvested and washed with PBS twice. Subsequently, they were stained with propidium iodide (PI) and Annexin V. Following 15 min of incubation in the dark at room temperature, cell apoptosis was detected by flow cytometry (FACSCalibur; BD Biosciences) using the CellQuest Pro software (version 3.3; BD Biosciences).

For cell cycle analysis, the cells were harvested and resuspended at 1×10<sup>6</sup> cells/ml in modified Krishan's buffer (0.1% sodium citrate, 0.3% NP-40, 0.02 mg/ml RNase and 0.05 mg/ml PI). The stained cells were detected by flow cytometry (FACSCalibur; BD Biosciences) using CellQuest Pro software (version 3.3; BD Biosciences).

**Transwell assay.** Migration and invasion assays were performed using a 24-well Transwell chamber (Sigma-Aldrich; Merck KGaA) with or without a Matrigel-coated membrane. The cells (5×10<sup>4</sup>/well) were plated in serum-free medium in the upper chamber with a non-coated membrane (24-well insert, 8-µm pore size) in order to assess migration, whereas a matrigel-coated membrane was used for the invasion assay. The lower chamber was filled with medium containing 10% fetal bovine serum (Thermo Fisher Scientific, Inc.). After 24 h of incubation, the cells on the upper surface were removed by a cotton swab and the cells on the lower surface were stained with 10% w/v aqueous Giemsa's solution (VWR International, LLC) for 30 min at room temperature. The stained cells were imaged and counted under a fluorescence microscope (Nikon Eclipse Ti-E; Nikon Corporation) in five randomly selected fields at x200 magnification.

**Dual-luciferase reporter assay.** TargetScan ([http://www.targetscan.org/vert\\_71/](http://www.targetscan.org/vert_71/)) and miRWalk (<http://zmf.umm.uni-heidelberg.de/apps/zmf/mirwalk2/>) online tools were used to predict target genes of miR-135b-5p. The predicted miR-135b-5p binding sequence (wild-type; WT) or a mismatch sequence (mutant type) in the 3'-untranslated region (3'-UTR)

of cyclin D2 mRNA were synthesized and cloned separately into the multiple cloning site of the luciferase miRNA expression reporter vector (Promega Corporation). The sequences of these synthesized oligonucleotides are as follows: Forward WT: 5'-GGCUCAGGUUUUGAGAAGCCAUC-3' and mutant 5'-GGCUCAGGUUUUGAGGUUAAGGC-3'. Subsequently, cells were co-transfected with pRL-TK-cyclin D2-WT or pRL-TK-cyclin D2-MT plasmid and miR-135b-5p agomir using Lipofectamine 3000 (Invitrogen; Thermo Fisher Scientific, Inc.). Additionally, cells were co-transfected with pRL-TK-cyclin D2-WT or pRL-TK-cyclin D2-MT plasmid and NC using Lipofectamine 2000 (Invitrogen; Thermo Fisher Scientific, Inc.), which was considered as the vector-control group. Meanwhile, cells transfected with pRL-TK-cyclin D2-WT or pRL-TK-cyclin D2-MT plasmid alone was considered as the control group. Following transfection, the cells were cultured for 6 h at 37°C and the transfection medium was removed and replenished with DMSO. At 48 h post-transfection, luciferase activity was measured using the dual-luciferase reporter assay system (Promega Corporation) and normalized to *Renilla* luciferase activity.

**Western blotting.** Total protein was extracted from the cells with lysis buffer (Cell Signaling Technology, Inc.). The lysates were centrifuged at 10,000 x g for 10 min at 4°C. The protein concentration was determined using a bicinchoninic acid protein quantification kit (Promega Corporation). Equal amounts of cell extracts (30 µg/well) were subjected to 10% SDS-PAGE and transferred to PVDF membranes (EMD Millipore) for antibody blotting. The membranes were blocked with 5% bovine serum albumin (Thermo Fisher Scientific, Inc.) for 1 h at room temperature and subsequently incubated with β-actin (1:1,000; dilution, cat. no. ab179467), Bax (1:1,000; cat. no. ab32503), cleaved caspase 3 (1:1,000; cat. no. ab2303), Bcl-2 (1:1,000; cat. no. ab59348), cyclin D2 (1:1,000; cat. no. ab207604), p27<sup>kip1</sup> (1:1,000; cat. no. ab32034), cyclin E1 (1:1,000; cat. no. ab33911), phosphorylated (p)-Akt (1:1,000; cat. no. 38449) and Akt (1:1,000; cat. no. ab8850) primary antibodies (all from Abcam) overnight at 4°C. Subsequently, the membranes were incubated with a horseradish peroxidase-conjugated anti-mouse (1:5,000; cat. no. ab97040; Abcam) or an anti-rabbit secondary antibody (1:5,000; cat. no. ab7090; Abcam) at room temperature for 1 h. Protein bands were visualized using an enhanced chemiluminescence reagent kit (GE Healthcare) on a Tanon 5200 Chemiluminescent imaging system (Tanon Science and Technology Co., Ltd.) according to the manufacturer's protocol. ImageJ software (v1.8.0.112; National Institutes of Health) was used for quantification of protein expression with β-actin as the reference protein.

**Tumor xenograft model.** A total of 24 female BALB/c nude mice (18-20 g, 4-5 weeks) were obtained from Vital River. The animals were housed at a temperature of 23±1°C, and a humidity of 50-60% for 4 days in a 12-h light/dark cycle prior to the initiation of the experiment. For induction of anesthesia, the mice were anesthetized by isoflurane (3% v/v) inhalation. Nude mice were inoculated with 17β-estradiol pellets 48 h before implantation of BT-474 cells. Subsequently, BT-474 cells (5x10<sup>6</sup> cells/ml in 100 µl PBS mixed with Matrigel matrix) were subcutaneously injected into the mice. Animal health and

behavior were monitored twice a day. All animal procedures were approved by the Seventh People's Hospital of Shanghai University of Traditional Chinese Medicine Committee in June 2019. The National Institute of Health Guide for the Care and Use of Laboratory Animals was followed (25). The mice were randomly divided into four groups (n=6) when the tumor volume reached 100 mm<sup>3</sup>: Control, miR-135b-5p agomir, trastuzumab and trastuzumab + miR-135b-5p agomir. The tumor volume was estimated according to the following equation: Volume= Length x Width<sup>2</sup>/2. Trastuzumab (10 mg/kg; ChemBest), miR-135b-5p agomir (50 nM) or the combination treatment of trastuzumab with miR-135b-5p agomir were administered as follows: Trastuzumab (10 mg/kg) was administered intraperitoneally once per week while miR-135b-5p agomir was directly injected into the implanted tumor at a dose of 50 nM (in 20 µl PBS) per mouse twice a week. After 3 weeks of treatment, the mice were euthanized using CO<sub>2</sub> at a displacement rate of 20% of the chamber volume/min (CO<sub>2</sub> flow rate, 2.5 l/min). The tumors were removed and weighted. The duration of the experiment is 6 weeks, and no mice died during the experimental process. Humane endpoints were determined as previously described (26). Criteria for judging the death of animals is continuous no spontaneous breathing for 2-3 min and no blink reflex.

**Immunohistochemistry (IHC) assay.** The tumor tissues were fixed in 4% paraformaldehyde for 48 h at room temperature, embedded in paraffin and cut into 4-µm sections. Deparaffinized tissue sections were incubated with Ki67 monoclonal antibody (1:500; cat. no. ab15580; Abcam) overnight at 4°C. Subsequently, the sections were incubated with biotinylated goat anti-rabbit immunoglobulin G (1:1,000; cat. no. ab6721; Abcam) at room temperature for 30 min. IHC reactions were visualized by using the IHC detection system (EnVision kit; Dako; Agilent Technologies, Inc.). The images were captured under a fluorescence microscope (Nikon Eclipse Ti-E; Nikon Corporation) at x200 magnification. ImageJ software (v1.8.0.112; National Institutes of Health) was used for quantification of protein expression.

**Statistical analysis.** All experiments were repeated three times and the results are expressed as the mean ± SD. One-way ANOVA followed by Tukey's post hoc test was performed to analyze differences among multiple groups (>2 groups). Statistical analysis was performed using GraphPad Prism (version 7; GraphPad Software, Inc.). P<0.05 was considered to indicate a statistically significant difference.

## Results

**miR-135b-5p agomir enhances the anti-proliferative effect of trastuzumab in HER-2-positive breast cancer cells.** To investigate the effects of miR-135b-5p agomir in BT-474 and SK-BR-3 cells, RT-qPCR assay was performed. The results indicated that the levels of miR-135b-5p in BT-474 and SK-BR-3 cells were significantly lower compared within the normal breast epithelial cell line MCF-10A (Fig. 1A). Moreover, miR-135b-5p agomir caused a significant upregulation in miR-135b-5p expression in these two breast cancer cell lines compared with the blank group (Fig. 1B). Subsequently,

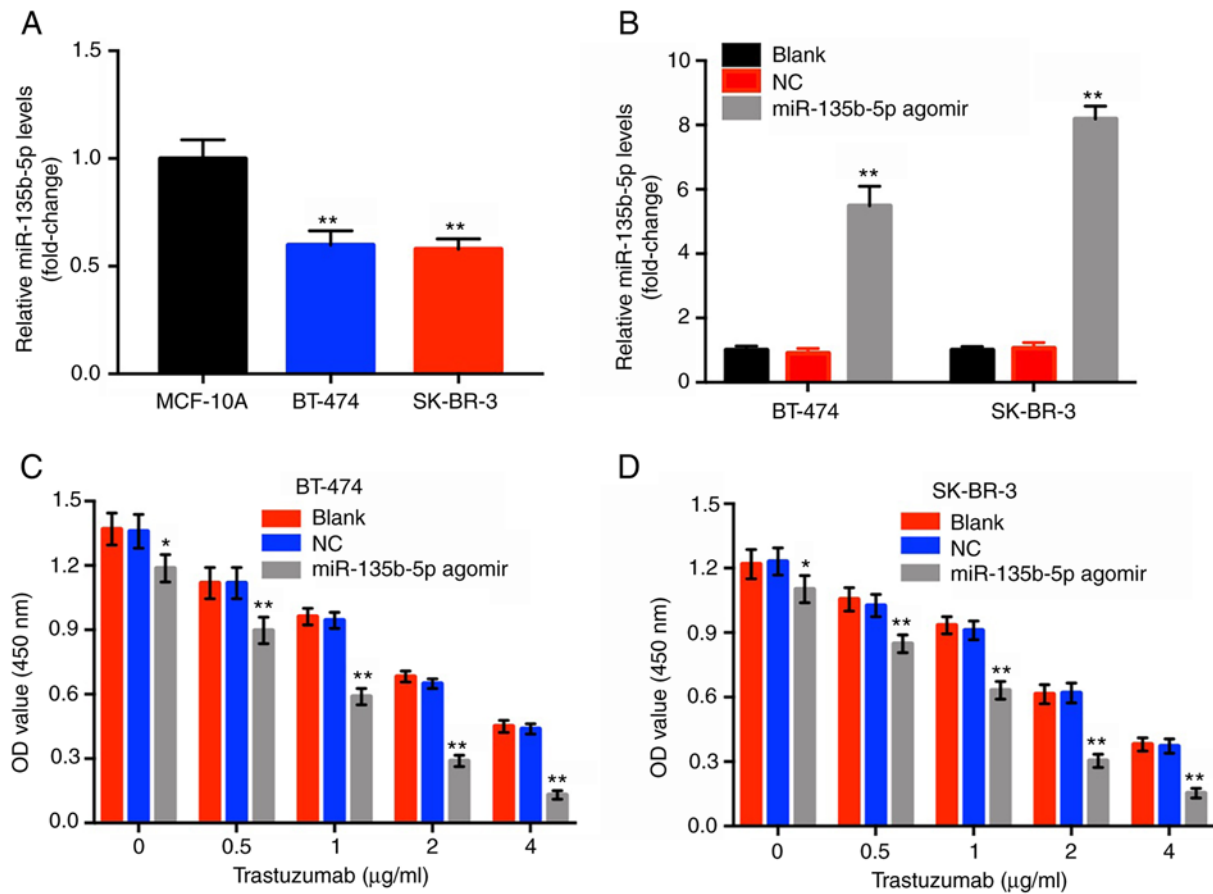


Figure 1. miR-135b-5p agomir enhances the anti-proliferation effect of trastuzumab in HER-2 positive breast cancer cells. (A) The levels of miR-135b-5p in BT-474, SK-BR-3 and MCF-10A cells were evaluated using RT-qPCR. \*\* $P < 0.01$  vs. MCF-10A. (B) The cells were transfected with either agomir or NC for 48 h. The levels of miR-135b-5p in BT-474 and SK-BR-3 cells after transfection was detected using RT-qPCR. \*\* $P < 0.01$  vs. blank. Breast cancer cells (C) BT-474 and (D) SK-BR-3 were treated with trastuzumab (0, 0.5, 1, 2 or 4 µg/ml) and miR-135b-5p agomir for 48 h. Cell viability was detected using a Cell-Counting Kit 8 assay. \* $P < 0.05$  and \*\* $P < 0.01$  vs. blank. miR, microRNA; RT-qPCR, reverse transcription-quantitative PCR; NC, negative control; OD, optical density.

a CCK-8 assay was performed to investigate the cytotoxic effects of trastuzumab or of the combination treatment (trastuzumab plus miR-135b-5p agomir) in HER-2-positive breast cancer cells. Trastuzumab alone inhibited the proliferation of HER-2-positive breast cancer cells in a dose-dependent manner, which was significantly enhanced in the presence of miR-135b-5p agomir compared with the blank groups (Fig. 1C and D). Since trastuzumab (1 µg/ml) combined with miR-135b-5p agomir induced ~50% inhibition of cellular growth, this dose was applied for subsequent cell assays. In addition, BT-474 cells were more sensitive to the combination treatment compared with SK-BR-3 cells and therefore this cell line was employed in the following experiments. The results suggested that miR-135b-5p significantly increased the anti-proliferative effect of trastuzumab in HER-2-positive breast cancer cells.

**Trastuzumab-induced apoptosis is enhanced by miR-135b-5p agomir in BT-474 cells.** Annexin V/PI staining was performed to detect the mechanism by which miR-135b-5p enhanced the anti-proliferative effect of trastuzumab. The results indicated that trastuzumab or miR-135b-5p agomir treatment significantly induced cell apoptosis compared with the control group (Fig. 2A). In addition, trastuzumab-induced apoptosis was significantly enhanced by miR-135b-5p agomir

in BT-474 cells (Fig. 2A). Moreover, the expression levels of apoptotic-associated proteins were examined in BT-474 cells by western blotting. The combination of miR-135b-5p agomir with trastuzumab significantly increased the expression levels of cleaved caspase 3 and Bax in BT-474 cells compared cells treated miR-135b-5p agomir or trastuzumab alone (Fig. 2B-D). Furthermore, trastuzumab-induced Bcl-2 downregulation was significantly enhanced in the presence of miR-135b-5p agomir (Fig. 2E). Taken together, the data demonstrated that miR-135b-5p agomir could enhance trastuzumab-induced apoptosis via activation of the intrinsic apoptotic pathway.

**Anti-migration and anti-invasion effects of trastuzumab is enhanced by miR-135b-5p agomir.** Following the initial investigation of the effects of miR-135b-5p on trastuzumab-induced apoptosis, its effect on the inhibition breast cancer cell migration and invasion were also investigated. Transwell assays indicated that trastuzumab significantly inhibited migration and invasion abilities in BT-474 and SK-BR-3 cells compared with the control group (Fig. 3A-D). Furthermore, the inhibitory effects of trastuzumab on the migration and invasion of BT-474 and SK-BR-3 cells were significantly enhanced in the presence of miR-135b-5p agomir (Fig. 3A-D). These results suggested that miR-135b-5p agomir could increase



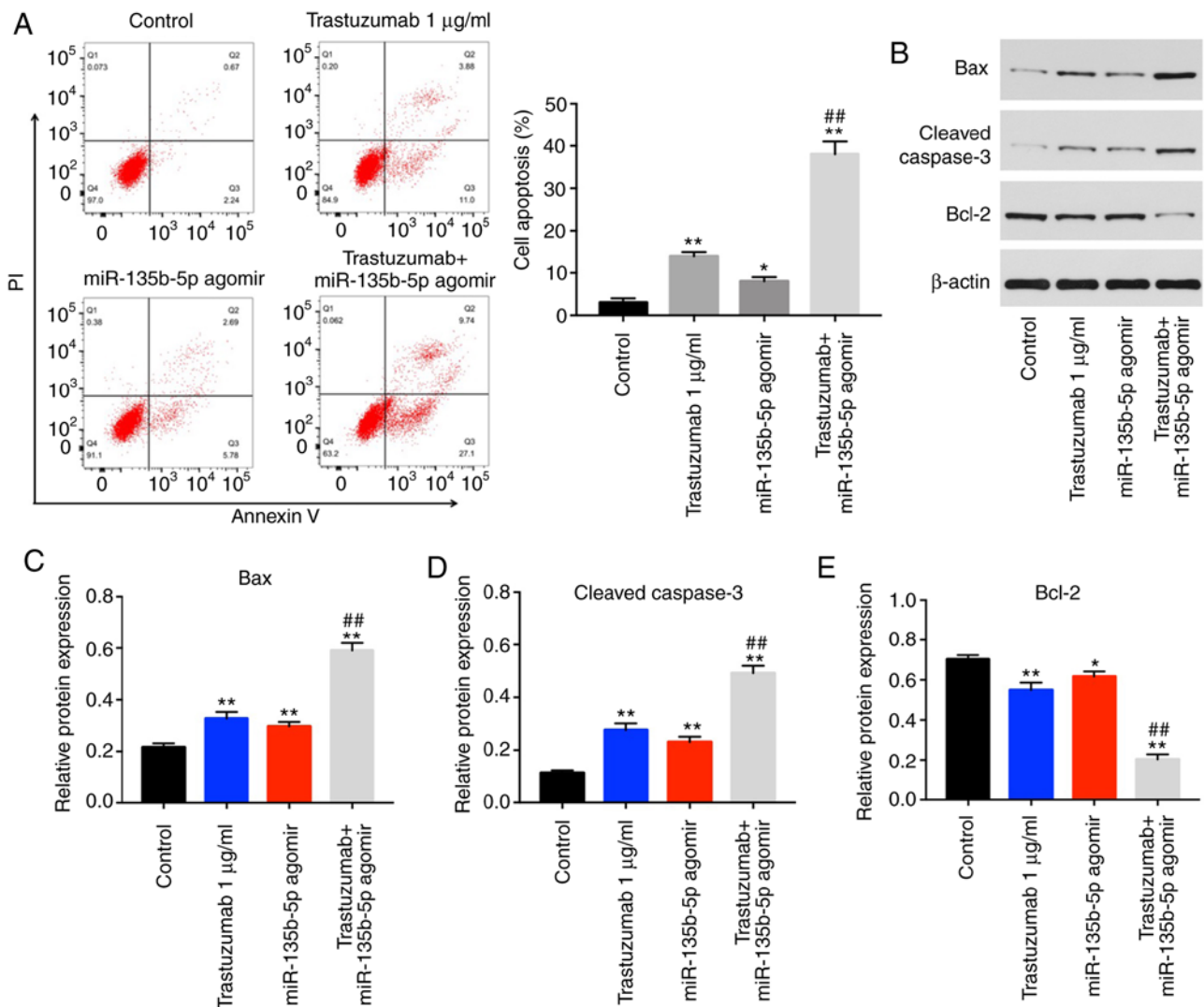


Figure 2. miR-135b-5p agomir enhances the anti-proliferation effects of trastuzumab via inducing apoptosis. (A) Annexin V/PI staining in BT-474 cells was detected by flow cytometry. (B) Expression levels of Bax, cleaved caspase 3 and Bcl-2 in BT-474 cells were detected using western blotting. The relative expression levels of (C) Bax, (D) cleaved caspase-3 and (E) Bcl-2 were quantified by normalizing to β-actin levels. \*P<0.05 and \*\*P<0.01 vs. control; ##P<0.01 vs. trastuzumab. miR, microRNA; PI, propidium iodide.

the anti-metastatic effects of trastuzumab in HER-2-positive breast cancer cells.

*Cyclin D2 is a direct target of miR-135b-5p in BT-474 breast cancer cells.* To investigate the underlying mechanisms by which miR-135b-5p agomir increased the antitumor effects of trastuzumab in breast cancer cells, TargetScan and miRWalk online tools were used. A putative miR-135b-5p-binding site was identified at a specific position in the 3'-UTR of cyclin D2 mRNA. To further confirm the direct interaction between miR-135b-5p and cyclin D2 mRNA, the predicted binding site of miR-135b-5p to cyclin D2 mRNA was cloned into a luciferase reporter vector. miR-135b-5p agomir significantly inhibited luciferase activity compared with the control group (Fig. 4B), suggesting that it could interact directly with the 3'-UTR of cyclin D2 mRNA. In contrast to these observations, miR-135b-5p agomir did not affect the luciferase activity of the mutated cyclin D2. In addition, the qPCR data indicated that cyclin D2 mRNA synthesis was significantly inhibited when the cells were treated with miR-135b-5p agomir, and cyclin D2

levels decreased over time following agomir transfection (Fig. 4C). Taken together, the data indicated that cyclin D2 was a direct target of miR-135b-5p in BT-474 breast cancer cells.

Since cyclin D2 mRNA was demonstrated to directly bind to miR-135b-5p, the cell cycle of miR-135b-5p agomir-treated cells was examined. miR-135b-5p-overexpressing BT-474 cells displayed a significant increase in the percentage of cells in the G0/G1-phase but significantly decreased proportions in S-phase cells compared with controls, while trastuzumab exhibited no effects (Fig. 4D and E). In addition, western blot analysis illustrated that trastuzumab treatment had no effect on the expression levels of cyclin D2, p27<sup>kip1</sup> and cyclin E1 in BT-474 cells. However, miR-135b-5p agomir or combination treatment significantly inhibited the expression levels of cyclin D2 and cyclin E1 and significantly increased p27<sup>kip1</sup> levels in BT-474 cells compared with the control group (Fig. 5A-D). Moreover, trastuzumab treatment significantly downregulated the expression of phosphorylated (p)-Akt in BT-474 cells, while overexpression of miR-135b-5p did not affect p-Akt activation in BT-474 cells (Fig. 5A and E).

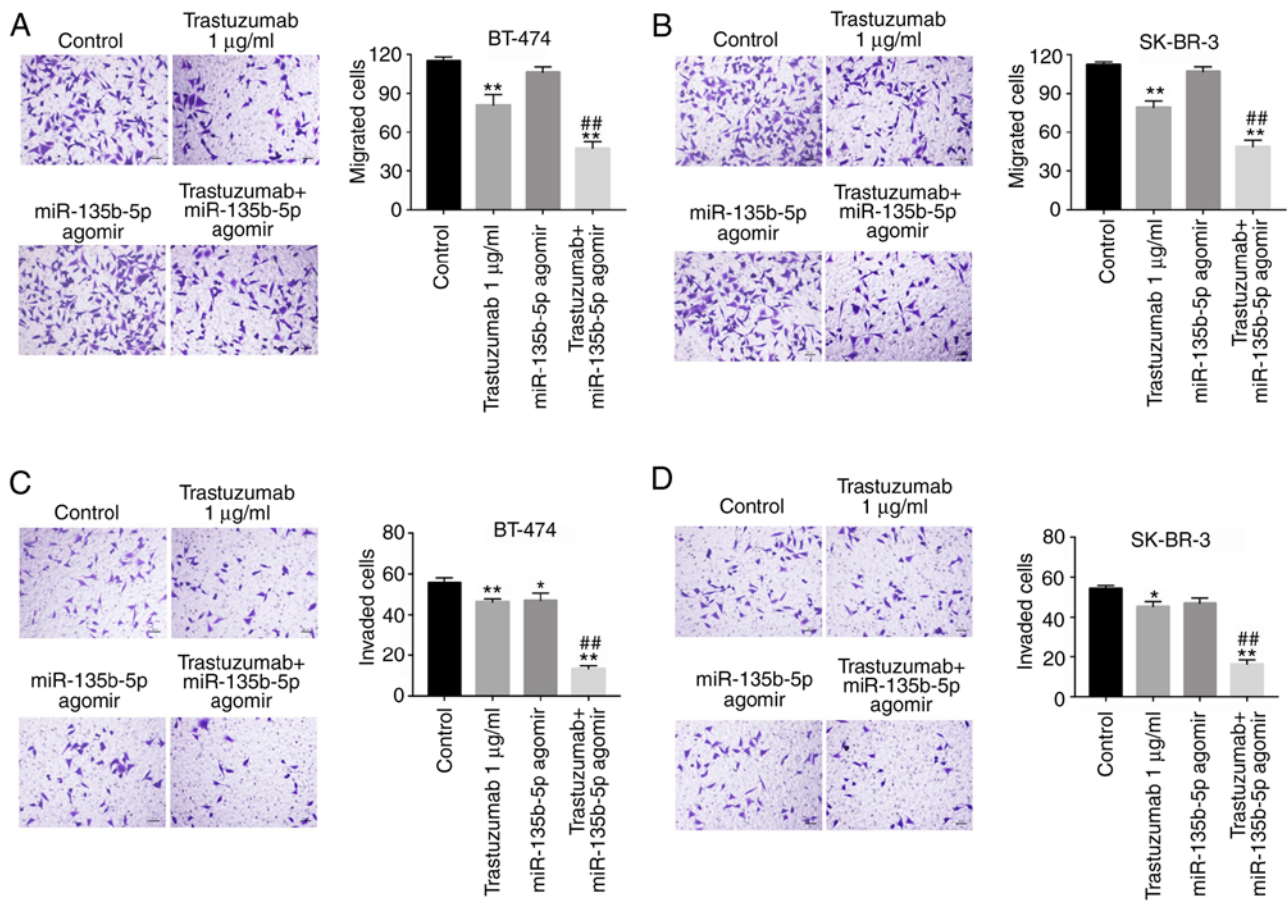


Figure 3. Anti-migration and anti-invasion effects of trastuzumab on breast cancer cells is enhanced by miR-135b-5p agomir. BT-474 and SK-BR-3 cells were transfected with 1  $\mu$ g/ml trastuzumab, 50 nM miR-135b-5p agomir or a combination of trastuzumab and miR-135b-5p agomir for 24 h. Cell migration assay was performed to evaluate the migration ability of (A) BT-474 and (B) SK-BR-3 cells. Magnification x200. Cell invasion assay was performed to evaluate the invasion ability of (C) BT-474 and (D) SK-BR-3 cells. Magnification, x200. \* $P < 0.05$  and \*\* $P < 0.01$  vs. control; ## $P < 0.01$  vs. trastuzumab. miR, microRNA.

Interestingly, trastuzumab-induced p-Akt downregulation was enhanced in the presence of miR-135b-5p agomir (Fig. 5A and E). Meanwhile, overexpression of miR-135b-5p did not show any significant effect on the expression of cyclin D1 in BT-474 cells (Fig. 5F and G). Taken together, the data demonstrated that miR-135b-5p agomir could induce G<sub>0</sub>/G<sub>1</sub> arrest in BT-474 cells by directly binding to cyclin D2. Furthermore, the overexpression efficiency of cyclin D2 was confirmed by western blotting (Fig. 5H and I). Overexpression of cyclin D2 significantly reversed the growth-inhibitory activity of miR-135b-5p in combination with trastuzumab in BT-474 cells (Fig. 5J). These data indicated that miR-135b-5p agomir enhanced the anti-proliferative effects of trastuzumab in BT-474 cells via downregulation of cyclin D2.

*miR-135b-5p agomir enhances the antitumor effect of trastuzumab in vivo.* To explore whether miR-135b-5p could increase the antitumor effects of trastuzumab *in vivo*, a breast cancer xenograft model was established. The results indicated that trastuzumab treatment alone was able to decrease tumor size compared with controls (Fig. 6A). Tumor weight and tumor volume were significantly reduced in the combination treatment group compared with mice treated with trastuzumab or miR-135b-5p agomir alone (Fig. 6A-C). However, miR-135b-5p agomir alone showed no significant difference

on tumor volume. In addition, the body weight changes indicated the safety and tolerability of the combination treatment (Fig. 6D). Furthermore, the expression levels of miR-135b-5p was significantly upregulated in the miR-135b-5p agomir or combination treatment group compared with the control group, indicating that miR-135b-5p is stably expressed in tumor tissues (Fig. 6E). Additionally, trastuzumab had no effect on cyclin D2 expression in tumor tissues, whereas the level of cyclin D2 significantly decreased in the presence of miR-135b-5p agomir compared with controls (Fig. 6F). IHC assay indicated that miR-135b-5p agomir or trastuzumab treatment significantly inhibited proliferation in tumor tissues, compared with the control group. As expected, combination treatment significantly inhibited proliferation in tumor tissues compared with the trastuzumab treatment group (Fig. 6G and H). These data were consistent with the *in vitro* results. Taken together, these results confirmed that miR-135b-5p agomir could enhance the antitumor effects of trastuzumab *in vivo*.

## Discussion

In the present study, the combined antitumor effects of trastuzumab with miR-135b-5p agomir were investigated *in vitro* and *in vivo*. miR-135b-5p agomir enhanced the

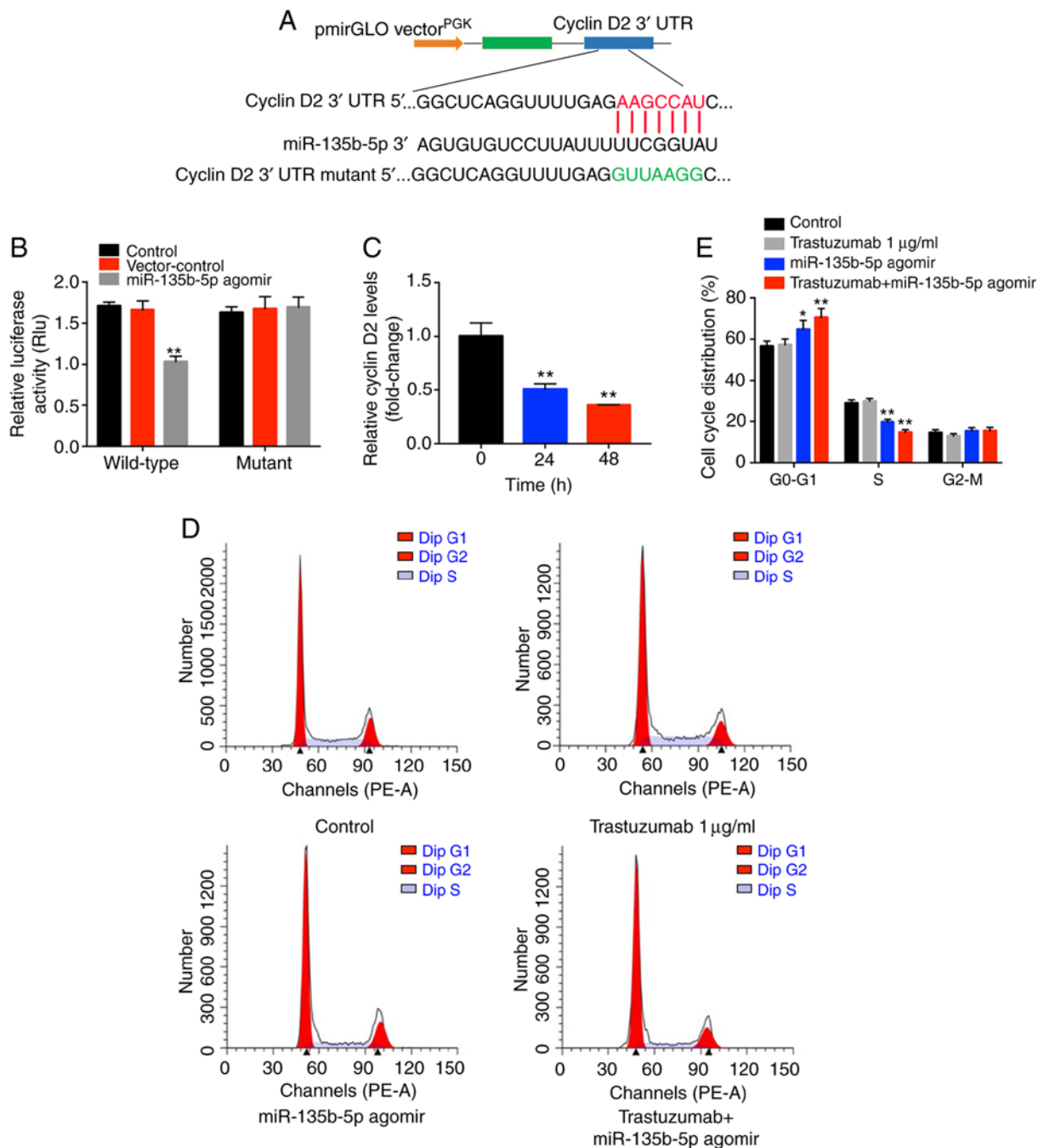


Figure 4. miR-135b-5p induces G<sub>0</sub>/G<sub>1</sub> cell cycle arrest in BT-474 cells via directly targeting cyclin D2. (A) The target sequences for miR-135b-5p within the 3'-UTR of cyclin D2. (B) Comparison of luciferase activity in cells of different groups that were co-transfected with wild-type or mutant 3'-UTR cyclin D2. \*\*P<0.01 vs. control. (C) The levels of cyclin D2 mRNA was evaluated using reverse transcription-quantitative PCR. \*\*P<0.01 vs. 0 h. (D) Cell cycle analysis was performed to determine the effect of miR-135b-5p agomir on cell cycle distribution. (E) Quantification of cell cycle distribution ratios. \*P<0.05 and \*\*P<0.01 vs. control. 3'UTR, 3'-untranslated region; miR, microRNA.

antitumor effect of trastuzumab in breast cancer cells *in vitro* and *in vivo*. A previous study reported that overexpression of miR-135b-5p suppressed migration and invasion in breast cancer (22). The present results indicated that miR-135b-5p agomir suppressed proliferation, migration and invasion in BT-474 and SK-BR-3 HER-2-positive breast cancer cells, consistent with previous reports (27,28). Agomir is a type of specially labeled and chemically modified double-stranded microRNA that regulates the biological function of target genes by mimicking endogenous microRNA (29). Compared with common miRNA mimics, miRNA agomir has a higher

affinity for the cell membrane (29). In addition, agomir is especially suitable for animal *in vivo* experiments and have higher stability (30). These data showed that miR-135b-5p exerted anti-proliferation, anti-migration and anti-invasion effects in HER-2-positive breast cancer cells. Moreover, miR-135b-5p agomir potentiated the antitumor effect of trastuzumab by inducing apoptosis. In addition, the inhibition of breast cell migration and invasion by trastuzumab was enhanced by miR-135b-5p agomir transfection. Tang *et al* (31) indicated that overexpression of miR-200c increased drug sensitivity to trastuzumab in HER-2-positive breast cancer cells, consistent

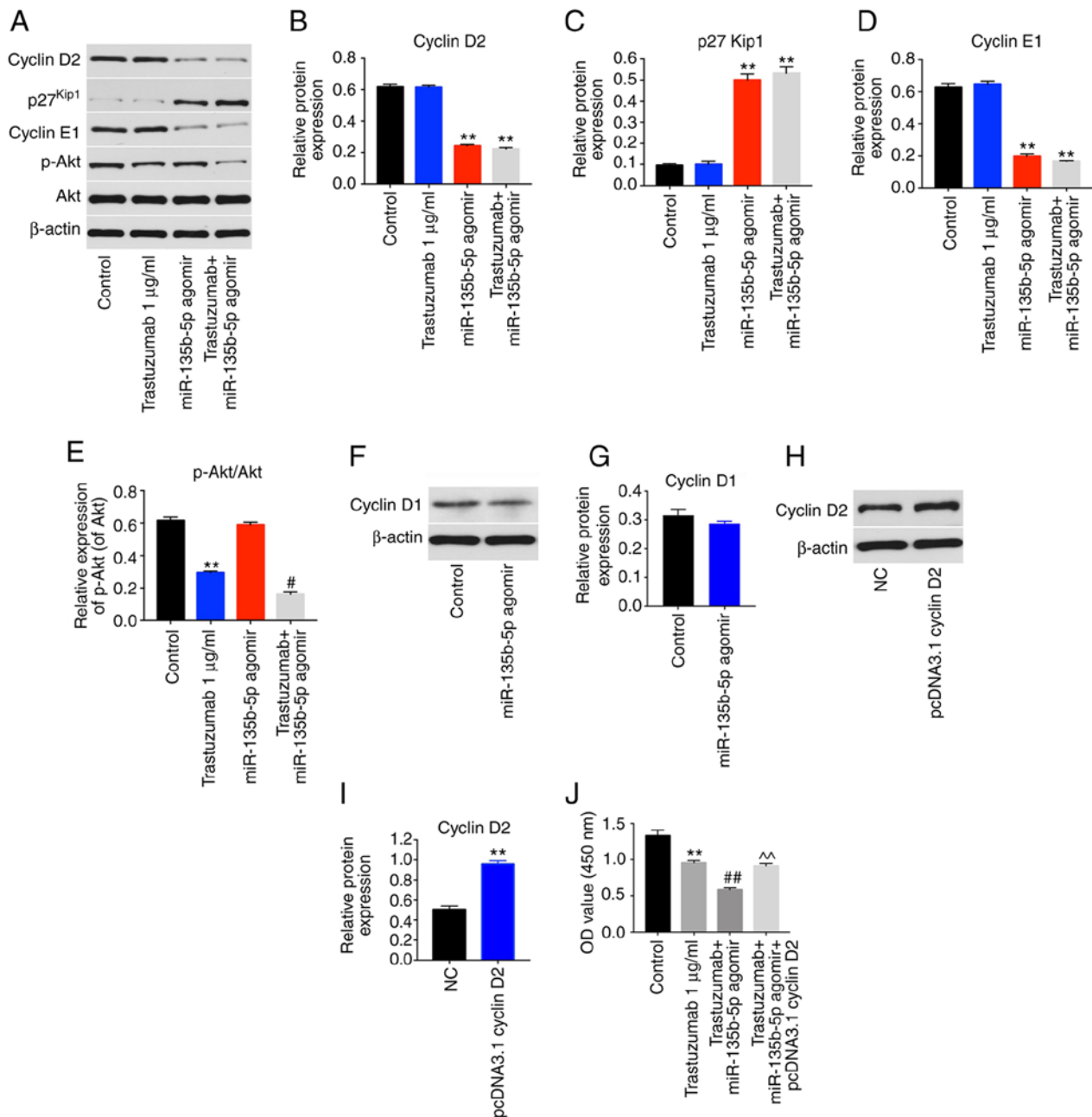


Figure 5. miR-135b-5p agomir regulates cyclin D2, p27<sup>Kip1</sup> and cyclin E1 signaling pathways. BT-474 cells were transfected with 1 μg/ml trastuzumab, 50 nM miR-135b-5p agomir or a combination of trastuzumab and miR-135b-5p agomir for 48 h. (A) Expression levels of cyclin D2, p27<sup>Kip1</sup>, cyclin E1 and p-Akt in BT-474 cells were detected with western blotting. The relative levels of (B) cyclin D2, (C) p27<sup>Kip1</sup>, (D) cyclin E1 and (E) p-Akt were quantified by normalizing to β-actin levels. (F) BT-474 cells were transfected with miR-135b-5p agomir or NC for 48 h. The levels of cyclin D1 in BT-474 cells was detected using western blotting. (G) Quantification of cyclin D1 expression. (H) BT-474 cells were transfected with NC or pcDNA 3.1-cyclin D2 for 48 h. (I) The level of cyclin D2 in BT-474 cells was detected with western blotting. (J) BT-474 cells were transfected with pcDNA 3.1-cyclin D2 and miR-135b-5p agomir for 48 h in the presence of trastuzumab. The cell viability was detected using a Cell Counting Kit-8 assay. \*\*P<0.01 vs. control; \*P<0.05 and ##P<0.01 vs. trastuzumab; ^P<0.01 vs. trastuzumab + miR-135b-5p agomir. p-Akt, phosphorylated Akt; miR, microRNA; OD, optical density; NC, negative control.

with the data reported here. miR-135b-5p agomir enhanced the antitumor effect of trastuzumab *in vivo* and *in vitro*.

The target predicting databases TargetScan and miRWalk were used to investigate the mechanism by which miR-135b-5p increases the antitumor effect of trastuzumab. Cyclin D2 was identified as a potential target of miR-135b-5p. Cyclin D2 protein is involved in the regulation of the cell cycle at the point of transition from G1 to DNA synthesis (32). Zhou *et al* (33) indicated that overexpression of miR-206 blocked G1/S transition and inhibited the proliferation of

breast cancer cells via downregulation of cyclin D2 expression. In the present study, miR-135b-5p agomir suppressed the proliferation of breast cancer cells by inhibiting cyclin D2 expression, which was consistent with the previous study. Zhang *et al* (23) reported that miR-135b-5p enhanced doxorubicin-sensitivity of ER-positive breast cancer cells by targeting anterior gradient 2. The differences noted on the targeting of proteins may be associated with the breast cancer subtypes, namely estrogen (ER) or HER-2, which are upstream regulators of miR-135b-5p. Therefore, in addition



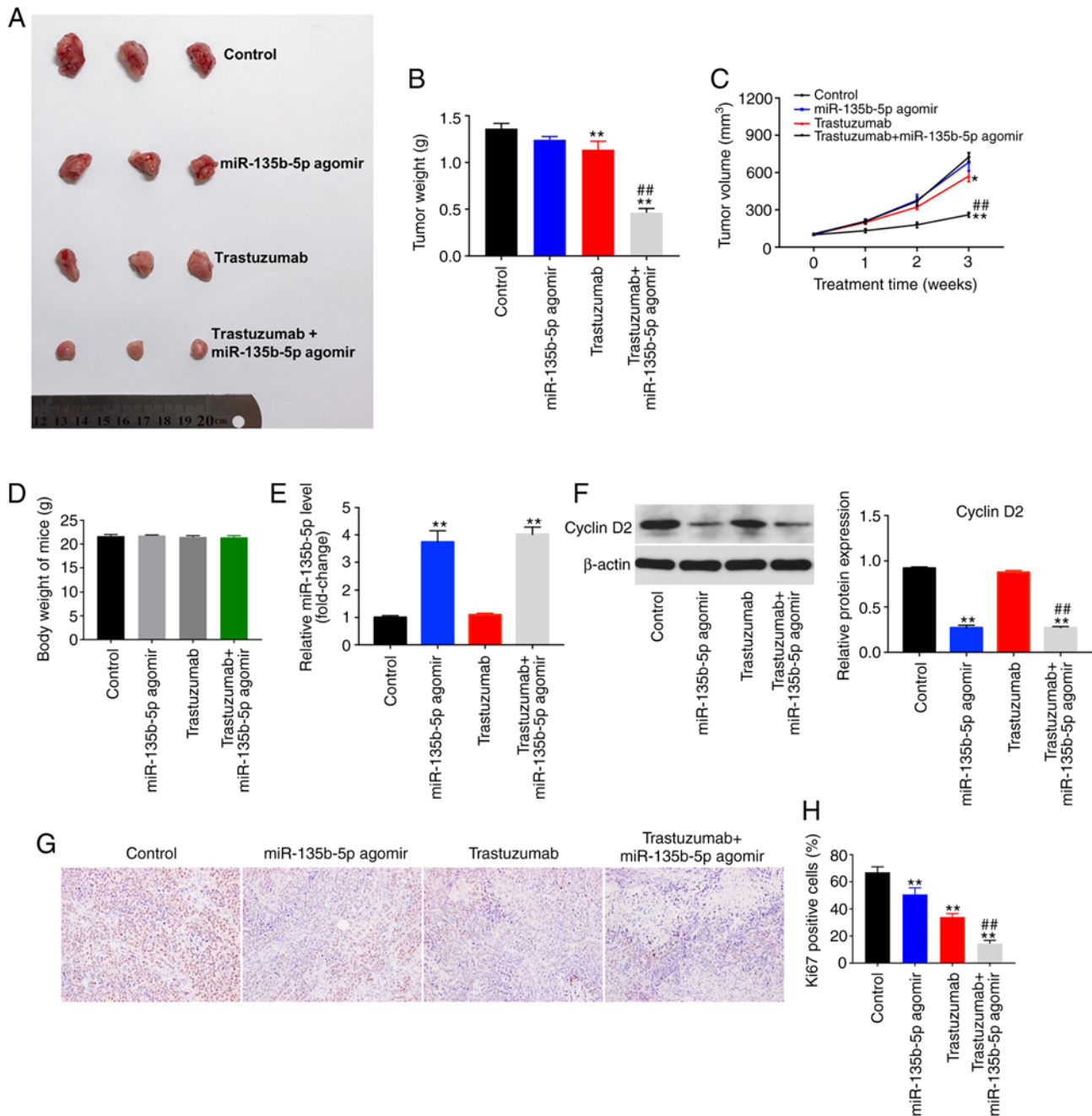


Figure 6. miR-135b-5p agomir enhances the antitumor effects of trastuzumab *in vivo*. (A) The mice were treated with saline solution (control), trastuzumab, miR-135b-5p agomir or a combination of trastuzumab and miR-135b-5p agomir for 3 weeks. (B) Tumor weight in each group was detected after 3 weeks. (C) Tumor volumes of nude mice were monitored weekly. (D) Body weight of each group of nude mice was evaluated at day 27. (E) The levels of miR-135b-5p in tumor tissues was detected using reverse transcription-quantitative PCR. (F) The expression levels of cyclin D2 in tumor tissues was detected using western blotting. Relative protein expression was quantified by normalizing to  $\beta$ -actin levels. (G) Immunohistochemistry images and (H) quantification of proliferation in tumor tissues. Magnification x200. \* $P < 0.05$  and \*\* $P < 0.01$  vs. control; ## $P < 0.01$  vs. trastuzumab. miR, microRNA.

to ER positive breast cancer, miR-135b also plays a role as a regulator of HER-2-positive breast cancer. These findings suggested that miR-135b-5p may act as regulator in different types of breast cancer.

Overexpression of ErbB2 (HER-2) leads to the potentiation of cyclin E-Cdk2 activity by sequestration of the cyclin-dependent kinase inhibitor p27<sup>Kip1</sup> (10). A previous study reported that trastuzumab inhibited mitogen-activated protein kinase and PI3K/AKT pathways, leading to cell cycle arrest (34). However, in the present study, the HER-2 antibody

trastuzumab exhibited limited effects on the expression of cyclin D2 or p27<sup>Kip1</sup>, while miR-135b-5p acted as a regulator of cyclin D2 and p27<sup>Kip1</sup> expression levels. The correlation between p27<sup>Kip1</sup> expression and the PI3K/Akt pathway in the combination treatment of miR-135b-5p with trastuzumab can be further explored in future studies. In addition, a number of miRNAs have been shown to suppress cyclin D2 expression in various tumor types (35-38), including miR-133, miR-203, miR-204 and miR-206. To the best of our knowledge, the present study was the first to demonstrate that miR-135b-5p

directly suppressed cyclin D2 expression in HER-2-positive breast cancer.

However, the present study had several limitations. The present study indicated that miR-770-5p agomir in combination with trastuzumab treatment blocked the migration and invasion capacities of HER-2-positive breast cancer cells *in vitro*. However, the correlation between miR-770-5p agomir in combination with trastuzumab and breast cancer metastasis *in vivo* is still unclear and need to be investigated. In addition, the present study provided evidence to support the anti-cancer properties of miR-135b-5p, but little is known regarding the clinical significance of miR-135b-5p in human breast cancer. Therefore, the clinical application value of miR-135b-5p in breast cancer should be investigated in the future. Moreover, the present study only detected the expression of p-Akt in HER-2-positive breast cancer cells. However, expression of other proteins associated with the PI3K/Akt pathway, such as PI3K and mTOR should be investigate using western blotting to demonstrate whether miR-135b-5p agomir enhances the antitumor effect of trastuzumab in HER-2-positive breast cancer cells via the PI3K/Akt pathway. Meanwhile, rescue experiments are needed to further prove whether miR-135b-5p agomir could enhance the antitumor effect of trastuzumab in HER-2-positive breast cancer cells via targeting cyclin D2.

In conclusion, miR-135b-5p agomir enhanced the anti-tumor effect of trastuzumab in BT-474 cells *in vitro* and *in vivo*. miR-135b-5p induced cell cycle arrest by binding to cyclin D2. Therefore, the combination of miR-135b-5p agomir with trastuzumab may be a potential strategy for the treatment of patients with HER-2-positive breast cancer.

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## Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

## Authors' contributions

ZL and YQ made major contributions to the conception, design and manuscript drafting of this study. PC, QL and HS were responsible for data acquisition, data analysis, data interpretation and manuscript revision. XJ made substantial contributions to conception and design of the study and revised the manuscript critically for important intellectual content. All authors agreed to be accountable for all aspects of the work. All authors read and approved the final manuscript.

## Ethics approval and consent to participate

All animal procedures were approved by the Seventh People's Hospital of Shanghai University of Traditional Chinese Medicine Committee. National Institutes of Health Guide for the Care and Use of Laboratory Animals was followed strictly.

## Patient consent for publication

Not applicable.

## Competing interests

The authors declare that they have no competing interests.

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