

MEIS1 and its potential as a cancer therapeutic target (Review)

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Abstract. Meis homeobox 1 (Meis1) was initially discovered in 1995 as a factor involved in leukemia in an animal model. Subsequently, 2 years later, *MEIS1*, the human homolog, was cloned in the liver and cerebellum, and was found to be highly expressed in myeloid leukemia cells. The *MEIS1* gene, located on chromosome 2p14, encodes a 390-amino acid protein with six domains. The expression of homeobox protein MEIS1 is affected by cell type, age and environmental conditions, as well as the pathological state. Certain types of modifications of MEIS1 and its protein interaction with homeobox or pre-B-cell leukemia homeobox proteins have been described. As a transcription factor, MEIS1 protein is involved in cell proliferation in leukemia and some solid tumors. The present review article discusses the molecular biology, modifications, protein-protein interactions, as well as the role of MEIS1 in cell proliferation of cancer cells and MEIS1 inhibitors. It is suggested by the available literature MEIS1 has potential to become a cancer therapeutic target.

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1. Introduction

The Meis homeobox 1 (*Meis1*) gene was initially discovered as a common viral integration site in the BXH-2 mouse model of leukemia in 1995 (1). Subsequently, after 2 years, the human homolog (*MEIS1*) was cloned in the liver and cerebellum, and was found to be highly expressed in myeloid leukemia cells (2). Two pro-viral integration clusters exist between two regions located 90 kb apart, and the *MEIS1* gene alone was found between the two clusters (3). The *MEIS1* gene encodes a 3.8 kb transcript with an open reading frame that encodes a homeodomain protein (4).

Homeobox protein MEIS1 belongs to the three amino acid loop extension (TALE) homeodomain transcription factor family, whose members also include MEIS2 and MEIS3 (5). MEIS2 protein comprises of 477 amino acids and has a molecular weight of 51,790 Da. This protein is coded by the *MEIS2* gene, that has a length of 212,108 bp. The MEIS3 protein is coded by the *MEIS3* gene with a length of 19,110 bp and is composed of 375 amino acids. The potential interplay between the two proteins and MEIS1, is illustrated in Fig. 1 (modified from www.string-db.org/).

MEIS1 is involved in a number of physiological and pathophysiological processes; most notably, in cell migration, apoptosis and metabolism. The present review article summarizes the results of previous studies that have implicated MEIS1 in cancer cell proliferation. The potential use of MEIS1 as a novel biomarker and therapeutic target in cancer was also discussed.

2. Molecular biology of MEIS1

DNA/RNA. *MEIS1* is located between LOC729348 and LOC100507073 on chromosome 2p14, and contains the anonymous markers, D2S134 and N1B1519 (2,6). It consists of 13 exons and 137,360 bp (6). The 3' untranslated region (3' UTR) of *MEIS1* is highly conserved during evolution (7). The structure of the *MEIS1* gene is illustrated in Fig. 2. In total, four MEIS1 isoforms, produced by alternative splicing, have been described: MEIS1a, MEIS1b, MEIS1c and MEIS1d (6). MEIS1a possesses all 13 exons, whereas MEIS1b lacks the 95-bp exon 12 (4). MEIS1c lacks 49 amino acids (Val 162 to Gln 210) and is expressed in mouse embryos (8). MEIS1d lacks exon 8 and is expressed in mouse colorectal cancer cells (9).

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A linear diagram of MEIS1 isoforms with their exons is presented in Fig. 3.

The full-length *MEIS1* mRNA consists of 3,198 bp (6). The expression of *MEIS1* mRNA in different human tissues, according to the Human Protein Atlas (<http://www.protein-atlas.org/>), is illustrated in Fig. 4. The highest expression levels of *MEIS1* mRNA are found in the endometrium, fallopian tubes and smooth muscle, whereas the lowest levels are found in skeletal muscle, liver and bone marrow.

Protein. Full-length MEIS1 consists of 390 amino acids and has a molecular weight of 43,016 Da (6). MEIS1 contains a pre-B-cell leukemia homeobox (PBX)1 interaction domain, serine/threonine-rich domain, aspartic acid/glutamic acid-rich domain, poly-aspartic acid domain, homeodomain and transcriptional activation domain (10). The conserved PBX interaction motif, homeodomain and C-terminal region are critical for leukemic transformation (11).

MEIS1 is localized mainly in the nucleus (12), although its expression has also been detected in the cytoplasm in certain tumor cells, such as in ovarian cancer cells (13). During pregnancy, MEIS1 localizes to the cytoplasm and the membrane of endometrial glandular epithelial cells (14).

Relative MEIS1 expression at the protein level in different human tissues, according to the Human Protein Atlas, is illustrated in Fig. 5. The highest expression of MEIS1 occurs in the fallopian tubes and smooth muscle. However, a previous study suggested that MEIS1 was expressed at highest levels in the endo- and myometrium among 87 types of normal tissues (15).

In addition to the pathological state, other factors such as cell type, age and environmental conditions also affect MEIS1 expression. MEIS1 is highly expressed in all cell lines with a mixed-lineage leukemia (MLL) mutation, but not in wild-type MLL cell lines (16). The expression of MEIS1 in solid tumors is somewhat paradoxical, as it is overexpressed in certain tumor types, whereas it is downregulated in others (5). The cell types from the different organ sites under investigation (5) and the types of metabolism (10) the cancer cells require may be responsible for these contradicting findings. MEIS1 tends to be expressed at higher levels in fetal hearts than in adult hearts (17); however, another study reported a lower MEIS1 expression in postnatal hearts than in adult hearts (18). Among pediatric patients with acute lymphoblastic leukemia (ALL), MEIS1 expression is more frequent in infant ALL than in childhood ALL (19). The discrepancies among these studies, particularly, the association of MEIS1 with age, requires further investigation. Hypoxia has been shown to reduce the expression of MEIS1 in the heart (20), and to downregulate MEIS1 expression in pulmonary artery smooth muscle cells in both primary culture and animals (21).

3. Modification

MEIS1 related modifications include DNA methylation and protein ubiquitination. These are discussed below.

DNA methylation. The hypomethylation of the *MEIS1* promoter mediated by DNA methyltransferase 3A (DNMT3A) has been observed in acute myeloid leukemia (AML) without MLL fusions (22). In AML involving the AML-RUNX1 partner

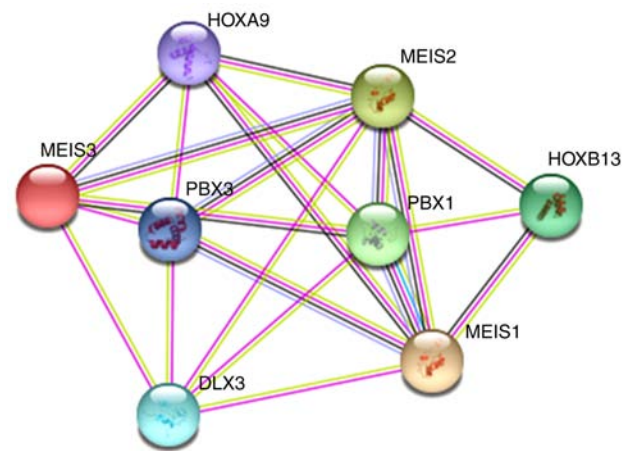


Figure 1. Potential interplay between MEIS2, MEIS3 and MEIS1. The data shown were modified from the STRING database (www.string-db.org). MEIS1, Meis homeobox 1; MEIS2, Meis homeobox 2; MEIS3, Meis homeobox 3; HOXA9, homeobox A9; HOXB13, homeobox B13; PBX1, pre-B-cell leukemia homeobox 1; DLX3, distal-less homeobox 3.

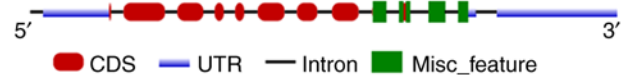


Figure 2. Structure of the *MEIS1* gene. *MEIS1*, Meis homeobox 1; CDS, coding sequence; UTR, untranslated regions.

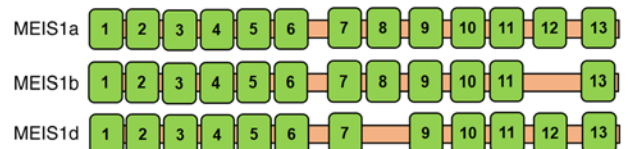


Figure 3. Linear diagram of MEIS1 isoforms with their exons (green boxes). MEIS1, Meis homeobox 1.

transcriptional co-repressor 1 (ETO) fusion, the hypermethylation of the *MEIS1* promoter causes the transcript level to decrease with time (23). *MEIS1* downregulation can be reversed by the combination of the demethylating agent, decitabine, and the histone deacetylation inhibitor, trichostatin A (23). Although no methylation of the *MEIS1* promoter has been observed in patients with AML (24), trimethylation mediated by SET domain bifurcated histone lysine methyltransferase 1 (SETDB1) or the complex of polymerase-associated factor 1 (PAF1) and SETDB1 (25), has been detected. Thus, modifications of the *MEIS1* gene differ across types of AML. *MEIS1* gene methylation has also been observed in solid tumors, such as bladder cancer (26). The methylation level of the *MEIS1* promoter is significantly associated with *MEIS1* downregulation in the B-raf proto-oncogene (BRAF)^{p.v600e} mutant human colon cancer tissues and colon cancer cell lines (27). However, another study using colorectal cancer tissues from 42 patients failed to detect the methylation of the *MEIS1* promoter (28).

Ubiquitination. Protein ubiquitination promotes its degradation (29). The exposure of hematopoietic stem cells to branched-chain amino acid (BCAA) has been shown to

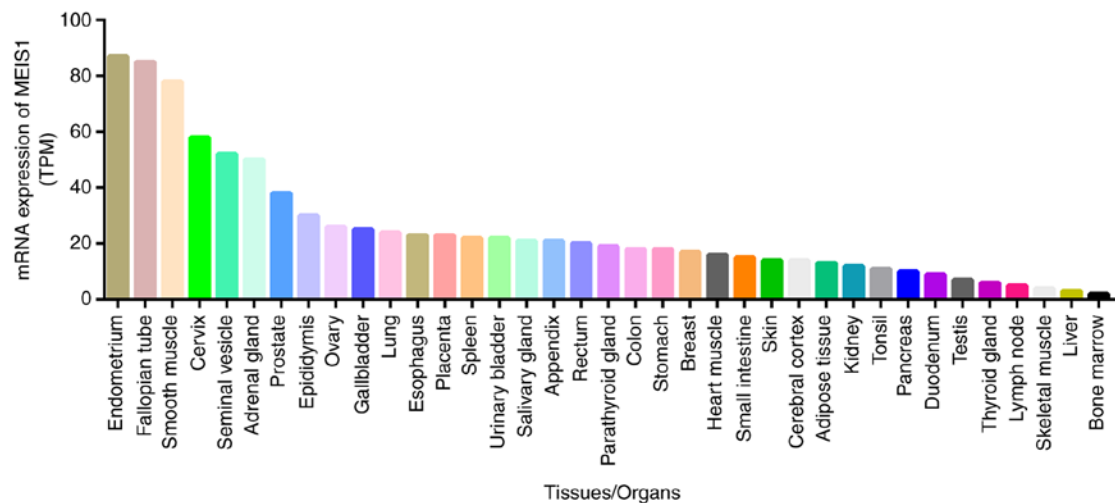


Figure 4. Expression of *MEIS1* mRNA in different human tissues. The data shown were derived from The Human Protein Atlas (<http://www.proteinatlas.org/>). *MEIS1*, Meis homeobox 1; TPM, transcripts per kilobase million.

increase the cell division cycle 20 (CDC20)-mediated ubiquitination of MEIS1 protein (30). Conversely, the competitive inhibition of E3 binding to MEIS1 by PBX3 prevents the ubiquitination of MEIS1 (31).

4. Protein-protein interactions

Protein-protein interactions play crucial roles in cellular functions and biological processes. MEIS1 is known to interact with several partners, including homeobox (HOX) and PBX proteins.

HOX protein family. MEIS1 binds to HOXA9 to form a heterodimeric complex (32), which in turn binds to target DNA sequences that contain the MEIS1 binding site (TGACAG) and an AbdB-like HOX site (TTTTACGAC) (33). HOXA10, HOXA11, HOXD12 and HOXB13 can also form DNA-binding complexes with MEIS1b (33). In mouse embryos, both HOXA13 and HOXD13 interact with MEIS1a C-terminal domain (18 amino acids) and MEIS1b C-terminal domain (93 amino acids) (34). HOX protein stabilizes the interaction between MEIS1 and its target DNA; as a result, target DNA dissociates at a markedly slower rate from the MEIS1-HOX complex than from MEIS1 alone (33). The overexpression of MEIS1 induces cell apoptosis through caspase-dependent processes, whereas the overexpression of MEIS1 and HOXA9 inhibits apoptosis and protects cells from the influence of apoptosis-inducing factors (35).

MEIS1 and HOX can also bind to a third protein to form trimers. For example, MEIS1 forms a trimer with HOXA9 and PBX2 in myeloid leukemia cells, which then binds to the target DNA (36). Trimers formed by MEIS1, HOXA10 and PBX2 mediate the expression of target genes in the human endometrium (37).

PBX protein family. PBX proteins located in the nucleus are significantly associated with MEIS1 expression (38). PBX1 and MEIS1 form a dynamic dimer, with each protein binding to its respective target DNA site (39). A cyclic adenosine monophosphate (cAMP)-responsive sequence (CRS1) in the bovine

cytochrome P450 family 17 (*CYP17*) gene is a transcriptional regulatory element that contains binding sites for PBX and MEIS1. PBX1 and MEIS1 bind cooperatively to CRS1 to regulate cAMP-dependent transcription, whereas neither protein can bind this element on its own (40). The dimers formed by PBX1 and MEIS1 can also bind to the PBX/MEIS binding site in the SRY-box transcription factor 3 (*SOX3*) promoter, where they regulate *SOX3* expression during development (41).

PBX-regulating protein-1 (PREP1) controls the expression of MEIS1 through post-transcriptional regulation: PREP1 inhibits the interaction between PBX1 and MEIS1 in mouse embryonic fibroblasts, destabilizing MEIS1 and inhibiting the interaction between MEIS1 and DEAD-box helicase 3 x-linked (DDX3x) and DDX5, and ultimately decreasing MEIS1 tumorigenicity (42). Dimerization with PBX3 stabilizes MEIS1, allowing it to upregulate target genes, such as FMS related receptor tyrosine kinase 3 (*FLT3*) and tribbles pseudokinase 2 (*TRIB2*), thereby enhancing HOX9-mediated transformation. MEIS1 protein that does not bind to PBX3 is prone to ubiquitination and subsequent degradation. Mutations in the PBX binding region in MEIS1 can also prevent the ubiquitination of MEIS1, as PBX3 and the responsible E3 ubiquitin ligase share common binding requirements within MEIS1 (30,43).

MEIS1 can form trimers with PBX protein, as well as HOX protein. MEIS1 in megakaryocytes binds to PBX1b and PBX2 to form MEIS1/PBX complexes, which in turn binds to the TGACAG sequence in tandem repeats of the MEIS1 binding element (TME) of the platelet factor 4 (*PF4*) promoter, inducing the expression of the gene. The simultaneous overexpression of MEIS1 and PBX2 potentiates the activation of the *PF4* promoter, and is abrogated when the MEIS1 binding site on the TME is destroyed (44).

5. Role of MEIS1 in the proliferation of cancer cells

As a transcription factor, MEIS1 functions as a positive regulator of the proliferation of leukemia cells (45), as well as in certain solid tumors, such as esophageal squamous cell carcinoma (46), malignant peripheral nerve sheath (47) and Ewing sarcoma (48); however, it has also been shown to function as a

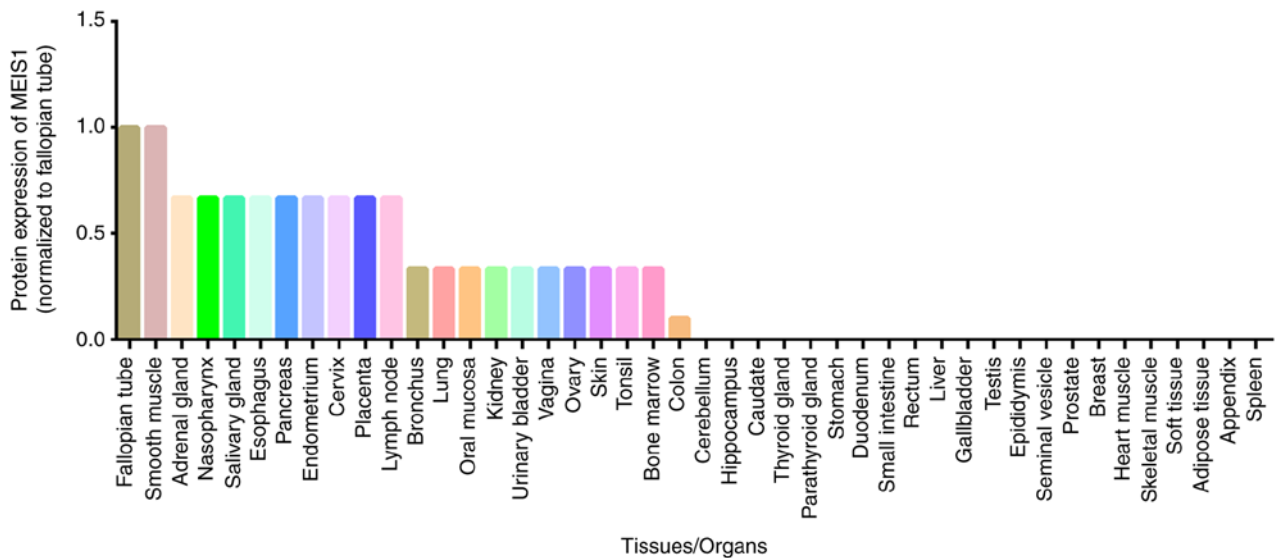


Figure 5. Relative MEIS1 expression at the protein level in different human tissues. The data shown were derived from The Human Protein Atlas (<http://www.proteinatlas.org/>). MEIS1, Meis homeobox 1.

negative regulator of several other solid tumors (49) (Fig. 6 and Table I). A schematic diagram of the promoting and inhibitory roles of MEIS1 in various types of cancer is presented in Fig. 6.

Leukemia. MEIS1 interacts with a variety of partners to promote the proliferation of leukemia cells. Recent studies have demonstrated that MEIS1 interacts with HOXA9 to promote cell proliferation in leukemia via synaptotagmin-like 1 (SYTL1) (50), cyclin D3 (CCND3) (51) or spleen tyrosine kinase (Syk) (52). In patients with acute AML, the interaction between MEIS1 and HOXA9 induces the proliferation of leukemia cells via the simultaneous overexpression of MEIS1 and HOXA9 (31,52,53), suppresses granulocyte colony-stimulating factor (G-CSF)-induced granulocytic differentiation and promotes cell proliferation via stem cell factor (SCF) (54). In addition, the complex formed by MEIS1 and HOXA9 is recruited to the *FLT3* promoter to turn on the gene (55), and *FLT3* induces mitogen-activated protein kinase (MAPK) phosphorylation, inhibits apoptosis and promotes leukemia cell proliferation (56). The MEIS1-HOXA9 complex also interacts with *TRIB1* to induce MAPK phosphorylation, and to stimulate leukemia cell proliferation (57). A recent study demonstrated that the MEIS1-HOXA9 complex bound to the endothelin receptor type A (*EDNRA*, a tumor promoter) promoter in leukemia cells, resulting in cell proliferation and in resistance to apoptosis (58). *HOXD13* also helps regulate the pro-proliferative function of MEIS1 in AML. The human leukemia-specific fusion gene *NUP98-HOXD13* (*ND13*) has myeloproliferative activity, although it does not directly induce AML in mice (59). However, when MEIS1 expression is upregulated, *ND13* can induce cell proliferation and generate AML in transplanted mice (59). *PBX* also functions as a cofactor of MEIS1 to regulate leukemia cell proliferation. Immortalized progenitor cells induced by HOXA9 typically do not undergo leukemic transformation; however, the co-expression of MEIS1 and *PBX* can induce *FLT3* expression, increase MAPK phosphorylation and promote leukemic transformation (60). The co-expression of MEIS1 and *PBX3* can transform normal

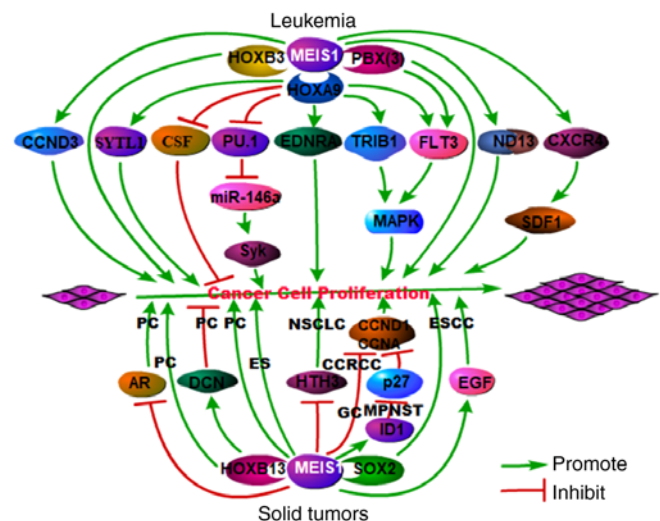


Figure 6. Schematic diagram of the promoting and inhibitory roles of MEIS1 in various types of cancer. Leukemia is illustrated in the upper panel, while solid tumors are presented in the lower panel. MEIS1, Meis homeobox 1; HOXB3, homeobox B3; HOXA9, homeobox A9; PBX3, pre-B-cell leukemia homeobox 3; CCND3, cyclin D3; SYTL1, synaptotagmin-like 1; CSF, colony-stimulating factor; PU.1, spl1; EDNRA, endothelin receptor type A; TRIB1, tribbles pseudokinase 1; FLT3, receptor tyrosine kinase 3; ND13, NUP98-HOXD13; CXCR4, C-X-C motif chemokine receptor 4; Syk, spleen tyrosine kinase; MAPK, mitogen-activated protein kinase; SDF1 (CXCL12), C-X-C motif chemokine ligand 12; PC, prostate cancer; ES, ewing sarcoma; NSCLC, non-small cell lung cancer; CCND1, cyclin D1; CCNA, cyclin A; CCRCC, clear cell renal cell carcinoma; ESCC, esophageal squamous cell carcinoma; AR, androgen receptor; DCN, decorin; HTH3, histone H3; GC, gastric cancer; MPNST, malignant peripheral nerve sheath tumor; ID1, inhibitor of DNA binding 1; EGF, epidermal growth factor; HOXB13, Homeobox B13; SOX2, SRY-box transcription factor 2.

hematopoietic stem/progenitor cells *in vitro* and induce AML (leukemia cell proliferation) in mice (61). MEIS1 is also involved in MLL. MEIS1 regulates the differentiation arrest, cycle activity, *in vivo* invasion and self-renewal of MLL cells, and thus represents a key limiting factor of MLL stem cell potential (62). The downregulation of MEIS1 and HOXA alters

Table I. Roles of MEIS1 in cancer cell proliferation.

Disease	Cell type	MEIS1 expression	Role of MEIS1 in proliferation	MEIS1 interacting partner	(Refs.)
Leukemia	H9M1	↑	Promote	HOXA9	(50)
Leukemia	Bone marrow cell lines	↑	Promote	-	(51)
Leukemia	Myeloid progenitors	↑	Promote	HOXA9	(52)
Leukemia	Myeloid clonogenic progenitor	↑	Promote	HOXA9	(32,53)
Leukemia	Bone marrow progenitors	↑	Promote	HOXA9	(54)
Leukemia	Hematopoietic progenitors	↑	Promote	HOXA9	(55)
Leukemia	Primary leukemia cells	↑	Promote	HOXA9	(57)
Leukemia	Bone marrow cell lines	↑	Promote	HOXA9	(58)
Leukemia	Myeloid clonogenic progenitor	↑	Promote	-	(59)
Leukemia	Hematopoietic progenitors	↑	Promote	PBX	(60)
Leukemia	Hematopoietic stem/progenitor cells	↑	Promote	PBX3	(61)
Leukemia	Precursor B-cell leukemic line, RS4	↑	Promote	-	(63)
Prostate cancer	Human prostate cancer cells, LNCaP and PC-3	↓	Inhibit	-	(49)
Prostate cancer	Human prostate cancer cells, LNCaP	↓	Inhibit	-	(64)
Prostate cancer	Human prostate cancer cells, DU145	↑	Promote	HOXB13	(65)
Prostate cancer	Human prostate cancer cells, DU145	↓	Inhibit	HOXB13	(66)
Non-small cell lung cancer	A549 cells	↓	Inhibit	-	(67)
Gastric cancer	Human GC cell line, MKN28	↓	Inhibit	-	(68)
Clear cell renal cell carcinoma	Human ccRCC cell lines, 786-O or Caki-1	↓	Inhibit	-	(69)
Esophageal squamous cell carcinoma	Human KYSE-30 ESCC cells	↑	Promote	SOX2	(46)
Esophageal squamous cell carcinoma	Human KYSE-30 ESCC cells	↑	Promote	-	(70)
Malignant peripheral nerve sheath	MPNST cell line, STS26T	↑	Promote	-	(47)
Ewing sarcoma	Ewing sarcoma cell lines, A673, SKNMC and TC32	↑	Promote	-	(48)

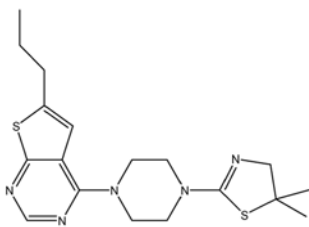
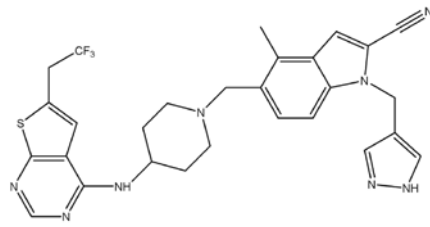
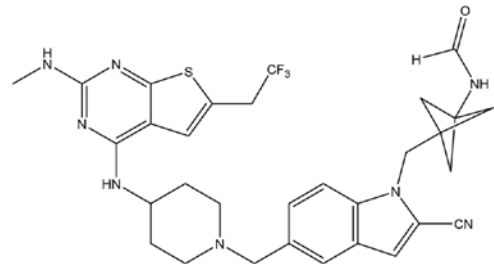
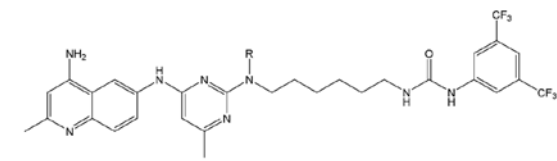
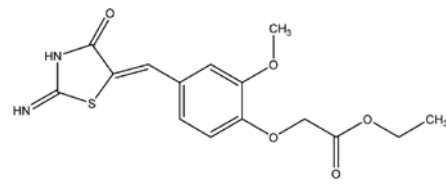
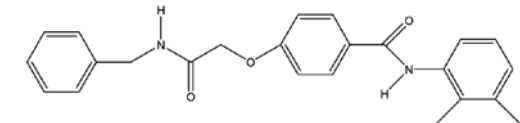
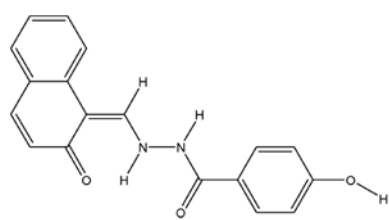
MEIS1, Meis homeobox 1; HOXA9, homeobox A9; PBX, pre-B-cell leukemia homeobox; SOX2, SRY-box transcription factor.

C-X-C motif chemokine receptor 4 (CXCR4)/stromal cell derived factor 1 (SDF-1) signaling to inhibit the proliferation of transplanted mutant MLL-rearranged acute leukemia cells (63), again supporting a pro-proliferative role of MEIS1 in MLL.

Solid tumors. MEIS1 has been associated with a variety of solid cancers, including prostate, non-small cell lung and gastric cancer, as well as clear cell renal cell carcinoma, esophageal squamous cell carcinoma, malignant peripheral nerve sheath tumors and Ewing sarcoma. The MYC-mediated downregulation of MEIS1 upregulates the androgen receptor (AR) to promote prostate cancer cell proliferation (64). AR transcription is inhibited by the overexpression of MEIS1 and is promoted by MEIS1 knockdown with small interfering RNA (49). A proposed mechanism is that MEIS1 interacts with AR to affect the trafficking of androgen between the cytoplasm and nucleus. In this manner, MEIS1 may inhibit prostate cancer

cell proliferation by regulating AR, since this receptor plays a key role in proliferation of human prostate cancer cells. MEIS1 also participates in prostate cancer through mechanisms independent of AR. Recent studies have indicated that MEIS1 regulates the proliferation of prostate cancer cells by interacting with MEIS-interacting domains in HOXB13 (65), which induces decorin (DCN), a multi-RTK inhibitor (66). MEIS1 knockdown has been shown to significantly increase the proliferation of non-small cell lung cancer cells through a mechanism related to DNA synthesis and histone H3 phosphorylation (67). MEIS1 expression is reduced in human gastric cancer and clear cell renal cell carcinoma, and MEIS1 overexpression inhibits cell proliferation by decreasing CCND1 and CCNA expression (68,69). In contrast to its inhibitory roles, MEIS1 maintains the stem cell status and cell proliferation through interaction with SOX2 in esophageal squamous cell carcinoma cells (46). The downregulation of MEIS1 upregulates epithelial markers and downregulates epidermal growth factor (EGF), a

Table II. Basic information about MEIS1 inhibitors.

Name/code	Chemical structural formula	Type of MEIS1 inhibited action	Effect	(Refs.)
MI-2		Indirect	Anti-proliferative effects in MLL cells	(71)
MI-503		Indirect	Inhibitory effects on MLL cells	(72)
MI-3454		Indirect	Anti-proliferative effect in acute leukemia cells and primary patient samples with <i>MLL1</i> translocations or <i>NPM1</i> mutations	(74)
Compound 9e		Indirect	Anti-proliferative activities against leukemia cells	(80)
CCI-007		Indirect	Cytotoxic activity against infant leukemia in <i>MLL-r</i>	(81)
MEISi-1		Direct	Modulated activity in hematopoietic stem cell via inhibiting MEIS1 directly	(82)
MEISi-2		Direct	Anti-proliferative activity in hematopoietic stem cell via inhibiting MEIS1 directly	(82)

MEIS1, Meis homeobox 1; MLL, mixed-lineage leukemia; NPM1, nucleophosmin 1; MLL-r, mixed-lineage leukemia-rearranged; MLL-FPs, mixed-lineage leukemia-fusion proteins.

marker of cell proliferation (70). In malignant peripheral nerve sheath tumors, MEIS1 expression is increased and promotes cell proliferation and maintains cell survival by inhibiting the

cell cycle suppressor, p27, via transcription factor inhibitor of DNA binding 1 (ID1) (47). In Ewing sarcoma, MEIS1 collaborates with EWS-FLI1 to stimulate cell proliferation (48).

6. MEIS1 inhibitors

Since MEIS1 plays an important pro-proliferative role in leukemia and certain solid tumors, it has the potential for use as a therapeutic target. MEIS1 inhibitors under development for treatment of cancer are reviewed and listed in Table II.

Menin-MLL inhibitors have an anti-proliferative function via MEIS1 in leukemia cells. MI-2 is a first-generation small molecule inhibitor of the Menin-MLL interaction, but has poor pharmacological profiles (71). The second-generation inhibitor, MI-503, is highly potent and orally bioavailable, and has been shown to exert profound anti-proliferative effects in MLL cells (72). The cytotoxic concentration of these compounds in cells was $>2 \mu\text{M}$ and the relatively modest effect *in vivo* suggested limited druggability (73). The third generation inhibitor, MI-3454, is well-tolerated and does not impair normal hematopoiesis in mice; however, it inhibits the proliferation and induces the differentiation of acute leukemia cells by downregulating MEIS1 and FLT3 indirectly (74). VTP-50469, another type of highly selective oral Menin-MLL inhibitor, appears to promote leukemia cell differentiation and inhibit cell proliferation through the indirect downregulation of MEIS1 (75-77). All Menin-MLL inhibitors mentioned above are reversible in nature. To achieve an optimal anti-leukemic activity, extended drug exposure is required (78). M-525, a highly potent and irreversible Menin-MLL inhibitor, has been shown to inhibit the proliferation of and suppress MEIS1 expression in MLL cells at a sub-nanomolar concentration (78), indicating that the downregulation of MEIS1 with Menin-MLL inhibitors may represent a promising therapeutic strategy for MLL. In addition to Menin-MLL inhibitors, certain other agents that downregulate MEIS1 indirectly can exert anti-proliferative effects in leukemia cells. For instance, the proton pump inhibitor, rabeprazole, selectively suppresses the proliferation and induces the apoptosis of leukemia cells harboring MLL fusion proteins by downregulating MEIS1 (79). The DOT1-like histone lysine methyltransferase (DOT1L) inhibitor compound, 9e (80), and the MLL-rearranged leukemia inhibitor, CCI-007 (81), also produce similar effects via MEIS1-dependent mechanism.

In a recent study, two small molecule MEIS1 inhibitors (MEISi-1 and MEISi-2) were identified using high-throughput *in silico* screening, and induced hematopoietic stem cell expansion (82).

7. Conclusions and future perspectives

The present review article summarized the characteristics of MEIS1, its role in cancer cell proliferation and the current status on the development of MEIS1 inhibitors as therapeutic agents. MEIS1 is upregulated and mediates cell proliferation in leukemia and certain solid tumors (e.g., esophageal squamous cell carcinoma and malignant peripheral nerve sheath tumors), whereas it inhibits cell proliferation in other solid tumors. Agents that inhibit MEIS1 directly or indirectly are being developed. MEIS1 is increasingly recognized as a marker of cancer diagnosis and a therapeutic target.

The paradoxical role of MEIS1 among solid tumors (stimulatory in some but inhibitory in others) warrants further

investigation. Illustrating the potential association between the Warburg effect, cancer cell proliferation and MEIS1 may resolve this contradiction, as the Warburg effect plays an important role in cell proliferation (83). Designing MEIS1 inhibitors based on the molecular structure of MEIS1 may expedite the developmental process of anticancer agents.

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Availability of data and materials

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Authors' contributions

MY drafted the manuscript. ZC conceived the study and participated in the manuscript preparation. ZY and KZ assisted in the literature search and edited the manuscript. YG revised the manuscript. ZC and MY confirm the authenticity of all the raw data. All authors have read and approved the final manuscript.

Ethics approval and consent to participate

Not applicable.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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