# HIF-1α: Its notable role in the maintenance of oxygen, bone and iron homeostasis (Review)

XINYI HUANG<sup>1,2\*</sup>, YILI ZHANG<sup>3\*</sup>, BAOYU QI<sup>1\*</sup>, KAI SUN<sup>1</sup>, NING LIU<sup>1</sup>, BIN TANG<sup>1</sup>, SHENGJIE FANG<sup>1</sup>, LIGUO ZHU<sup>1,4</sup> and XU WEI<sup>1,4</sup>

<sup>1</sup>Wangjing Hospital, China Academy of Chinese Medical Sciences, Beijing 100102; <sup>2</sup>School of Traditional Chinese Medicine, Beijing University of Chinese Medicine, Beijing 100029; <sup>3</sup>School of Traditional Chinese Medicine and School of Integrated Chinese and Western Medicine, Nanjing University of Chinese Medicine, Nanjing 210023; <sup>4</sup>Institute of Orthopaedics of Beijing Integrative Medicine, Beijing 100061, P.R. China

Received July 28, 2022; Accepted October 12, 2022

DOI: 10.3892/ijmm.2022.5197

Abstract. Hypoxia is a characteristic feature of numerous diseases, including metabolic bone disease, solid tumors, cardiovascular diseases, neurodegeneration and inflammation. It is also a risk factor for a poor prognosis in various diseases. Hypoxia-inducible factor-1 $\alpha$  (HIF-1 $\alpha$ ) is activated by hypoxia to regulate a series of pathophysiological pathways, which is of utmost significance for maintaining body homeostasis. The present review highlights the role of the HIF-1 $\alpha$  in oxygen, bone and iron homeostasis, and alludes on the biological complexity and dual functions of HIF-1 $\alpha$  regulation. In addition, the pathophysiological significance of HIF-1α in bone formation, bone absorption, angiogenesis, erythropoiesis, oxidative stress, energy metabolism, iron death, etc., is discussed An accurate understanding of all these processes may aid in the identification of possible therapeutic targets that may then be used in the treatment of related diseases. However, further studies are required to unravel the extensive complexity of HIF-1a regulation and to develop more precise treatment strategies.

\*Contributed equally

*Abbreviations:* Tf, transferrin; TFR, transferrin receptor; HO-1, heme oxygenase 1; VEGF, vascular endothelial growth factor; EPO, erythropoietin; PDK1, pyruvate dehydrogenase kinase 1; TCA, tricarboxylic acid; PHD, prolyl hydroxylase; BMSCs, bone mesenchymal stem cells; SDF-1, stromal cell-derived factor-1; CXCR4, c-X-c chemokine receptor type 4; MSCs, mesenchymal stem cells; GSH, glutathione; FGF23, fibroblast growth factor 23; GSNO, S-nitrosoglutathione

*Key words:* hypoxia, hypoxia-inducible factor- $1\alpha$ , oxygen homeostasis, bone homeostasis, iron homeostasis

# Contents

- 1. Introduction
- 2. Oxygen homeostasis
- 3. Bone homeostasis
- 4. Iron homeostasis
- 5. Summary and future perspectives

### 1. Introduction

Hypoxia-inducible factor (HIF) heterodimers consist of one of three  $\alpha$ -subunits (HIF-1 $\alpha$ , HIF-2 $\alpha$  and HIF-3 $\alpha$ ) and one  $\beta$ -subunit. HIF-1 $\alpha$ , a 120 kDa polypeptide subunit that heterodimerizes with HIF-1 $\beta$  (a 91 to 94 kDa polypeptide subunit), is a transcription factor regulated by hypoxia (1). Under normoxic conditions, HIF-1 $\alpha$  is hydroxylated to interact with von Hippel-Lindau (VHL) protein for ubiquitination and proteasomal degradation. HIF-1a is expressed in almost all cell types, whereas HIF-2a has a more limited distribution. Under hypoxic conditions, HIF-1 $\alpha$  plays a crucial role in the body's metabolic and functional adaptation to these conditions. All these observations have allowed the identification of HIF-1a as a critical factor in the regulation of homeostasis. It is worth noting that in the field of integrative physiology, research on baroreflex, chemoreflex, glucose regulation and temperature regulation is essentially the study of a series of homeostasis (2-4). Among these, oxygen, bone and iron homeostasis are involved in various critical functions of the body, including bone resorption and formation, mesenchymal stem cell (MSC) homing, angiogenesis, erythropoiesis, oxidative stress, iron metabolism and ferroptosis.

Bone homeostasis is maintained by a balance between osteoblast-mediated bone formation and osteoclast-driven bone resorption (5). Under hypoxic conditions, HIF-1 $\alpha$  exerts a series of direct and indirect effects on this balance (6). Further studies have indicated its critical role in the manipulation of bone mass accrual, bone material properties as well as micro-structures, including bone mineralization, bone collagen fiber formation and bone remodeling (7). Moreover, HIF-1 $\alpha$  is a master regulator of oxygen homeostasis in the body, which

*Correspondence to:* Professor Liguo Zhu or Professor Xu Wei, Wangjing Hospital, China Academy of Chinese Medical Sciences, 6 South Zhonghuan Road, Chaoyang, Beijing 100102, P.R. China E-mail: tcmspine@163.com E-mail: weixu.007@163.com

can induce the expression of angiogenic factors, promote glycolysis, increase the delivery of oxygen and nutrients (8). HIF-1 $\alpha$  also plays a key role in iron homeostasis by activating the transcription of iron metabolism genes, such as transferrin (Tf), transferrin receptor (TFR), ceruloplasmin and heme oxygenase 1 (HO-1) (9,10). Roxadustat, a HIF-prolyl hydroxy-lase inhibitor, has been shown to improve iron metabolism in phase 3 trials (11,12).

However, over the years, although the association between HIF-1 $\alpha$  and oxygen, bone and iron homeostasis has been the subject of increasing attention, no consensus has yet been reached on the role of HIF-1 $\alpha$ , at least to the best of our knowledge. Research into its effects on osteocyte apoptosis and osteocyte-mediated osteoclasts has also yielded non-univocal results (13-15). In addition, the local activation of HIF-1 $\alpha$  is required for chondrocyte survival in the center of the expanding growth plate; however, the cellular-intrinsic mechanisms remain unclear (16) The expression of the majority of HIF-1a-dependent genes contributes to the adaption of hypoxic environments in the human body. For example, the increase in the delivery of oxygen to hypoxic tissues is associated with the expression of erythropoietin (EPO) and glycolytic enzymes, which allows for the increased conversion of glucose to produce energy (17). However, HIF-1 $\alpha$  can also play a negative role in the hypoxic process.

Overall, considering the numerous processes in which HIF-1 $\alpha$  is involved and the yet not fully defined underlying mechanisms, the present review focused on the intimate association between HIF-1 $\alpha$  and bone homeostasis, oxygen homeostasis, as well as iron homeostasis. In addition, the pathophysiological significance of HIF-1 $\alpha$  in bone formation, bone absorption, angiogenesis, erythropoiesis, oxidative stress, energy metabolism, iron death, etc. is also discussed (Fig. 1). HIF-1 $\alpha$  is a promising target for the treatment of related diseases, and further information is required to determine the clinical utility of this factor.

#### 2. Oxygen homeostasis

HIF-1 $\alpha$ , mediating the expression of a series of genes, has been strongly established as a critical factor for maintaining oxygen homeostasis. The regulation of oxygen homeostasis is considered to be achieved by oxygen delivery and oxygen. Oxygen delivery is involved in the control of erythropoiesis, angiogenesis and vascular remodeling. Oxygen utilization is implicated in the regulation of glucose metabolism and redox homeostasis (18).

Oxygen delivery. Vascular endothelial growth factor (VEGF) is the most potent proangiogenic factor. EPO, a glycoprotein, is considered as the principal stimulator for erythropoiesis primarily. The expression of HIF-1 $\alpha$  is induced by a hypoxic environment, and it subsequently upregulates downstream key factors, such as EPO and VEGF, which promote angiogenesis to adapt to the environment and recover the oxygen content.

The primary cause of the ectopic overexpression of VEGF in tumors is the dysregulated expression of HIF-1 $\alpha$  involving the c-X-c chemokine receptor type 4 (CXCR4)/stromal-derived cell factor-1 (SDF-1) axis (19). The study by Li *et al* (20) conducted on cerebral ischemic rats, found that HIF-1 $\alpha$ 

attenuated neuronal apoptosis, partially by upregulating EPO expression. There is a novel molecular mechanism for the anti-angiogenic effects of peroxisome proliferator-activated receptor  $\alpha$ , which are achieved by inhibiting ischemia-induced EPC mobilization and homing through the inhibition of the HIF-1 $\alpha$ /SDF-1 pathway (21). Rankin *et al* (22) found that osterix-VHL mice with a deficiency in VHL in osteoblasts exhibited overexpressed HIFs, accompanied by a significant increase in circulating red blood cells. Gerri *et al* (23) reported that HIF-1 $\alpha$  regulated hematopoietic stem cells upstream of the Notch signaling pathway.

Oxygen utilization. HIF-1 $\alpha$ , in response to hypoxic irritation, participates in the regulation of glucose transporters and glycolytic enzymes, which are key genes in energy metabolism and exert critical effects on cell survival (24). Moreover, HIF-1 $\alpha$  inhibits pyruvate dehydrogenase by activating pyruvate dehydrogenase kinase 1 (PDK1), and thereby, pyruvate is redirected from the tricarboxylic acid (TCA) cycle and converted into lactate (25).

The overexpression of constitutive cardiac-specific HIF-1 $\alpha$  leads to changes in cellular metabolism and increased glucose utilization, subsequently resulting in cardiomyopathy in aging mice (26). On the other hand, the deletion of HIF-1a in cardiomyocytes results in decreased ATP, lactate and phosphocreatine levels, and inn an impaired myocardial contractility (27). Chondrocytes maintain an optimal energy balance during endochondral ossification, which is achieved by confined HIF-1 $\alpha$  signaling (28). However, it is only under hypoxic conditions that glucose uptake and bone resorption can be affected by HIF-1 $\alpha$ knockdown. HIF-1α promotes glycolysis during hypoxia; however, it also affects metabolism under normoxic conditions. A decreased HIF-1a activity also has effects on mitochondrial metabolism that results in mitochondrial loss and lipid accumulation, along with reduced oxidative phosphorylation and fatty acid metabolism (26,29). In addition, studies have demonstrated that HIF-dependent metabolic processes can also modulated by dimethyloxalylglycine, desferrioxamine, prolyl hydroxylase (PHD) and other small molecules (30,31).

Oxidative stress. HIF-1 $\alpha$  is an endogenous anti-oxidative stress modulator. The oxidative stress pathway induces the activation of HIF-1 $\alpha$ , and increases the production of mitochondrial complex II-mediated reactive oxygen species (ROS) (32,33). Moreover, it has been demonstrated that increased superoxide anion radicals induce PHD inactivation, resulting in the stabilization and accumulation of HIF-1 $\alpha$  (34). Under hypoxic conditions, HIF-1 $\alpha$ dynamically regulates glucose flux through the glycolytic pathway to resist the increased risk of ROS production and confers protection against apoptosis and renal injury in diabetes (35,36).

In recent years, increasing evidence has indicated that HIF-1 $\alpha$  can enhance antioxidant activity and neuroprotection (37,38). HIF-1 $\alpha$  has the ability to mitigate this toxicity or regulate redox homeostasis by limiting TCA activity, regulating the levels of NADPH and glutathione (GSH), and reducing mitochondrial mass through the upregulation of the



Figure 1. Role of the HIF-1 $\alpha$  in oxygen homeostasis, bone homeostasis, and iron homeostasis. +, indicates increased expression;  $\uparrow$ , promoting function;  $\downarrow$ , inhibitory function; HIF-1 $\alpha$ , hypoxia-inducible factor 1  $\alpha$ ; BNIP3, Bcl-2 interacting protein 3; CT-1, cardiotrophin-1; RANKL, receptor activator of nuclear factor  $\kappa$ B ligand; OPG, osteoprotegerin; ROS, reactive oxygen species; SDF-1, stromal cell-derived factor-1; CXCR4, c-X-c chemokine receptor type 4; MSC, mesenchymal stem cell; BMSCs, bone mesenchymal stem cells; VEGF, vascular endothelial growth factor; EPO, erythropoietin; PDK1, pyruvate dehydrogenase kinase 1; GSH, glutathione; TCA, tricarboxylic acid; Dcytb, duodenal cytochrome B; DMT1, divalent metal transporter 1; FPN1, ferroportin1; iASPP, inhibitor of apoptosis-stimulating protein of p53; Nrf2, nuclear factor erythroid 2-related factor 2; Tf, transferrin.

mitochondrial proteins, PDK1 and Bcl-2 interacting protein 3 (BNIP3) (39). Furthermore, HIF-1 $\alpha$  may be an indirect player in the promotion of mitochondrial-selective autophagy and may subsequently lower the mitochondrial mass, which inhibits the oxidation of both glucose and fatty acids, and reduces mitochondrial ROS production under hypoxic conditions (40). A previous study revealed that HIF-1 $\alpha$  can activate the nuclear factor erythroid 2-related factor 2 (Nrf2)/ARE pathway to protect against ischemia-reperfusion cardiac and skeletal muscle injuries (41).

#### 3. Bone homeostasis

Bone formation. Previous research has indicated that HIF-1 $\alpha$  may affect the osteogenesis of osteoblasts through the prevention of chondrocyte cell death in the growth plate, and also via direct or indirect actions on the delivery of oxygen and nutrients, together with metabolic adaptations (8). It has been reported that the overexpression of HIF-1 $\alpha$ , through its downstream marker, BNIP3, reduces the inhibitory effects of

dexamethasone on hypoxia-induced mitophagy and protects osteocytes from apoptosis (42). There is also evidence to suggest that miRNA-21, by upregulating the activation of HIF-1 $\alpha$  and p-Akt, can promote the osteogenic ability of bone MSCs (BMSCs) (43). HIF-1 $\alpha$  does not only promote osteogenesis, but also has repairing effects on bone (44). Moreover, it has been demonstrated that prolonged HIF-1 $\alpha$  signaling in chondrocytes, interfering with cellular bioenergetics and biosynthesis, results in skeletal dysplasia by collagen overmodification (27).

*Bone resorption.* The delicate balance between osteoblastic bone formation and osteoclastic bone resorption is a key factor in the regulation of mature bone tissue formation (45). Nevertheless, no consensus has yet been reached on the role of HIF-1 $\alpha$  in regulating osteoclast differentiation, at least to the best of our knowledge.

Promotion of osteoclast differentiation. HIF-1 $\alpha$  expression has been proposed to increase bone erosion in rheumatoid arthritis (46). HIF-1 $\alpha$  is involved in the increase

of osteoclastogenesis and bone resorption, since it has recently been shown to enhance the osteoclast-mediated stimulation of BMSC differentiation by secreting cardiotrophin-1 (47). Moreover, HIF-1 $\alpha$  functions by activating the JAK2/STAT3 pathway, promoting the expression of RANKL, and thus enhancing the differentiation of osteocyte-mediated osteoclastic in vitro (48). HIF-1α also plays a pro-apoptotic role in JNK/caspase-3 signaling pathway activation. Osteocyte-mediated osteoclastogenesis has been shown to be reduced with a concomitant decrease in HIF-1 $\alpha$  and caspase-3 expression (49). A previous study demonstrated that the deceleration of osteoclastogenesis occurred under conditions of HIF-1a deficiency, by inhibiting AMPK signaling under anoxic conditions (50). Of note, HIF-1a knockdown reduces bone resorption under both normoxic and hypoxic conditions. Thus, the targeting of HIF-1 $\alpha$  may prove to be of value in th treatment of osteoporosis (13).

Inhibition of osteoclast differentiation. Both the VHL/HIF and PHD/HIF signaling pathways in osteoblasts have been shown to reduce osteoclastogenesis by increasing osteoprotegerin expression and inhibiting sclerostin expression, resulting in increased bone formation and decreased resorption (14,51). In addition, it has been suggested that the activation of osteoblast HIF-1 $\alpha$  contributes to the inhibition of osteoclastogenesis, by increasing IL-33 expression (52).

Angiogenesis. A vital role in bone remodeling and vascularization is attributed to the HIF- $1\alpha$ /VEGF signaling pathway (53). The study by Kusumbe *et al* (54) demonstrated endothelial HIF- $1\alpha$  as a critical promoter of type H vessel formation in the metaphysis. HIF- $1\alpha$  has also been implicated in the increased number of type-H vessels and the restoration of bone mass (55-57). The miR-497B195 cluster has been proposed to regulate angiogenesis during coupling with osteogenesis, by maintaining endothelial Notch and HIF- $1\alpha$  activity (58). Furthermore, HIF- $1\alpha$  may have a dual function in the regulation of osteogenesis-angiogenesis coupling of long bone via the ROS-HIF $1\alpha$ /p53 axis (59).

MSC migration. HIF-1 $\alpha$  has also been demonstrated to trigger wound healing and functional recovery by regulating corresponding stem cells (60,61). BMSCs, a class of heterogeneous cells, have a series of feasible and diverse clinical values for generating stroma which can support hematopoiesis, bones, adipocytes and cartilage (62).

HIF-1 $\alpha$  regulates the expression levels of surface molecules, such as SDF-1, a downstream gene of HIF-1 $\alpha$ , which binds to its specific receptor, CXCR4, forming a pair of coupling molecules and promoting stem cell migration to ischemic and hypoxic sites (63). Guo *et al* (62) demonstrated that the HIF-1 $\alpha$ /SDF-1/CXCR4 axis enhanced BMSC migration, and alleviated neuronal damage and apoptosis. Moreover, there is evidence to suggest that the increased secretion of HIF-1 $\alpha$  induced by the hypoxic conditions of surrounding brain tissue accelerates the fracture repair process via chemotaxis due to the SDF-1/CXCR4 axis. In addition, the silencing of HIF-1 $\alpha$  has been shown to decrease MSC migration, as well as the mRNA and protein levels of SDF-1 and CXCR4 in MSCs (64).

#### 4. Iron homeostasis

*Iron metabolism*. HIF-1 $\alpha$  is a vital factor in iron metabolism by regulating the expression of iron-related proteins, such as divalent metal transporter 1, ferroportin 1, duodenal cytochrome B and TFR (65,66). An overload of iron has been found to be related to the dysfunction of MSCs and to the damage of the microenvironment that may be involved in the pathogenesis of myelodysplastic syndromes, and which may be achieved by the regulation of cytokine of MSCs through the ROS/HIF-1 $\alpha$ pathway (67). It has been demonstrated that HIF-1a induces TFR1 expression, thereby increasing iron uptake (68). In addition, it has been suggested that HO-1, induced by HIF-1 $\alpha$ , degrades heme into biliverdin, carbon monoxide and iron (69). Weinreb et al (70) and Guo et al (71) demonstrated that in various brain regions of adult mice, the upregulation of HIF-1 $\alpha$ by the iron chelator, M30, results in differentially induced levels of TFR, tyrosine hydroxylase and EPO (72).

*Ferroptosis*. Ferroptosis is a new form of regulated cell death as a result of iron-dependent lipid peroxidation (73). Moreover, HIF-1 $\alpha$  downregulation also promotes ferroptosis by inducing ferritin heavy chain degradation in RANKL-stimulated bone marrow-derived macrophages. A previous *in vitro* study demonstrated that the overexpression of inhibitor of apoptosis-stimulating protein of p53 inhibited ferroptosis through the Nrf2/HIF-1 $\alpha$ /TF signaling pathway (74). In another study, following pre-treatment with roxadustat (an inhibitor of HIF prolyl hydroxylase), the risk of ferroptosis was correspondingly reduced, along with increased levels of antioxidant enzymes and GSH, and decreased iron accumulation (75).

### 5. Summary and future perspectives

Constant oxygen supply is essential for proper tissue function, development and homeostasis. Hypoxia is a distinctive feature of diseases, including solid tumors, metabolic bone disease, cardiovascular diseases, neurodegeneration, inflammation and other chronic diseases (76,77). It is also a risk factor for a poor prognosis in various diseases. For example, hypoxia is responsible for the failure of the majority of solid tumors to respond to treatment, and promotes drug resistance (78). Under normoxic conditions, HIF is rapidly degraded by the high activity of PHDs. However, under hypoxic conditions, the shortage of oxygen results in the dimerization of non-hydroxylated and non-degradable HIF-1 $\alpha$  with HIF-1 $\beta$ , which binds to hypoxia-responsive elements in the regulatory regions of oxygen-sensitive genes. Given the ubiquitous localization of HIF-1 $\alpha$ , HIF-1 $\alpha$  acts as a main regulator in the expression of several thousand genes coding, including growth factors, enzymes, transcription factors, cytokines, hormones, receptors, solute transporters, ion channels and other essential regulators, which are involved in almost every cell function or dysfunction (79).

There is no doubt that bone, oxygen and iron homeostasis are of utmost significance to the human body, and the role of HIF-1 $\alpha$  in the maintenance of homeostasis cannot be ignored. Although HIF-1 $\alpha$  plays a beneficial role in the regulation of bone homeostasis, the degree of HIF-1 $\alpha$  pathway activation must be fine-tuned to avoid the disruption of

| Differential expression at protein/gene levels | Mechanisms  | Effects  | (Refs.) |
|--|---|--|---------|
| VEGF   | Chemokine receptor 4/stromal-derived cell factor 1 (CXCR4/SDF-1) axis                                       | Promotes tumor angiogenesis  | (19)    |
| EPO  | Upregulating EPO  | Attenuates neuronal apoptosis  | (20)    |
| SDF-1  | HIF-1a/SDF-1 pathway  | Anti-angiogenic effect   | (21)    |
| PDK1   | Inactivates pyruvate dehydrogenase (PDH)  | Increases ATP levels and prevents toxic ROS production                     | (39)    |
| NADPH and GSH                                  | Switches cells from oxidative to glycolytic<br>metabolism, to reduce mitochondrial<br>superoxide generation | Maintains redox homeostasis  | (40)    |
| Nrf2   | Nrf2/ARE pathway  | Protects against ischemia-reperfusion cardiac and skeletal muscle injuries | (41)    |
| BNIP3  | Reduces the inhibitory effects of DEX on hypoxia-induced mitophagy  | Protects bone cells from apoptosis   | (42)    |
| CT-1   | Enhances the osteoclast-mediated stimulation of BMSC differentiation  | Bone resorption  | (47)    |
| RANKL  | JAK2/STAT3 pathway  | Enhances the differentiation of osteocyte-mediated osteoclastic            | (48)    |
| IL-33  | Acts on bone marrow-derived monocytes   | Contributes to osteoclastogenesis inhibition                               | (52)    |
| SDF-1  | HIF-1α/SDF-1/CXCR4 axis   | Alleviates neuronal damage and apoptosis                                   | (62)    |

Table I. Mechanisms and effects of HIF-1 $\alpha$  on various diseases.

VEGF, vascular endothelial growth factor; EPO, erythropoietin; PDK1, pyruvate dehydrogenase kinase 1; DEX, dexamethasone; CT-1, cardiotrophin-1; IL-33, interleukin-33; SDF-1, stromal cell-derived factor-1; CXCR4, c-X-c chemokine receptor type 4; RANKL, receptor activator of nuclear factor kappa-B ligand; GSH, glutathione; BNIP3, Bcl-2 interacting protein 3; Nrf2, nuclear factor erythroid 2-related factor 2; RANKL, receptor activator of nuclear factor κB ligand.

homeostasis (8,80). Previous studies have confirmed that osteoblastic HIF-1 $\alpha$  affects bone formation (81-83); however, its role in osteoclasts remains controversial. An interesting aspect is that HIF-1a has minimal effects on osteoclast differentiation, although it predominantly functions as a regulator of osteoclast-mediated bone resorption (84). Similarly, the knockdown of HIF-1a does not affect the process of osteoclast differentiation, although it prevents the increased bone resorption under hypoxic conditions (6). Moreover, HIF-1 $\alpha$ has been demonstrated to regulate osteogenesis-angiogenesis coupling bidirectionally, and the effect is age-related (56). There is also evidence to suggest that HIF-1 $\alpha$  functions by increasing EPO levels directly or indirectly, inducing the expression and processing of fibroblast growth factor23 (FGF23), and thus affecting mineral homeostasis and vitamin D metabolism (8). Of note, increased serum FGF23 levels have been reported to be associated with the reduction of serum phosphate or 1,25(OH)2D, which in turn may alter bone homeostasis, although further confirmation is required (85,86).

Taken together, HIF-1 $\alpha$  expression is mainly induced by hypoxic stress and is common during the development of various diseases. The present review mainly focused on the role of HIF-1 $\alpha$  in regulating oxygen, bone and iron homeostasis. Although significant progress has been made in the understanding of the pathogenesis of diseases, such as atherosclerosis and emerging drug treatments, the current treatment options continue to have a number of deficiencies. Regulating the expression and signaling pathways of HIF-1 $\alpha$  may be a promising strategy for the treatment of diseases involving the pathophysiology of hypoxia (Table I). To date, a number of active ingredients of traditional Chinese medicine and natural products have been found to regulate the HIF-1 $\alpha$  content (87). At present, HIF-1 $\alpha$  inhibitors have been used to treat various diseases, such as tumors, leukemia, diabetes, ischemic cardiovascular and brain diseases, etc. Manassantin A and Manassantin B exert potent inhibitory effects on the secretion of hypoxia-induced VEGF, cyclin-dependent kinase inhibitor 1 and GLUT-1 genes (88,89). Lificiguat (YC-1) is a targeted HIF-1 $\alpha$  inhibitor, which can reduce HIF-1 $\alpha$  protein expression and is associated with the enhancement of EGFR degradation, thereby exerting antitumor effects (90). The benefit of S-nitrosoglutathione on traumatic brain injury is mediated by S-nitrosylation to stabilize HIF-1 $\alpha$  (91).

Notably, immense efforts and resources have been invested in identifying possible effective and specific small-molecules inhibitors of HIF-1 $\alpha$ . HIF-1 $\alpha$ , as a common pathophysiological mechanism of numerous diseases, plays an exploratory role in the treatment of comorbidities. However, there are several questions and challenges involved in translating the findings from mechanobiological studies into novel HIF-1 $\alpha$ -targeted therapeutics. The potential interaction network of multiple molecules regulates the expression of important genes. Important interactions between NF- $\kappa$ B and HIF-1 $\alpha$  (92-94) have been described recently. In addition, efficacy needs to be supported by high-quality clinical trial evidence.

HIF-1 $\alpha$  is a master regulator of homeostasis, and plays critical roles in physiological and pathological processes. Understanding the roles and regulation mechanisms of HIF-1 $\alpha$  in bone, oxygen and iron homeostasis may open a new era in the development of therapeutic strategies against a variety of pathologic conditions, such as metabolic bone disease, ischemic/hypoxic injuries, tumor growth, wound healing and cardiovascular remodeling.

### Acknowledgements

Not applicable.

### Funding

The present study was supported by the National Natural Science Foundation of China (grant nos. 82174416 and 82205140), the Innovation Team and Talents Cultivation Program of National Administration of Traditional Chinese Medicine (grant no. ZYYCXTD-C-202003), the Basic Research Program of Jiangsu Province (Natural Science Foundation; grant no. BK20220468), the National Natural Science Foundation of China (NSFC) Matching Project of Nanjing University of Chinese Medicine (grant no. XPT82205140), and the Fundamental Research Funds for the Central Public Welfare Research Institutes (grant no. ZZ13-YQ-039).

#### Availability of data and materials

Not applicable.

#### **Authors' contributions**

XW and LZ were responsible for the conceptualization of the study. XH was responsible for the research design. XH and YZ were responsible for the determination of the research design. XH and YZ wrote the manuscript. BQ, KS and NL were responsible for the literature search. BT and SF completed the references and were involved in document management and preparation. YZ and XH prepared the original draft. XW and LZ reviewed and edited the manuscript. All authors contributed to the article and have read and approved the submitted version. Data authentication is not applicable.

#### Ethics approval and consent to participate

Not applicable.

#### Patient consent for publication

Not applicable.

## **Competing interests**

The authors declare that they have no competing interests.

#### References

- 1. Greijer AE, van der Groep P, Kemming D, Shvarts A, Semenza GL, Meijer GA, van de Wiel MA, Belien JA, van Diest PJ and van der Wall E: Up-regulation of gene expression by hypoxia is mediated predominantly by hypoxia-inducible factor 1 (HIF-1). J Pathol 206: 291-304, 2005.
- 2. Bentley ER and Little SR: Local delivery strategies to restore immune homeostasis in the context of inflammation. Adv Drug Deliv Rev 178: 113971, 2021.
- Goldstein DS: How does homeostasis happen? Integrative physiological, systems biological, and evolutionary perspectives. Am J Physiol Regul Integr Comp Physiol 316: R301-R317, 2019.
- Suciadi LP, Henrina J, Putra ICS, Cahyadi I and Gunawan HFH: Chronic heart failure: Clinical implications of iron homeostasis disturbances revisited. Cureus 14: e21224, 2022.
- 5. Lee SY, Park KH, Yu HG, Kook E, Song WH, Lee G, Koh JT, Shin HI, Choi JY, Huh YH and Ryu JH: Controlling hypoxia-inducible factor-2α is critical for maintaining bone homeostasis in mice. Bone Res 7: 14, 2019.
- Knowles HJ: Distinct roles for the hypoxia-inducible transcription factors HIF-1α and HIF-2α in human osteoclast formation and function. Sci Rep 10: 21072, 2020.
  Chen S, Xiao L, Li Y, Qiu M, Yuan Y, Zhou R, Li C, Zhang L,
- Chen S, Xiao L, Li Y, Qiu M, Yuan Y, Zhou R, Li C, Zhang L, Jiang ZX, Liu M and Zhou X: Osteocytic HIF-1α pathway manipulates bone micro-structure and remodeling via regulating osteocyte terminal differentiation. Front Cell Dev Biol 9: 721561, 2021.
- Stegen S and Carmeliet G: Hypoxia, hypoxia-inducible transcription factors and oxygen-sensing prolyl hydroxylases in bone development and homeostasis. Curr Opin Nephrol Hypertens 28: 328-335, 2019.
- 9. Das NK, Schwartz AJ, Barthel G, Inohara N, Liu Q, Sankar A, Hill DR, Ma X, Lamberg O, Schnizlein MK, *et al*: Microbial metabolite signaling is required for systemic iron homeostasis. Cell Metab 31: 115-130, 2020.
- Galaris D, Barbouti A and Pantopoulos K: Iron homeostasis and oxidative stress: An intimate relationship. Biochim Biophys Acta Mol Cell Res 1866: 118535, 2019.
- 11. Ikeda Y: Novel roles of HIF-PHIs in chronic kidney disease: The link between iron metabolism, kidney function, and FGF23. Kidney Int 100: 14-16, 2021.
- Chen N, Hao C, Peng X, Lin H, Yin A, Hao L, Tao Y, Liang X, Liu Z, Xing C, *et al*: Roxadustat for anemia in patients with kidney disease not receiving dialysis. N Engl J Med 381: 1001-1010, 2019.
- Ni S, Yuan Y, Qian Z, Zhong Z, Lv T, Kuang Y and Yu B: Hypoxia inhibits RANKL-induced ferritinophagy and protects osteoclasts from ferroptosis. Free Radic Biol Med 169: 271-282, 2021.
- 14. Shao J, Zhang Y, Yang T, Qi J, Zhang L and Deng L: HIF-1α disturbs osteoblasts and osteoclasts coupling in bone remodeling by up-regulating OPG expression. In Vitro Cell Dev Biol Anim 51: 808-814, 2015.
- Meng X, Wielockx B, Rauner M and Bozec A: Hypoxia-inducible factors regulate osteoclasts in health and disease. Front Cell Dev Biol 9: 658893, 2021.
- 16. Maes C, Araldi E, Haigh K, Khatri R, Van Looveren R, Giaccia AJ, Haigh JJ, Carmeliet G and Schipani E: VEGF-independent cell-autonomous functions of HIF-1α regulating oxygen consumption in fetal cartilage are critical for chondrocyte survival. J Bone Miner Res 27: 596-609, 2012.
- Papandreou I, Cairns RA, Fontana L, Lim AL and Denko NC: HIF-1 mediates adaptation to hypoxia by actively downregulating mitochondrial oxygen consumption. Cell Metab 3: 187-197, 2006.
- Semenza GL: Hypoxia-inducible factor 1 and cardiovascular disease. Annu Rev Physiol 76: 39-56, 2014.
   de Nigris F, Crudele V, Giovane A, Casamassimi A, Giordano A,
- de Nigris F, Crudele V, Giovane A, Casamassimi A, Giordano A, Garban HJ, Cacciatore F, Pentimalli F, Marquez-Garban DC, Petrillo A, *et al*: CXCR4/YY1 inhibition impairs VEGF network and angiogenesis during malignancy. Proc Natl Acad Sci USA 107: 14484-14489, 2010.
- 20. Li J, Tao T, Xu J, Liu Z, Zou Z and Jin M: HIF-1α attenuates neuronal apoptosis by upregulating EPO expression following cerebral ischemia-reperfusion injury in a rat MCAO model. Int J Mol Med 45: 1027-1036, 2020.
- Wang Z, Moran E, Ding L, Cheng R, Xu X and Ma JX: PPARα regulates mobilization and homing of endothelial progenitor cells through the HIF-1α/SDF-1 pathway. Invest Ophthalmol Vis Sci 55: 3820-3832, 2014.

- 22. Rankin EB, Wu C, Khatri R, Wilson TL, Andersen R, Araldi E, Rankin AL, Yuan J, Kuo CJ, Schipani E and Giaccia AJ: The HIF signaling pathway in osteoblasts directly modulates erythropoiesis through the production of EPO. Cell 149: 63-74, 2012.
- 23. Gerri C, Marass M, Rossi A and Stainier DYR: Hif-1α and Hif-2α regulate hemogenic endothelium and hematopoietic stem cell formation in zebrafish. Blood 131: 963-973, 2018.
- 24. Jimenez-Blasco D, Busquets-Garcia A, Hebert-Chatelain E, Serrat R, Vicente-Gutierrez C, Ioannidou C, Gómez-Sotres P, Lopez-Fabuel I, Resch-Beusher M, Resel E, *et al*: Glucose metabolism links astroglial mitochondria to cannabinoid effects. Nature 583: 603-608, 2020.
- 25. Cerychova R and Pavlinkova G: HIF-1, Metabolism, and diabetes in the embryonic and adult heart. Front Endocrinol (Lausanne) 9: 460, 2018.
- 26. Hölscher M, Schäfer K, Krull S, Farhat K, Hesse A, Silter M, Lin Y, Pichler BJ, Thistlethwaite P, El-Armouche A, *et al*: Unfavourable consequences of chronic cardiac HIF-1α stabilization. Cardiovasc Res 94: 77-86, 2012.
- 27. Huang Y, Hickey RP, Yeh JL, Liu D, Dadak A, Young LH, Johnson RS and Giordano FJ: Cardiac myocyte-specific HIF-1alpha deletion alters vascularization, energy availability, calcium flux, and contractility in the normoxic heart. FASEB J 18: 1138-1140, 2004.
- 28. Stegen S, Laperre K, Eelen G, Rinaldi G, Fraisl P, Torrekens S, Van Looveren R, Loopmans S, Bultynck G, Vinckier S, *et al*: HIF-1α metabolically controls collagen synthesis and modification in chondrocytes. Nature 565: 511-515, 2019.
- 29. Ambrose LJ, Abd-Jamil AH, Gomes RS, Carter EE, Carr CA, Clarke K and Heather LC: Investigating mitochondrial metabolism in contracting HL-1 cardiomyocytes following hypoxia and pharmacological HIF activation identifies HIF-dependent and independent mechanisms of regulation. J Cardiovasc Pharmacol Ther 19: 574-585, 2014.
- Semenza GL: Pharmacologic targeting of hypoxia-inducible factors. Annu Rev Pharmacol Toxicol 59: 379-403, 2019.
- Knutson AK, Williams AL, Boisvert WA and Shohet RV: HIF in the heart: Development, metabolism, ischemia, and atherosclerosis. J Clin Invest 131: e137557, 2021.
- 32. Jiang L, Zeng H, Ni L, Qi L, Xu Y, Xia L, Yu Y, Liu B, Yang H, Hao H and Li P: HIF-1α preconditioning potentiates antioxidant activity in ischemic injury: The role of sequential administration of Dihydrotanshinone I and Protocatechuic aldehyde in Cardioprotection. Antioxid Redox Signal 31: 227-242, 2019.
- 33. Li X, Żhang Q, Nasser MI, Xu L, Žhang X, Zhu P, He Q and Zhao M: Oxygen homeostasis and cardiovascular disease: A role for HIF? Biomed Pharmacother 128: 110338, 2020.
- 34. Wu LY, He YL and Zhu LL: Possible Role of PHD Inhibitors as Hypoxia-mimicking agents in the maintenance of neural stem cells' self-renewal properties. Front Cell Dev Biol 6: 169, 2018.
- 35. Zheng X, Narayanan S, Xu C, Eliasson Angelstig S, Grünler J, Zhao A, Di Toro A, Bernardi L, Mazzone M, Carmeliet P, *et al*: Repression of hypoxia-inducible factor-1 contributes to increased mitochondrial reactive oxygen species production in diabetes. Elife 11: e70714, 2022.
- 36. Semenza GL: Hypoxia-inducible factors: Coupling glucose metabolism and redox regulation with induction of the breast cancer stem cell phenotype. EMBO J 36: 252-259, 2017.
- 37. Wu K, Zhou K, Wang Y, Zhou Y, Tian N, Wu Y, Chen D, Zhang D, Wang X, Xu H and Zhang X: Stabilization of HIF-1α by FG-4592 promotes functional recovery and neural protection in experimental spinal cord injury. Brain Res 1632: 19-26, 2016.
- 38. He Q, Ma Y, Liu J, Zhang D, Ren J, Zhao R, Chang J, Guo ZN and Yang Y: Biological functions and regulatory mechanisms of hypoxia-inducible factor-1α in Ischemic Stroke. Front Immunol 12: 801985, 2021.
- 39. Kim JW, Tchernyshyov I, Semenza GL and Dang CV: HIF-1-mediated expression of pyruvate dehydrogenase kinase: A metabolic switch required for cellular adaptation to hypoxia. Cell Metab 3: 177-185, 2006.
- Samanta D and Semenza GL: Maintenance of redox homeostasis by hypoxia-inducible factors. Redox Biol 13: 331-335, 2017.
- 41. Ji W, Wang L, He S, Yan L, Li T, Wang J, Kong AT, Yu S and Zhang Y: Effects of acute hypoxia exposure with different durations on activation of Nrf2-ARE pathway in mouse skeletal muscle. PLoS One 13: e0208474, 2018.
- 42. Xu K, Lu C, Ren X, Wang J, Xu P and Zhang Y: Overexpression of HIF-1α enhances the protective effect of mitophagy on steroid-induced osteocytes apoptosis. Environ Toxicol 36: 2123-2137, 2021.

- 43. Yang C, Liu X, Zhao K, Zhu Y, Hu B, Zhou Y, Wang M, Wu Y, Zhang C, Xu J, *et al*: miRNA-21 promotes osteogenesis via the PTEN/PI3K/Akt/HIF-1α pathway and enhances bone regeneration in critical size defects. Stem Cell Res Ther 10: 65, 2019.
- 44. Yu Y, Ma L, Zhang H, Sun W, Zheng L, Liu C and Miao L: EPO could be regulated by HIF-1 and promote osteogenesis and accelerate bone repair. Artif Cells Nanomed Biotechnol 48: 206-217, 2020.
- 45. Nakashima T, Hayashi M and Takayanagi H: New insights into osteoclastogenic signaling mechanisms. Trends Endocrinol Metab 23: 582-590, 2012.
- 46. Doi K, Murata K, Ito S, Suzuki A, Terao C, Ishie S, Umemoto A, Murotani Y, Nishitani K, Yoshitomi H, *et al*: Role of Lysine-Specific Demethylase 1 in Metabolically Integrating Osteoclast Differentiation and Inflammatory Bone Resorption Through Hypoxia-Inducible Factor 1α and E2F1. Arthritis Rheumatol 74: 948-960, 2022.
- 47. Tian Y, Shao Q, Tang Y, Li X, Qi X, Jiang R, Liang Y and Kang F: HIF-1α regulates osteoclast activation and mediates osteogenesis during mandibular bone repair via CT-1. Oral Dis 28: 428-441, 2022.
- 48. Zhu J, Tang Y, Wu Q, Ji YC, Feng ZF and Kang FW: HIF-1α facilitates osteocyte-mediated osteoclastogenesis by activating JAK2/STAT3 pathway in vitro. J Cell Physiol 234: 21182-21192, 2019.
- 49. Song X, Tang Y, Zhu J, Tian Y, Song Z, Hu X, Hong C, Cai Y and Kang F: HIF-1α induces hypoxic apoptosis of MLO-Y4 osteocytes via JNK/caspase-3 pathway and the apoptotic-osteocyte-mediated osteoclastogenesis in vitro. Tissue Cell 67: 101402, 2020.
- 50. Tang Y, Hong C, Cai Y, Zhu J, Hu X, Tian Y, Song X, Song Z, Jiang R and Kang F: HIF-1α mediates osteoclast-induced mandibular condyle growth via AMPK signaling. J Dent Res 99: 1377-1386, 2020.
- 51. Wu C, Rankin EB, Castellini L, Alcudia JF, LaGory EL, Andersen R, Rhodes SD, Wilson TL, Mohammad KS, Castillo AB, *et al*: Oxygen-sensing PHDs regulate bone homeostasis through the modulation of osteoprotegerin. Genes Dev 29: 817-831, 2015.
- 52. Kang H, Yang K, Xiao L, Guo L, Guo C, Yan Y, Qi J, Wang F, Ryffel B, Li C and Deng L: Osteoblast Hypoxia-inducible Factor-1α pathway activation restrains osteoclastogenesis via the interleukin-33-MicroRNA-34a-Notch1 pathway. Front Immunol 8: 1312, 2017.
- 53. Zou D, Han W, You S, Ye D, Wang L, Wang S, Zhao J, Zhang W, Jiang X, Zhang X and Huang Y: In vitro study of enhanced osteogenesis induced by HIF-1α-transduced bone marrow stem cells. Cell Prolif 44: 234-243, 2011.
- 54. Kusumbe AP, Ramasamy SK and Adams RH: Coupling of angiogenesis and osteogenesis by a specific vessel subtype in bone. Nature 507: 323-328, 2014.
- Peng Y, Wu S, Li Y and Crane JL: Type H blood vessels in bone modeling and remodeling. Theranostics 10: 426-436, 2020.
   Ding W, Xu C, Zhang Y and Chen H: Advances in the under-
- 56. Ding W, Xu C, Zhang Y and Chen H: Advances in the understanding of the role of type-H vessels in the pathogenesis of osteoporosis. Arch Osteoporos 15: 5, 2020.
  57. Ramasamy SK, Kusumbe AP, Wang L and Adams RH:
- S7. Ramasamy SK, Kusumbe AP, Wang L and Adams RH: Endothelial Notch activity promotes angiogenesis and osteogenesis in bone. Nature 507: 376-380, 2014.
   S8. Yang M, Li CJ, Sun X, Guo Q, Xiao Y, Su T, Tu ML, Peng H,
- 58. Yang M, Li CJ, Sun X, Guo Q, Xiao Y, Su T, Tu ML, Peng H, Lu Q, Liu Q, *et al*: MiR-497-195 cluster regulates angiogenesis during coupling with osteogenesis by maintaining endothelial Notch and HIF-1α activity. Nat Commun 8: 16003, 2017.
- 59. Shao J, Liu S, Zhang M, Chen S, Gan S, Chen C, Chen W, Li L and Zhu Z: A dual role of HIF1α in regulating osteogenesis-angiogenesis coupling. Stem Cell Res Ther 13: 59, 2022.
- 60. Tao L, Li D, Liu H, Jiang F, Xu Y, Cao Y, Gao R and Chen G: Neuroprotective effects of metformin on traumatic brain injury in rats associated with NF-κB and MAPK signaling pathway. Brain Res Bull 140: 154-161, 2018.
- Yao R, Hou W and Bao J: Complete oxidative conversion of lignocellulose derived non-glucose sugars to sugar acids by Gluconobacter oxydans. Bioresour Technol 244: 1188-1192, 2017.
- 62. Guo K, Yao X, Wu W, Yu Z, Li Z, Ma Z and Liu D: HIF-1α/SDF-1/CXCR4 axis reduces neuronal apoptosis via enhancing the bone marrow-derived mesenchymal stromal cell migration in rats with traumatic brain injury. Exp Mol Pathol 114: 104416, 2020.

- 63. Knerlich-Lukoschus F, von der Ropp-Brenner B, Lucius R, Mehdorn HM and Held-Feindt J: Spatiotemporal CCR1, CCL3(MIP-1α), CXCR4, CXCL12(SDF-1α) expression patterns in a rat spinal cord injury model of posttraumatic neuropathic pain. J Neurosurg Spine 14: 583-597, 2011.
- 64. Xue Y, Li Z, Wang Y, Zhu X, Hu R and Xu W: Role of the HIF-1α/SDF-1/CXCR4 signaling axis in accelerated fracture healing after craniocerebral injury. Mol Med Rep 22: 2767-2774, 2020.
- 65. Tacchini L, Bianchi L, Bernelli-Zazzera A and Cairo G: Transferrin receptor induction by hypoxia. HIF-1-mediated transcriptional activation and cell-specific post-transcriptional regulation. J Biol Chem 274: 24142-24146, 1999.
- 66. Yang L, Fan M, Du F, Gong Q, Bi ZG, Zhu ZJ, Zhu LL and Ke Y: Hypoxic preconditioning increases iron transport rate in astrocytes. Biochim Biophys Acta 1822: 500-508, 2012.
- 67. Hu J, Meng F, Hu X, Huang L, Liu H, Liu Z and Li L: Iron overload regulate the cytokine of mesenchymal stromal cells through ROS/HIF-1α pathway in Myelodysplastic syndromes. Leuk Res 93: 106354, 2020.
- 68. Lok CN and Ponka P: Identification of a hypoxia response element in the transferrin receptor gene. J Biol Chem 274: 24147-24152, 1999.
- 69. Lee PJ, Jiang BH, Chin BY, Iyer NV, Alam J, Semenza GL and Choi AM: Hypoxia-inducible factor-1 mediates transcriptional activation of the heme oxygenase-1 gene in response to hypoxia. J Biol Chem 272: 5375-5381, 1997.
- 70. Weinreb O, Mandel S, Youdim MB and Amit T: Targeting dysregulation of brain iron homeostasis in Parkinson's disease by iron chelators. Free Radic Biol Med 62: 52-64, 2013.
- Guo C, Hao LJ, Yang ZH, Chai R, Zhang S, Gu Y, Gao HL, Zhong ML, Wang T, Li JY and Wang ZY: Deferoxamine-mediated up-regulation of HIF-1α prevents dopaminergic neuronal death via the activation of MAPK family proteins in MPTP-treated mice. Exp Neurol 280: 13-23, 2016.
   Lim J, Kim HI, Bang Y, Seol W, Choi HS and Choi HJ:
- 72. Lim J, Kim HI, Bang Y, Seol W, Choi HS and Choi HJ: Hypoxia-inducible factor-1α upregulates tyrosine hydroxylase and dopamine transporter by nuclear receptor ERRγ in SH-SY5Y cells. Neuroreport 26: 380-386, 2015.
- 73. Dixon SJ, Lemberg KM, Lamprecht MR, Skouta R, Zaitsev EM, Gleason CE, Patel DN, Bauer AJ, Cantley AM, Yang WS, *et al*: Ferroptosis: An iron-dependent form of nonapoptotic cell death. Cell 149: 1060-1072, 2012.
- 74. Li Y, Cao Y, Xiao J, Shang J, Tan Q, Ping F, Huang W, Wu F, Zhang H and Zhang X: Inhibitor of apoptosis-stimulating protein of p53 inhibits ferroptosis and alleviates intestinal ischemia/reperfusion-induced acute lung injury. Cell Death Differ 27: 2635-2650, 2020.
- 75. Li X, Zou Y, Xing J, Fu YY, Wang KY, Wan PZ and Zhai XY: Pretreatment with Roxadustat (FG-4592) attenuates folic acid-induced kidney injury through Antiferroptosis via Akt/GSK-3β/Nrf2 Pathway. Oxid Med Cell Longev 2020: 6286984, 2020.
- Nakazawa MS, Keith B and Simon MC: Oxygen availability and metabolic adaptations. Nat Rev Cancer 16: 663-673, 2016.
- 77. Piccoli C, D'Aprile A, Ripoli M, Scrima R, Boffoli D, Tabilio A and Capitanio N: The hypoxia-inducible factor is stabilized in circulating hematopoietic stem cells under normoxic conditions. FEBS Lett 581: 3111-3119, 2007.
- 78. Lequeux A, Noman MZ, Xiao M, Van Moer K, Hasmim M, Benoit A, Bosseler M, Viry E, Arakelian T, Berchem G, *et al*: Targeting HIF-1 alpha transcriptional activity drives cytotoxic immune effector cells into melanoma and improves combination immunotherapy. Oncogene 40: 4725-4735, 2021.

- López-Barneo J and Simon MC: Cellular adaptation to oxygen deficiency beyond the Nobel award. Nat Commun 11: 607, 2020.
- Loots GG, Řobling AG, Chang JC, Murugesh DK, Bajwa J, Carlisle C, Manilay JO, Wong A, Yellowley CE and Genetos DC: Vhl deficiency in osteocytes produces high bone mass and hematopoietic defects. Bone 116: 307-314, 2018.
- Lappin KM, Mills KI and Lappin TR: Erythropoietin in bone homeostasis-Implications for efficacious anemia therapy. Stem Cells Transl Med 10: 836-843, 2021.
- Johnson RW, Schipani E and Giaccia AJ: HIF targets in bone remodeling and metastatic disease. Pharmacol Ther 150: 169-177, 2015.
- 83. Tao J, Miao R, Liu G, Qiu X, Yang B, Tan X, Liu L, Long J, Tang W and Jing W: Spatiotemporal correlation between HIF-1α and bone regeneration. FASEB J 36: e22520, 2022.
- 84. Hulley PA, Bishop T, Vernet A, Schneider JE, Edwards JR, Athanasou NA and Knowles HJ: Hypoxia-inducible factor 1-alpha does not regulate osteoclastogenesis but enhances bone resorption activity via prolyl-4-hydroxylase 2. J Pathol 242: 322-333, 2017.
- 85. Clinkenbeard EL, Hanudel MR, Stayrook KR, Appaiah HN, Farrow EG, Cass TA, Summers LJ, Ip CS, Hum JM, Thomas JC, *et al*: Erythropoietin stimulates murine and human fibroblast growth factor-23, revealing novel roles for bone and bone marrow. Haematologica 102: e427-e430, 2017.
- 86. Daryadel A, Bettoni Č, Haider T, Imenez Silva PH, Schnitzbauer U, Pastor-Arroyo EM, Wenger RH, Gassmann M and Wagner CA: Erythropoietin stimulates fibroblast growth factor 23 (FGF23) in mice and men. Pflugers Arch 470: 1569-1582, 2018.
- 87. Li RL, He LY, Zhang Q, Liu J, Lu F, Duan HX, Fan LH, Peng W, Huang YL and Wu CJ: HIF-1α is a potential molecular target for herbal medicine to treat diseases. Drug Des Devel Ther 14: 4915-4949, 2020.
- 88. Kasper AC, Moon EJ, Hu X, Park Y, Wooten CM, Kim H, Yang W, Dewhirst MW and Hong J: Analysis of HIF-1 inhibition by manassantin A and analogues with modified tetrahydrofuran configurations. Bioorg Med Chem Lett 19: 3783-3786, 2009.
- 89. Kwak SH, Stephenson TN, Lee HE, Ge Y, Lee H, Min SM, Kim JH, Kwon DY, Lee YM and Hong J: Evaluation of Manassantin A tetrahydrofuran core region analogues and cooperative therapeutic effects with EGFR inhibition. J Med Chem 63: 6821-6833, 2020.
- 90. Hu H, Miao XK, Li JY, Zhang XW, Xu JJ, Zhang JY, Zhou TX, Hu MN, Yang WL and Mou LY: YC-1 potentiates the antitumor activity of gefitinib by inhibiting HIF-1α and promoting the endocytic trafficking and degradation of EGFR in gefitinib-resistant non-small-cell lung cancer cells. Eur J Pharmacol 874: 172961, 2020.
- Khan M, Dhammu TS, Baarine M, Kim J, Paintlia MK, Singh I and Singh AK: GSNO promotes functional recovery in experimental TBI by stabilizing HIF-1α. Behav Brain Res 340: 63-70, 2018.
- 92. Lei R, Li J, Liu F, Li W, Zhang S, Wang Y, Chu X and Xu J: HIF-1α promotes the keloid development through the activation of TGF-β/Smad and TLR4/MyD88/NF-κB pathways. Cell Cycle 18: 3239-3250, 2019.
- 93. Feng S, Bowden N, Fragiadaki M, Souilhol C, Hsiao S, Mahmoud M, Allen S, Pirri D, Ayllon BT, Akhtar S, *et al*: Mechanical activation of hypoxia-inducible Factor 1α drives endothelial dysfunction at atheroprone sites. Arterioscler Thromb Vasc Biol 37: 2087-2101, 2017.
- 94. Wu D, Huang RT, Hamanaka RB, Krause M, Oh MJ, Kuo CH, Nigdelioglu R, Meliton AY, Witt L, Dai G, *et al*: HIF-1α is required for disturbed flow-induced metabolic reprogramming in human and porcine vascular endothelium. Elife 6: e25217, 2017.