

Insight into microbial extracellular vesicles as key communication materials and their clinical implications for lung cancer (Review)

JEONG YUN JANG^{1*}, JI HOON SEO^{2*}, JAE JUN CHOI³, HYUN JIN RYU^{4,5},
HYUNJUN YUN⁶, DONG MYEONG HA⁷ and JINHO YANG⁷

¹Department of Radiation Oncology, Konkuk University Medical Center, Konkuk University School of Medicine, Seoul 05030, Republic of Korea; ²Department of Environmental Health, Korea University, Seoul 02841, Republic of Korea; ³Department of Fire Disaster Prevention, Graduate School of Semyung University, Jecheon, Chungcheongbuk-do 27136, Republic of Korea; ⁴Department of Endocrinology and Metabolism, Kyung Hee University College of Medicine, Seoul 02447, Republic of Korea; ⁵School of Medicine, Kyung Hee University Hospital at Gangdong, Seoul 05278, Republic of Korea; ⁶The AI Convergence Appliance Research Center, Korea Electronics Technology Institute, Gwangju 61011, Republic of Korea; ⁷Department of Occupational Health and Safety, Semyung University, Jecheon, Chungcheongbuk-do 27136, Republic of Korea

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Abstract. The complexity of lung cancer, driven by multi-factorial causes such as genetic, environmental and lifestyle factors, underscores the necessity for tailored treatment strategies informed by recent advancements. Studies highlight a significant association between the lung microbiome and lung cancer, with dysbiosis potentially contributing to disease development via inflammation, immune response alterations and bacterial metabolite production. Furthermore, exposure to airborne bacteria may influence lung health by introducing pathogenic species or altering the human microbiome, thereby implicating certain dominant airborne bacteria in lung diseases, including the exacerbation of lung cancer. Extracellular vesicles (EVs) facilitate cell-to-cell communication, penetrating mucosal barriers to impact various organs, notably the lung. Epidemiological evidence suggests a strong relationship between the presence of microbial EVs (MEVs) in the air and chronic pulmonary diseases, with indications of a potential risk for lung cancer. MEVs play a significant role in pulmonary disease development by inducing airway inflammation and affecting lung function. The microbiome and MEVs offer considerable potential as novel tools in precision medicine for lung cancer. Biological data analysis and artificial intelligence technology

advancements are pivotal for fully realizing their diagnostic and therapeutic capabilities. These developments can potentially shape the future landscape of lung cancer diagnostics, therapeutics and prevention strategies.

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1. Introduction

Lung cancer is the most newly diagnosed cancer worldwide, comprising 12.4% (2.48 million cases) of cases and causing 18.7% (1.82 million deaths) of cancer-associated mortalities, and it is now the leading concern among males, with an incidence rate of 15.3% (1.57 million cases), surpassing prostate cancer in 2022 (1). In addition, domestic cancer statistics in South Korea demonstrate the highest fatality rates, with crude mortality rates of 36.8 per 100,000 individuals (18,902 deaths) in 2021, aligning with international trends (2). While lung cancer is traditionally classified into non-small cell lung cancer and small cell lung cancer, recent advancements such as next-generation sequencing (NGS) have revealed a significant heterogeneity within the disease. This has led to the identification of various pathological subtypes, highlighting the complexity of lung cancer and underscoring the need for tailored treatment strategies for individual patients (3-5).

Correspondence to: Professor Jinho Yang, Department of Occupational Health and Safety, Semyung University, 65 Semyung-ro, Jecheon, Chungcheongbuk-do 27136, Republic of Korea
E-mail: iamjinho@semyung.ac.kr

*Contributed equally

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Well-known risk factors for lung cancer include genetic polymorphisms, tobacco smoking, diet, alcohol consumption, chronic inflammation, exposure to ionizing radiation and occupational exposure to substances such as asbestos, chromium compounds, silica and diesel exhaust (6). Similar to most types of cancer, lung cancer is recognized to result from the interplay of various multifactorial causes. Among these factors, the microbiome has also emerged as a topic of growing interest in research (7). In the past, it was believed that the lung was sterile; however, recent research has uncovered a range of commensal microbiomes, including fungi, bacteria and viruses, all of which contribute to homeostasis (8,9). Numerous studies have investigated the association between these microbiomes and various lung diseases, including cancer (10). Although dysbiosis is not exclusively associated with cancer development, it has been found to correlate with innate immunity, suggesting potential therapeutic implications (11-13). Therefore, the present review aims to summarize the research on the influence of both human and environmental microbiomes on the occurrence and progression of lung cancer, as well as their potential clinical implications.

2. Association between the microbiome and lung cancer

Human microbiome. Multiple studies have highlighted significant associations between lung cancer and the human microbiome using fecal, sputum, tissue and saliva samples, as summarized in Table I. The lung microbiota is shaped by interactions with the gut and oral microbiome, as well as external exposures (14). In patients with lung cancer, *Streptococcus* was consistently found at higher levels in sputum, tissue and saliva compared to healthy individuals (15-17), while its abundance was significantly reduced in fecal samples (18). Conversely, *Faecalibacterium*, which is typically abundant in feces, was found in higher levels in fecal samples from patients with lung cancer but reduced in their saliva (17-19). Other pro-inflammatory bacteria, such as *Ruminococcus* and *Klebsiella*, were also elevated in fecal samples of patients with lung cancer compared to healthy controls (18,20). By contrast, beneficial genera such as *Bifidobacterium* and *Bacteroides* were observed in greater abundance in fecal samples from healthy individuals (19,21,22).

In addition to comparisons with healthy controls, studies have also evaluated differences in microbiota composition between patients with lung cancer and those with benign pulmonary conditions or other cancer types. For instance, higher levels of *Veillonella*, *Megasphaera*, *Atopobium* and *Selenomonas* were reported in patients with lung cancer compared to individuals with benign lung lesions (23). Similarly, *Haemophilus* levels showed significant variability between lung cancer and patients with esophageal squamous cell carcinoma (20). These findings suggest that the lung microbiota may interact with bacteria originating from the oral cavity or pharynx, further emphasizing the concept of a gut-lung axis (14).

Despite these advances, variability in study outcomes remains a challenge, often attributed to small sample sizes, diverse cancer subtypes, and differences in patient characteristics, such as age, sex and medical history. Furthermore, discrepancies arise from variations in sequencing approaches

and taxonomic databases. For example, targeting different 16S ribosomal RNA regions (such as V1-V3 vs. V3-V4) and using different platforms, such as Illumina MiSeq and Roche 454, yield differing results. Additionally, inconsistencies across databases [such as SILVA (<https://www.arb-silva.de/>), Ribosomal Database Project (RDP interface is no longer available), Greengenes (<https://greengenes2.ucsd.edu/>) and National Center for Biotechnology Information (<https://www.ncbi.nlm.nih.gov/>)] contribute to the variability (24-26). Nonetheless, several studies have consistently identified *Streptococcus* and *Faecalibacterium* as key genera associated with lung cancer, with the former being particularly prominent (14,17).

Dysbiosis, characterized by a shift in the microbiome that favors harmful over beneficial bacteria, plays a central role in cancer progression. It promotes inflammation, alters metabolic pathways and dysregulates immune responses, as observed in studies of colorectal cancer (14,27,28). In the lung, similar mechanisms are suspected but remain underexplored. Emerging evidence suggests that dysbiosis of the lung microbiota may facilitate lung cancer progression through bacterial metabolite release and activation of inflammatory pathways (29). Moreover, the gut-lung axis, influenced by gut microbiota such as *Lactobacillus reuteri* and *Clostridium*, shapes immune responses in the lungs, highlighting its potential role in lung cancer pathogenesis (30,31). Despite these findings, large-scale studies are needed to validate microbial biomarkers for lung cancer, which could pave the way for novel diagnostic and therapeutic strategies.

Environmental microbiome. A previous study investigated the relationship between environmental microorganisms and human health (Fig. 1). Humans are exposed to environmental microorganisms through the air, food, soil and water, which circulate and interact, influencing various ecological systems (32). These microbes can infiltrate the body via respiration, skin contact and ingestion, integrating into the complex interactions of these ecosystems. Subsequently, environmental microorganisms that have penetrated the body can directly impact human health by introducing pathogenic bacteria or indirectly by altering the human microbiome (33,34).

In particular, pulmonary diseases are related to exposure to airborne bacteria. Previous studies have identified some prevalent airborne bacteria, revealing that their composition varies depending on the characteristics of the outdoor environments. In outdoor air, the dominant genera varied according to meteorological conditions (32). In the indoor air of an office, the most dominant genera were *Methylobacterium*, *Enterobacteriaceae_unidentified genus*, *Exiguobacterium* and *Bacteroides* (32). Also, Shin *et al.* (35) reported that *Micrococcus*, *Paracoccus*, *Staphylococcus* and *Enhydrobacter* were the common genera in indoor air of childcare facilities.

Several studies have elucidated that airborne microbes are associated with lung diseases (36-40). Exposure to airborne microbes has been implicated in developing and exacerbating lung diseases, such as asthma and chronic obstructive pulmonary disease (COPD) (36). For example, *Pseudomonas aeruginosa* is common in patients with cystic fibrosis and COPD (37). It is known that exposure to bioaerosols, such as allergens, toxins and pro-inflammatory agents, induces airway inflammation, leading to respiratory symptoms (38). Asthma and bioaerosol

Table I. Clinical studies on microbiome genera alteration in lung cancer compared with healthy individuals and other lung diseases using 16S ribosomal RNA method.

Sample	Microbiome		(Refs.)
	Increase	Decrease	
Feces	<i>Haemophilus</i> , <i>Faecalibacterium</i>	<i>Neisseria</i> , <i>Fusobacterium</i> , <i>Treponema</i> , <i>Rothia</i> , <i>Burkholderia</i> , <i>Filifactor</i> , <i>Dialister</i> , <i>Mycoplasma</i> , <i>Catonella</i> , <i>Anaerovorax</i> , <i>Acholeplasma</i> , <i>Bacteroides</i> , <i>Peptococcus</i> , <i>Megamonas</i> , <i>Bradyrhizobium</i> , <i>TG5</i>	(19)
	<i>Eubacterium</i> , <i>Ruminococcus</i> , <i>Faecalibacterium</i>	<i>Streptococcus</i> , <i>Enterococcus</i> , <i>Roseburia</i>	(18)
	<i>Klebsiella</i> , <i>Streptococcus</i>	<i>Haemophilus</i>	(20)
	<i>Enterococcus</i>	<i>Bifidobacterium</i>	(21)
	<i>Ruminococcus</i>	<i>Lachnospira</i>	(124)
Sputum		<i>Faecalibacterium</i> , <i>Streptococcus</i> , <i>Bifidobacterium</i> , <i>Veillonella</i>	(22)
	<i>Parabacteroides</i> , <i>Eggerthella</i> , <i>Weissella</i>	<i>Haemophilus</i> , <i>Dialister</i> , <i>Burkholderia</i> , <i>WAL_1855D</i> , <i>Neisseria</i> , <i>Bulleidia</i>	(19)
	<i>Granulicatella</i> , <i>Abiotrophia</i> , <i>Streptococcus</i> ,	<i>Sphingomonas</i> , <i>Leptotrichia</i>	(15)
Tissue	<i>Streptococcus</i>	<i>Staphylococcus</i>	(16)
		<i>Corynebacterium</i> , <i>Halomonas</i> , <i>Lachnoanaerobaculum</i> , <i>Acidovorax</i>	(125)
		<i>Fusobacterium</i> , <i>Prevotella</i> , <i>Bacteroides</i> , <i>Faecalibacterium</i>	(126)
Saliva	<i>Veillonella</i> , <i>Streptococcus</i>		(17)

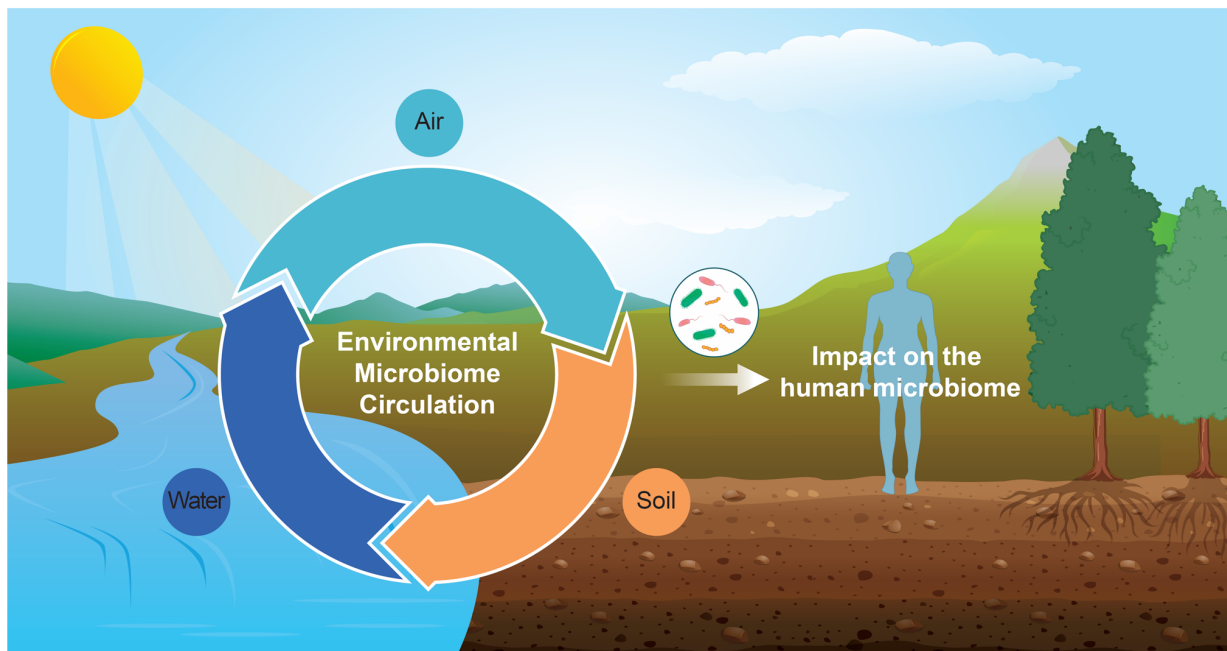


Figure 1. Relationship between the microbiomes of the environment and humans. Environmental microbiomes, circulating through air, water and soil, come into contact with humans, subsequently impacting the human microbiome.

exposure have been found to reduce lung function while increasing pulmonary inflammation (39). This series of lung function decline, increased inflammation and dysregulation

can contribute to the development of lung cancer. Additionally, a study reported a relationship between exposure to bioaerosols and the development of specific cancers, including pancreatic,

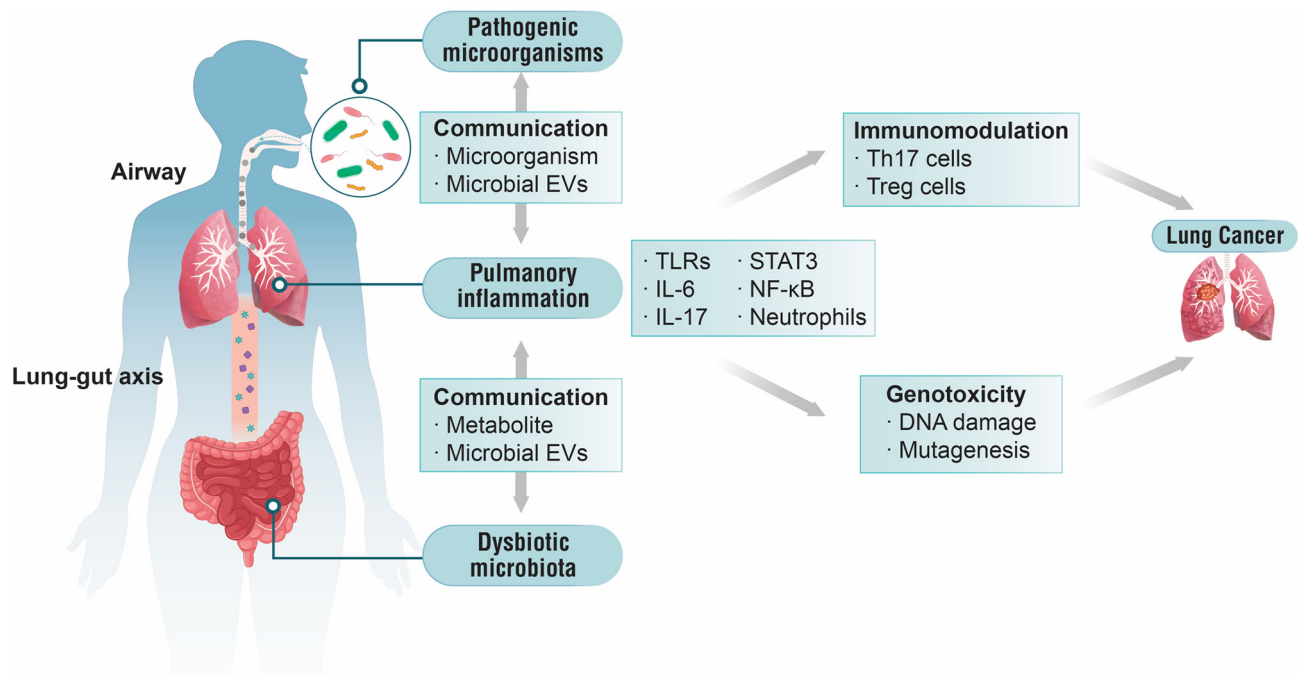


Figure 2. Microbiome as a significant factor in lung cancer carcinogenesis. Environmental microorganisms and gut microbiome contribute to pulmonary inflammation, which can lead to development of lung cancer. EVs, extracellular vesicles; TLRs, Toll-like receptors; Th, T helper cells; Treg, regulatory T cells.

liver and lung cancer (40). In summary, alterations in the pulmonary micro-environment and functions, which may contribute to the development of lung cancer, are increasingly acknowledged; however, research into the definitive impact of the microbiome on its pathogenesis remains limited.

3. Microbial extracellular vesicles as key communication materials between the microbiome and lung cancer

Exposure to microbial extracellular vesicles. Extracellular vehicles (EVs) are cell-to-cell communication materials enclosed in a lipid bilayer containing proteins, lipopolysaccharides (LPS) and nucleic acids, ranging from 20 to 200 nm in diameter. Microbial EVs (MEVs), found in all bacteria, are known as outer membrane vesicles (OMVs) in Gram-negative bacteria and membrane vesicles (MVs) in Gram-positive bacteria (41,42). Gut commensal microbes secrete MEVs, which penetrate the mucosal barrier and circulate throughout the body, reaching organs such as the lung, liver and skeletal muscle after oral administration (43). Additionally, dietary habits influence the microbiome and MEV composition, impacting human health and disease risk (44-46). For example, the microbial diversity in the feces of individuals on habitual Western diets was decreased compared with plant-based diets (44). Nanosized particles, including MEVs, are absorbed through inhalation and spread to various organs. These particles can accumulate in deep lung tissue, potentially affecting lung function over time (47).

Epidemiological studies have linked MEVs in indoor dust with chronic pulmonary diseases. A clinical study found that 63.6% of children with asthma had IgG1 sensitization to MEVs in indoor dust, suggesting a role in chronic lung diseases (48). Higher levels of anti-dust EV IgG antibodies were observed in patients with asthma, COPD and lung cancer compared

to healthy controls (49,50). In summary, while research is in its early stages, MEVs may pose a significant risk for lung disease, including cancer.

Pathogenesis of microbial extracellular vesicles in the development of lung diseases. As the EV membrane is embedded with surface ligands that interact with receptors on target cells, EVs can attach to and modify the physiological state of recipient cells (51,52). Furthermore, MEVs have recently been shown to be involved in the development of a wide variety of diseases, including cancer (24,53).

MEVs in beds were found to be mainly derived from pathogenic bacteria, such as *Pseudomonas*, *Acinetobacter*, *Enterobacter* and *Staphylococcus* (50). The prolonged exposure to MEVs in inhaled indoor dust induces significant airway inflammation, leading to severe asthma-like responses as well as emphysema. The induction of emphysema is of particular concern, as it is known to be a major factor in the development of irreversible airway obstruction (48). The exposure to MEVs during respiration and dysbiosis of gut microbiota constitute two primary pathophysiological mechanisms-the airway and gut-lung axis-contributing to disease development (Fig. 2).

When the parent cell is an extracellular gram-negative bacterium, OMVs induce T helper (Th)17 responses, leading to neutrophilic inflammation via the release of IL-17. This inflammation often causes airway hyperreactivity, fibrosis and conditions such as asthma and COPD, which may elevate the risk of lung cancer (54). A previous study has shown that OMVs from *Escherichia coli* trigger IL-17A-dependent neutrophilic inflammation and emphysema in mice, accompanied by elastase upregulation (55). Intraperitoneal injection of *E. coli* EVs induces lung dysfunction and mortality (56). Similarly, *P. aeruginosa* EVs exacerbate pulmonary inflammation through Toll-like receptor (TLR)2 and TLR4 activation, elevating in the

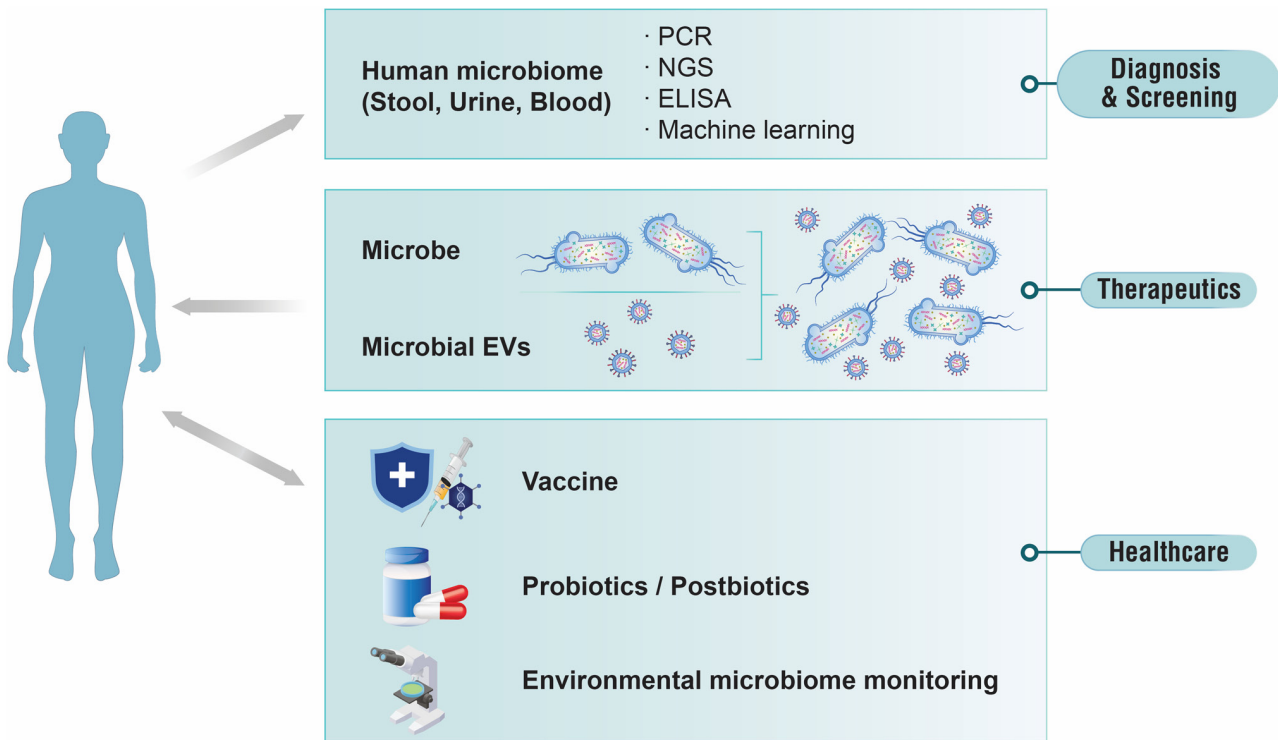


Figure 3. Clinical applicability of the microbiome: i) Diagnosis, including screening; ii) therapeutics; and iii) healthcare, such as monitoring and supplements. NGS, next generation sequencing; EVs, extracellular vesicles.

chemokines (CXCL1 and C-C motif ligand 2) and the cytokines (IL-1 β , TNF- α , IL-6 and IFN- γ), alongside neutrophil and macrophage infiltration (37). Moreover, indoor dust, including various bacterial components, has been associated with both Th1 and Th17 responses, leading to the induction of neutrophilic pulmonary inflammation (48,57). By contrast, MVs from intracellular Gram-positive bacteria primarily induce Th1 polarization via IFN- γ , leading to mononuclear inflammation and alveolar elastase production, which may cause emphysema (54). Although research on MVs has not been as extensive as on OMVs, a previous study has revealed immunological responses to some common MVs in the airway. For example, Repeated airway exposure to *Staphylococcus aureus* EVs triggers both Th1 and Th17 responses, increasing neutrophilic inflammation through TLR2 (58). These results suggest that the pathogenicity of MEVs is strongly related to lung diseases. Understanding these immunological pathways is crucial for advancing pulmonary health research.

As aforementioned, numerous immune responses are triggered by MEVs, and it is evident that the mechanisms of these responses in the airways vary according to the Gram type of the bacteria. For example, common airway OMVs, such as those derived from *P. aeruginosa*, *E. coli* and *Acinetobacter baumannii*, increase IL-6 levels and neutrophilic activity (37,56,59). Meanwhile, MVs, such as those derived from *S. aureus* and *Faecalibacterium prausnitzii*, have been reported to commonly increase IFN- γ levels (58,60).

4. Clinical implications of the microbiome for lung cancer

Recently, interest in the relationship between the microbiome and human health and efforts toward clinical application have

increased. Previous studies have suggested that the microbiome holds valuable information, demonstrating its potential as a material or biomarker for diagnosis, therapeutics and healthcare (Fig. 3) (24,61,62). After MEVs circulate throughout the body, they are excreted via feces, urine and exhaled air in their intact forms, unlike live microbes, which are restricted to the mucosal lumen or skin surface (43). Certain MEVs act as etiological agents of various diseases, while some MEVs have a protective role in disease pathogenesis. Therefore, circulating MEVs in our body provides us with noteworthy information for health and disease status (24).

Diagnostic potential of microbiome. Risk assessment, early diagnosis, treatment response prediction and disease monitoring are crucial for reducing mortality and enhancing quality of life in patients with cancer (63,64). There is a growing trend in developing diagnostic or screening technologies that utilize the microbiome-based quantitative polymerase chain reaction (qPCR), NGS, machine learning and enzyme-linked immunosorbent assay (Fig. 3). Most studies utilize fecal samples, which contain sufficient microbiomes for analysis, and therefore, research on microbiome-based diagnostics primarily targets gastrointestinal diseases. Previous studies on colorectal cancer diagnosis showed that a metagenomics algorithm achieved an area under the curve (AUC) of 0.89 (65), while qPCR demonstrated higher accuracy with an AUC of 0.93 (66). Additionally, this approach has been applied to lung cancer diagnostics using feces samples, with models using *Enterococcus*, *Streptococcus* and *Klebsiella* achieving an AUC of 0.96, a *Haemophilus*-specific model showing an AUC of 0.75 (20), and a model based on 13 OTU biomarkers demonstrating an AUC of 0.976 (22).

Recent studies have been conducted using diverse human samples, including urine, blood, saliva, bronchoalveolar lavage fluid (BALF) and sputum, with a focus on specific diseases, to development of diagnostic or screening technology based on microbiomes. For instance, to distinguish between benign lung disease and lung cancer using BALF, Kim *et al* (67) developed a prediction model based on *unclassified_SAR202_clade* (phylum Chloroflexi), achieving an AUC of 0.98, while a model using *Veillonella* and *Megasphaera* showed an AUC of 0.89 (23).

MEVs, rather than live microorganisms, are emerging as precise biomarkers for disease diagnostics using artificial intelligence (AI)-based analysis (61). McDowell *et al* (68) developed machine learning models using MEV metagenomes from serum, achieving AUCs of 0.93 for COPD, 0.99 for asthma and 0.94 for lung cancer. Antibodies against MEVs have also shown diagnostic potential, with IgG against MEVs derived from *S. aureus*, *Acinetobacter baumannii*, *Enterobacter cloacae* and *P. aeruginosa*, which are predominant in indoor dust, achieving AUCs of 0.78 for asthma, 0.79 for COPD and 0.81 for lung cancer (50).

MEV-based diagnostics have broad utility, with AUC of 0.95 for colorectal cancer using feces (69), 0.93 for brain tumor using blood (45), 0.87 for hepatocellular carcinoma using blood (70), 0.82 for gastric cancer using urine (71) and 1.00 for pancreatic cancer using blood (72). Furthermore, diagnostic models based on MEVs incorporating additional markers demonstrate improved performance. Combining MEV data with additional markers, such as metabolomics or tumor markers, significantly enhances diagnostic accuracy across various cancers (69,73). Thus, MEV-based diagnostic technologies, including composition assessment and immunoassays, can provide information on exposure to etiological agents (50). Additionally, MEVs derived from various samples can assist in the diagnosis of lung diseases.

5. Therapeutic potential of the microbiome

Live biotherapeutic products. Commensal bacteria are essential to human health, with growing recognition that humans are holobionts or supra-organisms. This means that the combined metabolic capabilities of both eukaryotic and prokaryotic components surpass those of each component alone (24). The U.S. Food and Drug Administration (FDA) has announced a new category called live biotherapeutic products (LBPs). The FDA has identified LBPs as biological products containing live organisms such as bacteria, which are used for disease prevention, treatment or cures, but are not vaccines (74). LBPs are administered in sufficient quantities to provide health benefits to the host (24,75).

Several studies have demonstrated the efficacy of LBP monotherapy. For examples, *Lactococcus lactis* inhibited lung cell proliferation (76,77). Other studies have shown that β -glucan, derived from *Saccharomyces cerevisiae*, can modulate immune responses and inhibit cancer cell viability in the lung cancer microenvironment (78,79). Short-chain fatty acids such as butyrate, propionate and acetate, when delivered from the gut to the lung, induce apoptosis in lung cancer cells (80). Although the exact mechanisms of LBPs remain unclear, immune system regulation and pathogen attachment interference are possible explanations.

LBPs can also be applied with regular treatments, such as conventional chemotherapy and immunotherapy, and have enhanced tumor suppression. Kotzampassi *et al* (81) found that the intake of *Lactobacillus* and *Bifidobacterium* reduced postoperative complications. In addition, Wada *et al* (82), demonstrated that the intake of *Bifidobacterium breve* during the chemotherapy period reduced the incidence of fever and decreased the need for intravenous antibiotics, thereby facilitating more effective therapy. Furthermore, combining *Lactobacillus* with cisplatin has been shown to reduce tumor size and increase immune responses in lung cancer models (83).

Microbial extracellular vesicles-based therapy as new-generation therapeutics. Recently, there has been a growing demand for developing new therapeutic targets distinct from conventional ones, suggesting the use of MEVs to address unmet medical needs as next-generation therapeutics. While the potential of using mammalian EVs for therapeutic purposes has been widely discussed, MEVs have yet to receive much attention thus far (84). Nevertheless, several studies have reported the beneficial effects of MEVs as therapeutic agents. EV derived from *Lactobacillus paracasei* significantly affects colorectal homeostasis in inflammation-mediated pathogenesis by attenuating LPS-induced inflammation in the intestine by activating endoplasmic reticulum stress (85). EVs derived from *Lactococcus lactis* can modulate airway inflammation by promoting a shift in immune responses from Th2 to Th1 by stimulating dendritic cells to produce IL-12, which offers a possible advantage for managing allergic asthma (86). Conversely, *Micrococcus luteus*-derived EVs alleviate neutrophilic airway inflammation by reducing IL-1 β and IL-17 levels in BALF and inhibiting group 3 innate lymphoid cells activation through upregulation of microRNA (miRNA) in airway epithelial cells, proposing them as a potential therapeutic for unresolved neutrophilic asthma (87). Additionally, *Lactobacillus plantarum*-derived EVs have been suggested to treat atopic dermatitis, decreasing skin inflammation and epidermal thickness (88). EVs derived from *Bifidobacterium longum* have been shown to reduce the occurrence of diarrhea, which can be a symptom of food allergy, by inducing mast cell apoptosis without affecting T cell-mediated immune responses (89). Therefore, therapeutic strategies can utilize beneficial MEVs as potential immunomodulators while suppressing harmful MEVs by inhibiting their production or function (90). In addition, postbiotics represent a new modality for next-generation therapy to complement current cancer treatment, including those for lung cancer, such as small molecules, proteins, monoclonal antibodies and cell-based therapeutics (24).

Therefore, LBPs and MEVs-derived treatments can be used independently or as adjuncts to chemotherapy or immunotherapy, potentially becoming a major component of lung cancer treatment in the near future.

6. Healthcare potential of the microbiome

It has been demonstrated that the human microbiome is strongly associated with health. Therefore, it can be utilized within healthcare systems for: i) Vaccines; ii) supplements such as probiotics and postbiotics; and iii) monitoring systems.

Vaccine. Cancer vaccines primarily target tumor-specific antigens but often lack sufficient efficacy. Researchers are exploring the potential of combining probiotics with cancer vaccines to enhance their effectiveness. *Plasmodium*, the malaria parasite, shows promise as an adjuvant for cancer vaccines, particularly in combination with DNA vaccines (91). Additionally, MEVs can deliver genetic materials of vaccine components into target cells, potentially improving vaccine efficacy (92,93). MEVs, with their bilayered lipids, primarily containing LPS and outer membrane lipids, could also potentially be used to deliver beneficial proteins, miRNAs and act as adjuvants in vaccine development (62,94). Due to the variety of glycolipids and glycoproteins in their composition, MEVs can introduce biological activity into cells, making them suitable as drug delivery vehicles for cyclic nucleotides, enzymes and antitumor drugs (95-97). Recently, nano and micro materials, such as virus-like particles and liposomal vesicles, are also being explored for vaccine delivery (98,99). Kim *et al* (100) demonstrated the successful modification of OMVs as multifunctional vaccine delivery vehicles to enhance immune responses against cancer cells. This approach aims to boost the immune response against cancer cells. Ongoing research is investigating these combinations' potential to improve cancer treatment outcomes.

Supplements such as probiotics and postbiotics. Probiotics, or LBPs, are live microorganisms that, when consumed in adequate amounts, provide beneficial effects to the host and are widely used in clinical practice. Numerous studies suggest that microbiome intake plays a role in cancer prevention (77,101-112), initially evidenced by Goldin and Gorbach (100) in 1980, which showed that *Lactobacillus acidophilus* supplementation reduced intestinal cancer incidence in rat models. Subsequent studies, particularly *in vitro* research, have demonstrated that probiotics can reduce cell proliferation, induce cell cycle arrest and trigger apoptosis (102-106). Strains such as *Lactobacillus plantarum*, *Lactobacillus rhamnosus* and *Bifidobacterium polyfermenticus* have been shown to reduce tumor incidence and progression in animal models (107-111). However, most research focuses on gastrointestinal cancers, with limited studies on lung cancer. Preclinical data on mice suggest that *Lactococcus lactis* can inhibit cancer cell proliferation and proinflammatory cytokine production, showing promise for lung cancer prevention (77,112). Despite limited research on the use of probiotics for lung cancer, the findings mentioned are promising, and future studies are expected to yield further positive outcomes.

Postbiotics, including MEVs, are soluble factors released by microbes or after microbial lysis that provide physiological benefits (113). MEVs are emerging as key postbiotics in precision medicine, facilitating intercellular communication through proteins and small molecules enclosed in a lipid bilayer (114). Cell-to-cell communication is tightly regulated, and its disruption prompts disease advancement. Soluble factors include proteins and small molecules, and cell-to-cell communication is performed by MEVs, which are packages of information from microbial cells enclosed by a cell membrane. Moreover, recent scientific evidence has shown that certain MEVs as postbiotics have protective effects against disease development or progression (62,85,115,116).

Therefore, we propose that the intake of probiotics and postbiotics holds significant potential in the prevention of lung cancer, offering a hopeful avenue for future research and preventive measures.

Monitoring system. A healthcare monitoring system can be employed to track human health biomarkers by analyzing the airborne microbiome (34). Considering the significant association between human health and air pollution, which includes particulate matter, bioaerosols and gaseous substances, the vigilant monitoring of atmospheric pollutants can contribute to disease prevention. In numerous countries, bioaerosol regulation has been implemented through measurements based on culturing techniques (117). Culture-based analysis can directly observe bacteria in the air and yield colony-forming units. However, this method faces several limitations: i) It can only measure bacteria counted >1% of the total in a solid medium agar plate (118); ii) it is restricted to analyzing specific bacterial species; and iii) it is incapable of analyzing unculturable bacterial material such as MEVs and dead bacteria. For these reasons, numerous studies are underway to enable real-time on-site monitoring of bioaerosols and biomarkers related to human health. Cho *et al* (119) developed the bioaerosol monitoring system based on ATP extracted from *E. coli* and demonstrated that this system can continuously monitor with high sensitivity in real-time. Additionally, a previous study has utilized reverse transcription-PCR to detect airborne bacteria (120). Furthermore, to facilitate the precise and rapid detection of bioaerosols, droplet digital PCR (ddPCR) has been employed extensively in various studies for pathogen diagnosis, mutation detection and transgenic research (120-122). For instance, airborne *Mycobacterium tuberculosis* was detected using ddPCR (123); however, this method currently cannot detect airborne bacteria in real-time on-site, indicating that the technology requires further improvement.

Through such monitoring systems, lung cancer surveillance can be enhanced. Furthermore, by observing microbiomes associated with lung cancer risk, these systems have the potential to utilize these microbiomes as biomarkers for the disease.

7. Conclusion

The present review explored the potential of microbiomes and MEVs as innovative tools in precision medicine for lung cancer. Disease patterns are linked to cellular aging and elevated reactive oxygen species, contributing to conditions such as inflammation, immune diseases and cancer. There is a growing shift toward promoting health through advanced diagnostics, safer therapeutics and prevention-focused healthcare systems. To support this shift, advancements in biological data analysis, including metagenomics and AI technologies such as machine learning, are essential for disease prediction and personalized therapies. While significant research on the microbiome exists, understanding the interactions between microbiota and host, particularly microbial products, remains limited. A deeper understanding of these interactions is key to developing beneficial microbial products. MEVs, unlike LBPs, can penetrate cells and target distant organs, offering significant advantages as diagnostic biomarkers and therapeutic candidates. We propose MEVs as next-generation technologies

for lung cancer, capable of replacing current biologics such as proteins, antibodies and genes.

Future research on MEVs is expected to enhance our understanding of their role in lung cancer and foster precision medicine approaches, including diagnostics and therapies utilizing MEVs from beneficial microorganisms.

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Availability of data and materials

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Authors' contributions

JYJ and JHS conducted the literature research, developed the methodology, generated the figures, and wrote the original draft. HJR, JJC, HY, and DMH contributed to the literature research and edited the manuscript. JY conceptualized the study, acquired funding, conducted the investigation, managed the project, supervised the research, visualized the results, and reviewed and edited the manuscript. All authors read and approved the final version of the manuscript. Data authentication is not applicable.

Ethics approval and consent to participate

Not applicable.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

References

- Bray F, Laversanne M, Sung H, Ferlay J, Siegel RL, Soerjomataram I and Jemal A: Global cancer statistics 2022: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* 74: 229-263, 2024.
- Park EH, Jung KW, Park NJ, Kang MJ, Yun EH, Kim HJ, Kim JE, Kong HJ, Im JS and Seo HG: Community of Population-Based Regional Cancer Registries: Cancer statistics in Korea: Incidence, Mortality, survival, and prevalence in 2021. *Cancer Res Treat* 56: 357-371, 2024.
- Zito Marino F, Bianco R, Accardo M, Ronchi A, Cozzolino I, Morgillo F, Rossi G and Franco R: Molecular heterogeneity in lung cancer: from mechanisms of origin to clinical implications. *Int J Med Sci* 16: 981-989, 2019.
- de Sousa VML and Carvalho L: Heterogeneity in lung cancer. *Pathobiology* 85: 96-107, 2018.
- Lim ZF and Ma PC: Emerging insights of tumor heterogeneity and drug resistance mechanisms in lung cancer targeted therapy. *J Hematol Oncol* 12: 134, 2019.
- Malhotra J, Malvezzi M, Negri E, La Vecchia C and Boffetta P: Risk factors for lung cancer worldwide. *Eur Respir J* 48: 889-902, 2016.
- Rahman MM, Islam MR, Shohag S, Ahasan MT, Sarkar N, Khan H, Hasan AM, Cavalu S and Rauf A: Microbiome in cancer: Role in carcinogenesis and impact in therapeutic strategies. *Biomed Pharmacother* 149: 112898, 2022.
- Dickson RP, Erb-Downward JR, Freeman CM, McCloskey L, Beck JM, Huffnagle GB and Curtis JL: Spatial variation in the healthy human lung microbiome and the adapted Island model of lung biogeography. *Ann Am Thorac Soc* 12: 821-830, 2015.
- Yu G, Gail MH, Consonni D, Carugno M, Humphrys M, Pesatori AC, Caporaso NE, Goedert JJ, Ravel J and Landi MT: Characterizing human lung tissue microbiota and its relationship to epidemiological and clinical features. *Genome Biol* 17: 163, 2016.
- Yagi K, Huffnagle GB, Lukacs NW and Asai N: The lung microbiome during health and disease. *Int J Mol Sci* 22: 10872, 2021.
- Ruan R, Deng X, Dong X, Wang Q, Lv X and Si C: Microbiota emergencies in the diagnosis of lung diseases: A meta-analysis. *Front Cell Infect Microbiol* 11: 709634, 2021.
- Jang HJ, Choi JY, Kim K, Yong SH, Kim YW, Kim SY, Kim EY, Jung JY, Kang YA, Park MS, *et al*: Relationship of the lung microbiome with PD-L1 expression and immunotherapy response in lung cancer. *Respir Res* 22: 322, 2021.
- Yang D, Xing Y, Song X and Qian Y: The impact of lung microbiota dysbiosis on inflammation. *Immunology* 159: 156-166, 2020.
- Xu N, Wang L, Li C, Ding C, Li C, Fan W, Cheng C and Gu B: Microbiota dysbiosis in lung cancer: Evidence of association and potential mechanisms. *Transl Lung Cancer Res* 9: 1554-1568, 2020.
- Hosgood HD III, Sapkota AR, Rothman N, Rohan T, Hu W, Xu J, Vermeulen R, He X, White JR, Wu G, *et al*: The potential role of lung microbiota in lung cancer attributed to household coal burning exposures. *Environ Mol Mutagen* 55: 643-651, 2014.
- Liu HX, Tao LL, Zhang J, Zhu YG, Zheng Y, Liu D, Zhou M, Ke H, Shi MM and Qu JM: Difference of lower airway microbiome in bilateral protected specimen brush between lung cancer patients with unilateral lobar masses and control subjects. *Int J Cancer* 142: 769-778, 2018.
- Zhang W, Luo J, Dong X, Zhao S, Hao Y, Peng C, Shi H, Zhou Y, Shan L, Sun Q, *et al*: Salivary microbial dysbiosis is associated with systemic inflammatory markers and predicted oral metabolites in non-small cell lung cancer patients. *J Cancer* 10: 1651-1662, 2019.
- Zhang M, Zhou H, Xu S, Liu D, Cheng Y, Gao B, Li X and Chen J: The gut microbiome can be used to predict the gastrointestinal response and efficacy of lung cancer patients undergoing chemotherapy. *Ann Palliat Med* 9: 4211-4227, 2020.
- Lu H, Gao NL, Tong F, Wang J, Li H, Zhang R, Ma H, Yang N, Zhang Y, Wang Y, *et al*: Alterations of the human lung and gut microbiomes in non-small cell lung carcinomas and distant metastasis. *Microbiol Spectr* 9: e0080221, 2021.
- Shen W, Tang D, Deng Y, Li H, Wang T, Wan P and Liu R: Association of gut microbiomes with lung and esophageal cancer: A pilot study. *World J Microbiol Biotechnol* 37: 128, 2021.
- Zhuang H, Cheng L, Wang Y, Zhang YK, Zhao MF, Liang GD, Zhang MC, Li YG, Zhao JB, Gao YN, *et al*: Dysbiosis of the gut microbiome in lung cancer. *Front Cell Infect Microbiol* 9: 112, 2019.
- Zheng Y, Fang Z, Xue Y, Zhang J, Zhu J, Gao R, Yao S, Ye Y, Wang S, Lin C, *et al*: Specific gut microbiome signature predicts the early-stage lung cancer. *Gut Microbes* 11: 1030-1042, 2020.
- Lee SH, Sung JY, Yong D, Chun J, Kim SY, Song JH, Chung KS, Kim EY, Jung JY, Kang YA, *et al*: Characterization of microbiome in bronchoalveolar lavage fluid of patients with lung cancer comparing with benign mass like lesions. *Lung Cancer* 102: 89-95, 2016.
- Yang J, Shin TS, Kim JS, Jee YK and Kim YK: A new horizon of precision medicine: Combination of the microbiome and extracellular vesicles. *Exp Mol Med* 54: 466-482, 2022.
- Castelino M, Eyre S, Moat J, Fox G, Martin P, Ho P, Upton M and Barton A: Optimisation of methods for bacterial skin microbiome investigation: Primer selection and comparison of the 454 versus MiSeq platform. *BMC Microbiol* 17: 23, 2017.

26. Balvočiūtė M and Huson DH: SILVA, RDP, Greengenes, NCBI and OTT-how do these taxonomies compare? BMC Genomics 18 (Suppl 2): S114, 2017.
27. Rajagopala SV, Vashee S, Oldfield LM, Suzuki Y, Venter JC, Telenti A and Nelson KE: The human microbiome and cancer. Cancer Prev Res (Phila) 10: 226-234, 2017.
28. Russo E, Taddei A, Ringressi MN, Ricci F and Amedei A: The interplay between the microbiome and the adaptive immune response in cancer development. Therap Adv Gastroenterol 9: 594-605, 2016.
29. Choi Y, Park H, Park HS and Kim YK: Extracellular vesicles, a key mediator to link environmental microbiota to airway immunity. Allergy Asthma Immunol Res 9: 101-106, 2017.
30. Karimi K, Inman MD, Bienenstock J and Forsythe P: Lactobacillus reuteri-induced regulatory T cells protect against an allergic airway response in mice. Am J Respir Crit Care Med 179: 186-193, 2009.
31. Atarashi K, Tanoue T, Shima T, Imaoka A, Kuwahara T, Momose Y, Cheng G, Yamasaki S, Saito T, Ohba Y, et al: Induction of colonic regulatory T cells by indigenous Clostridium species. Science 331: 337-341, 2011.
32. Yang J, Seo JH, Jee YK, Kim YK and Sohn JR: Composition analysis of airborne microbiota in outdoor and indoor based on dust separated by micro-sized and nano-sized. Aerosol Air Qual Res 23: 210231, 2023.
33. Panthee B, Gyawali S, Panthee P and Techato K: Environmental and human microbiome for health. Life (Basel) 12: 456, 2022.
34. Mbareche H, Morawska L and Duchaine C: On the interpretation of bioaerosol exposure measurements and impacts on health. J Air Waste Manag Assoc 69: 789-804, 2019.
35. Shin SK, Kim J, Ha SM, Oh HS, Chun J, Sohn J and Yi H: Metagenomic insights into the bioaerosols in the indoor and outdoor environments of childcare facilities. PLoS One 10: e0126960, 2015.
36. Wolf M and Lai PS: Indoor microbial exposures and chronic lung disease: From microbial toxins to the microbiome. Clin Chest Med 41: 777-796, 2020.
37. Park KS, Lee J, Jang SC, Kim SR, Jang MH, Lötvall J, Kim YK and Gho YS: Pulmonary inflammation induced by bacteria-free outer membrane vesicles from Pseudomonas aeruginosa. Am J Respir Cell Mol Biol 49: 637-645, 2013.
38. Kim KH, Kabir E and Jahan SA: Airborne bioaerosols and their impact on human health. J Environ Sci (China) 67: 23-35, 2018.
39. Baldacci S, Maio S, Cerrai S, Sarno G, Baiz N, Simoni M, Annesi-Maesano I and Viegi G: HEALS Study: Allergy and asthma: Effects of the exposure to particulate matter and biological allergens. Respir Med 109: 1089-1104, 2015.
40. Hayleeyesus SF, Ejeso A and Derseh FA: Quantitative assessment of bio-aerosols contamination in indoor air of University dormitory rooms. Int J Health Sci (Qassim) 9: 249-256, 2015.
41. Lee EY, Bang JY, Park GW, Choi DS, Kang JS, Kim HJ, Park KS, Lee JO, Kim YK, Kwon KH, et al: Global proteomic profiling of native outer membrane vesicles derived from Escherichia coli. Proteomics 7: 3143-3153, 2007.
42. Lee EY, Choi DY, Kim DK, Kim JW, Park JO, Kim S, Kim SH, Desiderio DM, Kim YK, Kim KP and Gho YS: Gram-positive bacteria produce membrane vesicles: Proteomics-based characterization of Staphylococcus aureus-derived membrane vesicles. Proteomics 9: 5425-5436, 2009.
43. Choi Y, Kwon Y, Kim DK, Jeon J, Jang SC, Wang T, Ban M, Kim MH, Jeon SG, Kim MS, et al: Gut microbe-derived extracellular vesicles induce insulin resistance, thereby impairing glucose metabolism in skeletal muscle. Sci Rep 5: 15878, 2015.
44. Conlon MA and Bird AR: The impact of diet and lifestyle on gut microbiota and human health. Nutrients 7: 17-44, 2014.
45. Yang J, Moon HE, Park HW, McDowell A, Shin TS, Jee YK, Kym S, Paek SH and Kim YK: Brain tumor diagnostic model and dietary effect based on extracellular vesicle microbiome data in serum. Exp Mol Med 52: 1602-1613, 2020.
46. Yang J, McDowell A, Kim EK, Seo H, Yum K, Lee WH, Jee YK and Kim YK: Consumption of a Leuconostoc holzapfelii-enriched synbiotic beverage alters the composition of the microbiota and microbial extracellular vesicles. Exp Mol Med 51: 1-11, 2019.
47. Son T, Cho YJ, Lee H, Cho MY, Goh B, Kim HM, Hoa PTN, Cho SH, Park YJ, Park HS and Hong KS: Monitoring in vivo behavior of size-dependent fluorescent particles as a model fine dust. J Nanobiotechnology 20: 227, 2022.
48. Kim YS, Choi EJ, Lee WH, Choi SJ, Roh TY, Park J, Jee YK, Zhu Z, Koh YY, Gho YS and Kim YK: Extracellular vesicles, especially derived from Gram-negative bacteria, in indoor dust induce neutrophilic pulmonary inflammation associated with both Th1 and Th17 cell responses. Clin Exp Allergy 43: 443-454, 2013.
49. Kim YS, Choi JP, Kim MH, Park HK, Yang S, Kim YS, Kim TB, Cho YS, Oh YM, Jee YK, et al: IgG sensitization to extracellular vesicles in indoor dust is closely associated with the prevalence of non-eosinophilic asthma, COPD, and lung cancer. Allergy Asthma Immunol Res 8: 198-205, 2016.
50. Yang J, Hong G, Kim YS, Seo H, Kim S, McDowell A, Lee WH, Kim YS, Oh YM, Cho YS, et al: Lung disease diagnostic model through IgG sensitization to microbial extracellular vesicles. Allergy Asthma Immunol Res 12: 669-683, 2020.
51. Raposo G and Stoorvogel W: Extracellular vesicles: Exosomes, microvesicles, and friends. J Cell Biol 200: 373-383, 2013.
52. Colombo M, Raposo G and Théry C: Biogenesis, secretion, and intercellular interactions of exosomes and other extracellular vesicles. Ann Rev Cell Dev Biol 30: 255-289, 2014.
53. Mishra S, Tejesvi MV, Hekkala J, Turunen J, Kandikanti N, Kaisanlahti A, Suokas M, Leppä S, Vihinen P, Kuitunen H, et al: Gut microbiome-derived bacterial extracellular vesicles in patients with solid tumours. J Adv Res 68: 375-386, 2025.
54. Yang J, Kim EK, Park HJ, McDowell A and Kim YK: The impact of bacteria-derived ultrafine dust particles on pulmonary diseases. Exp Mol Med 52: 338-347, 2020.
55. Kim YS, Lee WH, Choi EJ, Choi JP, Heo YJ, Gho YS, Jee YK, Oh YM and Kim YK: Extracellular vesicles derived from gram-negative bacteria, such as Escherichia coli, induce emphysema mainly via IL-17A-mediated neutrophilic inflammation. J Immunol 194: 3361-3368, 2015.
56. Park KS, Choi KH, Kim YS, Hong BS, Kim OY, Kim JH, Yoon CM, Koh GY, Kim YK and Gho YS: Outer membrane vesicles derived from Escherichia coli induce systemic inflammatory response syndrome. PLoS One 5: e11334, 2010.
57. Lundin JI and Checkoway H: Endotoxin and cancer. Environ Health Perspect 117: 1344-1350, 2009.
58. Kim MR, Hong SW, Choi EB, Lee WH, Kim YS, Jeon SG, Jang MH, Gho YS and Kim YK: Staphylococcus aureus-derived extracellular vesicles induce neutrophilic pulmonary inflammation via both Th1 and Th17 cell responses. Allergy 67: 1271-1281, 2012.
59. Jun SH, Lee JH, Kim BR, Kim SI, Park TI, Lee JC and Lee YC: Acinetobacter baumannii outer membrane vesicles elicit a potent innate immune response via membrane proteins. PLoS One 8: e71751, 2013.
60. Jafari B, Khavari Nejad RA, Vaziri F and Siadat SD: Evaluation of the effects of extracellular vesicles derived from Faecalibacterium prausnitzii on lung cancer cell line. Biologia 74: 889-898, 2019.
61. Yang J: Insight into the potential of algorithms using AI technology as in vitro diagnostics utilizing microbial extracellular vesicles. Mol Cell Probes 78: 101992, 2024.
62. Yang J, Kim EK, McDowell A and Kim YK: Microbe-derived extracellular vesicles as a smart drug delivery system. Transl Clin Pharmacol 26: 103-110, 2018.
63. Whitaker K: Earlier diagnosis: The importance of cancer symptoms. Lancet Oncol 21: 6-8, 2020.
64. Seo JH, Lee JW and Cho D: The market trend analysis and prospects of cancer molecular diagnostics kits. Biomater Res 22: 2, 2018.
65. Yang J, McDowell A, Kim EK, Seo H, Lee WH, Moon CM, Kym SM, Lee DH, Park YS, Jee YK and Kim YK: Development of a colorectal cancer diagnostic model and dietary risk assessment through gut microbiome analysis. Exp Mol Med 51: 1-15, 2019.
66. Yang J, Li D, Yang Z, Dai W, Feng X, Liu Y, Jiang Y, Li P, Li Y, Tang B, et al: Establishing high-accuracy biomarkers for colorectal cancer by comparing fecal microbiomes in patients with healthy families. Gut Microbes 11: 918-929, 2020.
67. Kim G, Park C, Yoon YK, Park D, Lee JE, Lee D, Sun P, Park S, Yun C, Kang DH and Chung C: Prediction of lung cancer using novel biomarkers based on microbiome profiling of bronchoalveolar lavage fluid. Sci Rep 14: 1691, 2024.
68. McDowell A, Kang J, Yang J, Jung J, Oh YM, Kym SM, Shin TS, Kim TB, Jee YK and Kim YK: Machine-learning algorithms for asthma, COPD, and lung cancer risk assessment using circulating microbial extracellular vesicle data and their application to assess dietary effects. Exp Mol Med 54: 1586-1595, 2022.

69. Kim DJ, Yang J, Seo H, Lee WH, Ho Lee D, Kym S, Park YS, Kim JG, Jang JJ, Kim YK and Cho JY: Colorectal cancer diagnostic model utilizing metagenomic and metabolomic data of stool microbial extracellular vesicles. *Sci Rep* 10: 2860, 2020.
70. Cho EJ, Leem S, Kim SA, Yang J, Lee YB, Kim SS, Cheong JY, Cho SW, Kim JW, Kim SM, *et al*: Circulating microbiota-based metagenomic signature for detection of hepatocellular carcinoma. *Sci Rep* 9: 7536, 2019.
71. Park JY, Kang CS, Seo HC, Shin JC, Kym SM, Park YS, Shin TS, Kim JG and Kim YK: Bacteria-derived extracellular vesicles in urine as a novel biomarker for gastric cancer: Integration of liquid biopsy and metagenome analysis. *Cancers (Basel)* 13: 4687, 2021.
72. Kim JR, Han K, Han Y, Kang N, Shin TS, Park HJ, Kim H, Kwon W, Lee S, Kim YK, *et al*: Microbiome markers of pancreatic cancer based on bacteria-derived extracellular vesicles acquired from blood samples: A retrospective propensity score matching analysis. *Biology (Basel)* 10: 219, 2021.
73. Kim SI, Kang N, Leem S, Yang J, Jo H, Lee M, Kim HS, Dhanasekaran DN, Kim YK, Park T and Song YS: Metagenomic analysis of serum microbe-derived extracellular vesicles and diagnostic models to differentiate ovarian cancer and benign ovarian tumor. *Cancers (Basel)* 12: 1309, 2020.
74. Ağagündüz D, Gençer Bingöl F, Çelik E, Cemali Ö, Özenir Ç, Özogul F and Capasso R: Recent developments in the probiotics as live biotherapeutic products (LBPs) as modulators of gut brain axis related neurological conditions. *J Transl Med* 20: 460, 2022.
75. Cordaillat-Simmons M, Rouanet A and Pot B: Live biotherapeutic products: The importance of a defined regulatory framework. *Exp Mol Med* 52: 1397-1406, 2020.
76. Lee NK, Han KJ, Son SH, Eom SJ, Lee SK and Paik HD: Multifunctional effect of probiotic *Lactococcus lactis* KC24 isolated from kimchi. *LWT Food Sci Technol* 64: 1036-1041, 2015.
77. Han KJ, Lee NK, Park H and Paik HD: Anticancer and anti-inflammatory activity of probiotic *Lactococcus lactis* NK34. *J Microbiol Biotechnol* 25: 1697-1701, 2015.
78. Peymaeei F, Sadeghi F, Safari E, Khorrami S, Falahati M, Roudbar Mohammadi S and Roudbary M: *Candida albicans* beta-glucan induce anti-cancer activity of mesenchymal stem cells against lung cancer cell line: An in-vitro experimental study. *Asian Pac J Cancer Prev* 21: 837-843, 2020.
79. Albeituni SH, Ding C, Liu M, Hu X, Luo F, Kloecker G, Bousamra M II, Zhang HG and Yan J: Yeast-derived particulate β -glucan treatment subverts the suppression of myeloid-derived suppressor cells (MDSC) by inducing polymorphonuclear MDSC apoptosis and monocytic MDSC differentiation to APC in cancer. *J Immunol* 196: 2167-2180, 2016.
80. Kim K, Kwon O, Ryu TY, Jung CR, Kim J, Min JK, Kim DS, Son MY and Cho HS: Propionate of a microbiota metabolite induces cell apoptosis and cell cycle arrest in lung cancer. *Mol Med Rep* 20: 1569-1574, 2019.
81. Kotzampassi K, Stavrou G, Damaraki G, Georgitsi M, Basdanis G, Tsaousi G and Giamarellos-Bourboulis EJ: A four-probiotics regimen reduces postoperative complications after colorectal surgery: A randomized, double-blind, placebo-controlled study. *World J Surg* 39: 2776-2783, 2015.
82. Wada M, Nagata S, Saito M, Shimizu T, Yamashiro Y, Matsuki T, Asahara T and Nomoto K: Effects of the enteral administration of *Bifidobacterium breve* on patients undergoing chemotherapy for pediatric malignancies. *Support Care Cancer* 18: 751-759, 2010.
83. Gui QF, Lu HF, Zhang CX, Xu ZR and Yang YH: Well-balanced commensal microbiota contributes to anti-cancer response in a lung cancer mouse model. *Genet Mol Res* 14: 5642-5651, 2015.
84. Zhou X, Xie F, Wang L, Zhang L, Zhang S, Fang M and Zhou F: The function and clinical application of extracellular vesicles in innate immune regulation. *Cell Mol Immunol* 17: 323-334, 2020.
85. Choi JH, Moon CM, Shin TS, Kim EK, McDowell A, Jo MK, Joo YH, Kim SE, Jung HK, Shim KN, *et al*: *Lactobacillus paracasei*-derived extracellular vesicles attenuate the intestinal inflammatory response by augmenting the endoplasmic reticulum stress pathway. *Exp Mol Med* 52: 423-437, 2020.
86. Lee DH, Park HK, Lee HR, Sohn H, Sim S, Park HJ, Shin YS, Kim YK, Choi Y and Park HS: Immunoregulatory effects of *Lactococcus lactis*-derived extracellular vesicles in allergic asthma. *Clin Transl Allergy* 12: e12138, 2022.
87. Sim S, Lee DH, Kim KS, Park HJ, Kim YK, Choi Y and Park HS: *Micrococcus luteus*-derived extracellular vesicles attenuate neutrophilic asthma by regulating miRNAs in airway epithelial cells. *Exp Mol Med* 55: 196-204, 2023.
88. Kim MH, Choi SJ, Choi HI, Choi JP, Park HK, Kim EK, Kim MJ, Moon BS, Min TK, Rho M, *et al*: *Lactobacillus plantarum*-derived extracellular vesicles protect atopic dermatitis induced by *Staphylococcus aureus*-derived extracellular vesicles. *Allergy Asthma Immunol Res* 10: 516-532, 2018.
89. Kim JH, Jeun EJ, Hong CP, Kim SH, Jang MS, Lee EJ, Moon SJ, Yun CH, Im SH, Jeong SG, *et al*: Extracellular vesicle-derived protein from *Bifidobacterium longum* alleviates food allergy through mast cell suppression. *J Allergy Clin Immunol* 137: 507-516.e8, 2016.
90. Chen J, Zhang H, Wang S, Du Y, Wei B, Wu Q and Wang H: Inhibitors of bacterial extracellular vesicles. *Front Microbiol* 13: 835058, 2022.
91. Guo H, Zhao L, Zhu J, Chen P, Wang H, Jiang M, Liu X, Sun H, Zhao W, Zheng Z, *et al*: Microbes in lung cancer initiation, treatment, and outcome: Boon or bane? *Semin Cancer Biol* 32: 1190-1206, 2022.
92. Kim SI, Ha JY, Choi SY, Hong SH and Lee HJ: Use of bacterial extracellular vesicles for gene delivery to host cells. *Biomolecules* 12: 1171, 2022.
93. Liu H, Zhang Q, Wang S, Weng W, Jing Y and Su J: Bacterial extracellular vesicles as bioactive nanocarriers for drug delivery: Advances and perspectives. *Bioact Mater* 14: 169-181, 2021.
94. Lee EY, Choi DS, Kim KP and Cho YS: Proteomics in gram-negative bacterial outer membrane vesicles. *Mass Spectrom Rev* 27: 535-555, 2008.
95. Papahadjopoulos D, Poste G, Schaeffer BE and Vail WJ: Membrane fusion and molecular segregation in phospholipid vesicles. *Biochim Biophys Acta* 352: 10-28, 1974.
96. Papahadjopoulos D, Mayhew E, Poste G, Smith S and Vail WJ: Incorporation of lipid vesicles by mammalian cells provides a potential method for modifying cell behaviour. *Nature* 252: 163-166, 1974.
97. Poste G and Papahadjopoulos D: Lipid vesicles as carriers for introducing materials into cultured cells: Influence of vesicle lipid composition on mechanism(s) of vesicle incorporation into cells. *Proc Natl Acad Sci USA* 73: 1603-1607, 1976.
98. Casal JI, Rueda P and Hurtado A: Parvovirus-like particles as vaccine vectors. *Methods* 19: 174-186, 1999.
99. Parmar MM, Edwards K and Madden TD: Incorporation of bacterial membrane proteins into liposomes: Factors influencing protein reconstitution. *Biochim Biophys Acta* 1421: 77-90, 1999.
100. Kim SH, Kim KS, Lee SR, Kim E, Kim MS, Lee EY, Cho YS, Kim JW, Bishop RE and Chang KT: Structural modifications of outer membrane vesicles to refine them as vaccine delivery vehicles. *Biochim Biophys Acta* 1788: 2150-2159, 2009.
101. Goldin BR and Gorbach SL: Effect of *Lactobacillus acidophilus* dietary supplements on 1,2-dimethylhydrazine dihydrochloride-induced intestinal cancer in rats. *J Natl Cancer Inst* 64: 263-265, 1980.
102. Ghoneum M and Gimzewski J: Apoptotic effect of a novel kefir product, PFT, on multidrug-resistant myeloid leukemia cells via a hole-piercing mechanism. *Int J Oncol* 44: 830-837, 2014.
103. Thirabunyanon M and Hongwittayakorn P: Potential probiotic lactic acid bacteria of human origin induce antiproliferation of colon cancer cells via synergic actions in adhesion to cancer cells and short-chain fatty acid bioproduction. *Appl Biochem Biotechnol* 169: 511-525, 2013.
104. Orlando A, Refolo MG, Messa C, Amati L, Lavermicocca P, Guerra V and Russo F: Antiproliferative and proapoptotic effects of viable or heat-killed *Lactobacillus paracasei* IMPC2.1 and *Lactobacillus rhamnosus* GG in HGC-27 gastric and DLD-1 colon cell lines. *Nutr Cancer* 64: 1103-1111, 2012.
105. Baldwin C, Millette M, Oth D, Ruiz MT, Luquet FM and Lacroix M: Probiotic *Lactobacillus acidophilus* and *L. casei* mix sensitize colorectal tumoral cells to 5-fluorouracil-induced apoptosis. *Nutr Cancer* 62: 371-378, 2010.
106. Kim Y, Lee D, Kim D, Cho J, Yang J, Chung M, Kim K and Ha N: Inhibition of proliferation in colon cancer cell lines and harmful enzyme activity of colon bacteria by *Bifidobacterium adolescentis* SPM0212. *Arch Pharm Res* 31: 468-473, 2008.
107. Park E, Jeon GI, Park JS and Paik HD: A probiotic strain of *Bacillus polyfermenticus* reduces DMH induced precancerous lesions in F344 male rat. *Biol Pharm Bull* 30: 569-574, 2007.
108. Ma EL, Choi YJ, Choi J, Pothoulakis C, Rhee SH and Im E: The anticancer effect of probiotic *Bacillus polyfermenticus* on human colon cancer cells is mediated through ErbB2 and ErbB3 inhibition. *Int J Cancer* 127: 780-790, 2010.

109. Gamallat Y, Meyiah A, Kuugbee ED, Hago AM, Chiwala G, Awadasseid A, Bamba D, Zhang X, Shang X, Luo F and Xin Y: Lactobacillus rhamnosus induced epithelial cell apoptosis, ameliorates inflammation and prevents colon cancer development in an animal model. *Biomed Pharmacother* 83: 536-541, 2016.
110. Hu J, Wang C, Ye L, Yang W, Huang H, Meng F, Shi S and Ding Z: Anti-tumour immune effect of oral administration of Lactobacillus plantarum to CT26 tumour-bearing mice. *J Biosci* 40: 269-279, 2015.
111. Walia S, Kamal R, Dhawan DK and Kanwar SS: Chemoprevention by probiotics during 1,2-dimethylhydrazine-induced colon carcinogenesis in rats. *Dig Dis Sci* 63: 900-909, 2018.
112. Jacouton E, Torres Maravilla E, Boucard AS, Pouderous N, Pessoa Vilela AP, Naas I, Chain F, Azevedo V, Langella P and Bermúdez-Humarán LG: Anti-tumoral effects of recombinant Lactococcus lactis strain secreting IL-17A cytokine. *Front Microbiol* 9: 3355, 2019.
113. Aguilar-Toalá JE, Garcia-Varela R, Garcia HS, Mata-Haro V, González-Córdova AF, Vallejo-Cordoba B and Hernández-Mendoza A: Postbiotics: An evolving term within the functional foods field. *Trends Food Sci Technol* 75: 105-114, 2018.
114. Fafián-Labora JA and O'Loughlin A: Classical and nonclassical intercellular communication in senescence and ageing. *Trends Cell Biol* 30: 628-639, 2020.
115. Liu Y, Defourny KAY, Smid EJ and Abee T: Gram-positive bacterial extracellular vesicles and their impact on health and disease. *Front Microbiol* 9: 1502, 2018.
116. Kang CS, Ban M, Choi EJ, Moon HG, Jeon JS, Kim DK, Park SK, Jeon SG, Roh TY, Myung SJ, *et al*: Extracellular vesicles derived from gut microbiota, especially Akkermansia muciniphila, protect the progression of dextran sulfate sodium-induced colitis. *PLoS One* 8: e76520, 2013.
117. Kim YJ, Lee BG, Shim JE, Lee H, Park JH and Yeo MK: Airborne bacteria in institutional and commercial buildings in Korea: Characterization with 16S rRNA gene sequencing and association with environmental conditions. *Aerosol Sci Technol* 58: 1281-1292, 2024.
118. Fahlgren C, Hagström Å, Nilsson D and Zweifel UL: Annual variations in the diversity, viability, and origin of airborne bacteria. *Appl Environ Microbiol* 76: 3015-3025, 2010.
119. Cho YS, Kim HR, Ko HS, Jeong SB, Chan Kim B and Jung JH: Continuous surveillance of bioaerosols on-site using an automated bioaerosol-monitoring system. *ACS Sens* 5: 395-403, 2020.
120. Gerdes L, Iwobi A, Busch U and Pecoraro S: Optimization of digital droplet polymerase chain reaction for quantification of genetically modified organisms. *Biomol Detect Quantif* 7: 9-20, 2016.
121. Biron VL, Kostiuk M, Isaac A, Puttagunta L, O'Connell DA, Harris J, Côté DW and Seikaly H: Detection of human papillomavirus type 16 in oropharyngeal squamous cell carcinoma using droplet digital polymerase chain reaction. *Cancer* 122: 1544-1551, 2016.
122. Brambati C, Galbiati S, Xue E, Toffalori C, Crucitti L, Greco R, Sala E, Crippa A, Chiesa L, Soriani N, *et al*: Droplet digital polymerase chain reaction for DNMT3A and IDH1/2 mutations to improve early detection of acute myeloid leukemia relapse after allogeneic hematopoietic stem cell transplantation. *Haematologica* 101: e157-e161, 2016.
123. Patterson B, Morrow C, Singh V, Moosa A, Gqada M, Woodward J, Mizrahi V, Bryden W, Call C, Patel S, *et al*: Detection of Mycobacterium tuberculosis bacilli in bio-aerosols from untreated TB patients. *Gates Open Res* 1: 11, 2018.
124. Lu X, Xiong L, Zheng X, Yu Q, Xiao Y and Xie Y: Structure of gut microbiota and characteristics of fecal metabolites in patients with lung cancer. *Front Cell Infect Microbiol* 13: 1170326, 2023.
125. Najafi S, Abedini F, Azimzadeh Jamalkandi S, Shariati P, Ahmadi A and Gholami Fesharaki M: The composition of lung microbiome in lung cancer: A systematic review and meta-analysis. *BMC Microbiol* 21: 315, 2021.
126. Greathouse KL, White JR, Vargas AJ, Bliskovsky VV, Beck JA, von Muhlinen N, Polley EC, Bowman ED, Khan MA, Robles AI, *et al*: Interaction between the microbiome and TP53 in human lung cancer. *Genome Biol* 19: 123, 2018.



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